

GA0182-0010

**A study on development of hypocholesterolemic foodstuff from
cereals**

I. :

II.

risk factor

가
phenolic compound가

tocotrienol, rutin

가 가

가

III.

1. *in vitro*

2. *in vivo*

3. 가

IV.

1. *in vitro*

HMG-CoA reductase

HMG-CoA reductase

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2.

in vivo

in vitro

HMG-CoA reductase

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가

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HMG-CoA reductase

G6PDH, malic enzyme

가

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3.

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가

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가 20%

가가

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SUMMARY

I. Title

A study on development of hypocholesterolemic foodstuff from cereals

II. Objective and Significance of Research

Although the rate of mortality has been considerably reduced, heart disease still remains as the leading cause of death. Data from well known epidemiologic studies have confirmed that hypercholesterolemia is one of the risk factor for development of coronary heart disease. Most of studies in this area are preventive in nature and focused on lowering plasma cholesterol as well as triglyceride. The rate of hepatic cholesterologenesis is largely determined by the activity of HMG-CoA reductase, which catalyzed the conversion of HMG-CoA to mevalonic acid. Epidemiological data indicate that population that consume diets rich in cereal grains tend to have a low incidence of cardiovascular disease. It is also well known that cereal grains contain various types of polyphenolic compounds such as tocotrienols, tannic acid and rutin. Cereals consumed in Korea are various in kinds and some of them have been reported to have hypocholesterolemic effects or not. But, it still remains to clarify the effect of cereal grains on lipid metabolism, and utilization of cereal grain as functional food is limited.

This study was performed to investigate effect of cereal grains on the activities for cholesterol biosynthesis, degradation and fatty acid synthesis in vitro and vivo, and to development the new processed food

III. Scope and Contents of Research

1. Screening of HMG-CoA reductase inhibitory activity from cereals
(*in vitro*)
2. Effects of some cereals on lipid metabolism (*in vivo*)
3. Examination of physicochemical characteristics of some cereals and development of processed food from cereals

IV. Results

1. Screening of HMG-CoA reductase inhibitory activity from cereals
(*in vitro*)

A study was conducted to screen the inhibitory activity of 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductase, which is known to be rate-limiting enzyme in cholesterol biosynthesis, from the extracts of 80% methanol and 70% ethanol of cereals. The strongest inhibitory activity was shown in the ethanol extract of sorghum among the ethanol extracts. The inhibitory activity of HMG-CoA reductase of prosomillet methanol extract was 73%, and highest among the methanol extracts. The inhibitory activity of 44.7% was observed in sorghum methanol extract. The methanol extracts of prosomillet, sorghum and black rice were further fractionated with hexane, chloroform, ethylacetate, butanol and water. Hexan fraction of these cereal methanol had the strongest inhibitory activity among five fractions, and the inhibitory activity was increased compared to each crude methanol extracts. It was also investigated the effect of methanol extracts and five fraction of cereals on cholesterol contents in HepG2 cell.

2. Effects of some cereals on lipid metabolism (*in vivo*)

Effect of sorghum, black rice and prosomillet, which had strong inhibitory activity of HMG-CoA reductase *in vitro* on lipid metabolism in rats fed high cholesterol diets or high fat diets. Sorghum methanol extracts decreased serum total cholesterol and increased fecal bile acid excretion in rats fed high-cholesterol diets. Black rice decreased significantly liver total cholesterol and triglyceride in rats fed high-fat diets. Prosomillet decreased significantly plasma total cholesterol, triglyceride, liver triglyceride and increased fecal cholesterol excretion. It was also found that liver cytosolic malic enzyme and G6PDH activities decreased by addition of prosomillet. These results indicate that certain active components in some cereals other than dietary fiber have potential to exert hypolipidemic effects via regulating hepatic cholesterologenesis and lipogenesis.

3. Examination of physicochemical characteristics of some cereals and development of processed food from cereals

Gelatinization characteristics of some cereals were determined in terms of amylograph, gel consistency, WAI, WSI and viscoelasticity. Black rice exhibited the lowest peak viscosity in Brabender amylograph among cereals tested, but gel consistency was significantly higher in black rice than the others.

Effect of addition of sorghum flour on the quality properties of wheat flour muffin were investigated. The sorghum flour addition at 10% level did not affect the volume of muffin, but the volume of muffin containing 30% sorghum flour was significantly decreased compared to that of control. Most of texture parameters such as hardness, adhesiveness,

gumminess and chewiness were not affected by the addition of sorghum flour up to 20% level. As results of sensory evaluation, appearance and flavor of muffins by addition of sorghum flour differed significantly from the control, but there were no differences among the muffins containing sorghum flour at 10, 20, and 30% levels. Texture of 10% sorghum flour muffin was similar to that of the control, but there was a significant difference in texture by 30% sorghum flour addition. However, taste and overall acceptability did not differ significantly up to 30% level.

It was also developed black rice beverage by examination of extraction condition, combination formula and sensory evaluation.

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- 5) Analysis of fecal lipids and bile acid

B. Results and discussion

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- 3) Serum lipids
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- 5) Liver lipids
- 6) Fecal lipids and bile acid

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o High-fat diet model

A. Materials and methods

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- 4) Analysis of lipids and enzymes

B. Results and discussion

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o High-cholesterol diet model

A. Materials and methods

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5) Protein assay

6) Statistical analysis

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2) Lipids of serum and liver

3) Fecal cholesterol and bile acid

4) Liver cholesterol 7- α -hydroxylase activity

5) Activity of liver G6PDH and malic enzyme

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A. Materials and methods

1) Animals and diets

2) Preparation of tissues for analysis and analysis of lipids and enzymes

3) Statistical analysis

B. Results and discussion

1) Body weight, organ weight

2) Serum lipids

3) Lipids and TBARS of liver

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B. Results and discussion

References

1

2 *in vitro*

1

1.

가.

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2.

가. diurnal cholestyramin 가

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2

1.

가.

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. HMG- CoA reductase

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2.

가.

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HMG- CoA reductase

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HMG- CoA reductase

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HMG- CoA reductase

3

1.

2.

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. .
2.

가.

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. .
. GOT GTP
. .

3

. High- fat

1.

가.

2.

가.

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. .
. HMG- CoA reductase
. Cholesterol 7- -hydroxylase
. Glucose- 6- phosphate dehydrogenase
. .

. High - cholesterol

1.

가. 가

- .
- .
- .
- .
- 2.
- 가.

cholesterol
Cholesterol 7- -hydroxylase

- 4
- 1.
- 가.

- 2.
- 가.

TBARS
malic enzyme G6PDH
Postprandial triglyceride

- 4 가
- 1
- 1.
- 2.
- 가. Amylogram

. Gel consistency, WSI, WAI

. Viscoelasticity

.

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2

1.

2.

가.

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.

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3

1.

2.

HMG- CoA
 (3- hydroxy- 3- methylglutaryl coenzyme A) mevalonate
 HMG- CoA reductase cholesterol
 HMG- CoA reductase LDL- receptor 가
 가
 compactin
 HMG- CoA reductase
 가
 가
 ,
 , oat bran
 , hemicellulose soluble
 dietary fiber
 가
 Qureshi , β- glucane

HMG- CoA reductase

tocotrienol

phenolic compound

가

가

in vitro

2 *in vitro*

1 *in vitro*

HMG- CoA reductase isotope

spectrophotometric assay

enzyme microsomal enzyme source

1.

가.

150- 200 g SD

cholestyramin 가

가 2

microsome

1) ether

10 ml saline

infusion

2) 3- 4 g 0- 4 buffer A(, pH

7.0, 50 mM, sucrose 0.2 M, dithiothreitol 2 mM) , homogenize

30- 40 ml

3) 15,000g 10

가

100,000g 75

microsome

pellet

4) 3) microsome buffer A 100,000g 60

microsome pellet

5) pellet suspension buffer (buffer A plus 50 mM EDTA, pH

7.0) biuret

50- 100 μ

6 N HCl 5 μ l 가 ,

mevalonic- lactone 30 37 .

3) 2) 10,000 rpm 5
 15 μ l TLC .

4) silica gel G plate (10x10, 20x10, 10 cm) 2-2.5
 cm band , benzene:acetone (1:1. v/v) .
 mevalonic lactone (Mev)
 가 apply .

5) Mev (Rf= 0.5)
 HMG- CoA () scintillation cocktail 5 ml 가
 TLC plate
 image analyzer band가 detect Mev

2.

<spectrophotometric method>

가. diurnal cholestyramin 가
 light/dark cycle 2
 10 dark cycle 5 microsome
 Table 2-1
 dark cycle 가 light cycle 2
 control middark
 cycle .

Table 2-1. Diurnal changes in HMG- CoA reductase

Specific activity	
Light	3,114 pmoles/min/mg protein
Dark	6,516 pmoles/min/mg protein

enzyme protein microsome
 2 , cholestyramin
 가 2 12 ,
 microsome . cholestyramin 가
 microsome HMG- CoA reductase activity 가 microsome
 protein 9 (Table 2-2).

Table 2-2. Effect of cholestyramin in HMG- CoA reductase activity

		Specific activity
Cholestyramin	가	702 pmoles/min/mg protein
Cholestyramin	가	6,647 pmoles/min/mg protein

, DMSO 가 , Table 2-3
DMSO가 가 가 10ml
DMSO
가
DMSO

Table 2-3. Effect of various solvent on HMG- CoA reductase activity

	methanol	ethanol	DMSO
0 ml		100 %	
10 ml	58 %	77 %	95 %
15 ml	61 %	71 %	88 %
20 ml	67 %	77 %	89 %

HMG- CoA reductase
activity microsome cholestyramin ,
middark cycle 12 microsome freeze- thaw

<Radiochemical method>

가. Microsome Protein

가

microsomal protein 660 μ g 가

60 μ l , 30 μ l

fig.2- 1

microsomal protein 가

가

660 μ g

가

가 protein 200- 400 μ g

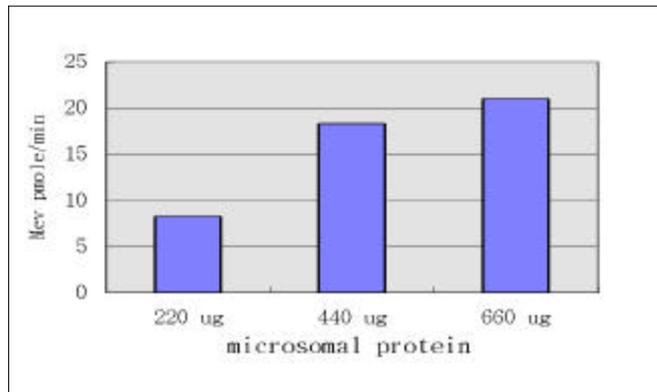


Fig. 2- 1 Effect of microsomal protein concentration on HMG- CoA reductase activity

30 60 μ l 15 30 ;

Fig. 2- 2

가

가

30 μ l

가

, 60 μ l

,

가

1

, , ,
, 2 g 100 mL
가 500 mL hexane 200 mL
10 1 hexane
hexane 200 mL 가 10 , 1 hexane
chloroform, ethylacetate, butanol
가 hexane hexane , chloroform
, ethylacetate , butanol ,
dimethylsulfoxide(DMSO)
가 assay .
200-220 g SD activation
cholestyramin 0.4% 가 10
11
cold buffer(, pH 7.0, 50 mM,
sucrose 0.2 M, dithiothreitol 2 mM) Potter-Elvehjem type
glass homogenizer homogenize 15,000g 10
가 100,000
g 70 microsome pellet buffer A
microsome 100,000 g 60 -20
-20 2 microsome pellet buffer
B(, pH 7.0, 50 mM, sucrose 0.1 M, dithiothreitol 2 mM, KCl
50 mM, EDTA 30 mM) 가 homogenize 30
100,000 g, 20 60
-70 .

. HMG-CoA reductase

HMG-CoA reductase Kleinsek ,
 , 1 mL Cuvette 20 μ L (control DMSO 20 μ L,
 (pH 7.0, 0.5 M) 100 μ L, DTT(20 mM) 100 μ L, NADPH(3 mM)
 100 μ L, 100 μ L 가 37
 HMG-CoA(3 mM) 100 μ L 가 340 nm
 5 . HMG-CoA 가
 blank

$$(\%) = \frac{[1 - (\text{control} - \text{control blank}) / (\text{sample} - \text{sample blank})]}{1} \times 100$$

Solubilized enzyme Lowry
 bovine serum albumin . .

2.

가.

10%

0.2% - 5%

22- 42%, 10- 23%

(Table 2- 4)

가 15.1%,

14.8%

, 2%

1%

17.5% 가

가 0.6% 가

Table 2-4. The yields of 70% ethanol and 80% methanol extracts of cereals and regumes.

Samples	Methanol extracts	Ethanol extracts
		(%)
Brown rice	1.9	1.8
Black rice	2.7	1.4
Prosomillet	2.9	2.0
Sorghum	2.8	2.4
Naked barley	4.0	2.9
barley	2.2	3.0
Job's tears	5.0	1.6
Glutinous millet	3.5	2.6
Buckwheat	2.6	3.5
Flavor rice	0.6	0.4
Red rice	2.5	2.0
Corn	7.0	4.9
Defatted soy bean	16.9	15.1
Defatted black soybean	17.5	14.8
Mungbean	9.1	8.1
Cowpeas	6.4	9.4
Small red bean	8.2	7.5
Black sesame	6.2	7.7

HMG-CoA reductase

100 μ g/assay 가

HMG-CoA reductase Table 2-5 .

가 41.1% 가 20.8%

16% - 17.3%

HMG-CoA reductase (Table 2-6),

73% 44.7%

가 27%, (26.2%), (18.8%), (16.2%)

. Qureshi

가 ,

β -D- glucans

(Hordeum vulgan, L) 가

, HMG-CoA reductase

가 tocotrienol

HMG-CoA reductase

65% 가

6%, 33%

HMG-CoA reductase 가

. HMG-CoA

reductase 26%

HMG-CoA reductase

HMG- CoA

reductase *in vivo*

가

Table 2- 5. The inhibitory effect of 70% ethanol extracts of cereals and legumes on the microsomal HMG- CoA reductase activity

Samples	Specific activity (nmole/ng protein/min)	Inhibition rate (%)
Control	3.53	
Sorghum	2.08	41.1
Prosonillet	3.18	9.8
Barley	2.97	16.0
Naked barley	3.37	4.6
Glutinous millet	4.02	-14.0
Buckwheat	3.53	0
Flavor rice	4.13	-17.0
Black rice	3.51	0.7
Red rice	3.48	1.3
Rice germ	2.75	22.1
Brown rice	4.09	-16.0
Job's tear	3.52	0
Black sesane	2.80	20.8
Mungbean	2.92	17.3
Defatted soybean	4.31	-22.0
Cowpeas	2.92	17.3
Small red bean	3.57	-1.0
Corn	2.94	16.7
Defatted-		

black soybean 3.53 0

* The concentrations of each sample tested was 100 µg/assay

Table 2-6. The inhibitory effect of 80% methanol extracts of cereals and regumes on the microsomal HMG-CoA reductase activity

Samples	Specific activity (nmole/ng protein/min)	Inhibition rate (%)
Control	4.15	
Sorghum	2.29	44.7
Prosonillet	1.13	72.7
Barley	4.45	-7.3
Naked barley	3.37	18.8
Glutinous millet	4.66	-12.2
Bucksheat	3.06	26.2
Flavor rice	3.35	19.2
Black rice	5.70	-37.4
Red rice	4.76	-14.7
Brown rice	4.15	0.0
Job's tear	3.93	5.2
Rice germ	4.77	-14.9
Black sesane	3.89	6.2
Mungbean	4.02	3.0
Defatted soybean	4.07	1.8
Cowpeas	4.34	-4.5
Snall red bean	5.69	-37.3
Corn	4.40	-6.1
Defatted- black soybean	4.23	-2.1

* The concentrations of each sample tested was 100 µg/assay

(Table 2-7)

41% 가

38%

56%,

29%, 7%

52%

가 18%

45% 가

가

가

4

가

Table 2-7. Yield of each solvent fraction

	Sorghum	Prosomillet	Black rice	Buckwheat
Hexane	38.2	55.6	18.0	14.0
Chloroform	4.5	1.4	5.6	5.5
Ethylacetate	1.1	0.8	2.1	4.3
Butanol	11.4	7.4	12.4	20.9
Water	39.4	28.6	48.0	45.0

HMG- CoA reductase

100ug/assay

가 HMG- CoA reductase

(Fig.2-4)

96.2%, 98%, 93.1%

3

가

(Fig. 2-5)

10%
HMG- CoA reductase

가

(Fig. 2-6)

88.7%
22.5%

(Fig. 2-7)

3

43.9%

HMG- CoA reductase

TLC, preparation HPLC

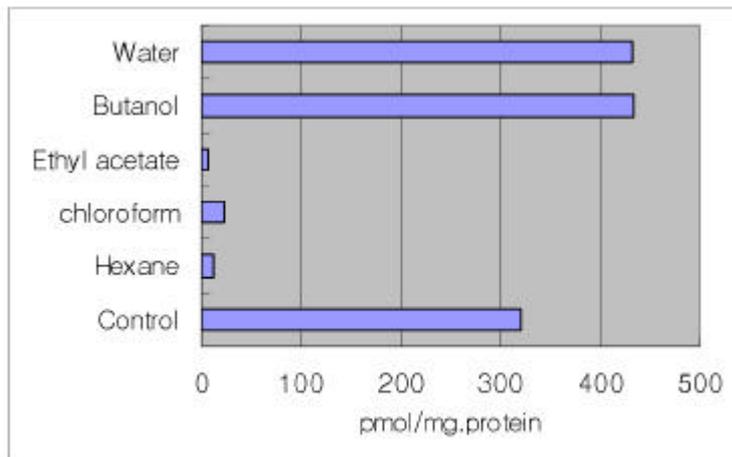


Fig. 2-4 Effect of various solvent fractions of prosomillet methanol extract on activity of liver microsomal HMG- CoA reductase

pmol of mevalonic acid/min/mg microsomal protein

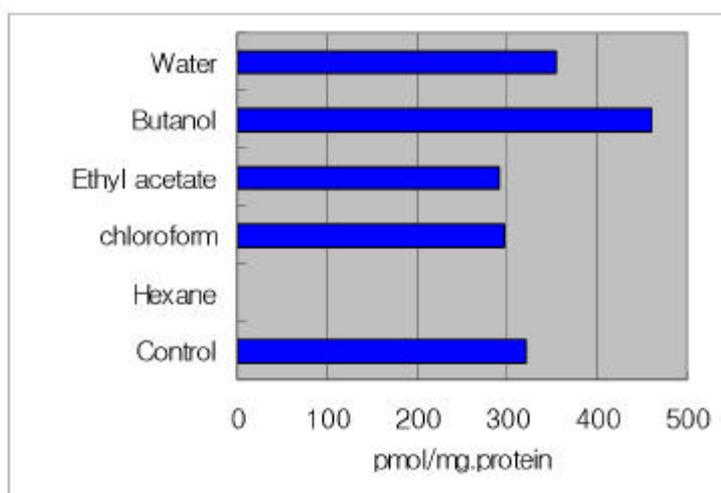


Fig. 2-5 Effect of various solvent fractions of sorghum methanol extract on HMG- CoA reductase

pmol of mevalonic acid/min/mg microsomal protein

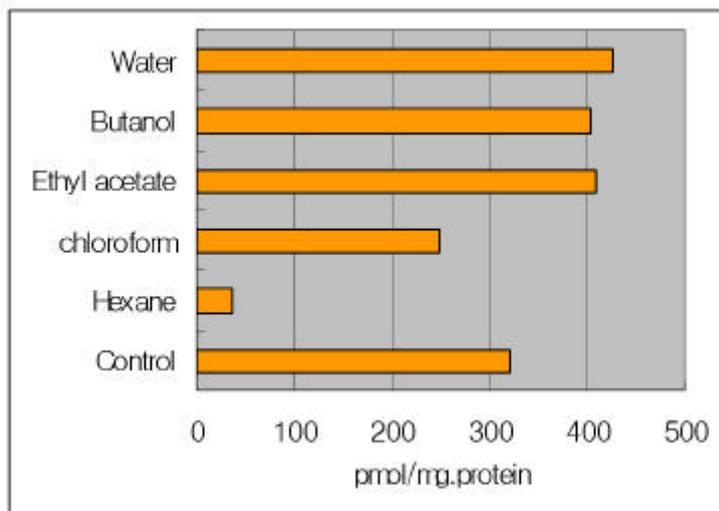


Fig. 2-6 Effect of various solvent fractions of black rice methanol extract on HMG- CoA reductase

pmol of mevalonic acid/min/mg microsomal protein

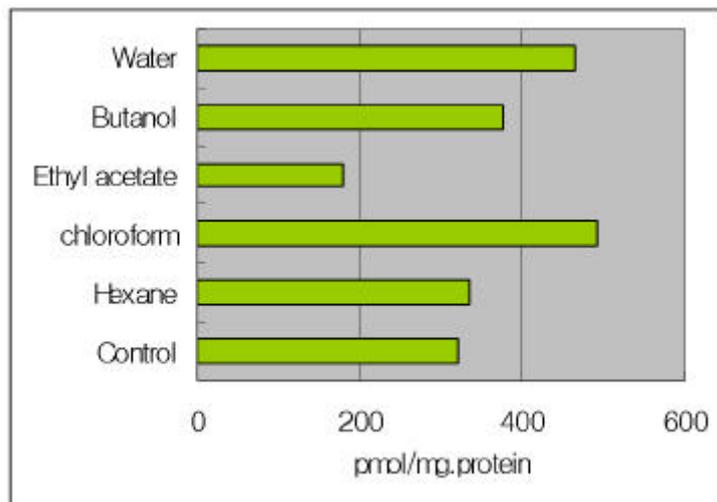


Fig. 2-7 Effect of various solvent fractions of sorghum methanol extract on HMG- CoA reductase

pmol of mevalonic acid/min/mg microsomal protein

, , ,
 , 가 25ug, 50ug, 100ug
 radioenzymatic method HMG-CoA reductase
 (2- 8,9,10,11,12). 100ug 가
 50ug 가 97%, 25ug 가 80%
 가 가 .
 100ug 가
 95.3%, 95.8%, 25ug 가 75%, 70.2%, 25ug 가 37.4%, 40.1%
 100ug 가
 81.3%, 50ug 가 53.4%, 25ug 가 30.5% 가 가
 25ug 가 가
 가

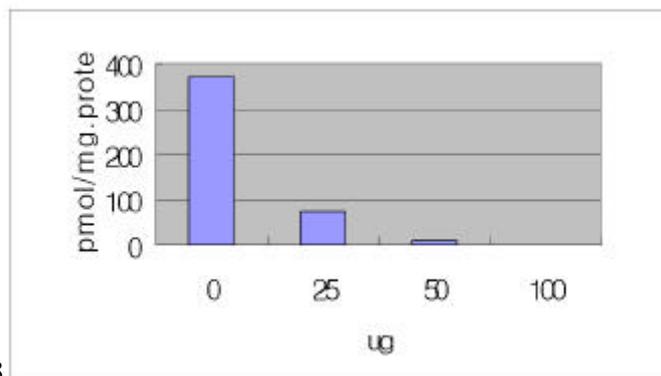


Fig. 2-8 Dose dependent inhibition of hexane fraction of sorghum methanol extract on HMG-CoA reductase activity

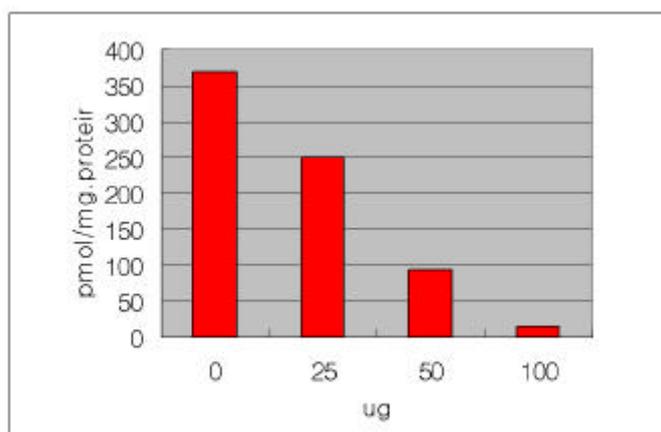


Fig. 2-9 Dose dependent inhibition of hexane fraction of prosomillet methanol extract on HMG-CoA reductase activity

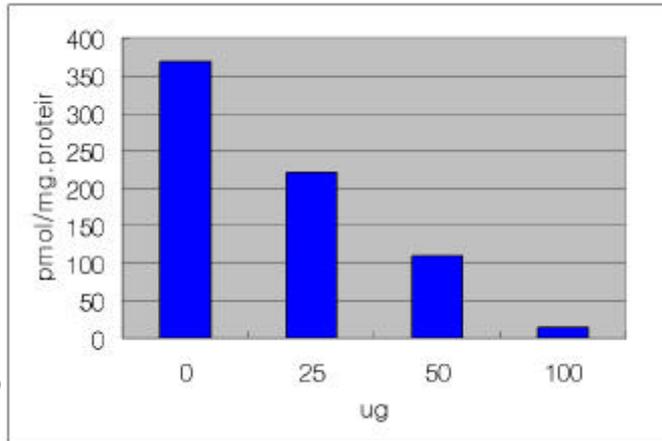


Fig.2- 10
D o s e
dependent
inhibition of chloroform fraction of sorghum
extract on HMG-CoA reductase activity

methanol

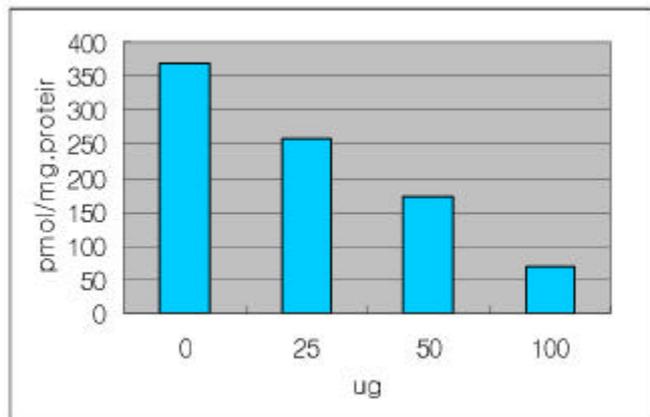


Fig. 2-11 Dose dependent inhibition of hexane fraction of black rice
methanol extract on HMG-CoA reductase activity

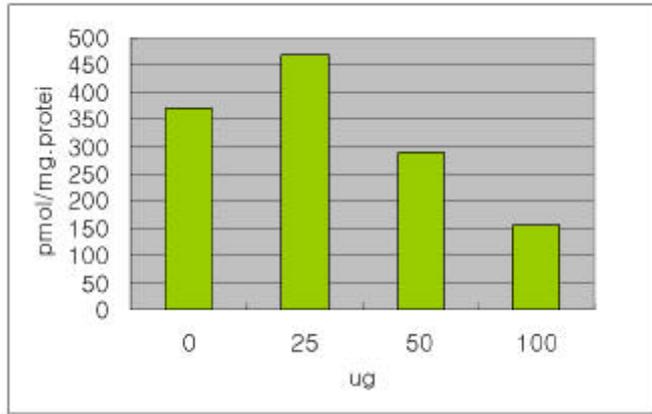


Fig. 2-12 Dose dependent inhibition of ethyl acetate fraction of buckwheat methanol extract on HMG- CoA reductase activity

HMG- CoA reductase		Table
2-8	HMG- CoA reductase NDGA가 85.6% 가	
	Tannic acid 65%	
		NDGA, tannic acid
	HMG- CoA reductase	rutin HMG- CoA
	tannic acid	

Table 2-8. Effect of various phenolic compounds on HMG- CoA reductase activity

Compounds	pmol/mg.protein	% of control
Control	455	100
Quercetin	477	105
Catechin	505	111
NDGA	65	14.4 *
Kampferol	436	96.2 *
Rutin	482	106
Tannic acid	161	35.5
Gallic acid	500	110
Ferulic acid	482	106

microsomal HMG- CoA reductase

가

HMG- CoA reductase inhibitor

mevastatin

가

3

TLC, preparation HPLC

3

HMG- CoA reductase

.

1.

가.

HMG- CoA reductase

microsome

2- 1

. SD

activation cholestyramin 가

microsome

HMG- CoA reductase

.

. HMG- CoA reductase

HMG- CoA reductase

Kleinsek

isotope

, , ,

tannin NDGA

.

Lowry

TLC plate spot

(CH3: MeOH: H2O = 65: 35: 10)

spot

UV- spectrum

2.

가.

HMG- CoA reductase

가 가

. 가

60 , 80 , 100

30 가

HMG- CoA reductase

25 () 30

100.3, 94.3, 95.4%

. 가

10

0 15 , 30 , 60

가

가

15

가

30

가

95.3%, 60 가

93.8%

HMG- CoA reductase

,

(Fig.2- 13, 14).

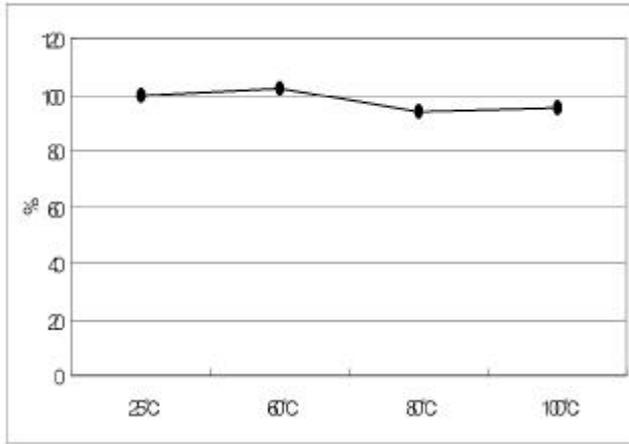


Fig. 2-13 Effect of hexane fraction of sorghum methanol extract heated to various temperatures on HMG-CoA reductase activity

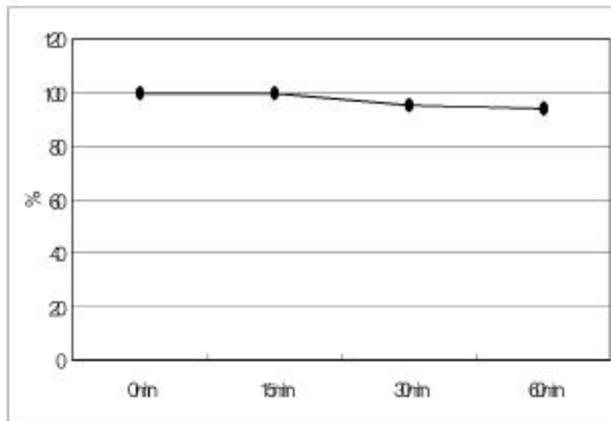


Fig. 2-14 Effect of hexane fraction of sorghum methanol extract

heated for various times at 100 on HMG- CoA reductase activity

, , , 25mg/ml 1ul 가
 (total 100ug/assay)
 100ug/assay 가
 . 2 가 가
 4 가 4 가
 가
 . 25mg/ml 2ul (total
 100ug/assay) 가 72.1%
 100ug/assay 가 53% (
), 63% ()
 HMG- CoA reductase tannin
 NDGA , 가
 . NDGA 가 84.6%
 NDGA 가
 , 가
 tannin 가
 74.8% . 68.9%
 tannin 가
 가
 가

가

. , TLC
4 spot Rf 0.75 spot
Rf 0.13, 0.25, 0.38, 0.5 4 spot
spot UV- spectrum flavonid, tannin

4

1.

가.

HepG2 Korea Cell Line Bank(KCLB)
fetal bovine serum 10% RPMI
1640(100unit/ml penicillin, 100unit/ml streptomycin)
100x100 mm dish 4x10⁶ cell seeding 가 monolayer

250ug/dish 가 4 HMG- CoA reductase
harvest . control

DMSO 가 . 4
100ug/dish add up 2

enzymatic 가 GC spectrophotometer
cold- PBS 4
washing harvest
hydrolysis 가 가 hexane , GC

. HMG- CoA reductase
cold- PBS washing 2 1
HMG- CoA reductase radio isotope
Lowry .

2.
가. uptake
가

uptake 가 100ug/dish 가 가 가
4 가 (Data not shown). Fig. 2- 15

uptake 가

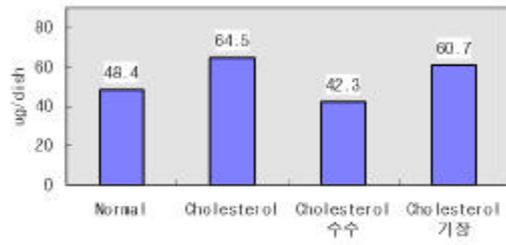


Fig. 2-15 Effect of sorghum and prosomillet methanol extract on cholesterol contents of HepG2 cell

HMG- CoA reductase
microsomal HMG- CoA reductase
spectrophotometer 가 radiochemical method
. Fig. 2-16 , (100ug/dish)
medium 가 4 microsome
HMG- CoA reductase
가 HMG- CoA reductase
60 pmol/min. mg protein 가
22 pmol/min. mg protein 1/3
가 none

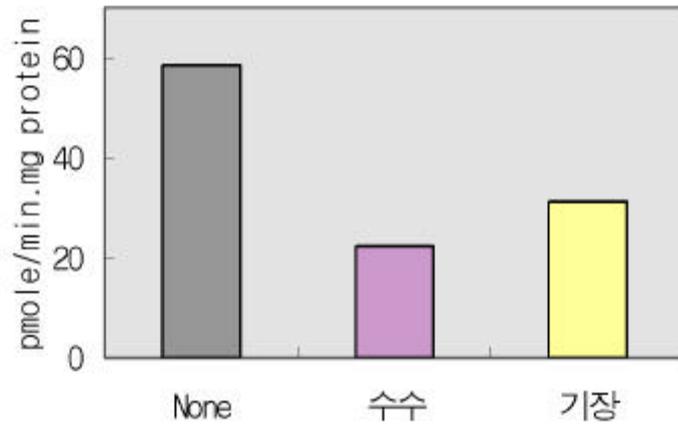
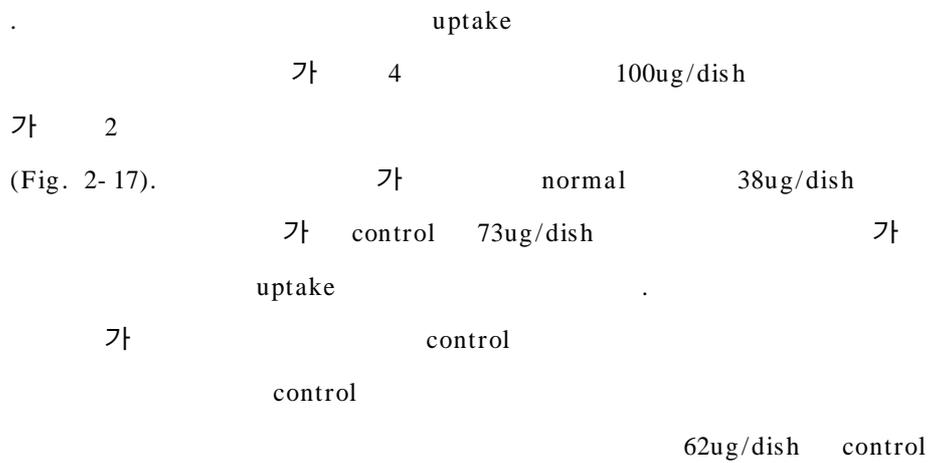
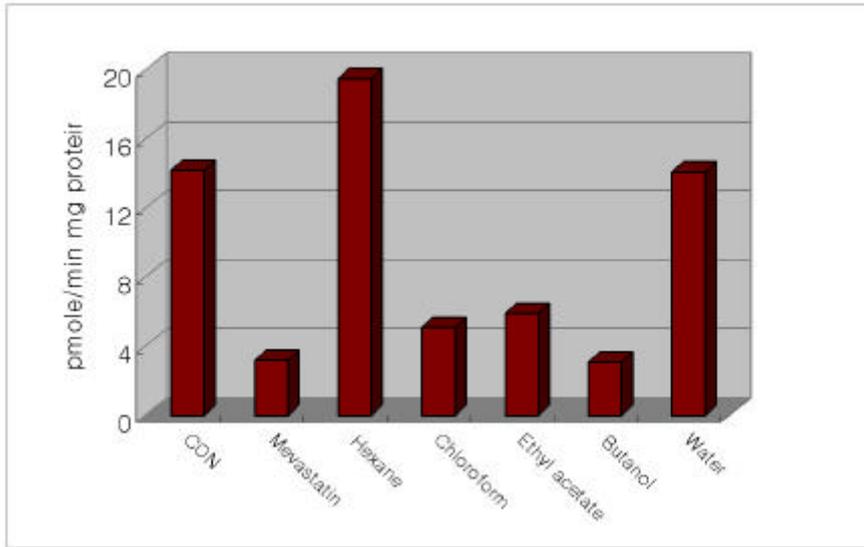


Fig. 2-16 Effect of sorghum and proso millet methanol extract on microsomal HMG-CoA reductase activity of HepG2 cell



pmole/min.mg.protein	control	
	5.1, 5.9 pmole/min.mg.protein	64%
	3.1 pmole/min.mg.protein	positive control
mevastatin		(Fig
2- 19)	control	
	9.8, 10.0, 10.9 pmole/min.mg.protein	29%
	1.94 pmole/min.mg.protein	
86%	mevastatin	

Fig. 2- 18 Effect of each solvent fraction of sorghum methanol extract on microsomal HMG- CoA reductase activity of HepG2 cell



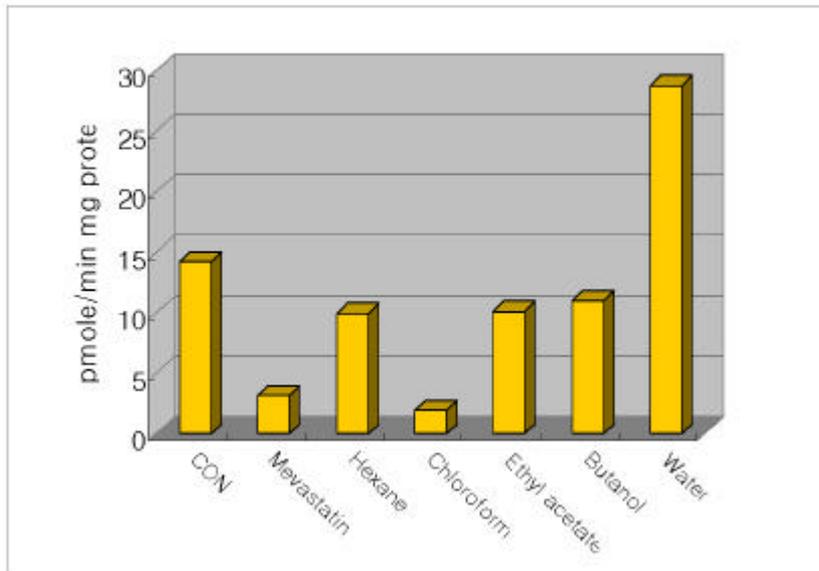


Fig.
2- 1
7

Effect of each solvent fraction of prosomillet methanol extract on microsomal HMG- CoA reductase activity of HepG2 cell

HMG- CoA reductase
uptake rat liver microsomal HMG- CoA

reductase . rat liver
microsomal , ,

가

rat liver microsomal

, ,

uptake

가

rat liver microsomal

.

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HMG-CoA reductase inhibitors. *Pharm. Res.*, 22, 555 (1990)

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3 *in vivo*

가 가 .
가 1
가 .
가 .
가 rice bran oil rice bran .
가 , ,
가 endogenous
가 . HMG-CoA reductase
가 가
가 .
rutin,
2 *in vitro* 3 , ,

1 가 High- cholesterol

1.

가.

200g SD
8- 10 4
, 0.4% , 0.8%
4
Table 3- 1 6
60%, 20%, 10%,
mix. 1%, mix 3.5%, 5% 가 mix.
mix. AIN- 76
4100 kcal/kg 4

4 - 70 가
,
,
2 2500 rpm
, trimming - 70
.

Table 3-1. The composition of experimental diet (g/1000 g diet)

Ingredient	Normal	Control	0.3% - extract	0.8% - extract
Casein	200	200	200	200
Soybean oil	50	50	50	50
Shortening	50	50	50	50
Mineral mix.	35	35	35	35
Vitamin mix.	10	10	10	10
Choline	2	2	2	2
Cholesterol	-	5	5	5
Taurocholate	1.2	1.2	1.2	1.2
Methionine	3	3	3	3
Cellulose	50	50	50	50
Sucrose	93.8	93.8	93.8	93.8
Corn starch	506.2	500	500	500
Sorghum- ext	-	-	3	8
Kcal/ 1000g	4100	4075	4075	4075

1)

, HDL-

CHCl₃/MeOH (2/1)

2)

500 mg

DeWeal

2.

가.

Fig. 3-1

299g

8

가

가

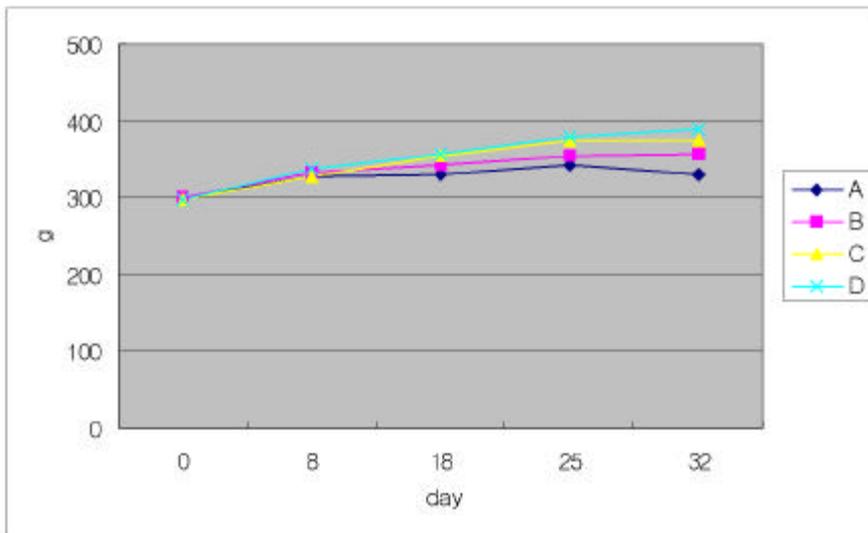


Fig. 3-1 Changes in body weight of experimental rats

(A:normal, B:control, C:0.3% ext, D:0.8% ext.)

Table 3-2

가	가	가	가
가	가	total cholesterol	가
가	가	0.3%	가
가	가	0.8%	가
가	HDL- cholesterol	가	가
0.3%	가	가	가
가	가	가	가

Table 3-2. Effect of sorghum methanol extracts on the concentrations of serum lipids of rats fed high cholesterol diets

Group	Total lipid	HDL cholesterol	Total cholesterol	Triglyceride
		(mg/100ml)		
A	345.95 ± 9.10c	60.74 ± 2.96a	69.03 ± 2.56c	44.81 ± 3.72
B	467.04 ± 24.91a	51.50 ± 1.72b	106.29 ± 5.82ab	56.91 ± 5.68
C	384.35 ± 10.98bc	45.88 ± 2.24b	93.92 ± 2.64b	51.53 ± 4.73
D	425.46 ± 20.85ab	37.63 ± 1.70c	114.13 ± 5.68a	55.45 ± 3.78

(A:normal, B:control, C:0.3% sorghum ext, D:0.8% sorghum ext.)

(Table 3-3)

가

0.3%

가

Table 3-3. Effect of sorghum methanol extracts on the weight of liver, kidneys and liver total lipid of rats fed high cholesterol diets

Group	Kidney weight	Liver weight	Liver total lipid (mg/g liver)
A	1.92 ± 0.036b	9.38 ± 0.237b	42 ± 0.12b
B	2.19 ± 0.060a	12.03 ± 0.200a	107 ± 0.67a
C	2.22 ± 0.089a	12.21 ± 0.135a	101 ± 0.69a
D	2.22 ± 0.059a	12.11 ± 0.357a	110 ± 0.91a

(A:normal, B:control, C:0.3% sorghum ext, D:0.8% sorghum ext.)

4

(Table 3-4, 5, Fig.3-2) 4

가 0.3%

가

가 .

cholesterol

2

가 . 0.3% 가 1.5
 . 3
 가 가
 가

Table 3-4. Effect of sorghum methanol extracts on the fecal cholesterol and weight of rats fed high cholesterol diets

Group	Fecal cholesterol		Fecal weight	
	mg/fecal g	mg/day	Dry (g/4days)	Wet (g/4days)
A	3.69 ± 0.337	4.95 ± 0.550	5.41 ± 0.460	6.70 ± 0.520
B	8.94 ± 0.464	11.86 ± 0.496	5.38 ± 0.294	6.87 ± 0.466
C	9.37 ± 0.707	15.67 ± 1.828	6.65 ± 0.531	8.57 ± 0.430
D	11.83 ± 1.123	15.87 ± 2.365	5.25 ± 0.377	0.34 ± 0.605

(A:nomal, B:control, C:0.3% sorghum ext, D:0.8% sorghum ext.)

Table 3-5. Effect of sorghum methanol extracts on excretion of fecal bile acids of rats fed high cholesterol diets

Group	Bile acid	
	mg/fecal g	mg/day
Normal	6.41 ± 0.24c	8.79 ± 0.86c
High- cholesterol control	23.69 ± 2.43b	30.23 ± 2.68b
0.3% sorghum ext.	25.78 ± 0.72ab	41.96 ± 3.93a
0.8% sorghum ext.	30.84 ± 1.89a	39.74 ± 1.90a

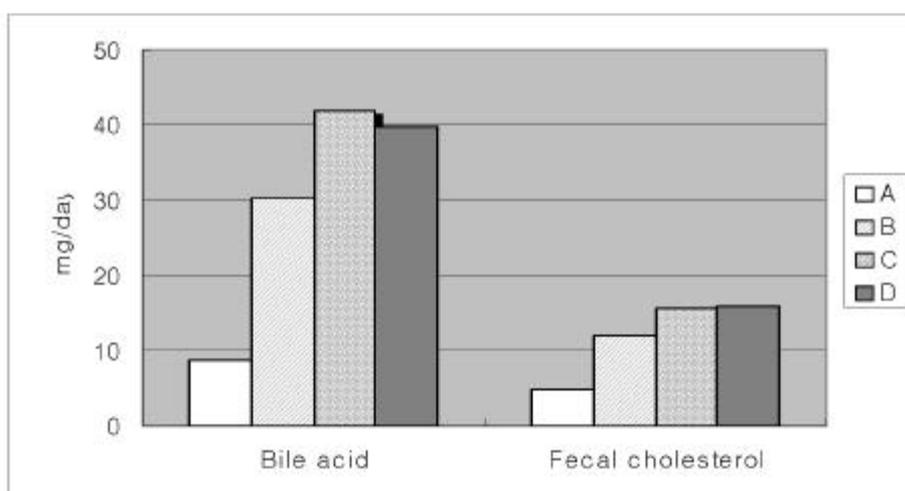


Fig. 3-2 Effect of sorghum methanol extracts on the excretion of fecal bile acid and cholesterol of rats fed high cholesterol diets

가

2 가 high- fat

1.

가.

40 10 80% 가 shaker
200rpm shaking
(Toyo No. 2) 5 80% 가
3
rotary evaporater

(SD rat, 4)

5

0.5% , 1% 가 , 15%, 30% 가
 AIN- 76 diet Table 3- 6
 casein 20% , lard 가
 AIN- 76 mixture .

5 .

Table 3- 6 . Compositions of experimental diets (g/1000 g diet)

Ingredient	Control (high- fat/ sucrose)	0.5% ext	1.0% ext	15% pow der	30% pow der
Casein	200	200	200	190	180
DL- Met	3	3	3	3	3
Starch	150	150	150	13	26
Sucrose	400	400	400	400	250
Cellulose	50	50	50	50	50
Lard	100	100	100	100	100
Corn oil	50	50	50	47	44
Min Mix	35	35	35	35	35
Vit Mix	10	10	10	10	10
Choline	2	2	2	2	2
BR- ext		5	10		
BR- Pow				150	300

12 dithyl ether
 2500 rpm 10
 가 trimming
 - 70
 3 - 70 가
 kit(Eiken, Tokyo,
 Japan) Triglyzyme- V,
 Cholestezyme- V, HDL-
 Folch Pearson
 TG Sale chloroform
 N2 gas triton X- 100
 ethanol kit
 200
 mg DeWeal KOH
 1 ml 3 autoclave 20% NaCl 1 ml 가
 20 ml 2 , c- HCl 0.2 ml 가
 20 ml 가 4
 MeOH:H₂O(5:1, v/v)
 kit(Kyokudo, Tokyo, Japan)

Cholesterol Pearson 550 nm .

program ANOVA Duncan Multiple range test SAS
($p < 0.05$)

2.

가.

5

Table 3-7

가 가
15%

Table 3-7. Effect of dietary black rice on the growth of rats fed high fat diet.

	Initial BW(g)	Weight gain(g)	Total intake(g)	FER
High fat control	281.1 ± 6.16ns	167.7 ± 5.74a	821.4 ± 14.5ns	0.18 ± 0.01ns
0.5% extract	281.0 ± 4.11	163.0 ± 9.27a	823.2 ± 15.77	0.20 ± 0.01
1% extract	281.3 ± 7.34	167.8 ± 6.49a	812.4 ± 9.52	0.21 ± 0.01
15% powder	281.4 ± 4.64	154.4 ± 6.56b	796.1 ± 6.99	0.19 ± 0.01
30% powder	281.5 ± 4.48	164.2 ± 10.04a	809.2 ± 6.7	0.20 ± 0.01

Table 3-8

가
1%

가

가

Table 3-8. Effect of dietary black rice on various organ weight of rats fed high fat diets

	Liver	Kidney	Spleen	Heart
				(g)
High fat control	14.87 ± 0.28 ^{ns}	2.86 ± 0.14 ^{ns}	1.10 ± 0.07 ^{ns}	1.25 ± 0.02 ^{ns}
0.5% extract	14.17 ± 0.63	2.89 ± 0.12	1.17 ± 0.08	1.25 ± 0.02
1% extract	15.57 ± 0.46	2.97 ± 0.07	1.15 ± 0.06	1.32 ± 0.23
15% powder	14.75 ± 0.44	2.79 ± 0.07	1.09 ± 0.06	1.28 ± 0.01
30% powder	14.62 ± 0.73	3.00 ± 0.10	1.05 ± 0.05	1.29 ± 0.04

	Triglyceride	Cholesterol	HDL cholesterol
		(mg/dl)	
High fat control	131.84 ± 14.15 ^{rs}	95.07 ± 6.41 ^{rs}	54.96 ± 7.09 ^{rs}
0.5% extract	124.92 ± 22.01	90.45 ± 19.60	58.90 ± 11.02
1% extract	121.08 ± 36.27	84.59 ± 21.02	53.68 ± 8.30
15% powder	132.17 ± 35.20	80.96 ± 23.61	60.73 ± 12.40
30% powder	134.25 ± 28.67	87.37 ± 11.79	58.68 ± 7.25

	GOT	GPT	GOT	GPT
			0.5%	
		가	가	
		1%		
	GPT			가

Table 3-10. Effect of dietary black rice on the activities of serum GOT and GPT of rats fed high fat diets.

	GOT	GPT
	(karmen)	
High fat control	19.02 ± 3.09ab	30.97 ± 4.55ns
0.5% extract	20.60 ± 4.58a	29.60 ± 4.25
1% extract	15.36 ± 3.91b	32.11 ± 5.23
15% powder	16.39 ± 6.50ab	33.54 ± 7.26
30% powder	16.57 ± 5.01ab	31.96 ± 7.97

Table 3- 11

가가

가

1% 30%
 가 가
 가
 가 가

가

가

Table 3- 11. Effect of dietary black rice on the contents of liver lipids of rats fed with high fat diets.

	Cholesterol	Triglyceride (mg/g.liver.wet.wt)
High fat control	3.96 ± 0.10a	19.52 ± 0.74a
0.5% extract	3.32 ± 0.07ab	16.12 ± 1.41ab
1% extract	2.98 ± 0.08b	14.71 ± 1.13b
15% powder	3.48 ± 0.09ab	16.51 ± 0.64ab
30% powder	3.02 ± 0.13b	16.85 ± 1.29ab

가

Table 3- 12

30%

가

30%

가 .

Table 3- 12. Effect of dietary black rice on the excretion of fecal bile acid, cholesterol and triglyceride of rats fed high fat diets.

	Fecal Weight (g/day)	Bile acid (mg/day)	Triglyceride (mg/day)	Cholesterol (mg/day)
High fat control	2.86 ± 0.09b	10.14 ± 1.69rs	6.28 ± 0.32b	13.95 ± 1.18b
0.5% extract	3.20 ± 0.29b	11.93 ± 1.98	6.66 ± 1.01b	16.90 ± 1.70a
1% extract	2.83 ± 0.10b	11.78 ± 0.35	6.41 ± 0.21b	13.20 ± 1.21b
15% powder	2.93 ± 0.17b	10.55 ± 1.87	6.72 ± 0.41b	13.10 ± 1.75b
30% powder	3.98 ± 0.24a	10.91 ± 1.51	8.41 ± 1.03a	16.74 ± 0.24a

3

I. High- fat

1.

가.

		6	6	Table 3-13
			가	()
		2		.
70-75%	methanol		diethyl ether	
		300-400 mesh		.
		15% (w/w)		(control)
4	(2	2)	
			5% (w/w)	
	normal			. 6
		60-65%,	20%,	mix 1%,
mix 3.5%,	5%			.
			casein	cellulose
	가			.
		4	100 g	Sprague-Dawley
			4-5	
8-9			4	.

Table 3-13 . Compositions of experimental diets (g/1000 g diet)

Ingredient	Normal (low fat/ starch)	Control (high-fat/ sucrose)	1% ext	0.3% ext	20% powder	70% powder
Casein	200	200	200	200	177	121
DL- Met	3	3	3	3	3	3
Starch	650	150	150	150	0	0
Sucrose	0	405	405	405	405	0
Cellulose	50	50	50	50	46	37
Lard	0	95	95	95	95	95
Corn oil	50	50	50	50	46	35
Min Mix	35	35	35	35	35	35
Vit Mix	10	10	10	10	10	10
Choline	2	2	2	2	2	2
Proso- ext			10	3		
Proso - Powder					200	700
Kcal/ _1000g	3850	4330	4240	4240	4080	4080

4 1-2

2 - 50

, 4 가 ,

saline perfusion

가 2500 rpm

0-4 buffer A(, pH 7.0, 50 mM, sucrose 0.2 M, dithiothreitol 2 mM) , homogenize (HMG- CoA reductase

) TEDKSF buffer (pH 7.4, Cholesterol 7-alpha hydroxylase

) homogenize 100,000g

60 microsome pellet - 70

Microsome , cytosol

- 70

1)

CHCl₃/MeOH (2/1)

2)

500 mg

DeWeal

3) HMG- CoA reductase Cholesterol 7- alpha hydroxylase

G6PDH

2 radiochemical assay

Microsome HMG- CoA reductase Cholesterol 7- alpha

hydroxylase mevalonate- lactone 7- alpha

hydroxycholesterol TLC scintillation counter

G6PDH cytosol

Lowry

bovine serum albumin

2.

가.

Fig. 3-3

6
0.3% M-ext 20 70%
가

Table 3-14

3

가가 normal,

control 1% M-ext 10g

가

100 g

control

가 , methanol

4 normal

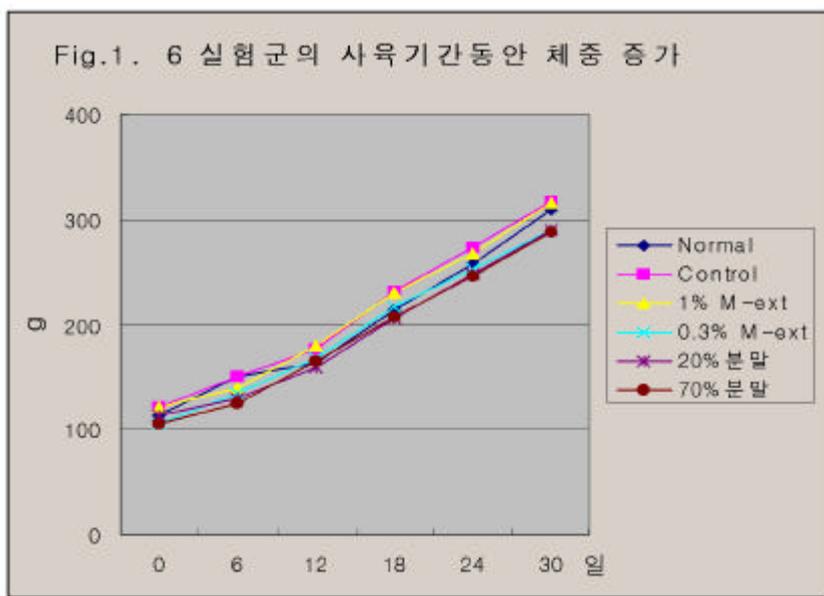


Fig 3-3. Changes of body weight during experimental period

Table. 3-14 Effect of dietary prosomillet on growth of rats fed high fat diet

	Normal	Control	1% M- ext	0.3% M- ext	20% powder	70% powder
Initial(g)	114	122	123	106	114	106
Final(g)	309	318	316	290	291	289
Gain(g)	195	196	193	184	177	183
Food intake(g)	18.6	17.7	17.8	17.7	17.8	16.3
FER	0.39	0.41	0.40	0.39	0.37	0.42
Liver wt. (g/100g bw)	4.51	4.81	4.51	4.59	4.49	4.14

Fig. 3-4 6 (TG), (T - Chol)
HDL- Chol . Fig. TG /
Control 가 , methanol
Control /
Normal . Control
methanol (M- ext)
0.3% M- ext 70% 가
70%
methanol Control
HDL- Chol T - Chol
HDL- Chol/T - Chol Table 3-15

가 / Normal

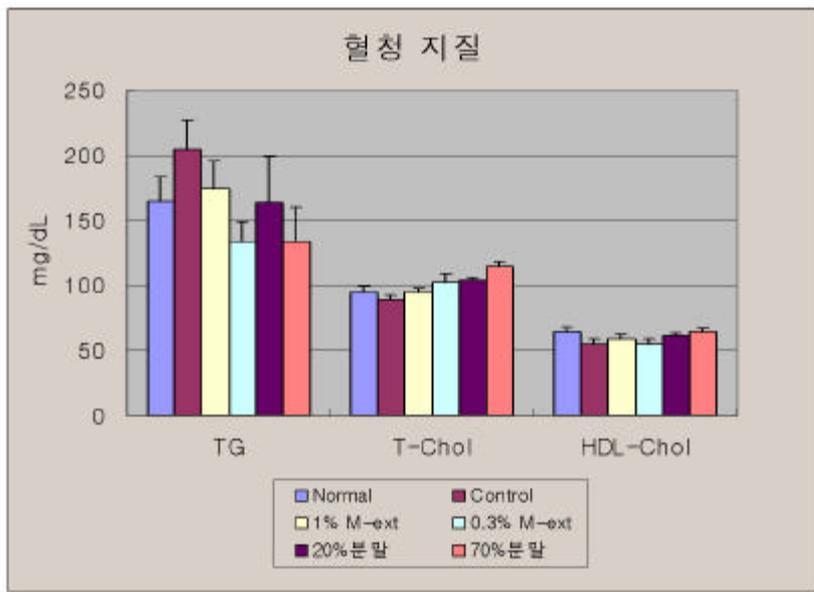


Fig. 3-4 Effect of prosomillet methanol extracts on serum lipids of rats fed high fat diets (Mean± SEM of 8 rats per group)

Table 3-15. Effect of prosomillet methanol extracts on serum HDL-cholesterol/total cholesterol ratio of rats fed high fat diets

Normal	Control	1% M- ext	0.3% M- ext	20% powder	70% powder
0.69±0.03	0.61±0.03	0.63±0.04	0.55±0.04	0.59±0.02	0.56±0.02

Fig. 3-5 가 control
 1.675 ± 0.115 mg/g 가 MeOH 0.3%, 1%
 가 20% 1,433 ± 0.062, 1.455 ± 0.058,
 1.436 ± 0.080 mg/g 70%
 가 1.642 ± 0.053 mg/g control
 MeOH
 가
 가
 가 MeOH
 Control, 1% MeOH, 0.3% MeOH 10.30 ± 1.13,
 10.53 ± 0.07, 10.55 ± 0.27 mg/g 가
 normal 7.29 ± 0.78 mg/g 30%, 20%
 6.99 ± 0.42 mg/g
 control 5
 가

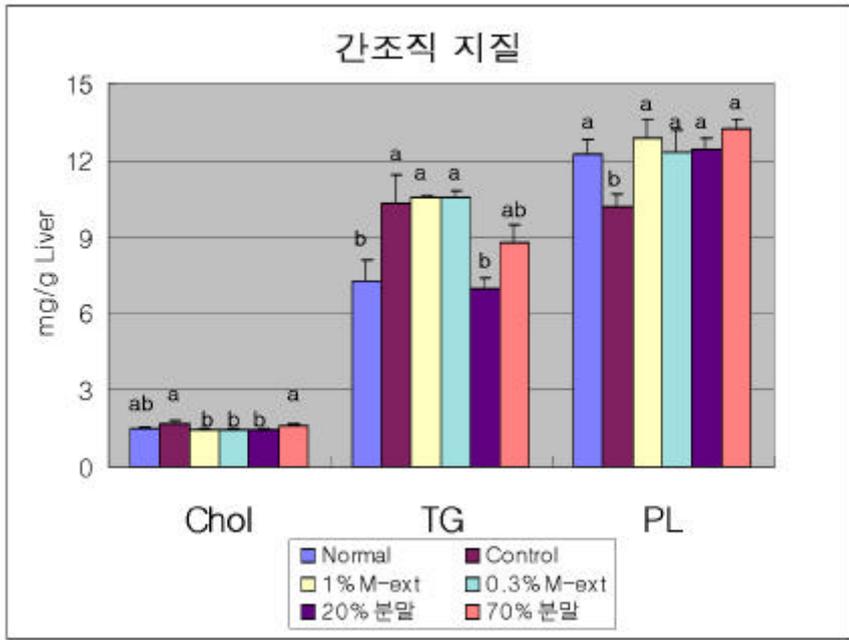


Fig. 3-5 Effect of prosomillet methanol extracts on liver lipids of rats fed high fat diets

(Mean± SEM of 8 rats per group)

HMG- CoA reductase

Fig. 3-6

methanol

Control

70%

methanol

가

(70%)

20%

, 0.3% M- ext

HMG- CoA reductase

Fig.

가

가

가

1% M- ext

HMG- CoA Reductase

normal

가

0.3%

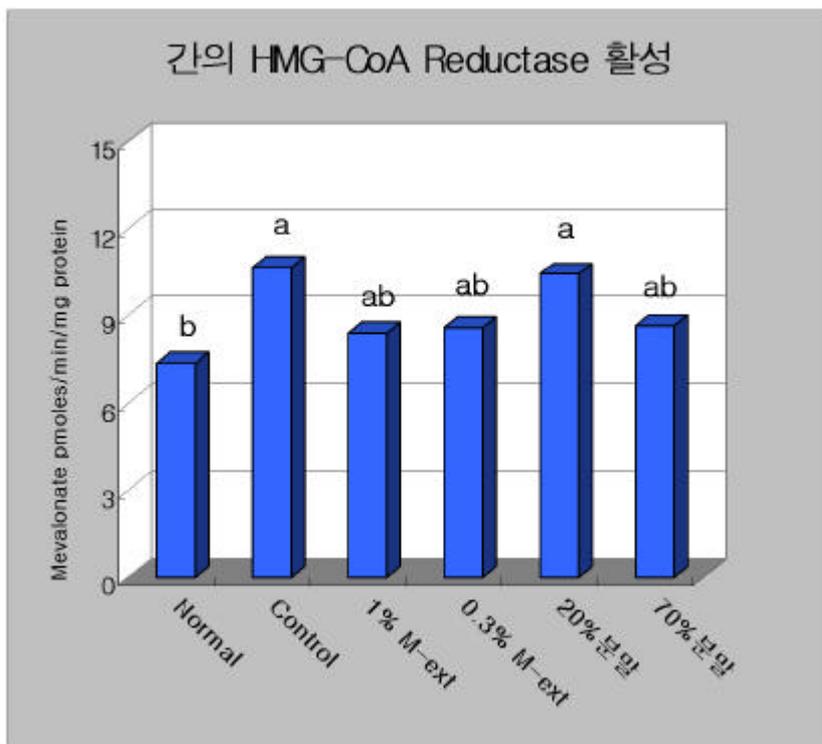


Fig. 3-6 Effect of prosomillet methanol extracts on activities of liver microsomal HMG-CoA reductase of rats fed high fat diets

(pmoles of mevalonic acid/min/mg microsomal protein
 Mean+ SEM of 8 rats per group
 Different alphabet letters in HMG-CoA reductase activity denotes significant different among groups)

Cholesterol 7- α -hydroxylase

Fig. 3-7 cholesterol cholesterol 7- α -hydroxylase

6 normal 20%

10.47 \pm 1.10, 9.95 \pm 1.20 pmol/min/mg protein

70%

13.72 \pm 1.59 pmol/min/mg protein

가 control 1% MeOH

11.27 \pm 0.59, 11.11 \pm 0.62 pmol/min/mg protein

0.3% MeOH 11.99 \pm 1.03 pmol/min/mg

protein

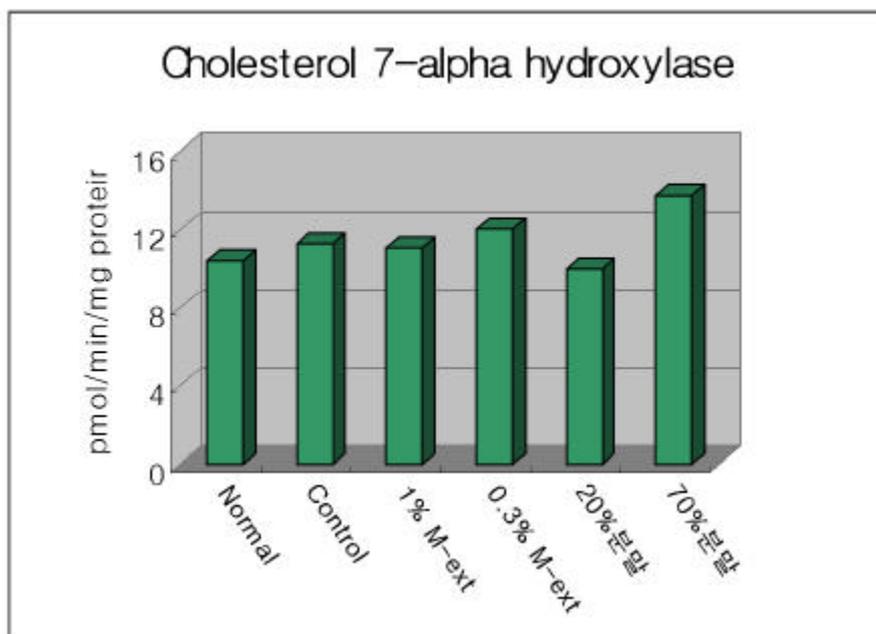


Fig. 3-7 Effect of prosomillet methanol extracts on activities of

liver microsomal cholesterol 7 α -hydroxylase of rats fed high fat diets
(Mean \pm SEM of 8 rats per group)

Glucose-6-phosphate dehydrogenase
NADPH
reducing equivalent
glucose-6-phosphate dehydrogenase (G6PDH) Fig. 3-8 6
cytosol glucose-6-phosphate
dehydrogenase (G6PDH) 가 Fig.
G6PDH control
3% MeOH 70%

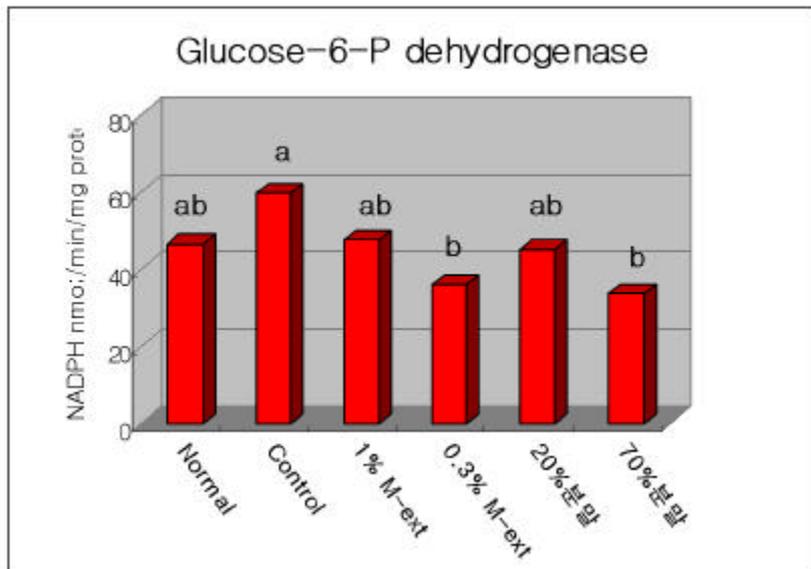


Fig. 3-8 Effect of prosomillet methanol extracts on activities of

liver microsomal cholesterol 7a-hydroxylase of rats fed high fat diets
(Mean± SEM of 8 rats per group)

Fig. 3-9

3

가

가

70%

Table 3-14

cellulose가 3.7%

가 1.3%

cellulose

가

methanol

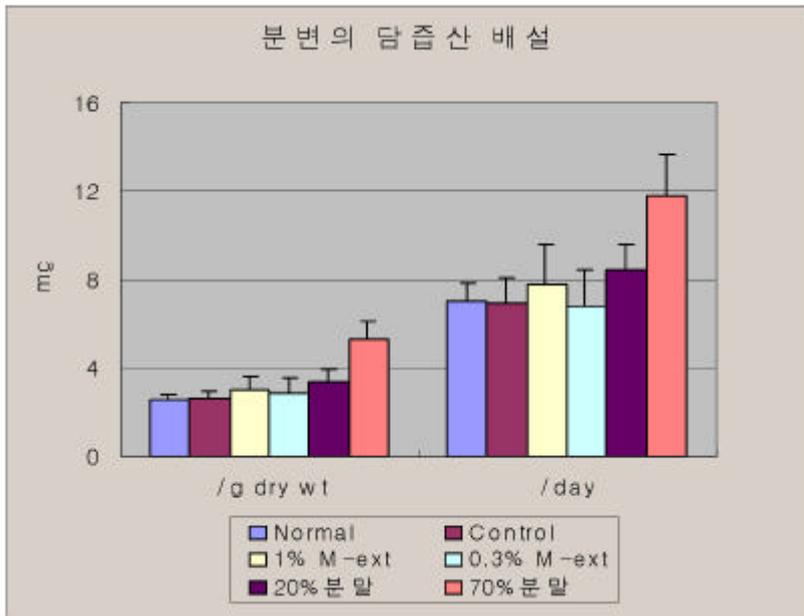
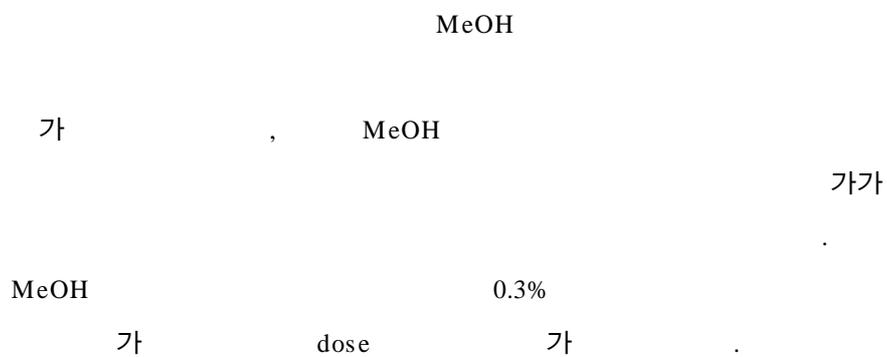


Fig. 3-9 Effect of prosomillet methanol extracts on fecal bile acid excretion of rats fed high fat diets
 (Mean+ SEM of 8 rats per group)



II. high- cholesterol

1.

가.

가
1% (w/w) 가
0.3, 0.6% (w/w) 가
15 30% (w/w) 가 4 ,
가 , 가
6 Table 3- 15
150- 170 g Sprague- Dawley
1 8- 9
4
4 1- 2
,

Table 3-15. General composition and contents of prosomillet powder and methanol extract of high cholesterol diet (g/1,000g)

Group Ingredient	C- Normal (free cholesterol)	C- Control (high- fat/ sucrose)	MeOH- ext		Powder	
			0.3%	0.6%	15%	30%
Casein	200	200	200	200	183	166
DL- Methionine	3	3	3	3	3	3
Starch	150	150	150	150	36	0
Sucrose	450	450	450	450	450	422
Cellulose	50	50	50	50	47	45
Lard	100	100	100	100	97	93
Mineral Mix	35	35	35	35	35	35
Vitamin Mix	10	10	10	10	10	10
Choline	2	2	2	2	2	2
Cholesterol		10	10	10	10	10
Na- tauro - cholate		3	3	3	3	3
MeOH- ext1			3	6		
Powder2					150	300
Kcal/1000g	4100	4047	4035	4023	3996	3765

1 : Prosomillet methanol extract

2 : Prosomillet powder

1)

Cholesterol 가 4 가
ether
cholesterol 가 4 16
ether heparin
2,500 rpm 20

cholesterol 가
saline perfusion , cholesterol 가
3 4 g
- 50

2)

Cholesterol 가 cholesterol
가 cholesterol
2 3 - 50

3)

liver cytosol, microsome 4

가) Cholesterol 7 - hydroxylase

4 300 mM sucrose TEDKSF buffer(300

mM sucrose pH 7.4, 40 mM Tris-HCl, 1 mM EDTA, 5 mM dithiothreitol, 50 mM KCl, 50 mM KF) Potter-Elvehjem type glass homogenizer homogenize

15,000 g 10 gauze

105,000 g 75

microsome pellet . microsome pellet TEDKF buffer(40 mM Tris-HCl, pH 7.4, 1 mM EDTA, 5 mM dithiothreitol, 50 mM KCl, 50 mM KF) suspension cholesterol 7 -hydroxylase 50- 100 $\mu\ell$ - 70

)

4 A(pH 7.0, KH₂PO₄ 50 mM, sucrose 0.2 M, dithiothreitol 2 mM) Potter-Elvehjem type glass homogenizer homogenize

15,000 g 10 gauze

100,000 g 75

cytosol lipogenic enzyme

- 70

1)

cholesterol, HDLc, TG kit

500, 555, 505 nm

Folch

T - chol Pearson

, PL Chen 550 nm, 660 nm

TG Sale chloroform

N2 gas triton X-100

ethanol kit .

2) cholesterol

2 freezer dryer(-70)

200 mg

DeWeal KOH 1 ml

3 autoclave 20% NaCl 1 ml 가 20 ml

2 , c-HCl 0.2 ml 가 20 ml 가 4

MeOH:H2O(5:1, v/v)

kit Crowell 500 nm .

Cholesterol Pearson 550 nm .

3) Cholesterol 7 -hydroxylase

Cholesterol 7 -hydroxylase [4-14C] cholesterol

Junker Story .

mixed soybean phosphatidyl-choline(asolectin)

. D- -phosphatidyl choline 5 g 100 ml,

BHT 0.5 mg asolectin

Substrate Tween 1.5 mg/assay, asolectin 0.5 mg/assay, cholesterol

0.1 μm/assay [180 μM] , [4-14C]- cholesterol 0.1 μCi/assay

TEDKF buffer 100 μl/assay 가 1-3 micropip

sonication bath sonication

. Cholesterol 7 -hydroxylase

500 $\mu\ell$ TEDKF buffer(pH 7.4), substrate 100 $\mu\ell$

(200- 500 μM cholesterol with 0.1 μCi [4- ^{14}C]- cholesterol/assay),

microsome 400 800 μg protein(20 30 mg/ml 10- 40 $\mu\ell$)

37 60 preincubation , NADPH, 0.6

nmole 가 50 , CHCl_3 :MeOH(2:1, v/v) 7.5 ml 가

. 1 ml 가 vortex 4

, , 3 ml

CHCl_3

80 $\mu\ell$ CHCl_3 ethylacetate:hexane(8:2, v/v)

TLC

TLC 7 -hydroxycholesterol

. 7-ketocholesterol 100 mg 6

ml methanol 100 mg NaBH_4 가 1

10 ml 가 . ether:benzene(1:1, v/v) 20 ml 2

5 ml 3 Na_2SO_4 가 .

, CHCl_3

. TLC image analyzer

scintillation counter .

1 microsome 1 mg

7 -hydroxycholesterol pmole .

4)

가) Glucose- 6- phosphate dehydrogenase

Cuvette tris buffer(pH 7.4) 37 mM, glucose- 6- phosphate disodium

salt 7.4 mM, NADP 0.07 mM, MgCl₂ 7.4 mM

cytosol 0.1 ml 가 2.7 ml

NADP가 G6P NADPH가 340 nm

) Malic enzyme

Cuvette triethanolamine(pH 7.4) 68 mM, L-malate 0.51 mM, MnCl₂
4.7 mM, NADP 0.14 mM cytosol 0.1 ml 가

2.95 ml NADPH 340
nm

bovine serum albumin

cytosol, cell microsome Lowry microsome Biuret

SAS ANOVA Tukey test

2.

가.

Table 3- 16

cholesterol (Powder)	(Control) cholesterol	가	가 (MeOH- ext) (Normal)
	cholesterol	가	Normal
	(g/100g bw)가	가	
	cholesterol	가	
Normal			가
Control			

Table 3-16. Effect of dietary prosomillet on growth of rats on high cholesterol diets

Group	Body weight (g)			Feed efficiency g/100g diet	Relative liver weight g/100g bw
	Initial	Final	Gain		
C- Normal1	197.3 ± 9.7	313.8 ± 6.4	116.5 ± 10.9	22.0 ± 0.02	2.67 ± 1.43
C- Control2	193.0 ± 9.1	320.0 ± 8.3	127.0 ± 5.3	24.9 ± 0.01	4.02 ± 2.62
MeOH- ext					
0.3%	190.5 ± 7.3	320.6 ± 4.1	130.1 ± 5.0	23.4 ± 0.02	4.17 ± 5.15
0.6%	193.5 ± 7.9	313.8 ± 8.2	120.3 ± 12.6	24.9 ± 0.02	4.20 ± 1.53
Powder					
15%	187.9 ± 7.2	308.8 ± 7.7	120.9 ± 12.9	24.0 ± 0.02	4.15 ± 2.61
30%	188.9 ± 8.4	300.6 ± 4.8	111.8 ± 6.4	18.9 ± 0.02	3.80 ± 0.14

1 : High fat-sucrose without cholesterol diet group

2 : High fat-sucrose with 1% cholesterol diet group

Mean ± SE of 8 rats per group

Table 3-17 T- chol, TG, HDLc, PL HDLc/TC ratio
 (C- Normal) T- chol cholesterol 가
 T- chol 0.3% (MeOH- ext) 가
 (Powder)
 0.6% MeOH- ext
 TG (C- Control) C- Normal
 HDLc cholesterol 가 5
 C- Normal 가
 PL C- Control
 가 HDLc/TC ratio C- Normal cholesterol
 가

Table 3-18 T- chol, TG cholesterol 가 5 cholesterol 가 C- Normal
 가 (Fig. 7)
 C- Control
 T- chol, TG
 가
 PL C- Control 가
 MeOH- ext Powder

Table 3-17. Concentrations of plasma lipids of rats on high cholesterol diets

Group	Tchol (mg/100ml)	HDLc	HDLc/T - chol	TG (mg/100ml)	PL
C- Normal1	100.6 ± 11.0c	45.1 ± 2.0a	0.488 ± 0.06a	34.1 ± 7.5ab	16.62 ± 0.86a
C- Control2	273.1 ± 47.3a	19.4 ± 2.4b	0.103 ± 0.03b	42.6 ± 6.8a	15.18 ± 2.74b
MeOH- ext					
0.3%	188.2 ± 43.1b	16.5 ± 1.5bc	0.088 ± 0.02b	20.4 ± 6.0b	15.67 ± 0.60ab
0.6%	285.3 ± 15.8a	13.6 ± 1.1c	0.070 ± 0.026b	26.9 ± 2.4ab	15.48 ± 1.12ab
Powder					
15%	249.0 ± 21.4ab	18.3 ± 1.5bc	0.079 ± 0.012b	22.1 ± 3.4b	16.19 ± 1.25ab
30%	218.6 ± 29.2ab	15.7 ± 1.9bc	0.083 ± 0.018b	23.5 ± 2.8b	14.82 ± 1.56ab

1 : High fat-sucrose without cholesterol diet group

2 : High fat-sucrose with 1% cholesterol diet group

Mean ± SE of 8 rats per group.

Values in the same column not sharing common superscript letters are significantly different at $p < 0.05$ by Tukey's test.

Table 3-18. The concentration of liver lipids of rats on high cholesterol diets

Group	Liver (mg/g liver)		
	cholesterol	TG	PL
C- Normal1	1.90 ± 0.31b	14.82 ± 1.24c	3.93 ± 0.16a
C- Control2	27.88 ± 4.39a	37.91 ± 4.8a	3.43 ± 0.12b
MeOH- ext			
0.3%	22.88 ± 4.02a	32.06 ± 4.10b	3.47 ± 0.10ab
0.6%	24.42 ± 2.25a	34.31 ± 2.97b	3.54 ± 0.17ab
Powder			
15%	25.06 ± 3.68a	29.97 ± 3.19b	3.72 ± 0.19ab
30%	25.28 ± 3.20a	35.62 ± 3.15ab	

1 : High fat- sucrose without cholesterol diet group

2 : High fat- sucrose with 1% cholesterol diet group

Mean ±SE of 8 rats per group.

Values in the same column not sharing common superscript letters are significantly different at $p < 0.05$ by Tukey's test.

cholesterol
 Table 3- 17 3 cholesterol
 (C- Normal) cholesterol 가
 5 cholesterol 가
 cholesterol (C- Control)
 가 (Powder)

Table 3- 19. Fecal excretion of bile acid and cholesterol of rats on high cholesterol diets

Group	Bile acid		Cholesterol	
	mg/g fecal	mg/day	mg/g fecal	mg/day
C- Normal1	10.12 ± 1.57c	11.18 ± 1.32c	9.94 ± 0.39b	11.50 ± 1.20c
C- Control2	35.16 ± 0.69ab	51.22 ± 3.26ab	65.11 ± 10.99a	92.70 ± 15.33b
MeOH- ext				
0.3%	38.5 ± 2.16a	56.13 ± 2.88ab	75.10 ± 3.20a	109.68 ± 4.97ab
0.6%	31.14 ± 2.37b	46.86 ± 2.90b	68.13 ± 2.01a	104.52 ± 8.15ab
Powder				
15%	37.77 ± 2.38a	59.56 ± 2.10a	75.21 ± 4.24a	118.90 ± 4.28a
30%	39.43 ± 3.69a	61.55 ± 9.30a	74.06 ± 3.32a	114.09 ± 9.20ab

1 : High fat-sucrose without cholesterol diet group
 2 : High fat-sucrose with 1% cholesterol diet group
 Mean ±SE of 8 rats per group.

Values in the same column not sharing common superscript letters are significantly different at $p < 0.05$ by Tukey's test.

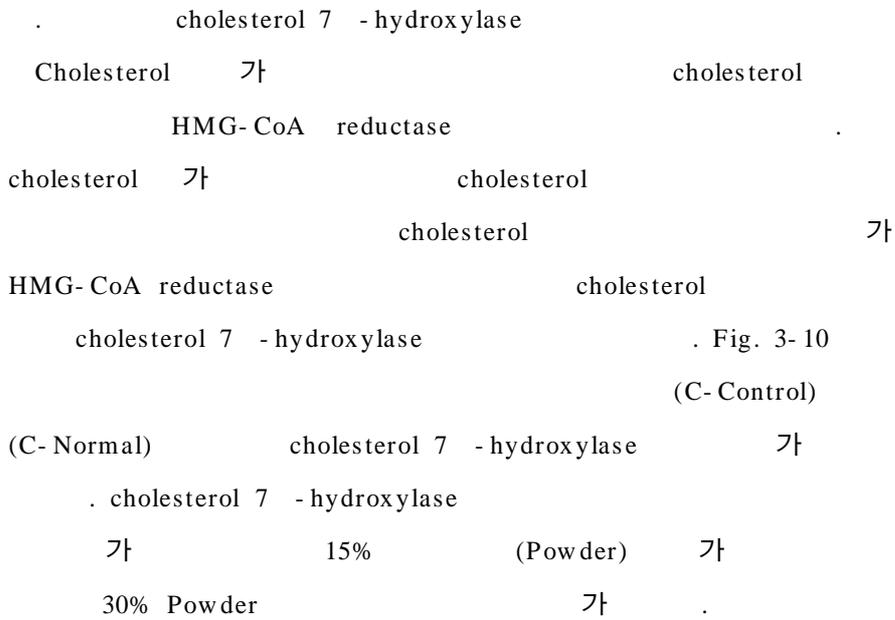
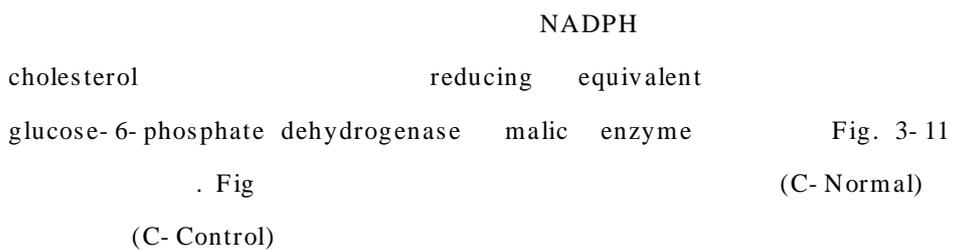
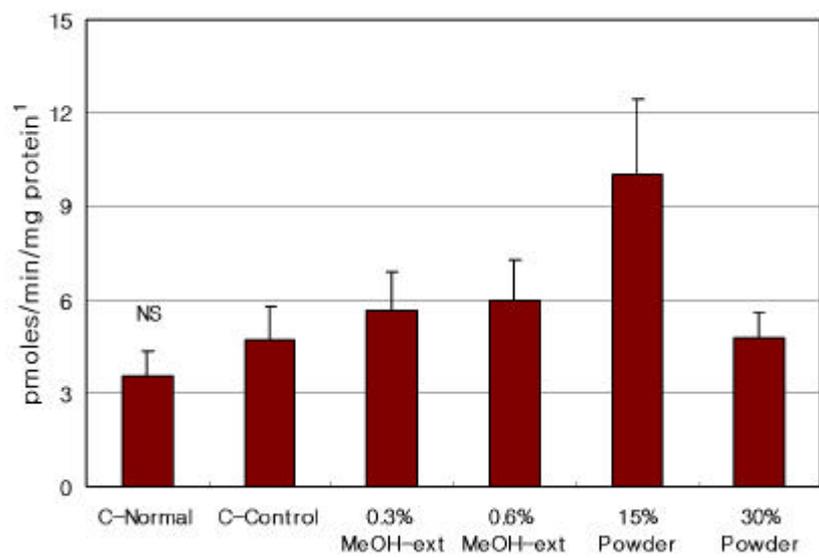


Fig. 3-10. Activities of liver microsomal cholesterol 7-hydroxylase of rats on high cholesterol diet



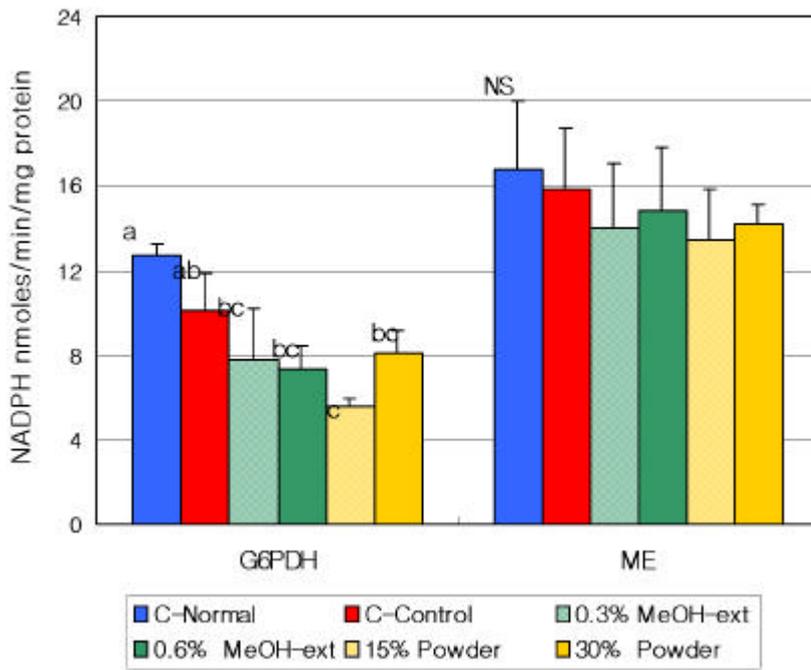


G6PDH

TG가

cholesterol 가

Fig. 3-11. Activities of liver cytosolic lipogenic enzyme of rats on



high cholesterol diet

4

2

(CA)

1.

가.

200-220g SD

8-10 4

, +0.15% CA , ,
+0.15% CA 4 .

3 1,2,3,

0.15% 가

4

.

total cholesterol

triglyceride .

.

T - test .

2.

가.

(Table 3-20, 21)

CA 가

가

.

.

Table 3-22

CA . ,

CA 가
 35% , 31% CA가
 가 . total cholesterol
 CA 가 . free
 cholesterol total cholesterol
 . free fatty acid
 30% CA
 . 2
 가
 가 . CA CA

Table 4-20. Effect of CA on body weight gain, food intake of experimental rats.

	High fat control	High fat 0.15% CA	High chol control	High chol 0.15% CA
Initial BW (g)	230.6 ± 7.41	230.4 ± 3.59	231.3 ± 2.43	235.5 ± 3.48
Final BW (g)	382.1 ± 8.98	373.4 ± 13.03	386.0 ± 6.08	389.5 ± 13.5
Total intake(g)	555.4 ± 15.1	547.1 ± 20.4	594.6 ± 10.4	614.1 ± 24.3
Weight gain (g)	148.4 ± 7.99	143.0 ± 14.0	154.6 ± 7.79	154.0 ± 11.2
FER	0.27 ± 0.01	0.26 ± 0.02	0.26 ± 0.01	0.25 ± 0.01

Table 4- 21. Effect of CA on organ weights of experimental rats

	Liver (g)	Kidney (g)	Spleen (g)
High fat control	17.22 ± 0.61	2.73 ± 0.08	0.90 ± 0.05
High fat +0.15% CA	16.57 ± 1.19	2.57 ± 0.12	0.81 ± 0.06
High chol control	13.96 ± 0.42	2.69 ± 0.04	0.80 ± 0.06
High chol +0.15% CA	14.49 ± 0.59	2.86 ± 0.04*	0.82 ± 0.03

Table 4- 22. Effect of CA on concentration of serum lipids of experimental rats

	High fat control	0.15% CA	High cholesterol control	0.15% CA
Triglyceride (mg/dl)	110.4 ± 11.8	71.95 ± 3.48*	141.56 ± 13.9	97.61 ± 5.98**
Total Cholesterol (mg/dl)	77.8 ± 3.59	74.71 ± 2.75	75.36 ± 4.51	62.62 ± 3.04*
HDL cholesterol (mg/dl)	37.8 ± 2.95	40.46 ± 2.29	40.41 ± 3.15	38.80 ± 2.35
Free cholesterol (mg/dl)	13.69 ± 1.14	13.93 ± 0.82	13.08 ± 1.24	10.26 ± 0.65*
Free fatty acid (µEq/L)	995.7 ± 47.2	697.4 ± 39.3***	914.0 ± 107.1	756.7 ± 26.92

* p<0.05

** p<0.01

*** p<0.001

TBARS

Table 4-23

CA

가

CA가

34%

CA

TBARS

Table 4-23. Effect of CA on contents of liver triglyceride, cholesterol and TBARS of experimental rats

	Cholesterol (mg/g liver)	Triglyceride (mg/g liver)	TBARS (MDA nmole/g liver)
High fat con	3.56 ± 0.19	18.22 ± 1.46	45.28 ± 2.14
0.15% CA	3.02 ± 0.12*	17.59 ± 2.20	35.86 ± 1.31**
High chol con	29.7 ± 1.53	25.89 ± 1.93	51.43 ± 3.54
0.15% CA	19.6 ± 1.45**	26.51 ± 1.51	37.30 ± 5.79

malic enzyme G6PDH
malic enzyme glucose-6-phosphate
dehydrogenase(G6PDH) CA Table 4-24
Table , CA
가 malic enzyme G6PDH Malic
enzyme G6PDH NADPH
cholesterol reducing equivalent
CA 가 TG free fatty
acid CA TG
CA
HMG- CoA reductase
level

Table 4-24. Effect of CA on activities of malic enzyme and G6PDH of liver in experimental rats

	Malic enzyme (nmoles/min mg protein)	G6PDH (nmoles/min mg protein)
High fat con	1.523 ± 0.03	2.718 ± 0.173
0.15% CA	1.128 ± 0.07**	2.130 ± 0.141*
High chol con	1.833 ± 0.06	6.06 ± 0.20
0.15% CA	1.562 ± 0.03**	2.73 ± 0.10***

. Postprandial triglyceride

CA 1 2 0 time
 soybean oil, 1ml/rat CA
 () 1
 TG 3 TG 가
 CA 가 2 가
 2 가 CA 6
 TG CA oil
 가

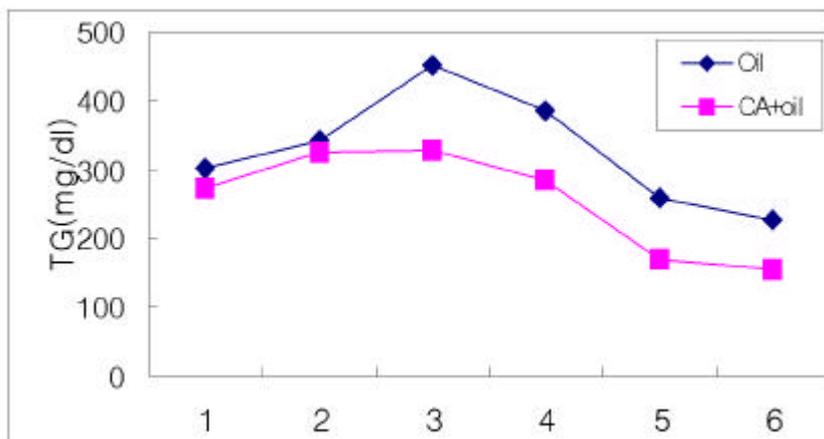


Fig. 3- 12 Effect of CA on postprandial triglyceride of rat

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4

가

, ,

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가

, ,

b- glucan

, phenolic

compounds,

rutin

.

2, 3

, , *in vitro*

HMG- CoA reductase

in vivo

가 .

가 가

가

가

가

.

1

1.

가.

in vitro
 , , ,
 , , ,
 cyclotec mill .

. Amylograph

amylogram Viskograph(Brabender, Germany)
 . 11% .

. WSI

(WSI) 50 mL 2 g
 25 mL 가
 36 1 15000 rpm 1
 1 mL (%) .

. WAI

(WAI) WSI
 (%) .

. Gel consistency

gel consistency amylograph 60
 water bath 10 40 oven
 (Bostwick consistometer, model Cento, U.S.A.) 50 g 30

. Viscoelasticity
 (black rice), (barley), (millet), (buckwheat),
 (sorghum) 5가 Advanced Rheometric
 Expansion System(2ARES- 11A, Rheometric Scientific, UK)
 . miller(cyclotec 1093 sample mill,
 Tecator, UK) (80 - 100mesh)
 가 50%
 30 가
 strain sweep(0.1 20%) strain
 10% frequency sweep(0.1 100 rad/sec)
 25 plate 25mm parallel plate
 .
 .
 100g . 16
 0 , 180 , 200 200 가
 160 가 180
 5, 10, 15 .

. Puffing

puffing 15 가 puffing
 , 가
 puffing
 , , , AOAC

puffing ,
CR-200 Chroma meter (Minolta
Inc., Japan) L(), a(), b() 5
Somoge- Nelson

2.

가. Amylogram

amylogram Table 4-1

가	1320 BU,	1110BU,	940BU,	755BU,
165BU	.		setback	
760BU	가	,	,	,
	가	가		.
				amylose
				.
73.5	가	가 69	, 64	, 63.5
가 62.6	가			.

Table 4-1. Amylograph characteristics for various cereals

	black rice	buckwheat	sorghum	prosomillet
Peak viscosity(B.U.)	165	1110	940	755
Hot paste viscosity(B.U.)	90	1035	430	225
Cold paste viscosity(B.U.)	100	1870	605	510
Breakdown (B.U.)	75	75	510	530
Total setback(B.U.)	10	835	175	285
Setback(B.U)	- 65	760	335	- 245
Initial pasting temp.()	64	63.5	69	73.5

. Gel consistency, WSI, WAI

Table 4-2 . Consistometer

	가 11.1 cm,	, 8.9 cm,	가
3.95 cm	0.3 cm	가	
가	가 59.51%	가	
46.92%	가		0.44%
가	, 0.2%		가

Table 4-2. Gel consistency, WAI and WSI of various cereals

	black rice	buckwheat	sorghum	prosomillet
Gel consistency(cm)	- *	0.3	11.1	8.9
WSI(%)	47.79	51.57	46.92	53.13
WAI(%)	0.26	0.20	0.24	0.44

*:

. Viscoelasticity

modulus), G'/G'' G' (storage modulus) G'' (loss modulus), $\tan \delta$, Eta^* Fig. 4- 1, 2, 3, 4 . frequency가 가 tan

가 , , frequency 가 가 tan 가 gel consistency index . G' G'' rubber like amylose glutinous rice .

L a b 가

(Table 4- 3).

가 15 10 . 가 , 가 (Table4- 4).

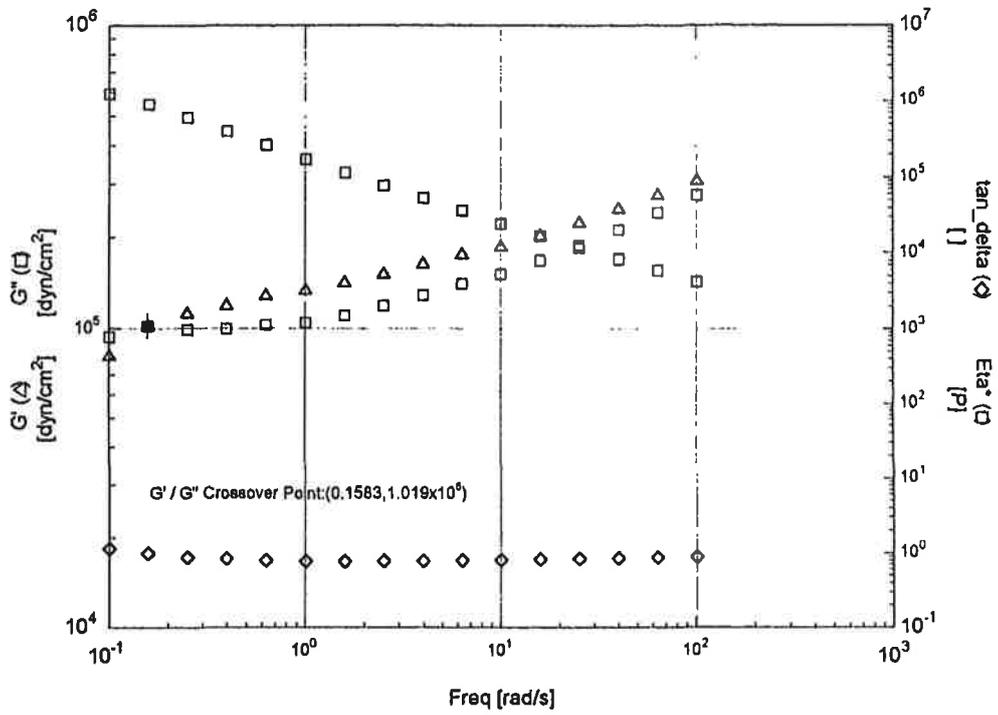


Fig. 4-1 Viscoelasticity of black rice flour

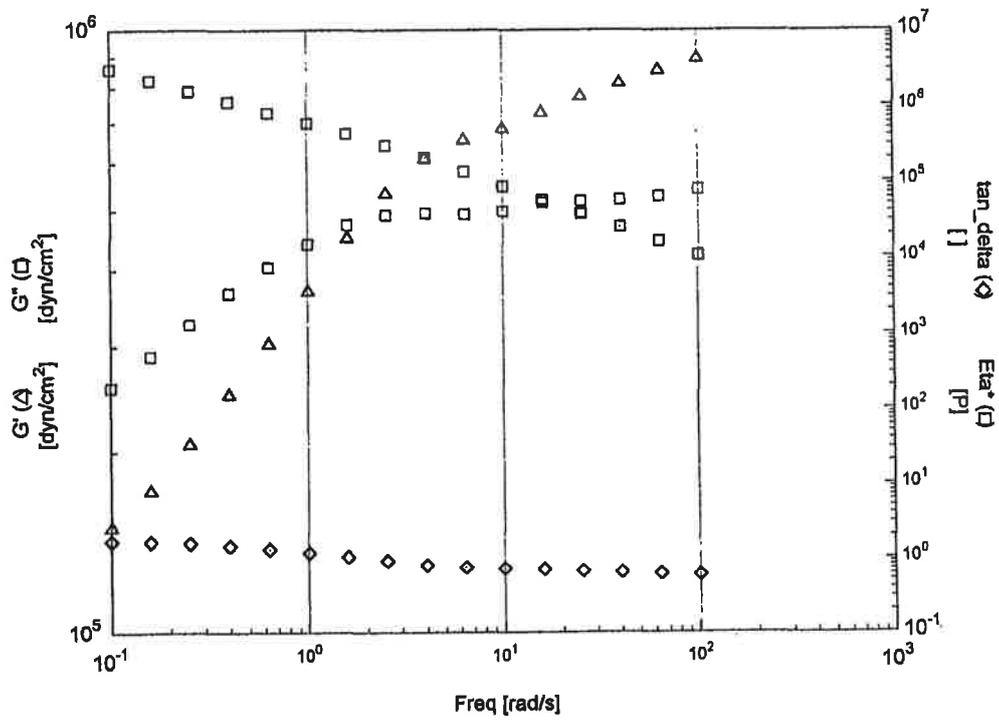


Fig. 4-2 Viscoelasticity of buckwheat flour

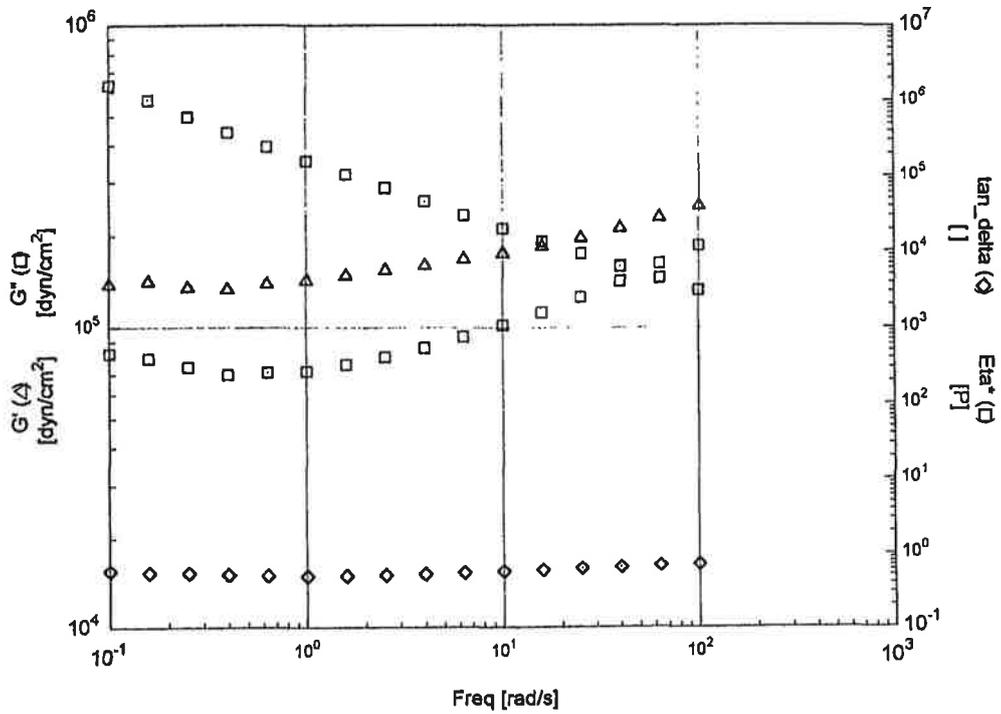


Fig. 4-3 Viscoelasticity of sorghum flour

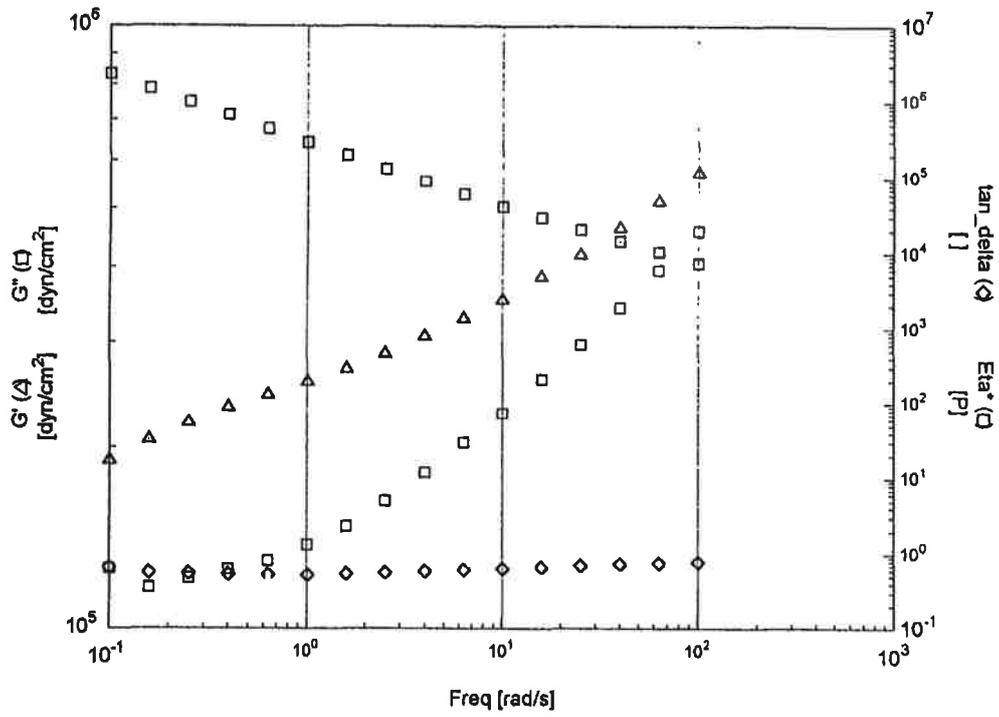


Fig. 4-4 Viscoelasticity of proso millet flour

Table 4-3 Changes in color values of various cereals during roasting at 180

		Grain			Powder		
		L	a	b	L	a	b
Prosomillet	5min	59.96	5.46	27.27	89.55	-0.92	14.24
	10	58.65	7.58	31.54	83.47	1.27	16.14
	15	50.29	9.42	35.73	79.53	2.18	17.71
Buckwheat	5min	20.73	0.20	3.09	87.55	0.33	11.06
	10	37.06	9.31	14.30	71.17	4.80	20.64
	15	60.26	5.00	28.13	63.75	5.85	21.84
Sorghum	5min	69.77	3.63	22.76	81.48	3.20	12.15
	10	44.94	9.42	21.68	73.36	4.25	16.67
	15	33.24	8.13	17.12	67.89	3.99	17.09
Brown rice	5min	60.16	6.73	28.73	82.57	0.87	15.04
	10	57.31	6.44	23.88	68.00	4.53	20.93
	15	31.21	6.15	12.18	65.22	4.53	21.44
Black rice	5min	56.70	7.26	30.13	62.92	2.51	6.10
	10	38.59	9.11	18.38	60.15	3.40	12.98
	15	21.20	0.09	2.84	57.19	3.71	15.10

Table 4-4. Changes in proximate composition and reducing sugar of various cereals during roasting

		Moisture	Protein	Fat	Ash	Reducing sugar
Prosomillet	5min	3.41	11.73	1.33	0.60	70.27
	10	1.54	12.54	1.16	0.72	80.15
	15	0.86	12.78	0.93	0.62	39.91
Buckwheat	5min	3.18	13.10	3.30	2.34	81.32
	10	0.84	14.16	3.29	2.39	81.92
	15	0.95	14.77	3.38	2.22	40.81
Sorghum	5min	5.93	10.63	3.21	1.31	53.73
	10	2.69	11.07	2.79	1.53	74.12
	15	1.14	11.36	3.00	1.29	55.14
Brown rice	5min	5.67	7.81	5.88	1.24	129.47
	10	1.77	8.13	2.08	1.23	90.28
	15	1.32	8.14	2.11	1.21	54.08
Black rice	5min	5.75	10.65	2.69	1.52	75.20
	10	2.05	11.33	2.62	1.97	100.68
	15	1.20	11.39	2.90	1.73	38.69

2

가

1.

가.

1997

40

(Food mixer, modle

Hanil FM707T)

40

가

1

가

가

AOAC

Prosky

total dietary fiber

kit(Sigma

Chemical Co. St. Louis, MO)

가

가

Brabender

Visco/Amylograph (Duisburg, Germany)

가

0, 10, 20, 30%

가

11% (w/w)

35

95

1.5 /min

가

95

30

50

2.4g, 226g, 20g, 42g, 180g, 566g, 340g, 340g
 가, hand mixer (National, Japan)
 1 20
 65g baking oven 190, 190
 20 baking 10, 20, 30% 가

baking 1

CR-200 Chroma meter (Minolta Inc., Japan) L(), a(), b() 5
 baking 1
 (5x5x3 cm) TA-XT2 Texture analyzer (Texture Technologies Corp., Scardale, NY)
 Texture profile analysis(TPA)parameter hardness, adhesiveness, springness, cohesiveness, gumminess, chewiness

46

. , 가 가 0, 10, 20, 30% 가
, , , 1() 9(
) 가 . 10
가
SAS ANOVA Student
Newman Keuls Test .

2.

가.

		가 (wet basis)			
가	9.1%	2.4%,	8.9%,	1.3%,	
가 3.2%	,	10.4%,	0.7%,	8.4%,	
가 0.9%					
	가				.

.

Table 4-5

		가		가	
		,		가 0%	(가 100%)가
550 B.U.		가 30%	가	690 B.U.,	가
100	830 B.U.		가	가	가
가	가 .	95	15		가
				가 0%	510 IU.
	가 100%		370IU.		.
가	가				.
	가 0%	(가 100%)	59.7		가
30%	67.3	가	가	가	가
	가	가			
가					가
	가		Carcea		.

Table 4-5. Amylograph characteristics for wheat flour substituted with sorghum flour 10, 20 and 30% levels

(11%, dry basis)

Sorghum flour contents(%)	Peak viscosity (B.U.)	15- min height (B.U.)	Final viscosity (B.U.)	Pasting temperature()
0%	550	510	1105	59.7
10%	575	500	1045	63.0
20%	570	445	920	64.2
30%	690	520	990	67.3
100%	830	370	550	69.2

가 가 가
 L a , b .(
 ;L 95.6, a - 0.76, b 7.04, 가 ;L 79.3, a 5.3, b 11.7).
 가 가 Table 4-6
 . L 가 가 가
 . a
 가 가 가
 b 가 가 20% 가 30% 가
 가 0% .
 가 가 가 가 ,
 anthocyanin, anthocyanidin . 가 가

(Table 4-7) 가 10% 가
 20% 가 140cc 130.5cc 30% 가
 125.5cc . Johnson 가
 가 가 가 가
 가 가 가 가
 10% 가 20, 30% 가
 가 가 가 가
 가 가 가 가
 가 가 가
 가 가

Table 4-6. Color values for the muffins with sorghum flour substitution at 10, 20 and 30% levels

Sorghum flour content(%)	L value	a value	b value
0%	77.01a	- 4.33d	22.07a
10%	70.93b	- 0.11c	19.67c
20%	64.93c	2.18b	19.88c
30%	60.17d	3.83a	20.74b

The same superscript letters in each column are not significantly different($p < 0.05$)

Table 4-7. Baking properties for muffins with sorghum flour substitution 10, 20 and 30% levels

Sorghum flour content(%)	Volume(cc)	Weight(g)	Width(cm)	Height(cm)
0%	140.0 ± 5.66a	58.9 ± 0.35	6.2 ± 0.17	5.7 ± 0.19a
10%	138.0 ± 0.00a	59.0 ± 0.17	6.3 ± 0.15	5.6 ± 0.17ab
20%	130.5 ± 0.71ab	59.3 ± 0.80	6.2 ± 0.11	5.4 ± 0.21b
30%	125.5 ± 4.95b	59.2 ± 0.30	6.2 ± 0.12	5.1 ± 0.13c

가 가

Hardness, adhesiveness, springness, cohesiveness, gumminess, chewiness

texture analyzer

(Table 4-8). Hardness

가 가 20%

30% 가

가 가

가

. Springness

가 가 0%

가

가

가 가

가 . Cohesiveness

가 가 20%

hardness

30% 가 hardness

Table 4-8. Texture profile analyse for muffins with sorghum flour substitution at 10, 20 and 30% levels

Sorghum flour content(%)	Hardness	Adhesive-ness	Spring-ness	Cohesive-ness	Gummi-ness	Chewi-ness
0 %	352.36b	- 3.30	0.895a	0.472a	166.36	148.84
10%	350.11b	- 5.19	0.853b	0.463a	162.13	138.39
20%	346.13b	- 5.78	0.853b	0.459a	159.13	135.61
30%	406.79a	- 8.91	0.820b	0.426b	173.57	142.37

가 0, 10, 20, 30% 가
 Table 4-9 가 7.37 가
 가 가 30% 가
 4.89 가 가
 가 가 가 10,
 20% 가
 가 가 가
 가 가
 10% 가 가
 가 가 가

Substitution Level	Adhesiveness	Gumminess	Chewiness	Hardness	Cohesiveness	TPA
30%	가	가	가	가	가	가
20%	가	가	가	가	가	가
10%	가	가	가	가	가	가

Table 4-9. Sensory characteristics for muffins with sorghum flour substitution at 10, 20 and 30% levels

Sorghum flour content(%)	Appearance	Flavor	Taste	Texture	Overall acceptance
0%	7.3a	6.6a	6.0	6.1a	6.0
10%	5.4b	5.8b	5.6	6.0a	5.8
20%	5.3b	5.7b	5.6	5.4ab	5.6
30%	4.8b	5.5b	5.4	4.9b	5.5

,
 가 0, 10, 20, 30% 가
 가 - 가
 가 가 가 가 가
 a 가 . 가 가 20%
 . Adhesiveness, gumminess, chewiness 가 가
 , hardness cohesiveness 20% 가
 가 . , , , 가
 가
 10% 가 가
 가 가 가 가 .
 가 가 가
 가 20% 가가 가

2
 2)
 50% 가 10
 가 2
 3)
 200 20 200g
 590nm OD
 (, , , , , ,)
 가
 , ,
 37
 가 , ,

2.

○

가

○

1

100g 500ml

가 가

(Brewers

amylliq T.S) 2-3 가 5 가

500ml 가

1000ml

1000ml

100g,

2g,

10g

가

2-3

가

hot

filling

10

가 . 9 가
 . 5
 . 37 2
 가 .
 가
 가
 가 .

○ 2

, ,

(Table 4- 10, 11).

200

20

150g

1500ml

가

가

가

Table 4- 10

B, C

A

B, C

A

B

C가 가

A가 가

B, C

A

가

A

B, C

가 10

가

3

가

Table 4- 11.

	A	B	C
	500	500	500
	37.5	-	-
	25	-	-
	12.5	15	15
	1.25	-	-
	12.5	15	15
	1	1.06	1.06
	-	20	20
	-	0.55	0.55
	-	0.165	-
(590nm OD)	17.19	1.64	1.52
(Brix)	9.1	10.8	10.6

Table 4- 12.

	A	B	C
	5.42b	7.50a	7.35a
	6.57a	4.23b	5.73a
	6.18a	4.52b	3.89b
	4.97b	7.12a	7.25a
	5.51ns	6.34	6.57

Table 4- 13. (g)

D	E	F	G
500	500	500	500
25	20	20	20
.	.	5	5
0.25	0.25	0.5	0.5
5	20	10	10
10	10	15	15
0.25	0.25	.	.
5	.	5	.
.	0.20	.	.
.	0.015	.	.
.	.	.	0.015

Table 4- 14.

	D	E	F	G
	5.0ab	7.25a	3.75b	6.75a
	5.75ns	7.25	4.75	6.50
	4.50b	4.50b	6.75a	7.25a
	6.0ab	4.25b	6.50ab	6.75a
	6.0b	4.0b	6.75a	6.75a

○ 4

2

Table 4- 15

가

H, I, J

H

J

가가

J가

가

가

가

37

10

J

가

4- 15. (g)

	H	I	J
	500	500	500
	10	10	10
	10	10	13.5
0.53			
25	25		17.5
1.25			
25	25		17.5
1	1		0.5
0.14			0.26
0.01			0.015

Table 4- 16.

H	I	J
7.37 ^{ns}	7.07	7.64
7.91 ^{ns}	7.41	8.35
7.03 ^{ns}	6.89	7.15
7.56 ^{ns}	6.96	7.97
7.55 ^b	7.35 ^b	8.23 ^a

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