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최 종 연구보고서

밤을 이용한 기능성 발효제품의 개발

Development of a Fermented Food Product
Using Chestnut

연구기관 전주대학교

농 림 부

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MIR1, M2R1, M2R6, M3R3, F1R4 .

2) MGG1, MGG2, NBL1, PAP1, PAP2 .

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fructose-6-phosphate phosphoketolase acetate/lactate

MIR1, M2R1, M2R6, M3R3, NBL1, PAP2 Bi fi dobactri um

F1R4, MCG1, MCG2, PAP1 Lactobaci ll us .

- 4 -

1) 0. 81-2. 11 ml (0. 1N NaOH) 2) 2% 32% 가 가 가 F1R4 PAP1가 가 , 4% 8% 가 **8**% 3) MIR1, MGG2, MGG1, F1R4, PAP1 8% 32% 8% 가 가 1) 가 가 0. 5-8% Lactobacillus 가 가

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0. 05% 0. 4% cysteine, 0.2% yeast extract Gel ysate peptone, Phyton peptone, Trypticase peptone 가 Phyton peptone 가 F1R4 PAP1 가 가 3) Lactose, maltose, sucrose glucose 4) 가 가 0. 5-8% 가 가 2. 0-8.0% 0.5-2% **4**% 5) 10% 가 F1R1, MGG1, PAP1가 가 , F1R4 , MGG1 , **PAP1** 가 6) 가 가 **48** 가 , 24

24 48 F1R4, MGG1, PAP1 가 가 7) 가) **8**% 24 8.05-9.05 가 MIR1 PAP1 9.05 9.03 PAP2 . 48 8. 28-9. 15 M2R1가 9.15, 8.99 가 24) 1: 1 **20ml** 0. 8ml 가 48 F1R4 3. 26, 3. 66 MGG1, MGG2, NBL1가 PAP1 9. 22, 9. 16, 9. 24 가 가 가가 가 가 가 8) 가 ()) 가 가 가 가

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F1R4, PAP1 9.05, 9.32 가 ,

5. 26, 5. 21 1. 30- 2. 45

F1R4 PAP1 가 .

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(5) 3 MGG1 MGG27 | 8.00 8.03

가 8.66 8.83 가 .F1R4

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. F1R4 PAP1 가 가

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рН

6. 76 4. 38 1000

pН 4) 가 10% , MGG1 1. 43 8. 61 , 23% 가 1.08 8.26 4. 42 8. 00 F1R4 4. 05 7. 56 . 12 MGG1 가 1.43 1. 13 7. 03 , F1R4 8. 14 4. 70 23% 3. 02 4. 40 3. 68 MGG1 F1R4 5) 1 10 가 6) 가) Lactobacillus PAP1

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Lactobacillus MGC2

가 3 pH 5. 17) PAP1 MGG2 , 5 1:1 3 2 3 6. 62 3.65 가 가 2 가 3 3 3.65 4. 60, 5. 88 7.64 가 .) Hansen YC-180(S. thermophilus L. bulgari cus) MSK B2(S. thermophilus, B. infantis aci dophi lus) , 24 Hansen

0. 5% . 0. 1%(MRS 18)

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가 가 .

Hansen

가 MSK

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가 1) 가 가(0.5%) 가 가 yeast extract Phytone 가 Phytone peptone peptone 0.8%, yeast extract 0.8%, glucose 0.5% 가 5. 11 9. 42 , glucose 0.5% yeast extract 가 가 0. 2% , Phytone pepton 0.4% 3.89 9.0 2) Phytone pepton Yeast Exract 가 가 Yeast Extract 가가 1/3 가 가 Phytone peptone 가 가 3) 40%(V/V) 가 가 4) 가 pectin(UnipectineTM AMP285) sodium alginate 가 al gi nate 0.5% 가 , pectin al gi nate가

- 11 -

5). 0.4%, 0. 2%, $0.5\%\!\!$, sodium alginate $0.2\!\!$, modified starch $2\!\!$ 가) 가 10%가 가 **5**% **5**%) 3. 0-2. 5) 0.08, 0. 01, 0. 01% 가 가 5%, 5%, 0. 08%, 3.0 0. 01%, 0. 01% 1) 가 가) 0. 2-1. 6% 가 가

0. 2%

) 0.8% , 0.4% 가) 0. 1-0. 8% 0.4% 가) 0. 2%, 0.4%, Lactobacillus 0.4% 가 MRS 가 2) 2. 18 9. 05 1.30 가 8.83 4.80 9. 33, 5. 53 8.83 가 **4**% 가 2. 가 가. , 가 paste block 4가 가 paste block 가 가

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. 기 Lactobacillus PAP1

Lactobacillus MGC2 ,

Hansen YC-180(S. thermphilus L. bulagaricus)

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- 20 -

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SUMMARY

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. Title

Development of a fermented food product using chestnut

. Objects and importance of the study

The annual amount of chestnuts produced in Korea, 30% of which is exported and the rest of which is consumed in domestic market, is about 100,000 tons and ranked second to the top production in the world. However all the chestnuts domestically available are not fully consumed and even left in the trees without harvest by the frustrated farmers because chestnut demand in the market is low. Low demand comes from the lack of the technologies to process the chestnuts into various commercial food products and the lack of the adequate technologies needed to harvest and peel. We tried to develope fermented food products using chestnuts in a way to increase usage of chestnuts which will increase the demand for the chestnuts in the market and prevent loss of chestnuts by the frustrated farmers who would be encouraged to harvest chestnuts in response to the increased demand caused by this technique.

Technically this study to develope fermented food product using chestnuts will (1) promote the development of various fermented plant

products by providing the basic techniques needed in fermentation of plant materials, (2) convert chestnuts with high content of energy, into the product with high organic acids and lactic acid bacteria, which will help consumer's health by correcting the balance of the gastrointestinal microflora, and (3) promote to develope technologies needed to make machines to harvest and peel the chestnuts. Finally this fermented products will make many Korean people, not familiar to fermented milk, more like the fermented foods because this products will be made of chestnuts familiar to most Koreans for thousands years.

. Results and Scopes of this development

There are two parts of studies in this research. One is microbial study, goal of which concerns the isolation of strains appropriate to ferment chestnut broth and the optimization of fermentation condition. The other is making process study, target of which deals with the optimum manufacturing process for the final products. The study results are as follows.

1. Microbial study

A. Isolation of the lactic strains

1) From human feces

Five strains which showed good fermenting properties were isolated from healthy men and women and named MIR1, M2R1, M2R6, MBR3, F1R4 for the time being.

2) From the commercial yogurts

Five strains, MGG1, MGG2, NBL1, PAP1 and PAP2 were isolated and named in the same way.

3) Identification of the isolated strains

Ten isolated strains were identified by the properties gathered from the biochemical test of the Rapid32A Kit, activity test of fructose-6-phosphate phosphoketolase and the ratio of acetate/lactate in the culture, which identified MIR1, M2R1, M2R6, MBR3, NBL1, PAP2 as *Bifidobactrium* and F1R4, MGG1, MGG2, PAP1 as *Lactobacillus*.

B. Properties in fermentation of chestnut broth by the isolated strains

1) Fermentation test in chestnut broth

When the 4% chestnut broths were fermented by the isolated strains, titrable acidities ranged from 0.81 to 2.11(ml, 0.1 N NaOH).

2) Effects of chestnut concentration on fermentation

Within the range of 2-32%, the higher were the concentrations of chestnut, the higher were the titrable acidities. The acidity increases were highest with F1R4 and PAP1. The acidity increase rate was highest when the concentration was increased from 4% to 8%, which means that 8% is the most economic.

3) Effects of chestnut kinds on fermentation

When the chestnut broths made separately of EunGee and EePyung from SanChong and the wild strains from Jeonbuk Province were fermented with each of MIR1, MGG2, MGG1, F1R4 and PAP1, the difference ranges of acidity were higher with different bacterial strains than with

different chestnut strains. This result means the difference of chestnut strains are not so important. However they may become so on manufacturing stage by the apparent difference of taste and flavour.

C. Examination on the fermentation conditions

1) Effect of sugar concentration

When glucose concentration were increased within the range of 0.5-8.0%, titrable acidities of the fermented broths were increased with *Lactobacillus* strains, but not with *Bifidobacterium* strains.

2) Effect of nutrient supplementation

When the chestnut broths were supplemented separately with 0.05% cysteine, 0.2% yeast extract, 0.4% each of Gelysate peptone, Phytone peptone, and Trypticase peptone, the acidities were increased with most of the strains, but the increase was highest with combination of F1R4 or PAP1 and Phytone peptone.

3) Effect of sugar kinds

Addition of disaccharides like Lactose, Maltose and Sucrose did not stimulate acid production higher than glucose.

4) Effect of seed culture size

The acidities of the broths were more increased with higher inoculation size within the range of 0.5-8.0%. However increase rates within the range of 2.0-8.0 were lower than the range of 0.5-2.0%.

5) Effect of vegetable and fruit extracts

When the extracts of apple, orange, tomato, pear, cucumber, carrot, and dropwort were added at 10%, the acidities of the broths were increased in most of the strains. The increase was highest with

combination of F1R4 and carrot(or dropwort), MGG1 and orange(or carrot), and PAP1 and cucumber(or carrot or dropwort).

6) Effect of fermentation time

When the turbities of the culture broths measured with time intervals were increased up to 48 hours. However, the increase approached the stationary phase by 24 hours. The turbidity were higher with F1R4, MGG1 and PAP1, but also the initial turbidity of the broth after inoculation were higher due to the higher turbidity of the inocula.

7) Effect of fermentation type

a. Fermentation of fluid type chestnut

The transparent liquids of 8% chestnut broth after removal of precipitates showed viable counts between 8.05 and 9.05(log10CFU/ml) after cultivation of 24 hours, in which MIR1 and PAP1, 9.05 and 9.03, were highest among the tested strains. The 48 hour cultivations showed viable counts between 8.28 and 9.15 with PAP2 and M2R1, 9.15 and 8.99, highest.

b. Fermentation of custard type chestnut

When the mixtures of cooked chestnuts and water in ration of 1:1, in which no liquid layers were shown, were inoculated with 0.8ml of seed cultures and cultivated for 48 hours, F1R4 and PAP1 were highest in acidities with 3.26 and 3,66, and MGG1, MGG2, and NBL1 were highest in viable counts with 9.22, 9.16 and 9.24.

8) Effect of chestnut hydrolysis

When the chestnut broth hydrolyzed by liquefying enzyme and Koji

showed higher acidities than those by malt extracts. The range of acidities of both were 4.39-14.67 and 3.35-9.52. When this results compared with those of glucose addition, the increases were beyond the sole sugar effects.

- D. Selection of the best strains among the isolates
- 1) Comparison of the growth abilities

The viable counts and acidities of the fermented broth of 8% chestnut and nutrient supplement were much higher with F1R4(9.05 and 5.26) and PAP1(9.32 and 5.21) than with any other tested strains.

2) Changes of viable counts during storage

The initial viable counts of MGG1 and MGG2(8.66 and 8.83) were reduced to 8.00 and 8.03 after storage of 3 weeks in 5 refrigerator. However those of F1R4 and PAP1(9.05 and 9.32) were reduced to 2.80 and 3.00. This result means the higher the acidity of the broth, the smaller the viable counts after storage.

3) Effect of pH on the changes of viable counts during storage

When the fermented broth were kept at 5 for 12 days after adjustment of pH to 5.17, the viable counts were 1000 times higher than the unadjusted broths.

4) Effect of the precipitates on the viable counts

The chestnut broth made through heating and homogenization has precipitates. The effect of precipitate was surveyed. The precipitate did not much influence the microbial growth and acidity production when MGG1 and F1R4 strains were used within the precipitate range of 0-23%. However in case of F1R4, which shows high acidity and low

viable counts after storage, the precipitate(23%) reduced slightly the lowering of the viable counts of the fermented broth after storage of 12 days at 5 .

5) Selection by sensory evaluation

When the fermented broths were evaluated sensuously by the panel of 10 personnels, the scores were all very low in acidity, flavor, taste and body sense. The causes of the overall bad scores were thought to be the reduction of chestnut flavor by the heat sterilization, bad flavors eluted from the fermenting vessels, the excess of acid flavor, unsatisfactory body sense by low viscosity.

6) Final selection

 a. Effect of the mixed cultures on viable counts and storage of custard type products

Based on the analysis of test results above, PAP1 and MGG2 were selected. The mixed cultures of both were better in viable counts than the cultures of single strains each. The mixed cultures in custard type products gave higher numbers of viable counts than single cultures after 3 week storages at 5 .

b. Effect of the mixed cultures on storage of fluid type products

The viable counts of fluid type products made by the mixed cultures decreased to 6.62 after 2 week storage and to 3.65 after 3 week storage. However, the viable counts after 3 weeks increased to 4.60, 5.88, and 7.64 by each supplementation after fermentation with equal volumes of sugar, chestnut broth and both.

c. Comparison between the selected strains and the commercial seed

cul tures

When the selected strains, Hansen YC-180(S.thermophilus+L.bulgaricus), and MSK B2(S. thermophilus+B. infantis+L. acidophilus) were compared on their ability to growth and acid production in chestnut broths, the selected strains and Hansen were better in viable counts than MSK. Acidity was the best with the selected strains. So Hansen was used in the making process study because of the ease in use while the selected strains in microbial study.

d. Examination on the possible inoculation size

When the inoculation sizes are lowered to 0.1% from 0.5%, the viable counts of the products did not decreased so much.

E Final selection of the fermentation conditions

1) Optimum concentration of nutrients

The optimum concentrations of Phytone peptone and yeast extract were 0.8 and 0.8 at 0.5% glucose and the acidity. On this condition, the acidity and the viable counts with mixed culture of PAP1 and MGG2 were 5.11 and 9.42. However, yeast extract more than 0.2% and Phytone peptone more than 0.4% were thought to be enough to get the acidity and viable counts no less than 3.89 and 9.00.

2) Substitution of supplements with industrial byproducts

When the industrial byproducts such as soybean meal, soywhey and beer yeast waste were examined to see the possibilities of substitution with Phytone peptone and yeast extract, the beer yeasts were 1/3 times active to yeast extract and found to be unable to replace yeast extract. However soybean meal and soywhey were active as much as phytone peptone.

3) Supplementation of health improving mushrooms extracts

Two kinds of healthy improving mushrooms, *Ganderma lucidum* and *Cordyceps militalis*, were added up to 40% and found not to inhibit the acidity and viable counts of the fermented broth.

4) Effects of stabilizers on viable counts

Two kinds of stabilizer, pectin and sodium alginate, were added up to 0.5% to see the effects on the acidity and viable counts. Sodium alginate increased both of the acidity and viable counts and pectin reduced both. This result indicated sodium alginate is the better choice to get higher viable counts.

5) Final selection of the fermentation condition and improvement of the sensory quality

From the accumulated data, final composition for the fermentation was determined, which consists of chestnut 8%, soybean meal 0.4%, yeast extract 0.2%, glucose 0.5%, sodium alginate 0.2%, and modified starch 2.0%. However the fermented products made from this composition need improvement.

a. optimum ratio of sugars

The products with 10% sweetners were favored most. The sensory values of sweetness were best with the combination of sucrose 4%, high fructose syrup 4% and honey 2%. However the economy considered, the combination of sucrose 5% and high fructose syrup 5% was selected as practical one.

b. optimum acidity

The sensory value of the acidity was improved best adjusting the titrable acidity of the product to 2.5-3.0.

c. optimum combination of flavors

The sensory value of the flavor was greatly improved by adding the mixtre of chestnut, yogurt, and vanila flavors at the concentration of 0.08, 0.01 and 0.01% respectively.

F. Development of cheap seed culture media

1) Utilization of soywhey

Soywhey, the byproduct of DooBoo(bean curd) making, was thought to be a good substitution to expensive media like Lactobacillus MRS broth because it contains much monosacchrides and oligosaccharides, which can supply energy source to fermenting bacteria and also contains minerals, which can neutralize the lactic acid produced.

a. Supplementation with sugars

When glucose were added to soywhey at the range of 0.2-1.6%, the viable counts of the products were best at below 0.2%.

b. Supplementation with soybean meal

When soybean meal were added at the range of 0-0.8%, the viable counts increased above 0.4%.

c. Supplementation with yeast extract

Supplementation with yeast extracts at the range of 0.1-0.8% did not increase the viable counts of the products with exception of 0.4%.

d. Titer of the developed media compared to MRS

When the two seed culture media, the soywhey which was supplemented

with 0.2% glucose, 0.4% soybean meal and yeast extract 0.4%, and MRS were used in growth of the selected strains(PAP1 and MGG2), both produced similar viable counts, though MRS produced a little more. So the supplemented soywhey was found out to be a good seed culture medium

2) Utilization of sweet potatoes

Sweet potato broths produced acidity of 2.18 and viable counts of 9.05 while chestnut broth 1.30 and 8.83 respectively. The sweet potato broths hydrolyzed by liquafying enzyme and Koji produced acidity of 4.80 and viable counts of 9.33 and the chestnut broth hydrolyzed by the same ways 5.53 and 8.83. So sweet potato seemed to be able to replace chestnut when it is used at small size to minimize the influence of the different flavour.

2. Study on the making processes

A. Selection of chestnut forms as raw materials

When four types of chestnut materials, peeled raw, cooked, sweetened cooked, and freezed paste block, were compared on the criteria based on availability, storage properties, convenience and cost, freezed paste block was the best material forms for the production of fermented chestnut products.

B. Comparison of the components in chestnuts from different origin and lands

The difference of the components and colors of the chestnuts originated from the difference of the kinds and the locations were insignificant and can not deserve attention because of the indiscriminated gathering and selling in the market.

C. Selection of the fermentation types for the products

The fluid type fermentation was selected from the facts that fluid type products are more favored 2.8 times by Korean consumers and more free from microbial contamination and quality consistency than custard type product.

- D. Composition and contents in raw ferment liquor
- 1) Problems from the first test fermentaion

Problems like low body sense in taste of products, precipitates, requirement of long fermentation time were detected on the first preliminary production using tentative composition and condition.

- 2) Trials to find the best compositions
- a. Selection of chestnut contents
 - (1) Sensory evaluation

When three contents of chestnut in the products, 5, 10, and 15 %, were evaluated by sensory tests, 10 and 15 % were significantly more favored than 5 %.

(2) Precipitate comparison

The higher the chestnut concentration, the more the precipitate. So the concentration of 8% was selected on practical reason and the data from microbial study.

b. Selection of sugar kinds

Though glucose produced more acid than high fructose syrup and sucrose, mixture of glucose and high fructose syrup were thought to be

better by consideration on cost and sweet feeling. Seed size of 50U/250kg was not much inferior in acid production than 100. However to achieve higher viable counts of the products, 100U/250kg was thought to be better choice.

c. Selection of sweetener ratios

When three combination of glucose and high fructose syrup, 6:7, 3:9, and 9:5, were compared, 6:7 was the best, though insignificant.

d. Supplementation with plant and mushroom extracts

When four kinds of the plant extracts, pine leaves, coal, wornwood, and mulberry leaves, and two kinds of the mushroom extracts, *Ganderma lucidum* and *Cordyceps militalis*, were added to the chestnut broth, the sensory scores with any of the supplements were lower than the score without any supplement. The score was in order of coal, *Cordyseps*, pine leaves, *Ganderma*, mulberry leaves, and wornwood with the wornwood the worst by changing the original chestnut taste.

e. Selection of best body sense

When the mixture of sodium alginate and modified starch were used as stabilizer at the concentration of 0.2 and 2.0% each, the increase in the viscosity of the product and the score of body sense and decrease in the precipitate were higher than the products added with each stabilizer separately.

- 3) Selection of the making processes
- a. Prevention of the precipitation

To prevent precipitates developed even in condition with stabilizers, the downsizing processes of colloidal particles were examined. The precipitation was prevented when the particle size was less than $50\,\mu$. However, with the burdens of work needed to decrease the size, $100\,\mu$ was thought to be enough as a practical size.

b. Selection of sterilization temperature

Sterilization above 105 generated bad flavors and decreased the original chestnut flavors, but not below 75. However working time considered, sterilization at 85 may be practical.

c. Maintenance of viable counts

(1) Regulations in the food sanitation law

The viable count of 10.0(logCFU/ml) is favored in circulation in the market to deal with the unfavorable storage temperature and period met in this real world. However this counts couldn't be achieved with chestnuts as material. So 9.0 was selected as standard viable count.

(2) Fermentation time to meet the regulations

The fermentation needed more than 19 hours to reach the viable count of 9.0(logCFU/ml) in products

d. Processes for the selected stains in this study

For Lactobacillus MG1 and Lactobacillus PAP1, isolated in this study, the medium composition for seed culture was soywhey with 0.02% glucose, 0.4% soybean meal, 0.4% yeast extract and the medium composition for main fermentation was chestnut 8, soybean meal 0.4, yeast extract 0.2, glucose 0.5, sodium alginate 0.2, modified starch 2.0, clean water 100. Fermentation each for 15 and 19 hours at 37 was enough to meet the required viable counts above 9.0.

To improve the taste and viable count, mixture of sucrose 5, high

fructose syrup 5, chestnut flavor 0.08, yogurt flavor 0.01, Vanila flavor 0.01, should be added to fermented liquor after fermentation and before homogenization.

The products made with the mixture of flavors had stable and unique taste and flavor enough to cover the bad flavors generated from the high temperature sterilization.

- e. Flow sheet of making processes
- (1) For the selected commercial strains

[cooking-->supermicro-colloidalization-->freezing into block--> mixing(chestnut 8%+clear water + glucose 6% + high fructose syrup 7% + stabilizer 2.2%) -->homogenization-->sterilization-->inoculation with seed culture -->fermentation-->homogenization-->filling and sealing-->preservation in refrigerator]

(2) For the selected strains in this study

[cooking-->supermicro-colloidalization-->freezing into block--> mi xi ng (chestnut 8%+clear water+glucose 0.5%+soybean meal 0.4%+yeast extract0.2%+stabilizer2.2%)-->homogenization -->sterilization --> cultivation of seed culture(chestnut 8%+glucose 0.2%+soybean meal 0.4%+yeast extract0. 4%) -->i nocul ati on wi th seed culture--> fermentation-->mixing(sucrose 5%+high fructose syrup 5%+chestnut 0.08%+yogurt flavor 0. 01%+vani l a flavor homogenization --> filling and sealing --> preservation in refrigerator]

4) Selection of the standard making processes

Standards and criteria in making process were presented in summery

and also quality standards were noted.

5) Selection of the standard circulation period in the markets

When the products were stored at 5 in a refrigerator on the same condition as commercial stores, the viable counts were decreased after 12 days and change in the taste was noted by one of the sensory evaluators. So the products should be kept below 5 and managed not to be sold beyond 12 days.

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, (Castana crenata) , 9 10

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, 61%, 1%, 1%, 8. 5%,

, (栗子) (氣)

(補胃) (腎氣) ,

(栗皮), (栗毛殼) (東醫寶

鑑).

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. 1968

2 3 ha 1978 14 ha , 79, 155ha (1996)

가 52, 697가 가

 (1996
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 (41%, 800
),
 (22%, 400
),
 (16%,

 280
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 80%
 ,
 (11.0%),

(10. 1%), (7. 8%) (1998).

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100, 000 1, 900 ((FA0

1993-1995), 30% 30,000 , 70% フト

, **70**%

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(1998). 가 .

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가 가

47%

. 70

가 .

가 . . .

가 ,

, (Hoverstad , 1984)

,

가 , (Rasic Kurmann,

1983).

· 가

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가 , , 가

가 (1989, 1991a, 1991b).

가 가 가 가 .

가 ,

가 . 가 가

. 가 가 , **pH**가

가 가 가 가 가 가 가 2 1991, 1997) (1994) (Mbk (Tomi naga Sato (1996, 1997, 1991),

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1990

1989)

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2. 2 1 가 . 1 9 9 (, , 가 , paste block)) (,) () , (가 (, pH 2 가

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2.

가 (:
, 가 , paste
block)

(,), (,)

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(1998) : Lactobacillus, Bifidobacteria ,

1

: L. acidophilus, L. casei, L. bulgaricus)

(, :

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Ca, 가 , pH)

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( 2 )
                   ( , ), ( , )
                   )
2
(1999)
 4
1. 1
   1
    . , , , 가 , paste block
             ( , ) ( , )
                            Lactobacillus
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Bi fi dobacteri a

Lactobacillus 가 가 : 가 가 pН 2. 2 : 1 가 2 가 yeast extract yeast paste, Phytone peptone () 1 lactose MRS broth 10가 1 2-3 가 가 , 가

- 61 -

.

, 가 가

, 가 .

, 가 , 가

. , , , Ph, Brix, 가,

,

, , Brix,

· : 가

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5		
3.		
	가	
	가	



2

1

(lactic acid)

•

(homofermentative)

7 ethanol (heterofermentative)

fructose bisphosphate aldolase

.

Streptococcus, Leuconostoc, Pediococcus, Lactobacillus, Enterococcus,

Lactococcus Leuconostoc

. Streptococcus 가

Lactococcus

Enterococcus Streptococcus

(Madigan, 1997).

Streptococcus(Lactococcus)

Lactobacillus

probiotic 가 . probiotic

anti bi oti c anti bi oti c probi oti c 가 Lilly Stillwell Fuller (Fuller, 1989). probi oti c 가 가 probiotic (Fuller, 1989). (microbial barrier, or interference) (Tanock 1981, 1984). (Sal vage, 1877), (short chain fatty acids: SCFA) (Rolfe, 1984) bacteri oci n 가 (Sal vage, 1877) 가 (Macfarlane Gibson, 1994) 가 deamination ammonia, decarboxylation deamination, transamination,

phenol, indol

dehydrogenati on

steroi d

deconjugation(Wostmann, 1981), desul furation, transformation(Eyssen Verhulst, 1984) (Gordon Pesti, 1971), , (Macfarl ane Gibson, 1994), (Corring , 1981) probi oti c tablet Lactobacillus GG(Oksanen L. acidophilus, L. bulgarius, 1990) Bifidobacterium bifidum, Streptococcus thermophilus (Black , 1989) acidophilus milk가 (Alm , 1983) 가 Lactobacillus (Floch Mbussa, 1998).

Allergy, lactose intolerance

가 (1997), , 가 (1997),

가

, 가

,

- 67 -

가 , , MRS 가 가 seed culture 가 가 가 2 1. 20-26 가 11 1 가 (Balmer & Wharton, 1989; BHI 37g, glycerol 100ml, cysteine 0.5g, resazurin 0.1% sol. 1ml, pH7) C02 가 - 60 10-1-10-6 $(\mathop{\mathsf{modi}}\nolimits \mathop{\mathsf{fi}}\nolimits \mathop{\mathsf{ed}}\nolimits$ MRS agar with Broncresol green) Tomato-juice agar , 37 C02 2-3

(Balow

, catal ase

coryne

1991).

3 Lactobacillus MRS
agar(Difco) CO2 Anaerobic jar 2-3
coryne , catal ase

2.

Rapid ID 32A (bioMeieux, France)
urease activity, - -galactosidase , -glucosidase
, fructose-6-phosphate phosphoketolase(F6PPK)
, HPLC(Shimadzu) acetate/lactate

Chevalier (1990) .

3. 2kg 30

1. 5kg .

.

seed culture 1-4% 37 1-2

•

4. pH 5ml 50ml

5ml 가 pH (TOA HM-20S) .

가 (Digitrate) 3 0. 1N NaOH 가 NaOH (Mitsuoka: Mitsuoka 1980, 1/4 가) 10 (Trypticase 10, Phytone 5, yeast extract 2.5. (25µl) PTY glucose 5, Tween 80, cysteine \cdot HCl 0.5, K2HP04 2, MgCl 2 \cdot 6H20 0.5, ZnS0 $4 \cdot 7H20 \ 0.25$, CaCl 2 0.15, FeCl 3 0.03 g/l) 3 7 , CO27 2-3 . Cream Image Analysis Bac **Program** col ony 1ml log Log10(CFU/ml) **5**. 10 5 (5= , 1= , 3=) 가 1 3 1. 가. catal ase

5

M1R1, M2R1, M2R6, M3R3, F1R4

.

, catalase , 1 ,

5 MGG1, MGG2, NBL1, PAP1, PAP2

•

107 rapid32A ki t

. 4 .

4.

test	M1R1	M2R1	M2R6	MBR3	F1R4	MGG1	MGG2	NBL1	PAP1	PAP2
URE	+	-	-	-	-	-	-	-	-	-
ADH	+	+	+	+	-	-	-	+	-	+
GAL	+	+	+	+	-	-	-	w	-	+
GAL	+	+	+	+	+	+	+	+	+	+
GP	w	w	w	w	-	-	-	+	-	w
GLU	-	-	-	-	+	w	+	-	w	-
GLU	+	+	+	+	w	+	+	+	+	+
ARA	-	-	-	-	-	-	-	-	-	-
GUR	-	-	-	-	-	-	-	-	-	-

(

(4)

NAG	+	+	+	+	w	w	+	+	w	+
MNE	+	+	+	+	+	+	+	+	+	+
RAF	-	-	-	-	-	-	-	-	-	-
GDC	-	-	-	-	-	-	-	-	-	-
FUC	-	-	-	-	w	+	w	-	+	-
NIT	-	-	-	-	-	-	-	-	-	-
IND	-	-	-	-	-	-	-	-	-	-
PAL	-	-	-	-	+	+	-	-	+	-
ArgA	w	w	w	w	w	w	+	w	w	w
ProA	-	-	-	-	+	+	+	-	+	-
LGA	w	w	w	w	-	w	-	w	-	w
PheA	+	+	+	+	+	+	+	+	+	+
LeuA	w	w	+	+	+	+	+	+	+	w
PyrA	+	+	+	+	-	w	-	+	w	+
TyrA	w	+	+	w	-	w	-	w	w	w
Al aA	w	w	w	w	w	+	w	w	+	w
GlyA	w	w	w	w	-	w	-	-	-	w
Hi sA	+	+	+	+	+	+	w	+	+	+
GGA	-	-	-	-	-	-	-	-	-	-
SerA	+	+	+	+	+	w	w	+	+	+

Bi fi dobacteri um
gal actosi dase (-GLL) - gl ucosi dase (-GLL) 7 }

Streptococcus urease, -GAL -gal actosi dase7\\ Lactobaci l l us -GAL , -GLU \(Bal ow , 1991 \).

MIR1, M2R1, M2R6, M3R3, NBL1, PAP2 F1R4, M3G1, M3G2, PAP1 . Cheval i er

(Chevalier , 1990)

F6PPK acetate/lactate 5

•

5. **F6PPK**

MIR1 M2R1 M2R6 M3R3 F1R4 M3G1 M3G2 NBL1 PAP1 PAP2

F6PPK + + + + + - - - + + - +

acetate
/lactate

1. 36 1. 80 1. 82 1. 92 0. 04 0. 01 0. 02 2. 50 0. 01 2. 10

MIR1, M2R1, M2R6, M3R3, NBL1, PAP2 acetic acid
F6PPK 7 Bifidobactrium , F1R4,
MGG1, MGG2, PAP1 acetic acid F6PPK 7
Lactobacillus .

2.

가.

10

10ml , MRS seed culture 4%
37 . 36 60 5ml
가 phenol phthal ein 2-3 가

0. 1 N NaOH 6 .

, . 60

가 가

48

MIR1 M2R1 M2R6 M3R3 F1R4 M2G1 M2G2 NBL1 PAP1 PAP2

36 1. 58 1. 59 1. 64 1. 37 1. 65 0. 84 0. 88 0. 81 1. 58 0. 72

60 2. 11 1. 69 1. 61 0. 72 1. 67 0. 71 0. 74 0. 78 1. 74 0. 82

: 4 g/100 ml

6.

*: 0. 1N NaOH (ml)

.

. 가

2% 32% 가

, 가 **F1R4**

PAP1가 가 4% 8% 가 가 .

8% 가

7.

(g/100ml) MIR1		M2R6			MGG1	MGG2	NBL1	PAP1	PAP2
2	1. 19	1. 10	1. 15	1. 23	1. 58	0. 80	1. 12	0. 79	1. 53	0. 55
4	1. 84	1. 94	1. 62	2. 12	1. 85	0. 78	0. 88	1. 01	2. 16	0. 96
8	2. 75	2. 12	2. 63	2. 28	4. 56	0. 99	1. 27	1. 48	4. 97	1. 28
16	3. 23	3. 02	3. 20	2. 76	8. 09	1. 35	1. 77	2. 29	7. 79	1. 85
32	3. 84	3. 72	4. 15	3. 78	9. 08	2. 47	2. 42	3. 23	8. 88	3. 22

•

, ,

.

MIR1, MGG2, MGG1, F1R4, PAP1

8 . 8% 32% 8%

가 .

•

가 .

8.

	MIR1	MGG2	MGG1	F1R4	PAP1	
	1. 88	0. 60	0. 66	2. 45	2. 39	_
C8	1. 73	0. 74	0. 63	2. 33	2. 44	
	1. 92	0. 72	0.65	2. 21	2. 29	
	3. 26	1. 46	1. 60	6. 18	6.06	
C32	3. 26	2. 22	1. 96	6. 94	6.80	
	2. 82	1. 81	1.82	5. 96	6. 18	
	2. 80	3. 35	3. 36	5. 85	5. 55	
C8S	2. 75	3. 36	3. 32	5. 63	5. 48	
	2. 92	3. 29	3. 27	5. 60	5. 62	

. , , .

C8: 8g/100 ml , C32: 32g/100 ml

C8S: 8g/100 ml + (glucose 0.5%, Phytone 0.4%, Yeast Extract

0.2%)

가. 가

0. 5% 8% 가 가

가

가

가 가

. F1R4, MGG1, MGG2, PAP1 Lactobacillus 가 Bifidobacteria 가 가

9.

(%)	MIR1	M2R1	M2R6	MBR3	F1R4	MGG1	MGG2	NBL1	PAP1	PAP2
0. 5	2. 37	1. 98	2. 29	1. 07	1. 80	3. 32	3. 12	0. 82	2. 49	0. 92
1. 0	2. 18	2. 01	2. 14	0. 81	1. 89	3. 66	3. 15	0. 92	2. 75	1. 05
2. 0	2. 16	2. 17	2.06	0. 81	2. 22	3. 71	3. 20	1. 03	3. 06	0. 91
4. 0	2. 20	2. 24	2. 33	1. 01	2. 76	3. 76	3. 60	0. 91	3. 57	0.83
8. 0	2. 26	2. 07	2. 35	1. 10	3. 12	3. 48	3. 31	0. 79	3. 69	0. 85

: 8 g/100 ml

potential . OR

potential .

가 3 peptone 가

peptone .

Lactobacillus Bifidobacteria peptone 가

Lactobacillus Bifidobacteria 가

yeast extract 0.2% 가

MIR1

F1R4 PAP1 가 .

	MIR1	M2R1	M2R6	MBR3	F1R4	MGG1	MGG2	NBL1	PAP1	PAP2
Bl ank	2. 83	1. 81	2. 09	1. 85	2. 38	3.00	2. 93	0. 86	2. 54	0. 92
Cys	2. 59	2. 24	2. 16	2. 23	1. 90	2. 70	2. 97	0. 92	2. 17	0. 78
GP CP	2. 80	2. 26	2. 25	2. 19	3. 72	3. 70	3. 82	1.06	3. 64	1. 23
PP	2. 91	2. 24	2. 30	2. 31	5. 53	3. 63	3. 64	1. 68	5. 92	1. 70
TP	2. 83	2. 28	2. 23	2. 17	5. 67	3. 71	3. 77	1. 66	3. 92	1. 82
YE	2. 68	2. 29	2. 34	2. 08	3. 34	3. 68	3. 75	1. 10	3. 33	1. 40

Cys: Cysteine 0.05%, GP: Gelysate Peptone 0.4%, PP: Phyton Peptone
0.4%, TP: Trypticase Peptone 0.4%, YE: Yeast Extract 0.2%

: 8 g/100 ml

.

-gal actosi dase

가 Lactose

0.5%

11 . 가

MGG2, NBL1, PAP2

lactose maltose sucrose

가

	MIR1	M2R1	M2R6	MBR3	F1R4	MGG1	MGG2	NBL1	PAP1	PAP2
	2. 47	2. 01	1. 94	1. 86	6. 05	3. 25	3. 36	1. 67	5. 74	1. 82
	2. 47	1. 97	1. 96	1. 90	3. 91	0. 97	3. 95	1. 78	3. 89	1. 88
	3. 06	2. 13	1. 99	1. 99	5. 84	1. 28	1. 97	1. 57	5. 62	1. 66
	2. 85	2. 27	2. 23	2. 05	5. 91	1. 05	1. 27	1. 77	5. 90	1. 63
:	,	t extra	_	;/100 ml		one 0.4	<u>10%</u>			
•										
1	2			フ	ŀ			가		0. 5-8%

0. 5-2%

가

,

. 4%

·

가

10% 가 13 .

MER1

. F1R1, MGG1, PAP1가 가 , F1R4 , MGG1 , PAP1 ,

, 가 .

12.

(%)	MIR1	M2R1	M2R6	MBR3	F1R4	MGG1	MGG2	NBL1	PAP1	PAP2
0. 5	2. 68	2. 02	2. 22	2. 15	5. 14	3. 33	3. 47	1. 47	5. 65	1. 84
1. 0	2. 47	2. 11	2. 00	2. 02	5. 85	3. 64	3. 48	1. 73	5. 52	1. 69
2. 0	2. 74	2. 03	2. 13	2. 22	5. 82	3. 65	3. 60	1. 76	5. 56	1. 90
4.0	2. 86	2. 33	2. 34	2. 34	6. 25	3. 60	3. 75	1. 76	6. 20	1. 80
8. 0	3.06	2. 28	2. 72	2. 43	6. 34	4. 05	4. 11	1. 93	6. 86	1. 98

: 8 g/100 ml

: yeast extract 0.2%, Phyton peptone 0.4%, glucose 0.5%

14 . 48 가 가

가 , 24 .

24 48

가										
	MIR1	M2R1	M2R6	MBR3	F1R4	MGG1	MGG2	NBL1	PAP1	PAP2
 가	2. 04	1. 90	1. 82	1. 65	2. 45	3. 17	2. 99	0. 89	2. 41	0. 88
	2. 46	1. 80	2. 03	1. 87	3. 29	4. 12	2. 84	0. 82	3. 30	0. 75
	2. 22	1. 82	1. 69	1. 62	4. 12	4. 94	3. 66	0. 78	4. 01	0.88
	2. 37	2. 06	2. 10	2. 01	4. 13	4. 51	3. 62	0.83	4. 17	0.83
	2. 67	1. 88	2. 05	1. 85	3. 53	4. 61	3. 18	1. 22	3. 49	1.09
	2. 64	1. 96	1. 99	1. 95	4. 94	4. 05	3. 42	1. 17	5. 07	1. 14
	2. 65	1. 78	1. 88	1. 77	5. 76	4. 92	3. 85	1. 42	5. 78	1. 34
	2. 81	2. 30	2. 20	2. 18	5. 38	3. 80	3. 69	1. 90	5. 42	1. 14

가 : 10%(V/V),

: 8 g/100 ml

: 0.5%

F1R4, MGG1, PAP1

가 가

F1R4 PAP1 seed

cul ture

						(550				
	MIR1	M2R1					MGG2			PAP2
0	0. 19	0. 17	0. 15	0. 16	0. 41	0. 41	0. 35	0. 15	0. 41	0. 14
8	1. 12	1. 24	0.87	1. 21	1. 16	1. 13	1. 11	0. 73	1. 14	0. 72
24	1. 30	1. 45	1. 40	1. 41	1. 78	1. 73	1. 61	0. 83	1. 84	0.83
48	1. 48	1. 59	1. 54	1. 55	1. 99	1. 93	1. 80	0. 99	1. 98	0. 99
72	1. 48	1. 59	1. 55	1. 55	2. 01	1. 94	1. 79	1. 00	1. 98	1. 02

: Phyton peptone 0.4%, yeast extract 0.2%, glucose 0.5% : 8 g/100 ml $\,$

.

1)

10가

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		MIR1	M2R1	M2R6	MBR3	F1R4	MGG1	MGG2	NBL1	PAP1	PAP2
		7. 27	7. 19	7. 15	7. 12	7. 24	7. 03	7. 18	7. 00	7. 37	7. 13
0		0. 28	0. 27	0. 50	0. 27	0. 44	0.48	0.39	0. 29	0. 25	0. 26
	pН	5. 00	5. 17	4. 46	5. 14	4. 56	4. 50	4. 63	5. 05	5. 19	5. 18
		0. 58	0. 45	1.00	0. 48	0. 65	0.60	0. 63	0. 50	0. 45	0. 45
		9. 05	9. 03	8. 80	8. 63	8. 64	8. 13	8. 05	8. 75	9. 03	8. 87
24		1. 03	0.90	0. 66	0.90	0. 70	0.65	0.65	0. 81	0. 78	0. 71
	pН	3. 73	3. 99	4. 36	4. 09	4. 26	4. 32	4. 31	4. 21	4. 09	4. 09
		2. 05	1. 25	0. 80	1. 15	0. 85	1. 03	0. 93	1. 00	1. 25	1. 00
		8. 28	8. 99	8. 67	8. 51	8. 70	8. 93	8. 72	8. 74	8. 73	9. 15
48		1. 15	1. 11	0.82	1. 01	0. 82	0. 93	0. 84	0.88	0. 87	0. 99
	pН	3. 77	3. 88	4. 10	4. 05	4. 26	4. 10	4. 17	4. 16	4. 07	3. 94
		2. 15	1. 73	1. 25	1. 28	0. 90	1. 20	1. 03	1. 03	1. 13	1. 40

= log(CFU/ml), : 8 g/100 ml

2)

.

•

. 1: 1 20ml

0. 8ml 7 2 16

. F1R4 PAP1 3. 26, 3. 66

MGG1, MGG2, NBL1 9. 22, 9. 16, 9. 24

가

가 가가 가

가

가 .

16.

MIR1 M2R1 M2R6 M3R3 F1R4 MGG1 MGG2 NBL1 PAP1 PAP2

7. 12 8. 06 7. 88 8. 00 8. 15 9. 22 9. 16 9. 24 7. 88 9. 09

 $1.\ 55 \quad 1.\ 46 \quad 1.\ 44 \quad 1.\ 51 \quad 3.\ 26 \quad 1.\ 35 \quad 1.\ 53 \quad 1.\ 35 \quad 3.\ 66 \quad 1.\ 60$

 $= \log(\text{ /ml}). \qquad \qquad 1g \qquad \qquad 9ml \quad \text{serial dilution}$ $, \qquad \qquad , \qquad \qquad 2g$

10ml .

. 가

가 .

•

가

,

가 . 가

.

7ト 100g 200ml , () 300μℓ 7ト 90 1 , 300ml () 7ト 55 2 . 12.5% 55 1

. 50g 200ml 55

. 17 .

1

가 가 4.39-14.67

가 malt extract 3. 35-9. 52

. 0.5% dextrose7

, 가 . 가

. 가

. 8g/100

1/3 1/3 가

가 .

가 가 가

, 가가 가

.

17. 가

						(0. 11	NaOH	ml)		
	MIR1	M2R1	M2R6	MBR3	F1R4	MGG1	MGG2	NBL1	PAP1	PAP2
	3. 00	2. 82	3. 19	2. 89	8. 18	1. 55	1. 74	2. 29	7. 42	2. 27
0. 5%Dex	3. 39	2. 80	3. 08	2. 89	6. 87	4. 63	4. 83	2. 47	6. 84	2. 71
M	4. 17	4. 05	4. 08	4. 22	9. 52	4. 79	6. 31	3. 31	9. 96	3. 35
1/2M	3. 06	2. 88	3. 42	3. 12	7. 77	2. 59	3. 30	1. 97	7. 42	1. 84
1/4M	3. 23	2. 21	2. 17	2. 17	3. 15	1. 42	1. 50	0. 92	3. 15	0. 85
K	6. 74	6. 49	6. 49	6. 53	13. 97	14. 67	13. 19	5. 35	14. 08	4. 39
1/2K	4. 76	4. 40	4. 46	4. 83	11. 00	11. 50	11. 65	2. 48	9. 81	2. 30
1/4K	3. 16	2. 46	2. 24	2. 44	5. 63	6. 23	6. 61	1. 45	5. 62	1. 45
:		=25g	g/100ml	, 0	. 5%Dex:	dextr	ose가 0	. 5%		, M
Malt extra	act	가			, 1/2N	f M	1/2	, 1/4	M M	1/4
, К:	,					, 1/2	ek: K	1/2	, 1	/4K: K
1/4	•									

4.

가

(1)

, (2)

. (3) 가

가 가

. 가

3가 .

가.

37 , **48**

TPY Plate Counting 18

. 8. 58-9. 32

F1R4, PAP1 9. 05, 9. 32

, 5. 26, 5. 21 1. 30-2. 45

F1R4 PAP1 가 .

18.

: Log10(CFU/ml), :

Factor

MIR1 M2R1 M2R6 M3R3 F1R4 M3G1 M3G2 NBL1 PAP1 PAP2

TA 2. 45 1. 70 1. 81 1. 62 5. 26 1. 37 1. 30 1. 76 5. 21 1. 83

Number 8. 60 8. 86 8. 98 8. 82 9. 05 8. 66 8. 83 8. 68 9. 32 8. 58

0. 1N NaOH

(ml)

- 88 -

(5)

3

3

8.00 8.03 가 8.66 8.83 가

. F1R4 PAP1 9.05 9.32 3 2.80 3.00

가 가 **MGG1**

MGG2가 가 .

19

19.

(Log10CFU/ml) () MIR1 M2R1 M2R6 MBR3 F1R4 MCG1 MCG2 NBL1 PAP1 PAP2 0 8.60 8.86 8.98 8.82 9.05 8.66 8.83 8.68 9.32 8.58 1 8.20 8.40 8.34 8.56 8.69 8.53 8.82 8.41 8.93 8.21 2 7.62 7.96 7.85 7.90 4.02 8.14 8.64 8.24 6.10 7.93 3 5.05 6.82 6.50 6.84 2.80 8.00 8.03 7.02 3.00 6.62

: 8 g/100 ml

. рН

F1R4 PAP1 가 3

·

가

MGG1 MGG2フト

가 pН 24 F1R4 48 가 pН 가 12 pН

20

20. pН

			()	
		0	6	12
]	рН	3. 64	3. 63	3. 59
		3. 25	2. 95	3. 23
		8. 45	8. 47	4. 38
]	Hc	3. 57	3. 56	3. 46
		4. 45	3. 93	3. 75
		8. 14	7. 78	4. 38
]	Но	5. 17	5. 71	5. 17
Н		1. 03	0. 52	0. 70
		7. 61	8. 00	6. 76

가 . 24 가 3.25 8.45 48 4. 45 8. 14 가 **48** pH 5. 17 pН 12 4. 38 6. 76 1000 pН 가 50%가 pol ymer mi cropocket 10% 가 0-23% 가 F1R4

21

MGG1

가

. MGG1

1. 43 8. 61 1. 08 8. 26 23%

가

F1R4 4. 05 7. 56 4. 42 8. 00

21.

				(%)	
		0	2. 3	10	23
	0	8. 61	8. 12	8. 63	8. 28
		1.43	1. 25	1. 20	1. 08
	3	8. 64	8. 34	8. 74	8. 76
		1.05	0. 98	1. 15	1. 05
MGG1	6	8. 15	8. 13	8. 82	8. 30
		1. 20	1. 60	1. 13	1. 21
	9	8. 18	8. 36	8. 60	8. 04
		1. 18	1. 10	1. 25	1. 15
	12	8. 14	7. 49	7. 38	7. 03
		1. 43	1. 15	1. 28	1. 13
				(

- 92 -

	0		8. 00	8. 78	7. 72	7. 56
			4. 42	4. 28	4. 10	4. 05
	3		7. 86	7. 51	7. 45	5. 90
			4. 53	4. 03	3. 80	4. 10
F1R4	6		4. 60	5. 60	6. 90	5. 30
			4. 65	4. 63	4. 38	4. 13
	9		3. 38	5. 30	5. 68	4. 90
			4. 40	4. 28	5. 08	4. 20
	12		3. 02	2. 90	5. 08	3. 68
			4. 70	4. 25	4. 13	4. 40
	10%,	4 ,	lo	g(1ml)		
12				MGG1		
가 1	. 43 8. 14	23%		1. 13 7.	03	, F1R4
	4. 70 3. 02	4. 40	3. 68		MGG1	
F1 R 4						
r i N4						
F1K4						
r i n/4						
. 1						
	· 가					

(21

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22

가

, , ,

,

가 , 가 , 가

.

가

22.

	MIR1	M2R1	M2R6	MBR3	F1R4	MGG1	MGG2	NBL1	PAP1	PAP2
рН	3. 14	3. 60	3. 52	3. 58	3. 01	3. 65	3. 67	3. 61	3. 00	3. 52
	2. 45	1. 70	1. 81	1. 62	5. 26	1. 37	1. 30	1. 76	5. 21	1. 83
	8. 60	8. 86	8. 98	8. 82	9. 05	8. 66	8. 83	8. 68	9. 32	8. 58
	1. 8	1. 9	1. 9	1. 9	1. 5	1. 9	1. 9	1. 9	1. 6	1. 9
	1. 2	1.4	1.4	1. 2	1. 2	1.6	1. 6	1. 4	1. 2	1. 4
	1. 6	1.8	1. 7	1. 7	1. 4	1.8	1.8	1.8	1. 4	1. 6
	1. 1	1. 4	1. 4	1.4	1. 1	1.4	1. 4	1. 4	1. 1	1. 4

10 5 1 3

가 가

가

가

1)

. PAP1, F1R4

MGG1, MGG2 . PAP1 MGG2

PAP1 MGG2 가

23 0.99 9.45

가 가

MGG2 PAP1 0.71 8.95 0.87

9. 39 .

가 50% 5 3 0.72 8.27 , MGG2

PAP1 0. 99 8. 03 0. 73 3. 00 .

3 .

pH 5. 17 3
0. 37 7. 64 , MGG2 PAP1 0. 72 7. 98 0. 28

4.00 . 3 pH

•

			0	1			2		3	
MCCO		0. 71	8. 95	0. 73	8. 82		1. 02	7. 90	0. 99	8. 03
MGG2	pН	0. 34	9. 02	0. 39	9. 17		0. 40	8. 02	0. 72	7. 98
DAD1		0. 87	9. 39	0. 83	8. 69		0. 82	7. 30	0. 73	3. 00
PAP1	pН	0. 45	9. 46	0. 43	9. 27		0. 49	7. 86	0. 28	4. 00
W D		0. 99	9. 45	0. 73	8. 89		0. 83	7. 91	0. 72	8. 27
M+P	pН	0. 41	9. 31	0. 38	8. 90		0. 43	7. 91	0. 37	7. 64
M+P: M	GG2+PAP1,	:	рН	, рН	:		pН	5. 17		
2)										
,	PAP1	MGG2						,	,	
			1: 1	, 5	i	3				
			24					2	3	
6.	62 3	. 65	가		2					
フ	ŀ			•						가
					:	,	,			
		가	3							
		3			3. 65		2	1 . 60 , 3	5. 88	7. 64
가										

					+
	рН	3. 32	3. 32	3. 32	3. 32
0	TA	1. 77	1. 77	1. 77	1. 77
	CFU	9. 0	9. 08	9. 08	9. 08
	pН	3. 36	3. 32	3. 48	3. 31
1	TA	1. 76	3. 13	1. 79	2. 53
	CFU	7. 39	7. 95	7. 74	9. 22
	pН	3. 48	3. 41	3. 58	3. 39
2	TA	1. 91	2. 40	2. 06	2. 57
	CFU	6. 62	6. 81	7. 29	8. 06
	pН	3. 37	3. 25	3. 49	3. 28
3	TA	1. 85	2. 66	1. 98	2. 80
	CFU	3. 65	4. 60	5. 98	7. 64

: 10% (, ,), : 8% , : 50ml 50ml 4

3)

가

		0	24
	рН	4. 60	3. 32
PAP1+MG2		0. 77	3. 54
		8. 27	9. 16
	рН	5. 98	3. 84
Hansen YC-180		0. 38	2. 00
		8. 15	9. 15
	рН	5. 89	3. 53
N		0. 44	2. 66
		8. 32	8. 51

Hansen 0.4U/kg , N 0. 2%(W/V) 25 Hansen 8. 15 0. 38, N 8. 32 0. 44 , PAP1 MGG2 8. 27 0. 77 , Hansen 가 N 가 가 . **24** Hansen 8.51 2.66 , 9. 15 2. 00, N 9. 16 3. 54 . Hansen 가 가 가 Hansen 4) **4**% 가 0.5% 26 0. 1%(MRS 18

- 99 -

(%)	рН		
0. 1	3. 89	2. 06	9. 42
0. 3	3. 90	2. 04	9. 42
0. 5	3. 90	2. 07	9. 49

5.

가. 가

(20)

가

Phytone peptone,

48

24

24

•

가

yeast extract glucose

. 27 가

가 가 가 . Yeast

extract Phytone peptone 7

. Yeast extract 가 Phytone

peptone 가가

Phytone peptone 0.8%, yeast extract 0.8%, glucose 0.5% 7 † 7 yeast extract 0.2%

, Phytone pepton 0.4% $$7\mbox{\ensuremath{\dag}}\ 9.0$

. 가

glucose 0.5, peptone 0.4, yeast extract

0. 2%

27. Phytone Peptone Yeast Extract 가

		Yeast Extract(%)									
Phytone (%)		0	0. 1	0. 2	0. 4	0.8	0	0. 1	0. 2	0. 4	0.8
` '		Sugar 0%				Sugar 0.5%					
	pН	3. 83	3. 87	3. 90	3. 95	4.03	3. 21	2. 98	2.95	3.07	3. 24
0		0.91	1. 19	1.39	1. 55	2. 30	2.87	3.33	3.85	4. 01	4. 56
		8.39	8. 45	8.48	8.49	9.06	8.79	8. 70	8.80	8.86	9. 07
	pН	4.04	3.99	3.97	4.05	4.06	3. 01	3.07	3. 14	3. 23	3. 24
0.2		1. 07	1. 23	1. 39	2. 60	2.04	3.71	3.83	3.98	4.04	4. 24
		8. 85	8.38	8. 51	8. 53	8. 54	8. 69	8.93	8.71	8.90	8. 81
	рН	4. 19	4.11	4. 12	4.09	4.09	3.08	3. 10	3.08	3. 17	3. 23
0.4	I					1. 99				4. 33	
		8.78	8.92	8.54	8. 77	8. 75	9. 21	8, 87	9. 14	8. 65	8, 80

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( 27
          )
       рH
            4.45 4.39 4.30 4.29 4.29
                                     3.25 3.32 3.33 3.36 3.48
                                     3.60 3.96 3.95
0.8
            1.47 1.46 1.54 1.67 2.37
                                                     4. 12 5. 11
            8.86 8.68 8.53 8.56 8.93
                                     9.10 9.05 8.99
                                                     9.35 9.42
                                     3.52 3.55 3.56 3.60 3.62
      pH 4.84 4.66 4.62 4.59 4.54
1.6
           1.56 1.74 1.69 1.85 2.22
                                     3.93 4.07 4.14
                                                     4.58 4.57
           9.23 8.84 9.23 8.84 9.16
                                     9.31 9.21 9.26
                                                     9.36 9.23
 Phytone pepton yeast exract 가
                           가
                                                      . Phytone
pepton soybean protein 가
                  ( : soywhey)
                                          , Yeast extract
               가
        가
            ( , 1989,
                               1989),
                 yeast extract
                                          28
                         가가 1/3
   yeast extract
```

가

가

Phytone peptone

가

가 28.

		DW	YE	YE+G	RY	RY+G
	pН	3. 89	3. 95	3. 33	3. 96	3. 24
DW		1. 07	1. 50	3. 53	1. 12	3. 28
		8. 66	8. 98	8. 99	8. 56	9. 05
	pН	4. 60	4. 49	3. 70	4. 57	3. 65
Pt		1. 33	2. 05	4. 08	1. 54	3. 81
		8. 93	8. 91	9. 46	8. 93	9. 18
	pН	4. 24	4. 21	3. 49	4. 26	3. 41
SW		1. 38	3. 84	3. 68	1. 19	3. 33
		9. 00	8. 92	9. 01	8. 84	8. 91
	pН	4. 12	4. 12	3. 45	4. 16	3. 36
SM	•	1. 12	1. 63	3. 67	1. 18	3. 34
		8. 98	8. 85	9. 08	8. 92	9. 21
DW:		ytone peptone , G: glucose (25%(V/V),	SM (,	0. 4%, YE

, 1%)

가

가

가 (, **1996**). 가

. 0.4% Phytone

pepton, 0.2% yeast extract, 0.5% gluocse

. 29 40%(V/V)

가 .

가

가가 가 4가

가 가 가가

. 4가 가

가가

·

. 가 가

. 가 , 가

가 .

29. 가

가 (%, v/v))		
	рН	3. 41	3. 22
0	TA	3. 31	3. 61
	CFU	8. 89	8. 99
	pН	3. 22	3. 31
5	TA	3. 80	3. 75
	CFU	9. 03	9. 04
	pН	3. 29	3. 34
10	TA	3. 70	3. 69
	CFU	9. 03	9. 04
	pН	3. 16	3. 32
20	TA	3. 80	3. 71
	CFU	9. 06	9. 07
	pН	3. 29	3. 34
40	TA	4. 02	4. 01
	CFU	9. 11	9. 14

: 7% : 10%

: Phytone peptone 0.4%, yeast exract: 0.2%, glucose: 0.5%

pectin sodium alginate

, pectin alginate

7 polymer .

7 pectin sodium alginate

0.5%

7 .

30 가 가 . pectin

, alginate

가 .

·

polymer .

30. 가

al ginate가

(%)		alginate Na	pectin
	рН	3. 32	3. 32
0. 00	TA	3. 48	3. 48
	CFU	8. 99	8. 99
	рН	3. 10	3. 35
0. 10	TA	3. 52	3. 41
	CFU	9. 01	8. 63

- 106 -

(30)
------	---

	pН	3. 12	3. 34
0. 20	TA	3. 57	3. 29
	CFU	8. 19	7. 94
	рН	3. 20	3. 44
0. 30	TA	3. 83	3. 39
	CFU	9. 12	7. 85
	рН	3. 23	3. 41
0. 40	TA	3. 85	3. 59
	CFU	8. 86	8. 18
	рН	3. 21	3. 39
0. 50	TA	3. 87	3. 89
	CFU	8. 85	8. 21

37 24 가 1 가 가 가 2% 가 가 1) 가 0. 5% 가 가 10%가 가). (), 가 31 가 가 1:1

(%)				
10	2. 3	3. 8	2. 5	3. 8
5 + 5	2. 5	3. 8	2. 5	3. 6
10	2. 1	3. 2	2. 1	3. 4
4 + 2	2. 7	4. 1	2. 8	3. 8
: 8%, sodium alginate 0.2%,	0. 5%,: 2%,	0. 4%, : 5	0. 2%,	
2)				
				가
가 가	3. 0-2. 5			32
		가		가
	가	가 .		
0. 5%				가
가	•			

3. 7	3.0	3	3. 8	2. 7	3. 8	3
3. 0	3.8	3	3. 9	2. 4	3. 9)
2. 5	3.8	3	3. 9	2. 4	3. 9)
2. 0	2.8	3	3. 9	2. 3	3. 8	3
	: 8%,	0. 5%,	0. 4%,	0). 2%,	
	sodi um al gi	nate 0.2%,	2%			
:	5% +	5%,	: 5			
:	: 5N NaOH solution	1				
3)						
	가		가	가	,	
			가			
			71			
				가		
		71	71			20
		가	가	•		33
	,	,	8: 1	: 1	가	
가						
·	<i>,</i>		- 0.		- 0.	0.000:
			5%,		5%,	0. 08%,
	0. 01%,	0. 0	01%		3. 0	

(x 0.0	01%)					
8+	2		3. 0	3. 8	3. 4	3. 5
8+	2		3. 4	4. 2	3. 7	3. 7
8+	1+	1	4. 1	4. 2	4. 1	4. 4
6+	2+	2	3. 9	4. 2	3. 9	4. 1
0. 2%,	: 8%, 2%,	0. 5%	6, O. 5% +	4 %, 5 %,	0. 2%, so : 5	odium alginate , : 3.0

5.

Lactobacillus MRS 가

pepton, yeast extract

•

가 .

가.

가 .

34

가 .

•

34.

	(mg/l)*	(mg/l)**
BOD	7, 325	3, 312
Acetic acids	15	226
Lactic acids	10	1, 080
Monosacchari des	980	*
Sucrose	5, 740	*
Raffinose	990	*
Stachyose	3, 080	*
Total Nitrogen	365	198. 5
Total Phosphorus	242	133
pН	5. 5	4. 2
Calcium	430	22. 3
Sodi um	223	24. 9

^{*:} Ji n, 1994 **: Kang , 1993

1)

 0.2-1.6%
 가

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35.	가
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(%)	рН		
0.0	4. 55	0. 78	8. 96
0. 2	3. 95	1. 43	9. 21
0.4	3. 95	1. 46	9. 12
0.8	3. 94	1. 49	8. 90
1. 6	3. 95	1. 47	8. 95

2)

0. 1-0. 8% 36 .

2% フト 0.4%

. 0.4%

가 .

36. 가

(%)	рН			
0. 0	4. 55	0. 78	8. 96	
0. 1	4. 57	0. 90	8. 95	
0. 2	4. 57	1.00	8. 97	
0. 4	4. 58	1. 11	9. 25	
0.8	4. 56	1. 55	9. 18	

3) 가 가

. 0. 1-0. 8%

가 37

. 가 0.4%

. 0.4%

가 .

37. 가

(%)	рН		
0.0	4. 55	0. 78	8. 96
0. 1	4. 55	0. 90	8. 91
0.2	4. 55	1. 01	8. 98
0.4	4. 53	1. 23	9. 22
0.8	4. 48	1. 66	8. 89

4) 가

0. 2%, 0. 4%,

0.4% 가 , Lactobacillus

MRS . 38 MRS

•

가

가 .

38.	MRS	

	рН		
MRS	4. 46	4. 29	9. 47
	3. 95	1. 43	9. 42

.

가 .

. 39

2. 18 9. 05 1. 30

8.83 , 가 4.80

9. 33, 5. 53 8. 83

가 , 4%

가 . 1% 가

•

39.

	가		가
1. 30	5. 58	2. 19	4. 80
8. 83	9. 36	9. 05	9. 33

가 : **25g/100ml**

4

1.

가.

MIR1, M2R1, M2R6, M3R3, F1R4

MGG1, MGG2, NBL1, PAP1, PAP2

•

10가 rapid32A kit

fructose-6-phosphate phosphoketolase acetate/lactate

MIR1, M2R1, M2R6, M3R3, NBL1, PAP2 Bi fi dobactri um
F1R4, M3G1, M3G2, PAP1 Lactobaci ll us .

2.

가.

4%

0. 81-2. 11 ml (0. 1N NaOH)

•

2% 32%

5

가 , 가

8% MIR1, MGG2, MGG1, F1R4, PAP1 32% 8% 8% 가 가 3. 가. 가 가 0. 5-8% Lactobacillus 가 가 가 Bi fi dobacteri a 가 0.05% Cysteine, 0.2% Yeast Extract 0.4% Gelysate Peptone, Phyton Peptone, Trypti case Peptone 7 , Phyton peptone 가 F1R4 PAP1 가 가 . Lactose, maltose, sucrose gl ucose

, 4%

가

가

F1R4 PAP1가 가

- 117 -

가 가 0. 5-8% 가 가 0. 5-8% 0. 5-2% **4**% 10% 가 F1R1, MGG1, PAP1가 가 , F1R4 , PAP1 , MGG1 가 **48** 가 가 가 , 24 24 48 F1R4, MGG1, PAP1 가 가 1)

- 118 -

24

8. 05-9. 05

8%

MIR1 PAP1 9.05 9.03 가 . 48 8. 28-9. 15 PAP2 M2R1가 9.15, 8.99 가 24 2) 1: 1 **20ml** 0. 8ml 가 48 F1R4 PAP1 3. 26, 3. 66 MGG1, MGG2, NBL1가 9. 22, 9. 16, 9. 24 가 가가 가 가 가 가 . 가 () () 가 가 가 가 0.5% dextrose가 가 4. 39- 14. 67 3. 35-9. 52 가 4.

- 119 -

가.

8. 58-9. 32

F1R4, PAP1 9.05, 9.32 가

5. 26, 5. 21 1. 30-2. 45

3

F1R4 PAP1 가

(5) 3 MGG1 MGG2가 8.00 8.03

8. 66 8. 83 가 . **F1R4** 가

PAP1 9.05 9.32 3 2.80 3.00 가

. **F1R4 PAP1** 가 가

가 가

pН

pH 5. 17 5 12

pН

6. 76 4. 38 1000

pН

가

, MGG1 10% 1. 43 8. 61 , 23% 가 1.08 8.26 . 12 F1R4 4. 42 8. 00 4. 05 7. 56 가 1.43 MGG1 8. 14 23% 1. 13 7. 03 , F1R4 4. 70 3. 02 4. 40 3. 68 MGG1 F1R4 . 1 10 가 1) Lactobacillus PAP1 Lactobacillus MGC2 가 3 pH 5. 17

- 121 -

2) PAP1 MGG2 1:1 , 5 3 2 3 6.62 3.65 가 가 2 가 3 3 3.65 4.60, 5.88 가 7.64 3) Hansen YC-180(S. thermophilus bul gari cus MSK B2(S. thermophilus, B. infantis) , 24 aci dophi lus Hansen 가 MSK 가 가 가 Hansen 4) 0.5% 0.1%(MRS 18) **5**. 가. 가 가(0.5%) 가 가 가 Yeast extract Phytone

Phytone peptone

peptone 가

```
가
0.8%, Yeast extract 0.8%, Glucose 0.5%
                                              5.11 9.42 가
                   , Glucose 0.5%
                                           Yeast Extract 0.2%
 , Phytone Pepton 0.4%
                               가
                                                 가
                                                         3.89
9.0
 Phytone pepton Yeast Exract 가
                                           가
                                     가가 1/3
                Yeast Extract
           Phytone peptone
                               가
                                     가
                                                       가
                가
                                                     40%(V/V)
               가
            가
   가
                   pectin sodium alginate 0.5%
          가
                                    al gi nate
        가
               , pectin
                   al ginate가
                                          8%,
                                                   0.4%,
    0.2%,
                0.5%, sodium alginate 0.2%, modified starch 2%
 1)
```

가

10%가 가

5% **5**% 2) 3. 0-2. 5 3) 0. 08, 0. 01, 0. 01% 가 가 5%, 5%, 3.0 0. 08%, 0. 01%, 0. 01% **5**. 가. 가 1) 가 0. 2-1. 6% 가 0.2% 2) 0.8% , 0.4% 가 3)

0.4%

- 124 -

0. 1-0. 8%

•

4) フト 0. 2%, 0. 4%,

0.4% 7 , Lactobacillus

MRS . 가

•

. 2. 18 9. 05 1. 30

8. 83 , 가 4. 80

9. 33, 5. 53 8. 83

가 4%

가 .



3 가

1

, 가

가 , , 가

,

가 . 가 가 .

가

,

2 ,

,

(, 1998, 1999) , 3

pilot (Hoyer, Denmark, 201/hr) .

1.

가

가.

40 . 40 , Y

, У

Wiesby, Culture System, Sanofi, Valio,
C. Hansen , ,

- 128 -

M		,	G, F
N		,	G, D
S		,	D, F
Н		,	D, N
В		,	D , F
Н		,	N, F
D		,	F,
Y	,	,	J,
G		,	
В		,	

N: New Zeal and, D: Denmark, F: France, J: Japan : (7 | cost)

 가
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 가
 (5)

(granule type) pellet 가 , 가 가 가 가(cost) 가 가 (Streptococus) 1) S. thermophilus 가 , S. lactis, S. cremoris . acetal dehyde, diacetyl **40-45c** . (Lactobacillus) L. acidophillus L. bulgaricus7 casei 가 . 가 0. 3-1. 6% L. acidophillus 가 35-38cC , *L. bul gari cus* 40-43cC 가

- 130 -

3) (Bi fi dobacteri um) B. bifidum 4) (Saccharomyces) , Kefir plain 가 가 가 가 Streptococus Lactobacillus 가 가 가

가 .

(C. Hansen's Laboratory, Denmark) S. thermophilus(TH-3), L bulgaricus(LB-12), S. thermophilus L. bul gari cus (YC-180) 3가 10.0% 90.0% starter 125cC 7.5 100U/250kg 가 , **42**cC 2 , 10ml sample

0. 1N NaOH ml

, 41 , S. thermophilus L. bulgaricus

가

41.

	0	1	2	3		5	6	7	8
S. thermo.	2. 0	2. 0	2. 3	3. 2	4. 0	4. 5	5. 2	5. 5	5. 8
L. bulgar.	2. 0	2. 0	2. 4	3. 0	3. 8	5. 1	6. 2	6. 5	7. 0
S+L	2. 0	2. 0	2. 2	3. 9	6. 0	7. 8	9. 5	9. 9	10. 5

S+L: S. thermophilus + L. bulgaricus

Lactobaci lus , Streptococcus acetal dehade 가 , 가 가 S. thermophilus L. bulgaricus 가 가 가 가 가 가 가 가 1 Bi fi dobacteri um Lactobacillus 2. 37-38 가. 가 가 가 가 가 , 가 가 . 가

가 가 Color Difference Meter (Hunter Colorimeter, D25-9, Sensor-D25Optical sensor) L. a. b. 1) 가) 12%, (sodium alginate) 0.2% 5%, 10%, **15**% , 12% 가 12-20% 가 14-16% (Arbuckle 1977), , 8-12%) (H), 가 8-16o Brix 12 (Rank Test: 1982) Duncan's

75cC

- 134 -

)

```
100kg/cm2
                         head
                                                        15ml
50ml
              (centrifuge cell)
 2)
                가
                                        , 가
                                                   72%),
(
                        (
          ),
 가
                   (110-114)> (100)>
                                              (70) (Catsberg &
Kenpen-van Domel en, 1990)
                                               3%
                                                        , 125cC
    7.5
                               50U/250kg
                                          100U/250kg
가
       42cC
                                        sampling
                                                    pН
                           1
                                   5ml
             digital pH Meter(Piccolo ATC pH meter, Padova,
 pН
Italy)
             (Herbal Extract) 가
 3)
                          가
                                             가
```

- 135 -

1 2 가 가 100 12%, 5%, 0.2% 0.3%, 0. 05%, 0. 5%, 0. 1%, 가 0.3% 5 , 가 가 1 5 가 (H , 1983) 4) 12% 가 7%, (2) , (1) **6**% + 3% + 가 9%, (3) 9% + **5**% 3 (1983) 5) (Body) 가 ((Modified Starch KYP,

0. 2% 2. 0%

가

Excell pro Inc., U.S.A)

. 15ml 3

, (Grindsted A/S, Brabrand, Denmark) .

.

1)

(Supermascolloider, Wet, Japan)

10, 30, 50, 100, 150 5 가 1 .

2)

, 가

, 가 가

500ml , 125cC 7.5 ,

D value

105cC, 95cC, 85cC, 75cC, 65cC , (異臭) .

.

가 () 1) (1.0 x 107/gr, 14-13-7) 0-6€ 3 10 6 sampling 1 5 1) 1 24 (20 가 2) 15 3 5 1

가 .

가

- 138 -

3

1.				
,	, 가	, paste	e block	
	[外皮]	[內皮]		(剝皮
栗), [生栗]	,			
: , 90 95	20			
0 6	:	50 Brix	가	12
0 30				
: 가 , 9	0 95	20		paste
40×6	60cm PE	-30		
i ce block ,			:	paste
block 47		가		
	,	,		,
4	,			가
. 42	,			paste
bl ock	가			
paste block	가			
,	loss가			
		가		
가 가				

- 139 -

				()		
	0.4(1)	1. 5(2)	0. 2(4)	0. 2(4)	2. 3	2 가
	0. 3(2)	0. 5(4)	0. 6(2)	0.6(2)	2. 0	
						가
가	0.1(4)	1.0(3)	0.8(1)	0.4(3)	2. 3	
n	aste 0. 2(3)	2 0(1)	0. 4(3)	0.8(1)	3. 4	가
block		2.0(1)	0.4(3)	0.8(1)	3.4	
	10	50	20	20	100	
*	: 1 :4	, 2 :3 ,	•	l :1		
	. (,			
2.	(,),	(,)		
	,			7	'L	
				,	Γ	
,	-					

- 140 -

, , ,

, 가

.

, 43

 $64. 33 \pm 0.05\%$, $2. 90 \pm 0.10\%$,

1. $05 \pm 0.00\%$, 11. $01 \pm 0.12\%$, 49. $55 \pm 1.25\%$, 62. $52 \pm 0.11\%$

3. $19 \pm 0.08\%$, 0. $83 \pm 0.01\%$, 8. $99 \pm 0.09\%$, 50. $65 \pm$

2. 05%

, $61.77 \pm 0.04\%$ $63.50 \pm 0.02\%$, $2.70 \pm 0.11\%$

 $2.82 \pm 0.12\%$, $0.85 \pm 0.02\%$ $1.04 \pm 0.02\%$, $9.28 \pm 0.04\%$

 $9.73 \pm 0.13\%$, $52.69 \pm 0.49\%$,

2... 2. - 2...

,

, L Value a b Value

, 60. 54, 60. 80, 60. 57 59. 06 L Value - 3. 34,

-3.34, -2.85 -2.90 a Value, 20.65 17.19,

19.44 19.24 b Value 가

7 11 10 71 10 101 10

a b Value

 $52.90 \pm 1.87\%$

가 .

	(%)					
	64.33 ± 0.05	62. 52 ± 0. 11	61.77 ± 0.04	63.50 ± 0.02		
	0.66 ± 0.05	1.08 ± 0.08	1.03 ± 0.00	0.63 ± 0.07		
	2.90 ± 0.10	3.19 ± 0.08	2.70 ± 0.11	2.82 ± 0.12		
	31.06 ± 0.12	32.38 ± 0.21	33.46 ± 0.14	32.20 ± 0.14		
	1.05 ± 0.00	0.83 ± 0.01	0.85 ± 0.02	1.04 ± 0.02		
	11.01 ± 0.12	8.99 ± 0.09	9.28 ± 0.04	9. 73 ± 0.13		
	49.55 ± 1.25	50.65 ± 2.05	52.69 ± 0.49	52. 90 ± 1. 87		
L Value	60.54 ± 0.78	60.57 ± 0.92	60.80 ± 0.52	59.06 ± 0.37		
a value	-3.34 ± 0.26	-2.85 ± 0.28	- 3. 34 ± 0. 21	-2.90 ± 0.22		
b value	20. 65 ± 0. 31	19.44 ± 0.60	17. 19 ± 0. 45	19.24 ± 0.55		

, 가

,

, 가

. **1998** 3 가

(44).

44. (1998)

		(%)						
62	. 43	3. 91	1. 39	1. 75	0. 12	30. 40		
± 1	. 16	± 0. 43	± 0. 21	± 0. 06	± 0. 01	± 1. 34		
61	. 59	3. 58	1. 37	2. 07	0. 13	31. 27		
± 0	. 95	± 0. 10	± 0.03	± 0. 10	± 0. 01	± 0. 90		

* 3 ±

,

,

가

,

, 가 ,

,

가

가 .

3. ,

,

가 가 . 가

가 , 가

2.8 . , 가 ,

•

,

가 .

,

, 가 , 가 .

,

4. 가. 1

, (1) paste block , (2) , (3)

, 5% 가 , 가가 lactose ,

Lactobacillus + Bifidobacterium

2:1 가

가 1 (45).

45 ,

plaste block 150mesh , 가 90 125 20 25 ,

7) 기

,

, (1) 가

bottom note top note 7 , body , (2)

, (3)

가

18

45. 1

(%)	
8	
23. 5	
6	+
2	
0. 5	L: B=2: 1
10	
15	
35	2 가
100	

L: Lactobacillus, B: Bifidobacterium

, 가 ,

가 .

.

•

1 ,

,

가 .

1) 7ŀ)

46

. 5, 10 15% フト , 10% 15% フト 5%

1%

, 10% 15%

46.

	(%)	*
5	-6.8	
10	1.7	
15	5. 1	

=1%

* : 1=0.85, 2=0, 3=-0.85

47.

(%))		
	4	16	24
5	+	++	++
10	++	+++	+++
15	+++	++++	++++

50ml

,

8%

- 148 -

2)

, 48 .

가 ,

. 100 50U/250kg 가

•

48. pH

	рН							
(U/250kg)	0	1	2	3	4	5	6	7
100	6. 6	6. 5	6. 4	6. 2	5. 6	5. 5	5. 1	4. 9
50	6. 6	6. 6	6. 5	6. 4	5. 7	5. 7	5. 3	5. 0
100	6. 6	6. 5	6. 4	6. 1	5. 7	5. 4	4. 9	4. 7
50	6. 6	6. 5	6. 4	6. 4	5. 7	5. 4	5. 1	4. 9
100	6. 6	6. 4	6. 1	5. 4	5. 1	4.8	4. 5	4. 5
50	6. 6	6. 5	6. 4	6. 2	5. 6	5. 3	5. 1	4.8

(P<0.05)

, 가 가 가 ,

,

.

가 50U/250kg 가

100U/250kg .

3)

가 6: 7, 3: 9, 6: 7

,

49 .

7% 가 가 9%

5% 가가 , 가 .

6% 7%

49.

*

G6+HFC7

2. 55

G3+HFC9

0. 85

G9+HFC5

-3. 4

G: , HFC:

*: 1=0. 85, 2=0, 3=-0. 85

4) 가

가 .

4가 가 가 가 가 () 가 **50** 가 가 가 가, 가 가 가 가 가 **50**. * 1 387 0.493 322 0. 134 4 248 -0.095 6 366 0.400 2 201 -0.551 7 0. 313 3 355

*:

5) (body) 8% (100) ,

0.114

5

58 .

285

, 가 .

, 가

0. 2% 2. 0% 가 2. 2% 가

, , body 51 .

가 가 74, 81

가 104 가 가 ,

. body

51. 가

	가 (%)	*	**	
(SA)	0. 2	74	++++	body
(MS)	2. 0	81	+++	
SA+MS	2. 2	104	+	

*: 100 , **: overnight

•

,

가 가 .

1)

가 . 52 50

가 , 100

가 **52**.

() 150 100 **50 30** 10

2) (異臭) 가

가

75

105 **53** 가

85cC

53.

()		
125	+++	가
105	+++	가
95	++	가
85	+	가
75	-	가
65	-	가

3)

가)

(14-13-7)

,

. , 107/gr

,

가 .

)

, 1010CFU/ml

•

,

•

9. O(logCFU/ml) ,

19 54 . 19 9.0

,

1x109 19

가 .

54.

	(hour)	(log10CFU/ml)
0		7. 85
6		8. 06
9		8. 39
12		8. 71
18		8. 99
19		9. 16

4)

Lactobacillus MGG1 Lactobacillus PAP1

7\ 7\ 7\ ... (55). 37 15 9.0(logCFU/ml) 55.

56 37 , 19 9.0 가 .

가 (57).

가

•

56.

8 0.4 0. 2 0.5 0. 2 2.0 100 **57**. 5 5 0.08 0.01 0.01 100

,

5)

가

.

가)

--> block --> (**8**% + +

6% + 7% + 2.2%)--> -->

--> --> -->

)

--> bl ock --> (**8**% + +

0. 5% + 0. 4% + 0. 2% + 2. 2%)--> -->

--> (+ 0.2% + 0.4% + 0.4%)-->

--> --> 7ト(5%, 5%, **0.08**%,

0. 01%, 0. 08%) - -> -->

.

1) ()

가 .

가)

58.

1	,	가 ,
2		100
3	bl ock	20 _c C
4		65cC, 8%, 6%, 7%, 0. 2%, 2. 0%,
5		100kg/cm2, plain head, 75oC
6		85cC
7		S. thermophilus + L. bulgaricus ready-to-use type, Hansen YC- 180, 100U/250kg
8		37aC, 19 , 1.0x109
9	curd	100kg/cm2, plain head
10	,	
11	,	0-6dC, 12-15 , 1.0 x 107

)

59.

1		,	가 ,	
2			100	
3	bl ock		20 cC	
4			65 c C, 8	%, 0. 5%
			0. 4%,	0. 2%,
			0. 2%,	2. 0%,
5			100kg/cm2, plain he	ad, 75oC
6			120cC	
7			, 0. 2%,	0. 4%,
			0. 4%, 37	, 15
8			Lactobacillus PAP1+	MGG2, 2%
9			37cC, 19 ,	1. 0x109
10		가	5%, 5	%, 0.08%
			0. 01%,	0. 08%
11	cur	rd	100kg/cm2, plain hea	ad
12	,			
13	,		0-64, 12-15	,
			1. 0x 107	

2)

60

60.

1 2 2.0%() 3.0% pН 3.85 4.60 3. 4-4. 0

15dBrix $1.\ 0x109CFU/ml$

1: Hansen YC-180 2: Lactobacillus PAP1 + Lactobacillus MGC2

()

가

show case

가 61 12

5

가 , 가 12-15

. 15

.

가

•

61.

()		
0	9. 16	G
3	9. 18	G
6	8. 91	G
9	8. 34	G
12	7. 93	В
15	7. 84	В

G: Good, B: Bad(12 15 5 1)

4

1.

, , 가 , paste block 4가

, , 4

가 paste block

가 , 가 , ,

(,), (,) 가 a b Value 가 가 3. 가 가 2.8 **4**. 가. 1 plaste block 150mesh , 5%, lactose 2% 가 , Lactobacillus + Bifidobacterium 2:1 가 가 90 125 20 25

2.

가

bottom note top note가 , body 1) 가) 10% 가 5% **15**%) 5-10%가 **8**% 2) 가 100 50U/250kg 가 가 , 가 100U/250kg 가 3) 가 6: 7, 3: 9, 9: 5 6:7 가 9:5 가

- 164 -

가 4) 가 **4**가 가 가 가 가, 가 가 (body) 5) 가 가 0.2% 2.0% 2. 2% 가 가 가 74, 81 가 가 가 , 104 , body 1) 가 가 **50** , 100 가 2) 가 (異臭) 105

가

75

3) 가) 가 $10.0(\log CFU/ml)$ 9.0) 9.0 19 4) Lactobacillus MGG1 Lactobacillus PAP1 0. 2%, 0.4% 가 , 37 15 9. 0(logCFU/ml) 8, 0. 4, 0. 2, 0. 5, 0. 2, 2. 0, 100 37 19 가 9.0 가 5, 5, 0.08, 0. 01, 100 0.01, 가

85cC

- 166 -

```
5)
가)
[ --> bl ock --> ( 8% + +
6% + 7% + 2. 2%)--> -->
--> -->
       -->
              --> ]
)
[ --> bl ock --> ( 8% + +
0. 5% + 0. 4% + 0. 2% + 2. 2%) --> -->
--> ( + 0. 2% + 0. 4% + 0. 4%)-->
--> --> 7ト( 5%, 5%, 0.08%,
0. 01%, 0. 01%) --> --> ]
•
            (
                         )
      show case
                   가 ,
      12
                    5
, 가 12-15
                         가
```

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-	168	-

4

1.					•	
2.				가		
			가		,	
	100					
3.			가		Lactobacillus	PAP1
	Lactobacillus M		,			
	Hansen YC-180(<i>S</i> .	thermi phi l us	L.	bul gari c	us)	
4.						
5.			•		가	
6.	•					
7.		·				
8.	·					
9.				_		
10.				•		

.

62.

1		, 가 ,				
2	100					
3	block	20cC				
4		1. :				
		65cC,				
		8%, 6%,				
		7%				
		0. 2%,				
		2.0%,				
		2. :				
		65aC, 8%,				
		0. 5%, 0. 4%,				
		0. 2%,				
		0. 2%, 2. 0%,				
5		100kg/cm2, plain head, 75oC				
6		1. : 85cC				
		2. : 120cC				
		(

7 , 0.2%,

0. 4%,

0. 4%

37 , 18

8 1. S. thermphilus +

L. bulgaricus

(Hansen YC-180)

100U/250kg

2. :

Lactobacillus PAP1+MGC2,

2%

9 37dC, 19 ,

1. 0x109

10 가 5%, 5%,

0. 08%, 0. 08%

0. 01%,

11 curd 100kg/cm², plain head

12 ,

13 , 0-6cC, 12-15 ,

1. 0x 107

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- 171 -

	170	
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1: 257-279.

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2.				
3.	가			

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