

최 종
연구보고서

636.413

L 2937

GOVP1200101815

거세돈의 도체 품질 개선을 위한 영양, 호르몬,
면역학적 기술 개발

Nutritional, Hormonal and Immunological Approaches for
Improving the Carcass Quality of Finishing Barrows

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농 립 부

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2000 . 10 . 28 .

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I.

, ,

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(boar-taint)

가 가 .

androsterone

110kg

가 .

가

가 .

.

,

anabolic steroids

implanting

.

,

anabolic steroids implantation

insulin-like growth factor(IGF) system

IGF system

NRC

() 85%

IGF-binding protein-3(IGFBP-3)

RIA

IGF system

component

acid-labile(ALS) cDNA fragment cloning

2

60kg 2[

(80%)] × 2[] × 2[no

implantation anabolic steroids implantation] factorial

105kg RNA

3

3 , estradiol, testosterone, IGF-I,

IGFBP-3 , 1 cloning ALS cDNA fragment

probe ALS mRNA level

(adrenocorticotropic hormone; ACTH)

(active

immunization)

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IV.

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1.

가.

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가

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(

가

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2.

가.

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3. Revalor(trenbolone + estradiol pellets) implantation

가.

. Trenbolone

(estrogen implantation

가 가

)

4. IGF system

가. IGFBP-3

1) IGFBP-3

RIA

- 2) **IGFBP-3**
- . **ALS cDNA fragment**
- 1) **300-bp ALS cDNA fragment cloning & sequencing**
 - 2) **cDNA clone**
- . , **anabolic steroids IGF system**
- 1) **IGF-I: Revalor implantation 가**
- &**
- 2) **IGFBP-3 :**
- Revalor implantation**
- 3) **ALS mRNA :**
가 가
 - 4) **IGF-I IGFBP-3**
가
 5. **ACTH**
 - 1) **ACTH 가**
 - 2) . ()

SUMMARY

I. Title

Nutritional, hormonal and immunological approaches for improving the carcass quality of finishing barrows

II. Objectives and Significance

Boars grow faster and also utilize the feed more efficiently than gilts, but the carcass quality of the former is inferior to that of the latter because of the boar-taint. The boar-taint is caused mainly by androstenone which is secreted from testes and accumulated in adipose tissues. Under the current situation where market pigs are slaughtered at 110kg of body weight, it is inevitable to castrate the boar because of the boar-taint.

Barrows have no boar-taint because their testes have been removed; however, they accumulate much more fat and utilize the feed less efficiently than gilts or boars, and moreover, their carcass grade is likely to be inferior to that of gilts. As for the way for preventing the excessive fat deposition of barrows, restriction of energy intake and administration of exogenous hormonal agents have been reported. Restricted feeding is well known as a means to limit energy intake, but this is not widely used in the field because of a few practical limitations. Use

of low energy diets, which also is not widely used in the field, has been known as a second choice for limiting the energy intake. As a third line of approach, implantation of anabolic steroid(s), has been introduced in European countries, but this has not been even tried domestically.

The present project was undertaken to investigate the effects of restricted feeding, feeding a low-energy diet and implantation of anabolic steroids on fat deposition and carcass quality and the expression of insulin-like growth factor(IGF) system components in finishing barrows and thereby to derive and make public practical means to prevent the excessive fatness of barrows and also to find insights into hormonal regulation of body composition and the role of the IGF system in somatic growth.

III. Experimental Approaches and Scope

The present study can be divided into three main steps: preparation, feeding trial and analyses of experimental samples of the trial. The preparation step includes formulation of a NRC-based control diet and a low-energy diet containing 85% energy level of the former and a preliminary feeding trial of the experimental diets. Also performed during the first step were a development of IGFBP-3 RIA following purification of the protein from porcine serum and raising its antiserum in a

rabbit and cloning of a cDNA fragment of acid-labile subunit(ALS), a major component of the circulating IGF system. At the next step, a feeding trial was performed using finishing barrows weighing 60kg under a 2[ad libitum vs restricted(80% ad libitum) feeding] × 2[control vs low-energy diet] × 2[no implantation vs implantation of anabolic steroids] factorial arrangement of treatments. Animals were slaughtered at 105kg, after which liver samples were taken for RNA extraction and carcass quality and physicochemical characteristics of the longissimus muscle section were analyzed. Blood samples were taken at the beginning day of the experiment and subsequently at three weeks intervals. The final step involved determination of concentrations in sera of estradiol, testosterone, IGF-I and IGFBP-3 and also determination of hepatic ALS mRNA abundance using the ALS cDNA fragment as probe that had been cloned during the first step. Also included in the final step was an initiative trial where effects of active immunization against ACTH, which is believed to play a significant role in increasing the body fat ratio, during the growing and finishing period of barrows on growth was investigated.

IV. Key Results and suggestions for Their

Utilization

Following are main results and their utilities of the

present research project.

1. Low-energy diet

- 1) Developed a low-energy diet containing a high percentage of grain by-product for finishing barrows.
- 2) Above low energy diet was confirmed to reduce the backfat thickness: this diet can be useful for excessively fat barrows.
- 3) The feed mill that participated in this project registered the low energy diet and made public the utility of it.
(Distribution of this report is considered to be a good channel of publication.)

2. Restricted feeding

- 1) Suppressed fat deposition and growth rate.
 - 2) Considered to be useful for small-scale production units.
3. Revalor(trenbolone + estradiol pellets) implantation
- 1) Suppressed fat deposition and weight gain and increased feed efficiency.
 - 2) Found not to be suitable for practical application because implanted barrows were judged as boars at the slaughter house, apparently resulting from an androgenic effect of trenbolone on development of external genitalia
(However, implantation of estrogen only deserves further investigation.)

4. Studies on the IGF system

- 1) IGFBP-3

- (1) Purified IGFBP-3, raised a polyclonal antiserum against it and developed its RIA.
 - (2) The antiserum can be distributed at request for non-profit research purposes.
- 2) ALS cDNA fragment
- (1) Cloned and sequenced a 300-bp porcine ALS cDNA fragment.
 - (2) This fragment can be distributed at request for non-profit research purposes.
- 3) Interactions of the IGF system with nutrition, feeding and anabolic steroids
- (1) IGF-I:
 - serum concentration increased by Revalor implantation.
 - exhibited no relation to feed intake or dietary energy.
 - had positive and negative correlations with growth rate and backfat thickness, respectively.
 - (2) Serum IGFBP-3 concentration:
 - found to have a positive correlation with growth rate.
 - found not to be related to feed intake, energy content of the diet, or Revalor implantation.
 - (3) Hepatic ALS mRNA abundance:
 - no apparent change by any treatment
 - (4) IGF-I and IGFBP-3 concentrations are considered to be useful as indexes of growth potential.
5. Active immunization against ACTH

- 1) no apparent effects of the active immunization on growth up to mid finishing period.
- 2) further observations/measurements up to marketing required(planned) as an initiative trial

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1

1

1.

· , , 1)

2) , 3) insulin-like growth factor(IGF) system IGF system

2.

(boar-taint) 가 가 (, 1996).

androstenone

(, 1996). androgen(

)

90kg 110kg 가

가 가

가

가

가

.

.

.

,

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가 .

anabolic steroids implanting

.

,

,

가

,

가

가

가

. 가

가

가

.

가

가 .

가

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3.

1)

가)

. Leymaster Mersmann(1991)

	85%	15%	
,	9%	7%	.
(1994)			80%
	14%	6.5%	
,	A, B	25.5%	.

)

. (1985) 가 3.4Mcal /kg

3.0Mcal /kg 가 11%

.

.

가 ,

가 (Baldwin , 1982) .

가

가 가 .

2)

가)

(growth hormone; GH) (Etherton Kensinger, 1984; Lee, 1988). Insulin-like growth factor-I(IGF-I) GH

7.5-kDa peptide (Jones Clemmons, 1995; , 1996; Liu LeRoith, 1999). 가

IGF가 IGF IGF-binding protein(IGFBP) IGF 40 45-kDa IGF-binding protein-3(IGFBP-3) 85-kDa acid-labile subunit(ALS) ternary complex (Baxter , 1989; Lee Rechler, 1995a; Rechler Clemmons, 1998). Ternary IGF:IGFBP-3:ALS complex IGF

IGF (Ueki , 2000). IGF-I, IGFBP-3

ALS IGF system 3 IGF-I peptide cDNA 가 IGFBP-3[porcine IGFBP-3(pIGFBP-3)] (Walton , 1989), pIGFBP-3 가 ALS protein cDNA cloning (Baxter , 1989; Leong , 1992; Dai Baxter, 1992; Baxter Dai, 1994; Delhanty , 1996; Lee Rechler, 1995b; Boisclair , 1996; Rhoads , 2000)

) Anabolic steroid .

anabolic steroids trenbolone
acetate[androst-4,9(10),11 trien-3-one, 17 β acetate; TBA]
estradiol-17 β 가 . TBA androgen anabolic steroid
testosterone anabolic activity가
(androgenercity) androgen
(Heitzman , 1977; Heitzman, 1979; Galbraith Topps,
1981; Lee, 1988). TBA Roussel UCLAF
Finaplix , estradiol
Revalor(TBA plus estradiol) .
anabolic steroids
Grandadam (1975) TBA(140mg) plus estradiol (14mg)
implanting 15% 10%
가 .
TBA plus estradiol anabolic action
. TBA plus estradiol IGF-I
, cortisol (Lee , 1990)
anabolic steroids IGF-I
testosterone
(cortisol) .
TBA plus estradiol IGF-I 가 IGFBP-3
ALS 가

) (active immunization)

insulin glucocorticoid(cortisol)가 . Glucocorticoids
adrenocorticotrophic hormone(ACTH)
catabolic hormone

(Henricks , 1984).

ACTH glucocorticoid hormone

가

. Silence (1992)

ACTH

37%

가

(passive immunization) 가

. ACTH glucocorticoid

가?

가

가?

가

ACTH

가

glucocorticoid

,

가

가?

,

somatotropin

가

(Kirby , 1993)

glucocorticoid hormone

2

implantation ACTH , anabolic steroids
IGF system . Table 1-1

Table 1-1. Yearly objectives and scope

Year	Objectives	Scope
1st (1997)	<ul style="list-style-type: none"> · Development of a low-energy diet · IGFBP-3 purification · ALS cDNA cloning 	<ul style="list-style-type: none"> · Formulation of a NRC-based control and a low-energy(85% NRC) diets · IGFBP-3 purification from porcine serum cation exchange chromatography IGF-I affinity chromatography C18 reverse-phase HPLC · Cloning of a porcine ALS cDNA fragment (PCR) & sequencing
2nd (1998)	<ul style="list-style-type: none"> · Development of IGFBP-3 RIA · Feeding trial 	<ul style="list-style-type: none"> · N-terminal amino acids sequencing · Production of antiserum in a rabbit · Development of pIGFBP-3 RIA · Feeding trial using a 2(ad libitum vs restricted feeding) x 2(energy levels) x 2(none vs anabolic steroids) factorial arrangement of treatments · Measuring live wt & blood sampling · slaughtering & carcass analysis
3rd (1999)	<ul style="list-style-type: none"> · Analyses of samples · Active immunization against ACTH 	<ul style="list-style-type: none"> · Serum glucose concentration, RIAs(IGF-I, IGFBP-3, estradiol, testosterone), Ligand blotting(total IGFBPs) · Hepatic ALS mRNA abundance: RNA extraction, Northern blotting RNase protection assay · ACTH-carrier coupling & injection to growing pigs · Blood sampling & analysis: Antibody titering, cortisol concentration · Analysis of growth and body composition

2 : , IGFBP-3 IGF-

I RIAs ALS Cloning

1

1.
가.

(過肥)

NRC

(1988, 1998)

Table 2-1

NRC

가

(digestible energy; DE) 103%

85%

15%

NRC

40

Table 2-2

110kg

가

(P

= 0.11)

(1985)

Table 2-1. Composition of experimental diets

	Control diet ¹	Low E diet ²
Ingredients		
corn, ground	62.68%	30.48
wheat	5.38	13.44
wheat bran		35.00
rice bran, polishings	3.00	3.00
soybean meal, sol.	17.76	9.64
rapeseed meal	2.00	2.00
limestone	0.48	0.88
dicalcium phosphate	1.80	1.62
salt	0.30	0.30
vitamin premix	0.30	0.30
mineral premix	0.26	0.26
tallow	3.04	
molasses	3.00	3.00
lysine-HCl		0.08
total	100.00	100.00
Chemical composition³		
crude protein	15.0	15.0
crude fat	6.2	3.2
crude fiber	3.4	5.2
crude ash	5.0	5.9
Ca	0.85	0.98
P	0.67	0.83
Lysine	0.75	0.75
Met. + Cys.	0.54	0.56
DE(kcal/kg)	3,509	2,946

¹Contains 103% NRC(1998) digestible energy(DE) requirement.

²Contains 87% NRC(1998) digestible energy(DE) requirement.

³Calculated values

Table 2-2. Growth performance and carcass measurements of finishing barrows fed the low-energy diet

Items	Control diet(n=30) ¹	Low E diet(n=27) ^{1,2}
Initial wt, kg	70.7 ± 1.2	72.0 ± 1.3
Final wt, kg	107.7 ± 1.3	103.6 ± 1.4*
ADG, gm	756 ± 22	661 ± 23*
Carcass wt, kg	69.7 ± 1.1	68.4 ± 1.2
Backfat(measured), mm	17.3 ± 0.64	15.2 ± 0.67*
Backfat(corrected), mm	17.8 ± 0.64	16.2 ± 0.67
A, B grade ratio, %	90	93

Data are LS means ± SE.

²Three stunted animals were excluded during the experiment.

*P<0.05.

90% A, B

2

2. IGFBP-3 RIA

가. IGFBP-3

1)

250ml centrifuge bottle

JA-14 rotor 3,000rpm(1,600 x g)/4, C 30

IGFBP-3

-20, C

IGFBP-3 Martin

Baxter(1996)

Zapf (1988)

Walton (1989)

2) SP Sephadex C-25 cation-exchange chromatography

가) 50gm SP Sephadex C-25 cation-exchange resin 1 M acetic acid(pH

3.0) equilibration 48 swelling

) cation-exchange resin swelling 1l 1l 2 M

acetic acid/150mM NaCl pH가 3.0 10

250ml centrifuge bottles rotating shaker 3

) centrifuge bottle 35ml swollen SP Sephadex C-25 resin

24 rotating shaker

) Resin 가 cheese cloth Whatman filter

10 N NaOH pH 6.5

) 40ml centrifuge tube JA-20 rotor 39,000

x g(18,000rpm)/4, C 1

) Whatman filter 2 IGF-I affinity column

loading

3) IGF-I affinity column (Lee Rechler, 1995a)

가) 2mg recombinant IGF-I 10mM HCl 1ml 2ml coupling

buffer[0.1 M NaHCO3/0.5 M NaCl (pH 8.0)] 가

) 0.6gm CNBr-activated Sepharose-4B resin 1mM HCl 50ml 15

swelling

) 1mM HCl 100ml swollen resin sintered glass filter washing

5ml coupling buffer 가 IGF-I 가:

coupling reaction .

-) Rocker platform 2 coupling
-) Reaction tube 5ml blocking buffer[50mM

Tris-HCl, pH 8.0]

-) 10ml blocking buffer 가 3 blocking
-) 4. C resin 15ml column 0.1 M sodium

acetate/0.5 M NaCl (pH 4.0) 20ml 0.1 M NaHCO₃/0.5 M NaCl (pH 8.3) 20ml

washing; 3

-) 50mM sodium phosphate(pH 6.5) equilibration

4) IGF-I affinity chromatography

4. C .

- 가) 2) IGF-I affinity column

30 60ml / (loading)

-) 1 0.5 M NaCl (70ml /) washing

-) Affinity column IGF-binding proteins(IGFBPs) 15ml 0.5 M

acetic acid(pH 3.0) elution

-) Centri con-30 ultraconcentrator 4. C

5) SDS-PAGE and electro-elution of IGFBP-3

- 가) IGF-I affinity chromatography total IGFBPs Laemli
- buffer 12.5%SDS-PAGE lane Coomassie staining IGFBP-3

-) IGFBP-3 band Mbl el 422 electro-eluter(Biorad)

5 IGFBP-3 elution(9mA/barrel; SDS-PAGE buffer)

) Centri con-30 - 20. C

*1 0.5 1mg IGFBP-3

1) 5)

6) IGFBP-3

total IGFbps electro-elution IGFBP-3 1 silver
staining(Pierce) (Fig. 2-1) total IGFbps
SDS-PAGE gel PVDF membrane transfer IGFBP-3
automated Edman degradation NH2
. Sequencing ---AVXTGPV
IGFBP-3 cDNA (Shimasaki , 1990)

IGFBP-3 .

. IGFBP-3

IGFBP-3 Lee Rechler(1996)가

1)

가) 0.2ml pIGFBP-3 0.4ml 50mM PBS(pH 6.75) (pH; 7.25)

0.6ml Freund' s Complete Adjuvant

) New Zealand White rabbit(5kg) preimmune blood
1ml (19 G needle) pIGFBP-3

Fig. 2-1. Silver staining of IGFBP-3 purified from porcine serum by SP Sephadex C-25 cation exchange chromatography, IGF-I affinity chromatography and electro-elution following SDS-PAGE. Lane 1, protein molecular mass standards representing 37, 43, 56 and 66kDa from the bottom to top, respectively. Lanes 2-4, 50, 200 and 800ng BSA, respectively. Lanes 5 and 6, purified IGFBP-3 after electro-elution representing 8.3 and 1.4ml serum equivalents, respectively. Lane 7, IGFBP's mixture(5ml serum equivalent) prior to electro-elution(after IGF-I affinity chromatography). Note 40-45kDa IGFBP-3 doublet bands in lanes 5 and 6 co-migrating with the 43kDa molecular mass standard.

2) Booster injection

가) pIGFBP-3	PBS	Freund's Incomplete Adjuvant	2
) Booster injection		1	56, 70,
84		8ml	
)		98	

3) IGFBP-3 RIA

가) pIGFBP-3 iodination(Lee Henricks, 1990)

- (1) 1 μ g pIGFBP-3 20 μ l 0.1M acetic acid 50 μ l 0.5M phosphate buffer(pH 7.0) 0.5nCi Na¹²⁵I (pH 10.0; 5 μ l) 가
- (2) Chloramine-T(1.2mg/ml) 10 μ l 가 20 iodination
- (3) Sodium metabisulfite(6mg/ml) 20 μ l 가 iodination
- (4) Sephadex G-50 chromatography(1.5 x 12cm column) free Na¹²⁵I (buffer: 50mM PBS, pH 7.4/0.02% BSA, specific activity: 100 200 μ Ci/ μ g)
- (5) -70. C

) Antiserum titration: 70 & 98 1: 300, 900, 2,700, 8100
[¹²⁵I]IGFBP-3 binding percentage (1:1500)

) RIA conditions : validation

- (1) (Walton Etherton, 1989)
- (가) 0.3ml total assay volume RIA buffer[50mM PBS, pH 7.4/0.5% fatty acid-free BSA/0.02% NaN₃], 1:1500 , 15,000cpm [¹²⁵I]IGFBP-3 non-antigen peptide 가
- () 4. C 16h 0.1ml 1:10 goat anti-rabbit IgG + 0.1ml 1:30 normal rabbit serum 가 1
- () 1ml ice-cold 6% PEG-8000 가 (3,000rpm/30) (aspiration) pellet γ -counting
- (2)

IGFBP-3 RIA

(3)

(가)

RIA non-specific binding(NSB) B0 5.3
± 0.3% 36.8 ± 2.8% .

Unlabeled pIGFBP-3 [125I]IGFBP-3 displacement curves
가 (parallelism) RIA가 (Fig. 2-2)
pIGFBP-3 IGF-I, hIGFBP-3 5%>

()

:
(Owens , 1999) (Fig. 2-3)

3. IGF-I RIA

가. : IGF-I RIA IGFBPs

- 1) 0.2ml + 1.3ml 1% aqueous trifluoroacetic acid(TFA) ()
- 2) C18 Sep-Pak cartridge preconditioning: CH3CN H2O aqueous 0.1% TFA
- 3) loading 3ml 0.1% aqueous TFA washing
- 4) Bound IGFs 2ml 0.1% TFA CH3CN elution
- 5) 0.2ml aliquots RIA 4. C

. IGF-I iodination

IGFBP-3 Na125I chloramine-T 200-400mCi/ug IGF-I
labeling Sephadex G-50 column free Na125I free

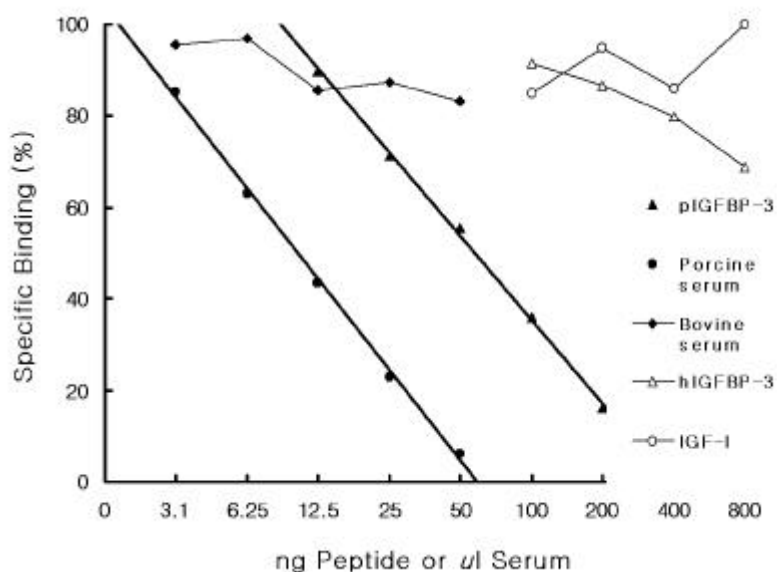


Fig. 2-2. Dose-response displacements of [125 I]IGFBP-3 from porcine IGFBP-3(pIGFBP-3) antibodies by unlabeled pIGFBP-3, a pooled porcine serum, bovine serum and other non-antigen peptides.

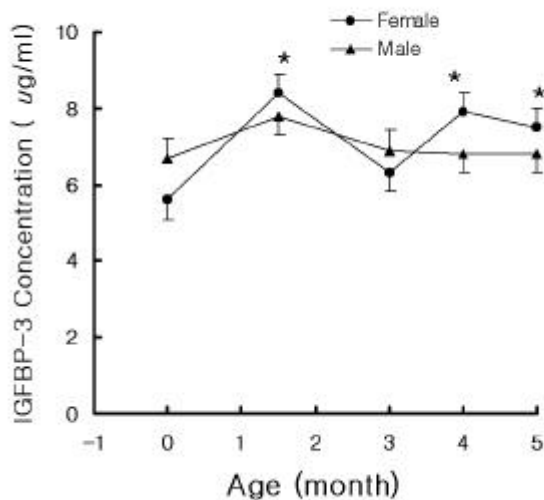


Fig. 2-3. Serum IGFBP-3 concentrations in gilts and barrows during development. At each sex \times age, mean \pm SE of eight different animals is indicated. *Different ($P < 0.05$) from the mean at birth within sex.

. RIA conditions : validation

1)

dried Sep-Pak eluate 0.1M 1:104,000
(Gropep, Adelaide, Australia), 30,000cpm [¹²⁵I]IGF-I, 가
Sep-Pak eluate RIA buffer[30mM sodium phosphate/10mM EDTA/0.2%
protamine sulfate/0.05% Tween-20/0.02% NaN₃ pH 7.5) 가
0.4ml pre-treated unlabeled IGF-I standard
dose-response [¹²⁵I]IGF-I displacement curves (parallelism)

2)

Sep-Pak chromatography
IGFBPs 2μl serum 0.1M acetic acid
Sep-Pak eluate RIA .

1)

Fig. 2-4 Unlabeled IGF-I
[¹²⁵I]IGF-I displacement curves가 (parallelism) RIA가

2)

Fig. 2-5 IGF-I
가 Lee (1991)
RIA

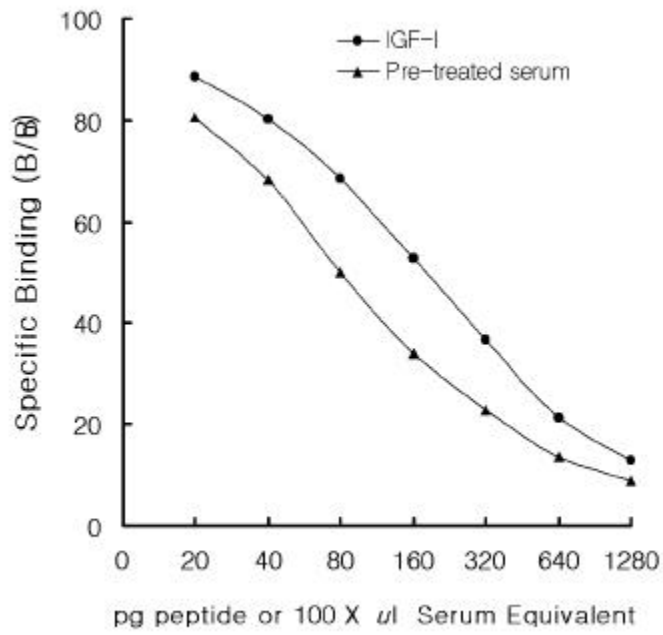


Fig. 2-4. Dose-response displacements of [125 I]IGF-I from IGF-I antibodies by unlabeled IGF-I and a pre-treated pooled porcine serum. Serum was subjected to acidic CB Sep-Pak chromatography, after which IGFBP-free Sep-Pak eluate was dried and reconstituted in 0.1M acetic acid prior to IGF-I RIA.

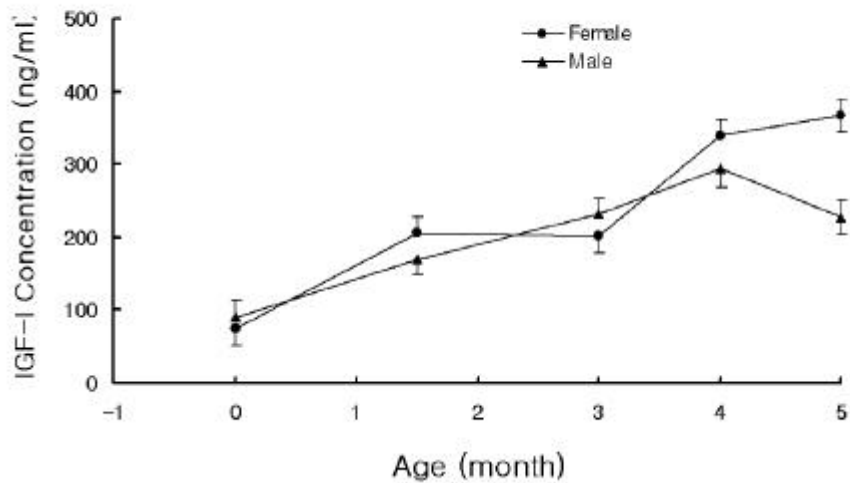


Fig. 2-5. Serum concentrations of IGF-I in gilts and barrows during postnatal development. At each sex \times age, mean \pm SE of eight different animals is indicated. Effects of sex, age and a sex \times age interaction were significant ($P < 0.05$).

4. ALS cDNA fragment cloning ALS mRNA

pALS cDNA liver RNA primer pairs
reverse transcription-polymerase chain reaction(RT-PCR)
TA-cloning , Northern blotting RNase
protection assay ALS mRNA abundance
가 .

가. RNA

Trizol total cellular RNA

, poly(A)+ RNA standard procedure(Sambrook , 1989) oligo(dT) cellulose chromatography .

. (reverse transcription; RT)

poly(A)+ RNA 100ng cDNA Cycle kit(Invitrogen)

20 μ l H₂O .

. polymerase chain reaction(PCR; Innis , 1989)

1) PCR primers

ALS cDNAs

304F 303R primer pair . ALS cDNA numbering

304F 303R primers nt104 120 390 406 . PCR

pGem 3Z plasmid vector cloning primers

Eco RI site Bam HI site .

Primer 304F: 5' AGCTGAATTCCTGGGTGGCACTGGGCC 3'

Primer 303R: 5' AGTCGGATCCGCCTGTGGCTCCAGGCT 3'

2) PCR

(liver RT products) 2 μ l, 304F & 303R primers

25pmoles High Fidelity PCR reagents(Boehringer Mannheim)

[94 \circ C 2 (94 \circ C 1 , 60 \circ C 1 , 72 \circ C 2) \times 35cycles 72 \circ C 10

4 \circ C] PCR agarose gel ALS cDNA

fragment가 .

. ALS cDNA fragment cloning & sequencing

- 1) PCR 300-bp ALS cDNA fragment Wizard DNA Purification kit agarose gel
- 2) cDNA fragment pCR 2.1 TA-cloning vector kit(Invitrogen) ligation & transformation blue/white screening
- 3) Positive colony plasmid [³⁵S]dATP Sequenase version 2.0(USB) standard dideoxy chain termination (Sanger, 1977) cDNA fragment (Figs. 2-6 & 7)

```

tggcaggcac ggagcccggg gcgccatcgg acgccgaggg cctgccgtgc 50
ccggtgcctt gctcctgagg ccacgacgac tacacggagc agctcagcgt 100
cttctgcagc tcccggaacc tcacgcagct gcccgacggc atcccagacg 150
ccgccagggc cctgtggctg gacagcaaca acttctctc cgtccccgcg 200
ggggctttcc gtaacctctc cagcctgggc ttctcaacc tgcagggcag 250
c

```

Fig. 2-6. Base sequence of a pALS cDNA fragment amplified by RT-PCR. Sequence homology to human cDNA is 80.8%.

```

AGTEPGAPSD AEGLPCPAAC SCGHDDYTDE LSVFCSSRNL TQLPDGIPDA 50
ARALWLDENN FSSVPAGAFR NLSSLGFLNL QCSG 84

```

Fig. 2-7. Deduced amino acid sequence of a pALS cDNA fragment amplified by RT-PCR. Sequence similarity to human ALS is 84.5%; identity is 69.0%.

4) Directional cloning of the ALS cDNA fragment into pGem-3Z plasmid

RNAse protection assay *in vitro* transcription riboprobe
TA-cloning vector ALS cDNA fragment
pGem-3Z plasmid vector cloning .
가) ALS cDNA fragment가 TA-cloning vector Eco RI Bam HI
agarose gel
) 300-bp ALS cDNA fragment gel Eco RI/Bam HI-cut
pGem-3Z(Pronega) plasmid vector standard procedures ligation &
transformation blue/white screening , transformed
plasmid Eco RI/Bam HI digestion agarose gel
ALS cDNA fragment가 .

. ALS mRNA abundance

ALS mRNA abundance Northern
blotting solution hybridization/RNase protection assay .

1) Northern blotting

가) Trizol total RNA
oligo(dT)-cellulose chromatography poly(A)+ RNA
) 20ng poly(A)+ RNA 2. 2M formaldehyde- 1. 5% agarose gel
) Nylon membrane overnight capillary transfer(20× SSC)
) Prehybridization: 2h/42. C, formamide-containing buffer
) Hybridization: nick-translated pALS cDNA fragment(1 x 10⁶ cpm/ml, 42
 . C/overnight)
) Washing: 2× SSC 30min/ 0.2× SSC 15min/50. C
autoradiography

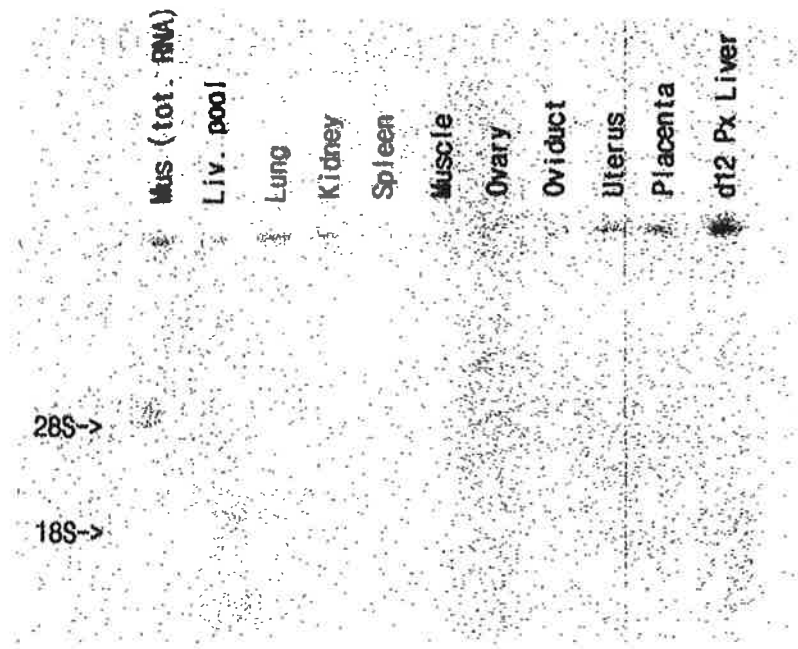


Fig. 2-8. ALS Northern blot analysis. In each lane 20 μ g total or poly(A)⁺ RNA was loaded. See text for details.

사) 결과(Fig. 2-8): 간에서만 2.2-kb ALS mRNA 가 발견되었으나 intensity가 너무 낮아 ALS mRNA abundance 측정법으로는 부적합할 것으로 판정됨

2) Solution hybridization/RNase protection assay

가) pGem-3Z plasmid에 들어있는 ALS cDNA fragment clone을 Eco RI digestion하여 linearization→agarose gel 전기영동→linearized DNA 추출

나) 0.5 μ g DNA를 50 μ Ci ³²P-UTP 존재하에서 SP6 promoter를 이용하여 37 °C에서 1시간 동안 *in vitro* transcription

-) hybridization: 1×10^5 cpm + 0.5 μ g poly(A)+ RNA/45. C/overnight
-) RNase T1 digestion(template): 37. C/30min
-) Free nucleotide : Sephadex G-50 chromatography
-) RNase (single strand RNA): 56. C/1h
-) EDTA, ammonium acetate yeast RNA 가()
-) Phenol/chloroform extraction ethanol precipitation
-) 6% polyacrylamide gel electrophoresis autoradiography
-) (Fig. 2-9)
- (1) 가 ALS mRNA
- (2) ALS mRNA
- (3) ALS mRNA abundance

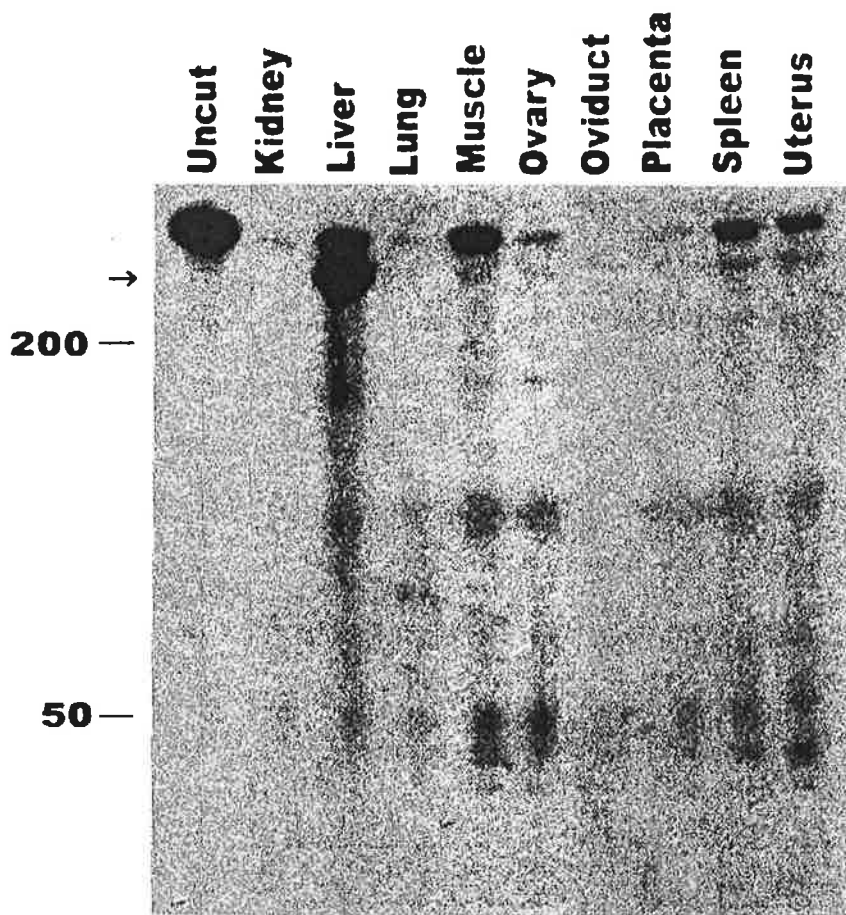


Fig. 2-9. Solution hybridization/RNase protection assay. Note the ~300-bp protected band indicated by an *arrow* on the left of the autoradiogram. See text for details.

3

1.

3

NRC

(3,400kcal DE/kg; 1988, 1998) 85%

3,400kcal DE/kg

NRC

103% 87%

2.

IGF system

IGF system

가 RIA

cDNA

IGF system components 3

IGF-I IGFBP-3

RIAs, ALS cDNA fragment cloning

3

IGFBP-3

IGFBP-3

IGFBP-3

2 3

IGFBP-3

, IGFBP-3 IGF-I RIAs

IGFBP-3

IGFBP-3

IGFs

IGFBP-3 antigen

. IGFBP-3 IGF-I RIAs

antigen standard dose-response displacement

curves parallelism IGF-I & IGFBP-3

RIAs가

ALS cDNA fragment

clone cloning DNA

가 .

IGFBP-3 ALS cDNA fragment

clone 가

, IGF-I RIA procedure

validation 가

.

3

1

, IGF system

IGF system

anabolic steroids implantation

, IGF system

.

(adrenocorticotropi c hormone; ACTH)

2

IGF-I & IGFBP-3 RIAs ALS cDNA fragment cloning
ALS mRNA abundance

2

1.

LYYD (Table 3-1) 50kg 118±7
8 2 × 2 × 2 factorial
2

Table 3-1 Experimental design of the feeding trial

Feeding	Ad libitum				Restricted			
	Control		Low Energy		Control		Low Energy	
Steroids	None	Impc	None	Imp	None	Imp	None	Imp
Number of animals	8	8	8	8	8	8	8	8

Given 80% ad libitum feed intake of the corresponding diet × steroids. For example, the fifth group from the left received 80% intake of the first.

Contains 85% digestible energy (DE) of the control diet (3509 kcal DE/kg).

Composition of the diets is shown in Table 2-1.

Implanted with Revalor (140mg trenbolone acetate + 14mg estradiol-17β).

steroids implantation

UCLAF, France) implantation .

Revalor H (Roussel

diet × steroids

3

18

19 gauge

needle

4. C

4. C 3,000rpm

30

1ml

- 20. C

50

4

105kg

real-time ultrasonic scanner (Aloka SSD-500V, Aloka Co.)

10

RNA

-70. C

2.

4. C

Chromameter(Minolta Co. CR 301)

30

Rheometer(CR 100, Japan)

chart

speed 120mm/min, maximum load 2000g,

20mm,

25mm,

adapter No. 4(13mmØ)

pH

10g

90ml

Polytron

homogenizer 14,000rpm 1

A. O. A. C. (1993)

3.

IGF-I

IGFBP-3

2

radioimmunoassays(RIAs)

IGF-I

IGFBP-3 RIAs

intra- and inter-assay coefficients of variation

15.3% & 19.3%

13.5% & 10.6%

estradiol-17β

testosterone

RIA kits(Diagnostic

Products Corporation, Los Angeles, CA)

RIAs 1 estradiol-17 β RIA
 intra-assay coefficient of variation 4.4% .
 IGFBPs (Hossenlopp , 1986; Lee ,
 1991) 1 μ l SDS-PAGE nitrocellulose membrane(0.45 μ m
 pore size) electro-transfer [¹²⁵I]IGF-II Western ligand
 blotting (semi-quantitation) . GLU-P strip
 DRI-CHEM 3000(Fuji Photo Film Co., Tokyo, Japan)

4. ALS Northern blotting & RNase protection assay

-70 $^{\circ}$ C total RNA Puissant
 Houdebine(1990) guanidinium-phenol-chloroform extraction
 (Lee , 1993) . ALS Northern blot
 analysis solution hybridization/RNase protection assay 2
 30 μ g 10 μ g total RNA .

5.

SAS(1986) general linear
 models procedure .
 (feeding), (diet) implantation(steroids) (main effects),
 3 (interactions) feeding \times diet \times steroids가
 interactions
 animal (nested within feeding \times diet \times steroids) 가
 main effects main effects animal (nested

within feeding × diet × steroids) error term

. (loin muscle area; LMA) National

Swine Improvement Federation(NC State Univ., 1995)

$$\begin{aligned}
 & 110\text{kg} \\
 & = +[(\quad - \quad) \times \quad \div \\
 & (\quad - b)]; \text{ b}=+30 \text{ for barrows}(\quad : \quad)
 \end{aligned}$$

$$\begin{aligned}
 \text{LMA} & = \text{LMA} + [(\quad - \quad) \times \quad \text{LMA} \div \\
 & (\quad +155)]; \quad \underline{\hspace{2cm}}
 \end{aligned}$$

3

1.

4

(Table 3-2). anabolic steroids
implantation .

가

steroids implantation

. , ultrasonic scanning

. A, B

가 가 ,

Table 3-2. Effects of restricted feeding, low energy diet and implantation of anabolic steroids on growth performance and carcass traits in finishing barrows

Item	<i>Ad libitum</i> feeding				Restricted feeding ¹				P<0.05
	Control diet		Low E2		Control diet		Low E2		
	None	Imp3	None	Imp3	None	Imp3	None	Imp3	
Initial wt (kg)	58.0 ±2.5	59.9 ±2.5	60.0 ±2.5	60.1 ±2.5	61.0 ±2.5	60.6 ±2.5	57.0 ±2.5	56.9 ±2.5	
Final wt (kg)	112.7 ±2.4	107.5 ±2.3	108.5 ±2.3	110.0 ±2.3	105.7 ±2.3	102.3 ±2.3	102.8 ±0.23	98.2 ±2.3	Feeding
ADG (kg)	0.96 ±0.04	0.81 ±0.04	0.86 ±0.04	0.79 ±0.04	0.78 ±0.04	0.67 ±0.04	0.74 ±0.04	0.61 ±0.04	Feeding Imp
Carcass wt(kg)	82.7 ±1.87	78.3 ±1.75	78.1 ±1.75	79.3 ±1.75	76.8 ±1.75	74.5 ±1.87	72.2 ±1.75	70.1 ±1.75	Feeding Diet
Dressing (%)	75 ±1.0	72 ±0.9	72 ±0.9	71 ±0.9	72 ±0.9	72 ±1.0	70 ±0.9	71 ±0.9	Diet
BF thickness ⁴ (mm)	22.3 ±1.58	17.5 ±1.47	20.0 ±1.47	18.0 ±1.47	22.7 ±1.47	18.4 ±1.58	15.7 ±1.47	15.3 ±1.47	Diet Imp
LMA ⁵ (cm ²)	39.9 ±1.05	38.3 ±0.99	39.3 ±0.99	40.0 ±0.99	39.0 ±0.99	37.3 ±0.99	37.2 ±0.99	37.1 ±0.99	Feeding
A, B grade ratio ⁶	6/7	·	8/8	·	6/8	·	5/8	·	not applicable
Feed/gain ⁷	3.37	3.14	4.61	3.59	3.29	3.15	4.02	3.66	not applicable

¹No two-way or three-way interactions observed

²180% *ad libitum*

³285% control diet ME

⁴Revalor(140mg trenbolone acetate + 14mg estradiol-17)

⁵Corrected for 110kg body weight

⁶Estimated by ultrasound and corrected for 110kg body weight

⁷Implanted animals excluded(misjudged as boars at the slaughter house)

⁸Mean values, all the other values: means ± SE

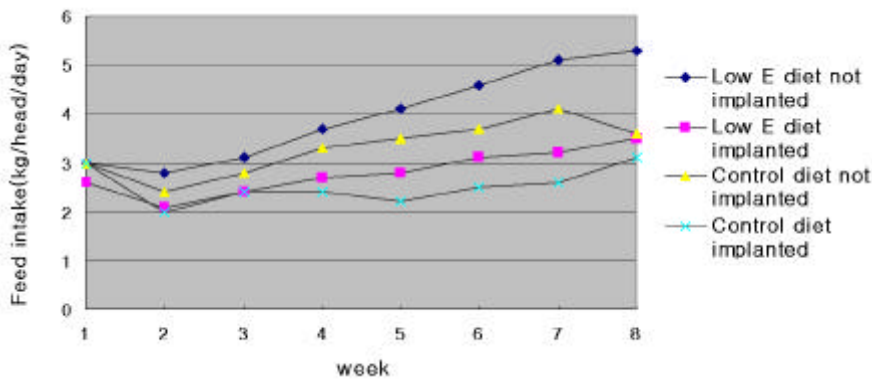


Fig. 3-1. Daily feed intake with increasing number of weeks of experiment in *ad libitum* groups

steroids implantation

Table 3-2 . 가

steroids implantation non-implanted

20 30% (Fig. 3-1).

(Table 3-3).

L*

가

a*

b*

가

steroids implantation

가 ,

pH

steroids

implantation

가

가

0.2 unit

L*

가

a*

b*

steroids implantation

. Steroids implantation

b*

Table 3-3. Effects of restricted feeding, low energy diet and implantation of anabolic steroids on physicochemical characteristics of the longissimus muscle section in finishing barrows¹

Item	Ad libitum feeding				Restricted feeding ^a				P<0.05		
	Control diet		Low Eb		Control diet		Low E				
	None	Imp ^c	None	Imp	None	Imp	None	Imp			
Back fat	Hardness (g/cm ²)	506 ± 59	633 ± 54	524 ± 59	503 ± 47	553 ± 59	546 ± 59	471 ± 54	515 ± 54		
	Color	L*	75.0 ± 0.79	75.6 ± 0.72	74.5 ± 0.79	75.2 ± 0.62	75.6 ± 0.79	74.6 ± 0.79	74.6 ± 0.72	74.1 ± 0.72	
		a*	3.82 ± 0.52	3.17 ± 0.48	3.17 ± 0.52	3.78 ± 0.41	3.35 ± 0.52	4.96 ± 0.52	4.63 ± 0.48	4.01 ± 0.48	Feeding
		b*	4.48 ± 0.46	5.03 ± 0.42	4.54 ± 0.46	4.69 ± 0.37	5.32 ± 0.46	5.77 ± 0.46	5.69 ± 0.42	5.56 ± 0.42	Feeding
Meat	Moisture (%)	72.3 ± 0.53	74.2 ± 0.49	73.3 ± 0.53	74.3 ± 0.42	74.0 ± 0.53	74.7 ± 0.53	73.7 ± 0.49	75.2 ± 0.49	Feeding Imp	
	Fat (%)	3.86 ± 0.37	3.20 ± 0.34	3.09 ± 0.37	3.04 ± 0.29	2.91 ± 0.37	2.37 ± 0.37	3.02 ± 0.34	2.09 ± 0.34	Feeding Imp	
	pH	5.48 ± 0.08	5.56 ± 0.07	5.52 ± 0.08	5.53 ± 0.06	5.57 ± 0.08	5.86 ± 0.08	5.47 ± 0.07	5.69 ± 0.07	Feeding Imp	
	Color	L*	50.4 ± 2.2	51.1 ± 2.0	46.5 ± 2.2	47.2 ± 1.7	52.0 ± 2.2	47.9 ± 2.2	52.6 ± 2.0	46.0 ± 2.0	
		a*	8.60 ± 0.84	7.26 ± 0.77	6.62 ± 0.84	6.72 ± 0.66	7.73 ± 0.84	6.58 ± 0.84	9.38 ± 0.77	6.67 ± 0.77	Imp
	b*	5.50 ± 0.59	4.89 ± 0.54	4.50 ± 0.59	4.84 ± 0.46	5.67 ± 0.59	4.01 ± 0.59	6.44 ± 0.54	4.22 ± 0.54	Imp	

¹Dara represent means ± SE.

^aFed 80% ad libitum feed intake.

^bContains 85% energy of the control diet(3509kcal DE/kg).

^cImplanted with Revalor(140mg trenbolone acetate + 14mg estradiol-17 β).

가 (P

value of feeding × steroids interaction<0.05).

2.

Table 3-4 estradiol, IGF-I, IGFBP-3

가 . testosterone 0.02ng/ml

. Estradiol steroids implantation

가 implanted non-implanted

IGF-I implantation 가(non-implanted vs implanted = 187 vs 226ng/ml, SE = 7ng/ml), (vs = 213 vs 200ng/ml, P = 0.16) (vs = 210 vs 202ng/ml, P = 0.39) . IGF-I 가 (P<0.01; Fig. 3-2).

IGFBP-3

IGF-I 가 가 (vs 21 vs 42 = 3.0 vs 3.3 vs 3.4ng/ml, SE = 0.05ng/ml; P<0.01). IGFBP-3 [¹²⁵I]IGF-II Western ligand blotting 가 가 (Fig. 3-3).

IGF-I IGFBP-3

Table 3-5 . IGF-I IGFBP-3 가 .

steroids-implanted non-implanted

Table 3-4. Effects of restricted feeding, low energy diet and implantation of anabolic steroids on serum concentrations of glucose and hormones in finishing barrows

Item	<i>Ad libitum</i> feeding				Restricted feeding ^a				P<0.05
	Control diet		Low Eb		Control diet		Low E		
	None	Impc	None	Imp	None	Imp	None	Imp	
E ₂ d (pg/ml)		47.1 ± 10.5		23.8 ± 10.5		35.0 ± 10.5		15.5 ± 10.5	Imp
IGF-Ie (ng/ml)	198 ± 15	243 ± 13	175 ± 13	224 ± 13	175 ± 13	214 ± 13	187 ± 13	209 ± 13	Imp
IGFBP-3 (ng/ml)	3.3 ± 0.15	3.4 ± 0.14	3.1 ± 0.14	3.3 ± 0.14	3.0 ± 0.14	3.3 ± 0.14	3.3 ± 0.14	3.1 ± 0.15	
Glucose (ng/dl)	91 ± 3.0	93 ± 2.7	90 ± 2.7	92 ± 2.7	88 ± 2.7	90 ± 2.7	91 ± 2.7	87 ± 2.7	

^aFed 80% *ad libitum* intake.

^bContains 85% energy of the control diet(3509kcal DE/kg).

^cImplanted with Revalor(140mg trenbolone acetate and 14mg estradiol-17 β).

^dNon-implant groups were at the detection limit(3.34pg/ml).

^eIncreased with increasing weeks of experiment(growth).

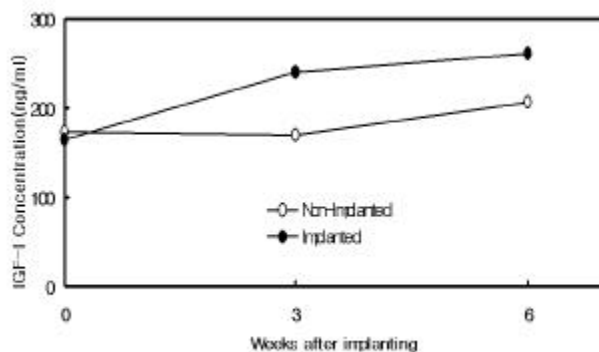


Fig. 3-2. Mean serum concentrations of IGF-I after implantation of Revalor(140mg trenbolone acetate + 14mg estradiol-17 β).

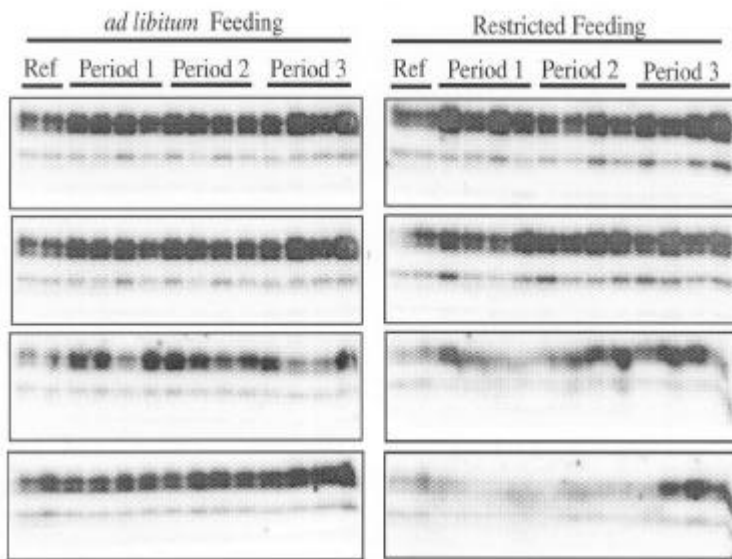


Fig. 3-3. Ligand blotting of serum IGFBPs. Four animals out of each group were randomly selected and 1 μ l serum out of each animal at each experimental period was subjected to 12.5% SDS-PAGE, electro-transfer onto nitrocellulose membrane, blotting with [125 I]IGF-II followed by autoradiography. On Ref lanes, same amounts of a pooled reference serum taken from market pigs were loaded; periods 1, 2 and 3 correspond to 0, 3 and 6 weeks after implantation of Revalor(140ng trenbolone acetate + 14ng estradiol- β). *Upper two row panels*, fed a control diet; *lower two row panels*, fed the low energy diet; *1st and 3rd row panels* from the top, non-implanted; *2nd and 4th panels*, implanted with Revalor(140ng trenbolone acetate + 14ng estradiol- β). On the blot, top two doublet bands correspond to glycosylation variants of IGFBP-3; middle and bottom dim bands are a mixture of unidentified IGFBPs and deglycosylated IGFBP-4, respectively.

(non-implanted , 21 42
: 0.17, 0.40 & 0.54, P=0.34, 0.03 & <0.01,
). IGF-I IGFBP-3
가 . steroids-implanted
IGF-I 가
non-implanted
21 IGF-I (r=0.69, P<0.01)
IGFBP-3 (r=0.50, P<0.01) (
). IGF-I 42 IGF-I
, 42 IGF-I
non-implanted (r=-0.45, P=0.01).

steroids implantation

(Table 3-2)

non-implanted
(r=0.48, P=0.16) (r=0.34, P=0.33)
가 ().

Table 3-5. Pearson correlation coefficients between serum concentrations of IGF-I and IGFBP-3 and selected carcass traits

	Hormone		Growth and Carcass traits				
	IGF- I	IGFBP- 3	ADG	Backfat	Dressing	LMAa	LM fatb
Correlation with hormone concentration at day 0 of Exp.							
IGF- I	.	0.19 (p=0.13)	0.21 (p=0.10)	0.02 (p=0.86)	-0.21 (p=0.10)	0.21 (p=0.09)	-0.02 (p=0.90)
IGFBP- 3	.	.	-0.07 (p=0.55)	0.01 (p=0.91)	-0.04 (p=0.74)	-0.09 (p=0.48)	0.22 (p=0.15)
Correlation with hormone concentration at day 21 of Exp.							
IGF- I	.	0.44 (p < 0.01)	0.16 (p=0.20)	-0.25 (p=0.05)	0.03 (p=0.83)	-0.05 (p=0.72)	0.05 (p=0.73)
IGFBP- 3	.	.	0.22 (p=0.09)	-0.04 (p=0.75)	0.07 (p=0.62)	-0.06 (p=0.63)	0.12 (p=0.48)
Correlation with hormone concentration at day 42 of Exp.							
IGF- I	.	0.57 (p < 0.01)	-0.02 (p=0.88)	-0.47 (p < 0.01)	-0.18 (p=0.17)	-0.17 (p=0.18)	-0.18 (p=0.33)
IGFBP- 3	.	.	-0.01 (p=0.93)	-0.7 (p=0.20)	0.02 (p=0.90)	-0.26 (p=0.04)	0.26 (p=0.09)
Growth and carcass traits							
ADG	.	.	.	0.24 (p=0.06)	0.23 (p=0.08)	0.33 (p < 0.01)	0.47 (p < 0.01)
Backfat	0.35 (p < 0.01)	0.31 (p=0.01)	0.33 (p < 0.02)
Dressing	0.14 (p=0.27)	0.46 (p < 0.06)
LMA	0.01 (p=0.97)
LM fat

aLongissimus muscle area at the 10th rib.

bFat content of the whole longissimus muscle.

3. Northern blotting and RNase protection assay

Fig. 3-4 & 5 ALS Northern blotting solution hybridization/RNase protection assay . ALS mRNA abundance 가 , 가 가 .

_____ <i>Ad libitum</i> Feeding _____		_____ Restricted Feeding _____	
__ Control_Diet __	__ Low_E __	__ Control_Diet __	__ Low_E __
__ None __	__ Imp __	__ None __	__ Imp __

Fig. 3-4. Northern blot analysis of hepatic ALS mRNA abundance. Thirty micrograms of total RNA were subjected to Northern blot analysis using a porcine ALS cDNA fragment as probe. Only the 2.2-kb ALS mRNA band is shown. See Section 4 of Chapter II for details. Ref, reference liver sample.

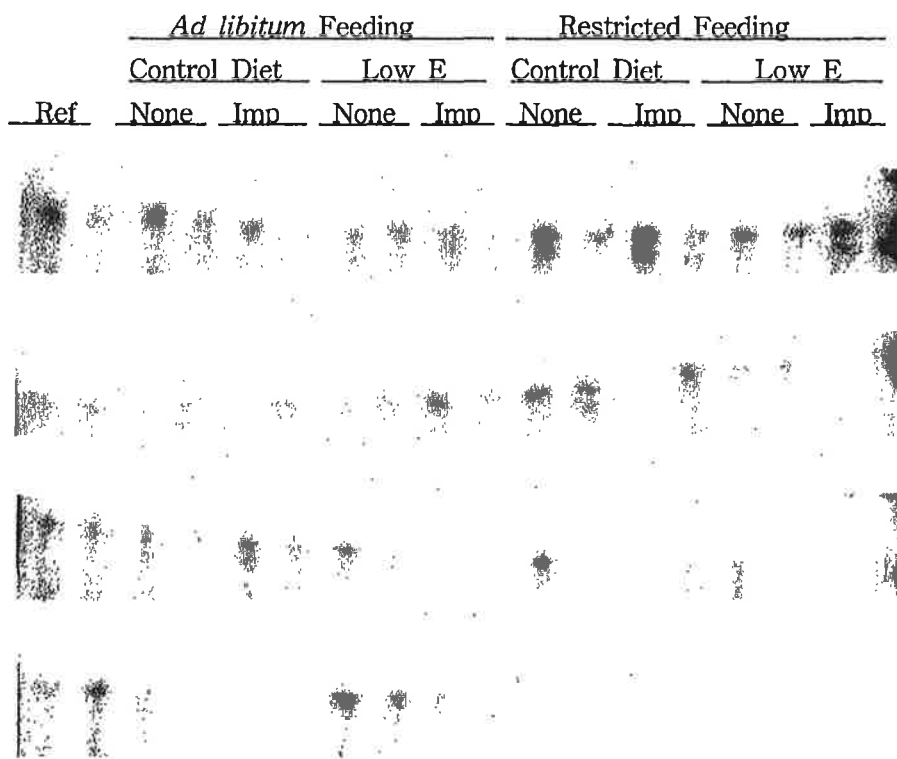


Fig. 3-5. Solution hybridization/RNase protection assay of hepatic ALS mRNA abundance. Ten micrograms of total RNA were subjected to the assay using a ³²P-labeled, 303-nucleotide(nt) porcine ALS riboprobe. Only the 303-nt protected band is shown. See Section 4 of Chapter II for details. Ref, reference liver sample.

4

1.

가.

(Leynaster Mersmann, 1991; ,
1994)

.

.

가

가

.

.

(1985)

,

.

가

가 .

. Anabolic steroids implantation

Anabolic steroids

가 (Galbraith Topps, 1981)

가

steroids implantation

androgen

trenbolone acetate estradiol

Revalor

implantation

가

Revalor implantation

가

Grandadam (1975)

De Wilde Lauwers(1984)

Revalor implantation

Revalor implantation

implantation

implantation

20

30%

steroids implantation

Revalor implantation

가

Revalor implantation

가

Revalor-implanted

가

(>90%) implanted

Revalor-implanted

Revalor implanting pellets
trenbolone

가가 (Heitzman, 1977)
androgenicity .
가
/
가
가
가
가
가 , Revalor
implantation implanting pellet
trenbolone 가 .
anabolic steroids
가 가
estrogen implantation
가 가 .
가
20mm 1998
A, B
가
가
가

가 .

2.

IGF system ,

anabolic steroids

가 가 . Non-implanted

IGF-I IGFBP-3

Owens (1999) IGF-I 가

가 . IGF-I

IGF-I

anabolism

(Gluckman , 1991; Lager , 1993; Jones Clemons, 1995).

Revalor implantation IGF-I 가

(Lee , 1990)

Revalor [trenbolone (androgen) + estrogen] anabolic

action IGF-I .

trenbolone estradiol steroid가 IGF-I

가 가 .

Revalor implantation

IGF-I 가 .

inplanted steroids

IGF-I 가 inplanted steroids

IGF-I 가 .

anabolic stroids IGF-I IGF-I

anabolic action .

IGF-I IGFBP-3

non-inplanted

IGF-I IGFBP-3

(Hall , 1999),

IGF-I IGFBP-3

가

IGF system

components 가 .

insulin system

IGF system components anabolism

5

: ACTH

1.

가

insulin glucocorticoid(cortisol)

(Eherton Kensinger, 1984). Glucocorticoids

adrenocorticotrophic hormone(ACTH)

catabolic hormone

(Henricks

, 1984).

ACTH

glucocorticoid hormone

가

,

Sillence

(1992)

ACTH

37%

가

ACTH

,

가

ACTH

(passive immunization)

가

ACTH

2.

가.

3-maleimido benzoyl chloride (MBS; Sigma)
 glutaraldehyde cross-linker keyhole limpet
 hemocyanin (KLH; Sigma) human histone carrier protein
 ACTH (Sigma) cross-linker: carrier
 cross-linking 8 2 × 2 × 2
 factorial (Table 3-6)

Table 3-6 Experimental design for the active immunization against ACTH

Cross-linker	MBS				Glutaraldehyde			
	KLH		Histone		KLH		Histone	
Carrier	None	ACTH	None	ACTH	None	ACTH	None	ACTH
Hapten	None	ACTH	None	ACTH	None	ACTH	None	ACTH
# of animals	8	8	8	8	8	8	8	8

. Hapten coupling

Hapten MBS glutaraldehyde carrier
 protein coupling .

1) MBS cross-linker

가) 70mg KLH human histone 4.4ml PBS(pH 7.4)
) Rocker 875ml MBS (25mg/4.16ml N,N-
 dimethyl-formamide(DMF; Sigma)] 가
 30
) Frit column loading
 eluate Centri con-30

-) P-6 column(Biorad; 10ml bed volume) loading 1ml
fractions 2 4(void volume)
-)) 가 suspension
-) 70ng ACTH[in 13.2ml PBS, pH 7.4) (pH 7.0)
[non-hapten coupling PBS]
-) stirring plate 3
-) (cut-off MW: 1000) 1 PBS 12
4. C 3
-) PBS(pH 7.4) 가 가 40ml 5 aliquots
- 70. C

2) Glutaraldehyde cross-linker

- 가) 70ng KLH histone 15ml PBS(pH 7.4)
-) Fume hood 70ng ACTH 가
[non-hapten coupling ACTH 가)
-) 15ml 0.2% glutaraldehyde 15ml 가 1
-) 50mM PBS(pH 7.2) 1M glycine 가
(glycine : 200mM) 1 (blocking)
-) 1) 5 aliquots - 70. C

. ACTH
8 carrier-conjugated ACTH conjugated
carrier aliquots Freund's complete
adjuvant(Signa) immunogen .

25kg LYD 64
 immunogen 2ml/
 8
 immunogen 4 immunogen
 Freund's incomplete adjuvant immunogen
 booster injection booster injection 2
 80kg real-time ultrasonic
 scanner(Aloka SSD--500V, Aloka Co.) 10

3.

Table 3-7 80kg ultrasonic
 scanner
 coupler coupler carrier
 ACTH hapten
 ACTH
 titer, cortisol

Table 3-7. Growth parameters of barrows actively immunized against ACTH with different couplers and carriers.

Item	MBS				Glutaraldehyde				P<0.05
	KLH		Histone		KLH		Histone		
	None	ACTH	None	ACTH	None	ACTH	None	ACTH	
Initial wt (kg)	28.4 ± 1.4	25.0 ± 1.6	25.4 ± 1.4	28.4 ± 1.4	26.1 ± 1.4	25.9 ± 1.4	25.1 ± 1.4	23.9 ± 1.4	
Final wt (kg)	83.0 ± 3.2	77.6 ± 3.2	79.0 ± 3.0	77.5 ± 3.0	81.3 ± 3.0	86.4 ± 3.0	81.6 ± 3.0	76.4 ± 3.2	
ADG (gm)	805 ± 41	767 ± 44	801 ± 38	733 ± 38	804 ± 38	903 ± 38	843 ± 38	785 ± 41	coupler
Backfata (mm)	10.2 ± 0.8	11.8 ± 0.8	11.4 ± 0.8	10.8 ± 0.7	9.5 ± 0.7	10.1 ± 0.7	11.6 ± 0.7	10.3 ± 0.8	
LMA _{c,b} (cm ²)	29.9 ± 1.3	30.9 ± 1.3	34.4 ± 1.3	31.2 ± 1.2	31.4 ± 1.2	31.9 ± 1.2	35.8 ± 1.2	34.8 ± 1.3	coupler carrier

^aCorrected for 80kg body weight.

^bLoin muscle area corrected for 80kg body weight.

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3. Revalor(trenbolone + estradiol pellets) implantation

가.

. Trenbolone

(estrogen

implantation

가 가

)

4. IGF system

가. IGFBP-3

1) IGFBP-3

RIA

2)

IGFBP-3

. ALS cDNA fragment

1) 300-bp ALS cDNA fragment cloning & sequencing

2) cDNA clone

. , anabolic steroids IGF system

1) IGF-I: Revalor implantation 가

&

2) IGFBP-3 :

, Revalor implantation

3) ALS mRNA :

가 가

4) IGF-I IGFBP-3

가

5. ACTH

1) ACTH 가

2) . ()

2 ()

1. . 2000. Insulin-like growth factor system ,

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2. , . 2000.

insulin-like growth factor(IGF-I) IGF-binding protein-3(IGFBP-3)

: IGFBP-3 , IGFBP-3 IGF-I RIAs .

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