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최 종  
연구보고서

극호염성 및 호산성 유산균의 탐색을 통한  
배추의 신가공 기술 개발

New process for Chinese cabbage fermentation by screening  
of halophilic and acid tolerant lactobacilli

연구기관  
생명공학연구소

농 립 부

“ 가 ”

2000. 12. .

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:  
(1) :  
(1) :  
(2) :  
(2) :  
(3) :  
(3) :

I.

가

II.

,

.

*Bifidobacterium*

*Lactobacillus, Streptococcus,*

*Leuconostoc*

100-1000

,

가

.

*pediococcus, Lactobacillus,*

*Lactococcus*

*Leuconostoc*

,

.

가

가

.

Sauerkraut, Pickles

가

가

가

1 가

가가

가

,

.

III.

,

가

- 1.
- 2.
- 3.
- 4.

IV.

- 1.

: 가

FDA 가 Lactobacillus

, 가, 200 4,000

, 1

, 6 6 Streptococcus sp. 72 , Lactobacillus sp.

98 , Bifidobacterium sp. 84 가 1

, api kit , GC

, Lactobacillus sp. HL-48 ,

KCTC 8808P(1997.6.12)

, Bifidobacterium longum Bifidobacterium longum

BO-11

: 12%

가 , 5%

가 가

가 가 ,

: Lactobacillus sp. HL-48 Tryptone

0.1%, sucrose 1% , Fermenter 1vvm,  
200rpm 15 7.0×10<sup>7</sup>/ ml . *Bifidobacterium longum* BO-11  
skim milk 8%, yeast extract 0.3%, soytone 0.3%, dextrose 3%, L-cystein  
Hcl 0.05%, MgSO<sub>4</sub>·H<sub>2</sub>O 0.01%, Tween 80 0.1% , 18 9.7X 10<sup>9</sup>  
7가 , 0.01M phosphate buffer  
100ml lactose 5g, peptone 1g, MSG 0.1% 93.1% .  
가

2. 가

가가

가

3.

가 가

, PET-JDSF

가

20

## SUMMARY

Kimchi is a traditional fermented food necessary to our food life and important as a food which affects the bacterial growth in intestine. But, this have disadvantage, short storage period, which is caused by over-ripeness as a result from fermentation of various microorganisms. To improve this, the halophilic and acid-resistant microorganisms were isolated from human intestine with screening , characterized and used for investigation of Kimchi-fermentation technology as a starter. We conducted the development of lactic acid-salt and fermented Kimchi with Chinese cabbage and seasoning. We obtained following results;

Isolates were identified and named as halophilic *Lactobacillus* sp. HL-48 and *Bifidobacterium longum* BO-11.

1. HL-48 grew in 12 % NaCl and BO-11 was acid and bile resistant.
2. We developed the new method for Kimchi manufacture, which is to make Kimchi fit for individual favor immediately using salted Chinese cabbage and seasoning stored separately.

3. We developed the lactic acid-salt improving sanitation of Kimchi with prevention of contaminants, numerous and various microorganisms in initial step of fermentation.

4. PEP-JDSF was the most effective package material for preservation of Kimchi.

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가.	.....	29
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2.	.....	39
3.	.....	39
3	.....	43
1 .	.....	43

1.	.....	43
가.	.....	43
.	.....	43
.	.....	44
.	.....	52
2.	Bifidobacterium sp. ....	57
가.	.....	57
.	.....	59
2	가 .....	71
1.	.....	71
2.	.....	75
가.	pH .....	75
.	.....	78
.	.....	80
.	.....	84
3.	.....	86
4.	.....	90
가.	.....	90
.	가.....	92
5.	.....	112
.	.....	132
.	.....	139
3	.....	146
1.	.....	146
2. 가	가.....	158

가. . . . . 158  
 . . . . . 160  
4 . . . . . 162

# 1

가

가

1960 , 1970 , KIST,

*Leuconostoc*, *Lactobacillus* *Streptococcus*

1980

가 , 1988

(1995),

(1990),

(1991),

(1992),

(1995)

(1995)

(1995)

*pediococcus*, *Lactobacillus*, *Lactococcus* *Leuconostoc*

150

가 1,500

70%

1 가 가가

가

가

, 20% 가

600

가

,

가가

가

가

가

가(2-4),  
가(5-7), 가(8-9), 가(10), , 가(11),  
가(12)  
, 가 가 530  
, 가 2,400,000 M/ T  
3,700,000 M/ T 가 3 ,  
17 가 가 가  
가 , 가  
Sauerkraut, Pickles 가 ,  
, 가 가  
. 가 가  
(1). (1)  
가 .  
, 가  
가 가  
가 가  
, Bifidus  
가 가 .  
가가



# 2

## 1

1.

: 가 100 50  
 50 200 Fig. 1 1  
 10% NaCl MRS(Difco) agar 37 48 , colony  
 CaCO<sub>3</sub>가 MRS agar Teeth picking 37 48 , colony  
 CaCO<sub>3</sub>가 Ca(OH)<sub>2</sub> CaCO<sub>3</sub> → 2H<sup>+</sup> Ca(OH)<sub>2</sub> → CO<sub>2</sub>  
 CaCO<sub>3</sub>가 Ca(OH)<sub>2</sub> . 2 3  
 Gram staining Gram positive rod  
 catalase negative , L-, D- Lactic dehydrogenase L. D.  
 Lactic acid .  
 : TS, EG, BL  
 (Non-selective medium) 3 DHL, PEES, TATAC, LBS, BS, VS, NBGT,  
 ES, NN (Selective medium) 9 , 12 .  
 , , Table 1 .  
 Fig. 2. 6 6 12 ,  
 (transport medium) BHI broth , O<sub>2</sub>-free  
 CO<sub>2</sub> gas , Diluent B 10 .



0.05ml 1/ 3-1/ 4 ,  
 Spreader . TS, DHL, PEES 37°C (Aerobic culture)  
 , EG, BL, BS, ES, VS, NN, LBS Anaerobic jar Steel wool method  
 (Anaerobic culture)(37°C, 48h) .  
 , colony Morphology ,  
 , colony slide glass , Gram stain  
 . *Streptococcus, Lactobacillus, Bifidobacterium* BL (pure  
 culture) . 1000 cell morphology

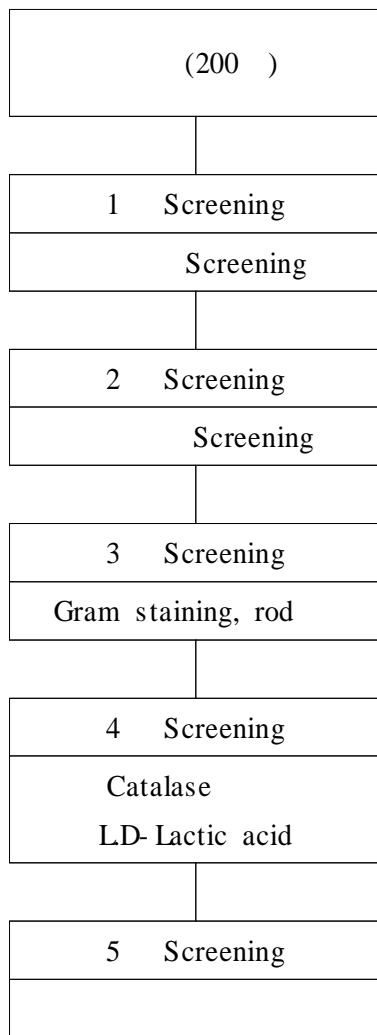


Fig. 1.

Table 1. The media and cultural method used in this study for comprehensive investigation of intestinal flora

Medium	Organisms usually enumerated	Incubation method	Incubation time(days)
Non-selective media			
EG agar	Anaerobes	Steel wool method	2
BL agar	Anaerobes	Steel wool method	2
TS agar	Aerobes	Aerobic culture	1
Selective media			
BS agar	<i>Bifidobacterium</i>	Steel wool method	2
ES agar	<i>Eubacterium</i>	Steel wool method	2
NBGT agar	Bacteroidaceae	Steel wool method	2
VS agar	Veillonella	Steel wool method	2
NN agar	<i>Clostridium</i>	Steel wool method	2
LBS agar	<i>Lactobacillus</i>	Steel wool method	2
DHL agar	Enterobacteriaceae	Aerobic culture	2
PEES agar	<i>Staphylococcus</i>	Aerobic culture	2
TATAC agar	<i>Streptococcus</i>	Aerobic culture	2

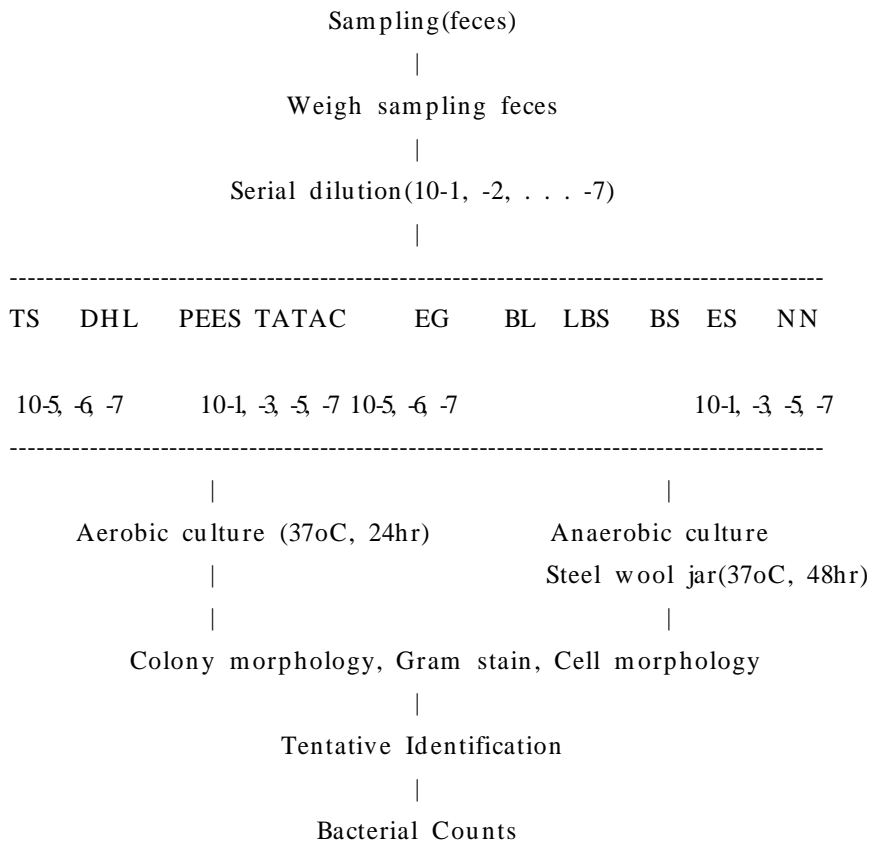


Fig. 2 Scheme for the isolation methods of the normal intestinal microflora from the feces of humans.

2.

api kit ,  
Bergy's Manual , api computer data base

3.

*Streptococcus, Lactobacillus, Bifidobacterium*

EGF broth 48 . Diluent A pH 3.0  
pH 4.0 vial 2 ml , 0.05 ml .  
37 °C 3 hr , , BL  
, , Colony  
가

*Streptococcus, Lactobacillus, Bifidobacterium* EGF broth 48  
Diluent A 0.15% oxgall(pH 7.0) 가 , vial 2  
ml , 0.05 ml . 37 °C 3 hr ,  
BL , , Colony

*Lactobacillus* sp. HL-48

*Bifidobacterium longum* BK11 , *Bifidobacterium longum* BK-11  
, , *Bifidobacterium longum*

*Lactobacillus* sp. HL-48 .

*Lactobacillus* sp. HL-48

ICR 40 (19.2 ± 1.5 g) .  
( : 23 ± 2) ( : 55 ± 3%) 가  
(GOG Environmental Control Unit, , )  
5 (420W x 260D x 190H

mm) 2 가 (

, ) 2 .

, ICR 10 ,

(kg 107 CFU) (kg 109 CFU) 10

*Lactobacillus* sp. HL-48 0.5ml 14

. ICR 10

, (kg 107 CFU), (kg 109 CFU) .

0.5 ml

AND Electronic Balance FA-6000 ( ) .

Eppendorf tube

30 5,000 rpm, 10

deep freezer (-70°C)

, Shimazu

Automatic Biochemistry Analyzer ( ) ,

AST, ALT, ALP,

BUN, ALB, CREA, GLU, TBIL, TP, ALB, CPK, CA ,

(Combur 9

Test RL) pH, , ,

. NaCl MRS

broth NaCl 5%, 10%, 15%가 가

30

. 8

positive , 8

weak positive ,

negative .

4.

*Lactobacillus* sp. HL-48

MRS carbon source c-source  
test tube 4.5ml . Glucose, Fructose, Galactose, Maltose,  
Sucrose, Lactose, Molasses, Cellobiose, Mannose, Trehalose. Xylose, Glycerol, Arabinose,  
Starch, Mannitol Carbon source 100mM , test  
tube 0.5ml 24 *Lactobacillus* sp. HL-48 50 $\mu$ l . 30  
24 OD(600nm) . Nitrogen source  
carbon source 가 Sucrose c-source nitrogen  
source n-source 100mM Beef extract, Bacto peptone,  
Casamino acid, Defated soybean, Polypeptone, Tryptone, Yeast extract, Ammonium  
sulfate, Ammonium phosphate, Ammonium chloride n-source  
. Mineral source , , 가 ,  
가 , , , , , , , , ,  
, , , m-source  
. *Bifidobacteria*  
7가 , 5가 12가

## 2 . 가

1.

15% 가 0.5 Ml 가  
30, 55 가  
, pH pH meter (Fisher Scientific AccumetR pH Meter  
915) . ,

가

12

2.

(UP)

(LP)

, UP, LP UP+LP

(5-25%)

, pH

, MRS, DHL

pH가 2.5가

가 ,

5

, ,

가 .

가

12

.

(flavor

profile method)

(acidic taste),

(salt taste),

(fresh cabbage taste)

(chewiness),

(crispness)

5가

(overall eating quality)

Table 2.



Table. 2.

: ,  
 : \_\_\_\_\_  
 : 1999

3 scoring test .

	A	B
1.		
(acidic taste)		
2.		
(fresh cabbage taste)		
3.		
(salt taste)		
4.		
(crispness)		
5.		
(chewiness)		
6.		
(yeast moldy taste)		
7. overall earing quality		
8. Comments		

3.

15%, pH 2.57†

25 4 85% 1%, 가 3.5%, 1.5%,  
2%, 0.5%, 6%, 4. 0.5%  
, ,  
25 .

4.

: Bifidobacterium

*Bifidobacterium longum* BK-11 MRS broth 9Mℓ , 37

24 , steel wool method

500g

(mixer)

1500 rpm 10

(filter paper)

4 test tube 30 Mℓ

, pH 3.0, pH 4.0, pH 5.0, pH 6.0

, membrane filter(0.45μm

size)

6Mℓ

test tube

. pH 3.0, 4.0, 5.0, 6.0

4

Bifidobacterium

(4.8×10<sup>9</sup> cfu/ MRS broth 1Mℓ

0.3Mℓ

, 1

. , 2 4 25

1 , 3 , 5 , 7

Bifidobacterium

(non-selective medium) BL

0.1ml

9.9ml diluent A

100

100, -2, -4, -6 BL

0.05Mℓ

, 37 48

steel

wool method

, colony

가 :

*Bifidobacterium longum*

가.

*Bifidobacterium longum*

*Bifidobacterium longum* 0.05% cysteine

가 MRS broth seed 37 48 (7,000rpm, 40min)

가

: (100g), (10g), (2g), (0.6g), (1.4g), (2g), 가 (1g), (2g), (1g), (1.5g), (1.5g), (0.5g), (0.5g), MSG(0.3g)

- (0.25% +15% )

15 3

:

가

106 g

가

4±1

5

4

centrifugation(7,000rpm, 1hr)

ultracentrifugation(30,000rpm, 2hr)

: 100g blender 3

- Plate count agar; - YM agar, (*Lactobacillus sp.*):  
modified LBS agar, *Leuconostoc sp.*- PES agar; - DHL agar

30

2가

1ml

Blood-BL , Anaerobic jar(BBL 社) 48

*Bifidobacterium* Fructose-6-phosphate phosphoketolase

(F6PPK) assay

: plate count agar(PCA, Difco Lab. U. S. A.; Bacto tryptone  
5g, Bacto yeast extract 2.5g, Bacto dextrose 1g, Bacto agar 15g, 1,000ml, pH 7.0)

***Lactobacillus spp.***: LBS(BBL Co. U.S.A.) m-LBS (Pancreatic digest of  
casein 10.0g, Yeast extract 5.0g, Dextrose 20.0g,  $\text{KH}_2\text{PO}_4$  6.0g, Ammonium citrate 2.0g,  
Polysorbate 80 1.0g, Sodium acetate hydrate 25.0g,  $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$  0.575g,  $\text{MnSO}_4$  0.12g,  
 $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$  0.034g, Acetic acid 2.5ml, Bacto agar 15g, 1000ml)

***Leuconostoc spp.***: Phenylethyl alcohol sucrose(PES, Bacto peptone 5.0 g , yeast extract  
0.5 g, sucrose 20.0 g,  $(\text{NH}_4)_2\text{SO}_4$  2.0 g,  $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$  0.5 g,  $\text{KH}_2\text{PO}_4$  1.0g, agar 15g,  
phenylethyl alcohol 2.5ml, 1,000ml, pH 6.8)

: YM agar(Difco Co. U.S.A.) m-YM

(modified YM agar ; Bacto yeast extract 3g, Bacto malt extract 3g, Bacto peptone 5g,  
Glucose 20g, Bacto agar 15g, Chloramphenical 100mg, 1,000ml)

***Bifidobacterium*** :

1)

*Bifidobacterium* BL(glucose-blood-liver) (Bacto beef extract 2.4g,

Bacto proteose peptone No.3 10g, Bacto tryptone 5g, Bacto soytone 3g, Bacto yeast extract 5g, Liver extract 3.2g, Dextrose 5g, Lactose 5g, Soluble starch 0.5g, Potassium phosphate monobasic 1g, Potassium phosphate dibasic 1g, Magnesium sulfate 0.2g, Ferrous sulfate 0.01g, Sodium chloride 0.01g, Manganese sulfate 0.007g, Antifoam A 0.2g, Polysorbate 80 1g, L-Cysteine hydrochloride 0.5g, Agar 15g, Distilled water 1,000ml)

2) F6PPK assay of *Bifidobacterium* cells

Centrifugation(12,000rpm, 5min) . pellet 0.05M phosphate buffer  
 (2 ) buffer 가 sonication NaF, K iodoacetate soln,  
 fructose-6-phosphate 가 37 30 . Hydroxylamine  
 HCl(pH6.5) 가 10 가 15% TCA, 4M HCl 가 . 5%  
 FeCl<sub>3</sub>·6H<sub>2</sub>O 가 .

**pH :** pH pH meter(Jenco Electronics LTD Model 1671)

: 4, 10,000rpm 10 DNS  
 . 20 100μℓ 200μℓ DNS (Dinitrosalicylic acid ;  
 NaOH 14g, DNS 7.5g, Potassium sodium tartrate 216g, Phenol 5.4ml, Sodium  
 pyrosulfite 5.9g, 1,000ml) 5 가 . 가  
 100μℓ 1,000μℓ 가 200μℓ 562nm  
 . glucose glucose  
 (mg/ ml)

**C:** C 4, 14,000rpm 10 DNP  
 . 200μℓ 5% metaphosphoric acid 1,800μℓ 가

2ml DCP(2,6-dichlorophenolindophenol) 1ml 가 1  
pink 가 thiourea metaphosphoric  
acid 2ml 가 pink , DNP (2,4-dinitro phenylhydrazine)  
1ml 가 60 90 ,  
85% H<sub>2</sub>SO<sub>4</sub> 5ml 가 .  
30 400μℓ 562nm .  
C L-ascorbic acid (Sigma. Co.) .  
**Acetic acid Lactic acid** : 1ml  
10ml가 10 1 vortexing .  
(12,000rpm, 1hr) 0.45 μm C18 SEP-PAK  
cartridge(Millipore Co.) . HPLC(Column : Aminex  
HPX-87H(300mm×7.8mm, Bio-Rad, Mobile phase : 4mM Sulfuric acid, Flow rate : 0.6Mℓ  
/min, Detector : UV detector at 206nm, Detector Temperature : 37) acetic  
acid lactic acid .

10

9 가 .

5.

가.

20

0%15%

, pH

가

pH

24.5

pH, , .

pH 2 , 5% 5% 가

가 가

hedonic scale 1( ) 2 , 5( )

Lactic acid pH 4.0 glucose

0.5%, potassium phosphate 0.01% 가

Lactobacillus sp. HL-48 BK-11

105/ ml

DHL (Eiken Chemicals, Tokyo)

Lactobacilli (Difco, Detroit)

37 48

가

: 가

150 g 2 , 5%

500 Mℓ 가 Lactic acid PURACR

: *Lactobacillus*

sp. HL-48

BA-8

10%

MRS

(Difco)

37

2

vial

-20

**pH**

: Lactic acid

pH 4.0

glucose 0.5%,

potassium phosphate

0.01% 가

*Lactobacillus* sp. HL-48

BA-8

105

:

DHL

(Eiken Chemicals, Tokyo)

Lactobacilli

(Difco, Detroit)

37

48

가

:

2

hedonic scale

10

1( )

5( )

, ,

:

20

가

pH 4

*Lactobacillus* sp. HL-48

BA-8

hedonic scale

5

2

20

B.O.D INCUBATOR(VISION SCIENTIFIC

VS-1203P5N)

pH



0%15%

, pH 가 pH 24.5

pH 2 , 5%

, 5% 가 . 1.5

5%, pH 2 1 ,

가 . pH, ,

2

, warring blander 2

가

pH, ,

Colorimeter (SPECTRO COLORIMETER JS555,

TOKYO JAPAN)

:

가

가

가 , ,

가

. Table 3

(S1)

가 (S2),

가 (S3),

가 (S4),

Lactobacillus

가 (S5)

가

pH, ,

. 100g

25

1M $\varnothing$  3 9M $\varnothing$  pH ,  
(TS medium) (MRS medium),  
(DHL medium) pouring culture method

HPLC , Table 4

solid phase extraction cartridge 0.2  $\mu$ m pore disc-type  
filter

가 : 가 2% 10% 2%, 1%, 1%  
, lactic acid 0, 0.4, 0.6, 0.8% 가  
가 20 pH,  
100g  
20

ml 3 9 ml pH ,  
(MRS medium) pouring culture method

Spectro colorimeter(color JS555, Japan)

Table 3. Composition of Kimchi-seasoning

<b>Materials</b>	<b>S* 1</b>	<b>S2</b>	<b>S3</b>	<b>S4</b>	<b>S5</b>
Radish	10**	10	10	10	10
Red pepper powder	2	2	2	2	2
Garlic	2	2	2	2	2
Ginger	1	1	1	1	1
NaCl	-	1	-	-	-
S. shrimp***	-	-	1	-	-
sugar	-	-	-	1	-
Starter	-	-	-	-	0.5

\* : Seasoning

\*\* : Ratio (%)

\*\*\* :

Table 4. Conditions of liquid chromatography for organic acid analysis.

---

---

Instrument	Waters 501 HPLC Pump
	Waters 486 HPLC Detector
Column	Supelco TM C-610 column
	30 cm × 7.8 mm ID
Mobile phase	0.1% phosphoric acid
Flow rate	0.5 mL/min.
Detector	Ultraviolet 210 nm
Injection volume	20 microliter

---

### 3 .

1.

: 가 ,

10<sup>6</sup>-10<sup>8</sup>/

가 . 150g 5% 150 가

Lactic acid PURAC (Japan) .

5% .

PET+JDSF

HDPE(high

density polyethylene)

LDPE(low density polyethylene

: , *Lactobacillus* sp.

HL-48 . *Lactobacillus* sp. HL-48 glucose Gram

10% MRS

(Difco) 30 2 . vial -20

**pH** : Lactic acid pH 4.0

potassium phosphate (K<sub>2</sub>HPO<sub>4</sub> : ATP ) 0.01% 가 .

*Lactobacillus* sp. HL-48 (hemocytometer)

10<sup>6</sup>-10<sup>8</sup> . pH 4.0

pH가 4.0 가 가

:

Table 5 MRS 30

, (E. coli) DHL (Eiken Chemical Tokyo)

37 . 0.85% 10-1,

10-2, 10-3, 10-4, 10-5, 10-6, 10-7 .

:

가

20 .

20 3

PET+JDSF

HDPE (high density polyethylene)

LDPE (low density polyethylene

20 . 10

, pH, , ( , , ) . 가 3

가

:

(SEKISUI SS-31A, japan)

,

homogenizer .

가 TAXTei texture analyzer (MHK trading co,

England) (hardness) .

Table 5. MRS            DHL

M R S		D H L	
Bacto Proteose Peptone No.3	10g	Bacto Beef Extract	3g
Bacto Beef Extract	10g	Peptone	20g
Bacto Yeast Extrat	5g	Lactose	10g
Tween80	1g	Glucose	10g
Ammonium Citrate	2g	Bile Salt(No.2)	1g
Sodium Acetate	5g	Sodium thiosulfate	2.2g
Magnesium Sulfate	0.1g	Feric ammonium citrate	1g
Manganese Sulfate	0.05g	Sodium citrate	1g
Dipotassium Phosphate	2g	Neutral red (pH indicator)	0.03g
agar	1.5g	agar	15g

2.

(Table 8)

가 가 .  
 Table 8 가 ,  
 , 가 , ,

Table 8

가 (flavor), (taste), (texture), (overall  
 acceptability) 4가 , 5 (scoring test) (Table 6).

3.

가

(Table 8)

가 가 .  
 Table 8 가 ,

, , 가 , ,

Table 8

. 가 (flavor), (taste), (texture),  
 (overall acceptability) 4가 , 5 (scoring test) (Table  
 6).



Table 6.

: 男, 女 ( ):

: 2000

:

( , , , ) 가 .

- 1
- 2
- 3
- 4
- 5

가	a	b	c	d	e	f	g	h
(flavor)								
(taste)								
(Texture)								
(Overall acceptability)								
Comments :								

Table 7.

: , : _____ : 1999 12 1				
5 scoring test .				
가 (1 )				
(2 )				
(3 )				
(4 )				
(5 )				
	365	375	385	395
1.				
(acidic taste)				
2.				
(fresh cabbage taste)				
3.				
(salt taste)				
4.				
(crispness)				
5.				
(chewiness)				
6.				
(yeast moldy taste)				
7.				
(overall eating quality)				
8. Comments				

Table 8. (total 16가 )

(Chinese cabbage)	1.	A. 15%	3	
		B. 15%	3	
(Chinese cabbage)	2.	A. 5%	5	
		B. 5%	, 0.25%	가
(Seasoning)	1.	A. 가		
		B. 0.4%	가	
(Seasoning)	2.	A.	10	
		B. 0.4%	가	10
Sample No.				
+	1	1,A-	1,A	
	2	1,A-	1,B	
	3	1,B-	1,A	
	4	1,B-	1,B	
+	5	2,A-	1,A	
	6	2,A-	1,B	
	7	2,B-	1,A	
	8	2,B-	1,B	
+	9	1,A-	2,A	
	10	1,A-	2,B	
	11	1,B-	2,A	
	12	1,B-	2,B	
+	13	2,A-	2,A	
	14	2,A-	2,B	
	15	2,B-	2,A	
	16	2,B-	2,B	

### 3

1 .

1.

가.

200	10% NaCl	MRS	colony	4,000
		2 5% NaCl	0.4% CaCO <sub>3</sub> 가	MRS
500		3	4 catalase	
Table 9		. 10% NaCl	MRS	
Coccus		yeast		, 가 rod
catalase negative	4000			.

4		2	API kit
	Table 10		HL-48 arabinose.

ribose, galactose, glucose, fructose, mannose, N-acetyl glucosamin, melibiose, trehalose

Lactic acid bacteria *Lactobacillus fermentum*, *Leuconostoc lactis*, *Leuconostoc mesenteroides*, *Lactobacillus brevis*

identification %가 35% *Lactobacillus fermentum* *Leuconostoc lactis*

, HL-48 lactic acid D. L. mixture  
*Lactobacillus fermentum*, identification percent가 Table 11

HL-48

*Lactobacillus fermentum*

identification

percent 가 38.5%

HL-48 *Lactobacillus fermentum*

가 가 *Lactobacillus* sp. HL-48

NaCl :

Table 12

12%

가

, 5%

*Lactobacillus* sp.

5%

가 , , , , ,

가

:

*Pseudomonase aeruginosa* positive, negative control

*Lactobacillus* sp. HL-48

가

, *Pseudomonas aeruginosa* , 1 2

가

*Lactobacillus* sp. HL-48

1 가 . 3 , 1 (*Lactobacillus bulgaricus*,

), 2 (*Pseudomonas maeruginosa* ) (*Lactobacillus* sp.

HL-48, 107,109 CFU/ )

10

, 0.5

14

, *Pseudomonas aeruginosa*

. 3 , 1, 2,

(Table 13),

(Table 14)



Table 9. Distribution of microorganisms isolated from soil and Korea pickled food

Characterization Cell morphology	Acid Secretion	Gram staining	Percentage from total population
Coccus shape	+ / -	+	> 90%
Yeast	+ / -	+	< 10%
Irregular shape	-	-	≈ 0.1%
Rod shape	+ / -	+	rarely

Table 10. Carbon utilization of isolated strain HL-48

Factor	Result	Factor	Result
Glycerol	-	Esculine	-
Erythritol	-	Salicine	<b>W</b>
D-Arabinose	-	Cellobiose	-
L-Arabinose	+	Maltose	+
Ribose	+	Lactose	-
D-Xylose	-	Melibiose	+
L-Xylose	-	Saccharose	<b>W</b>
Adonitol	-	Trehalose	+
β Methyl-xyloside	-	Inuline	-
Galactose	+	Melezitose	-
D-Glucose	+	D-Raffinose	-
D-Fructose	+	Amidon	-
D-Mannose	+	Glycogene	-
L-Sorbose	-	Xylitol	-
Rhamnose	-	β Genitobiose	-
Dulcitol	-	D-Turanose	-
Inositol	-	D-Lyxose	-
Mannitol	-	D-Tagatose	-
Sorbitol	-	D-Fucose	-
α Methyl-D-mannoside	-	L-Fucose	-
α Methyl-D-glucosamin	-	D-Arabitol	-
N Acetyl glucosamin	+	Gluconate	-
Amygdaline	-	2 ceto-gluconate	<b>W</b>
Arbutine	-	5 ceto-gluconate	-

W : Green color

-, W : Redish green color



Table 11. Comparisons of taxonomic similarity of HL-48 and type strains

Type strains	identification percent(%)
<i>Lactobacillus fermentum</i>	38.5 ± 0.53
<i>Leuconostoc lactis</i>	36.2 ± 0.50
<i>Leuconostoc mensenteroides</i>	15.3 ± 0.47
<i>Lactobacillus brevis</i>	9.4 ± 0.45
<i>Lactobacillus collinoides</i>	0.2 ± 0.30

Table 12. Growth rate and resistance of *Lactobacillus* sp. HL-48 according to sodium chloride concentrations in media.

Concentration of NaCl	Growth rate	Resistance
0 %	+5	+
5 %	+8	+
10 %	+2	+
12 %	+1	+
15 %	0	+
18%	0	+

Table 13. Serum biochemistry in mice before and after 7-day treatment of *Lactobacillus* sp. HL-48.

Treatment	Group	GOT (IU/L)	GPT (IU/L)	ALP (IU/L)	BUN (mg/dL)	GLU (mg/dL)	ALB (g/dL)	PRO (g/dL)	CHO (mg/dL)	BIL (mg/dL)	CRN (mg/dL)
Before	Control	164.2 ±24.2	62.9 ±6.8	743.5 ±123.4	32.3 ±15	94.2 ±27.6	3.7 ±0.2	5.7 ±0.3	98.4 ±1.1	0.1 ±0.0	0.7 ±0.1
	Low	150.8 ±19.8	52.1 ±5.8	738.5 ±27.7	32.5 ±4.7	111.3 ±5.1	3.9 ±0.5	4.8 ±1.1	82.7 ±14.4	0.1 ±0.0	0.6 ±0.2
	High	168.8 ±25.8	61.9 ±23.0	728.5 ±13.7	33.6 ±2.8	102.2 ±20.0	3.5 ±0.3	4.5 ±0.4	84.4 ±1.3	0.0 ±0.0	0.5 ±0.2
After	Control	162.4 ±30.2	65.0 ±6.5	631.2 ±1.2	32.0 ±2.6	121.0 ±18.9	3.9 ±0.3	5.7 ±0.3	90.8 ±7.8	0.1 ±0.0	0.7 ±0.1
	Low	142.9 ±29.0	46.1 ±2.0	570.3 ±67.0	31.4 ±5.2	164.2 ±12.8	3.6 ±0.3	4.8 ±1.1	97.3 ±4.2	0.0 ±0.0	0.7 ±0.1
	High	135.5 ±36.2	65.3 ±21.8	572.9 ±72.5	41.5 ±8.5	138.2 ±16.6	3.7 ±0.4	4.5 ±0.4	87.2 ±16.7	0.1 ±0.0	0.7 ±0.1

GOT: glutamic oxaloacetic transaminase, GPT: glutamic pyruvic transaminase, ALP: alkaline phosphatase, BUN: blood urea nitrogen, GLU: glucose, ALB: albumin, CHO: cholesterol, BIL: bilirubin, PRO: protein, CRN: creatinine.

Table 14. Urinalysis in rats after 7-day exposure to *Lactobacillus* sp. HL-48.

( n=10 No. of animals )

Items +	Degree	Control		Low		High	
		Pre-exposure	1- week	Pre-exposure	1- week	Pre-exposure	1- week
pH	6.0	0	0	1	0	0	0
	6.5	0	0	0	1	1	0
	7.0	7	7	6	7	8	8
	7.5	2	1	2	2	1	1
	8.0	1	0	1	0	0	1
	8.5	0	0	0	0	0	0
Protein	negative	6	4	4	6	6	5
	30	4	6	6	4	4	5
	100	0	0	0	0	0	0
	300	0	0	0	0	0	0
Glucose	normal	8	10	10	10	10	10
	50	2	0	0	0	0	0
	100	0	0	0	0	0	0
	300	0	0	0	0	0	0
	1000	0	0	0	0	0	0
Ketone body	negative	7	6	9	8	8	9
	+	3	4	1	2	2	1
	++	0	0	0	0	0	0
	+++	0	0	0	0	0	0
Bilirubin	negative	9	10	10	10	10	9
	+	1	0	0	0	0	1
	++	0	0	0	0	0	0
	+++	0	0	0	0	0	0
Urobilinogen	normal	10	10	10	10	10	10
	10	0	0	0	0	0	0
	40	0	0	0	0	0	0
	80	0	0	0	0	0	0
Occult blood	negative	9	10	10	9	10	10
	+/-	1	0	0	1	0	0
	+	0	0	0	0	0	0
	++	0	0	0	0	0	0

protein : mg/ d urobilinogen : mg/ L, glucose : mg/ dL

extract 0.1% Fig. 3 , Yeast  
 Fig. 4 Tryptone, Fig. 5 , Sucrose 가  
 Calcium sulfate , Sucrose 10g, Tryptone 1g, Calcium  
 sulfate 0.1g, Disodium phosphate 0.05g, NaCl 5g 1 .  
 (Fermenter)  
*Lactobacillus* sp. HL-48 1%1  
 1 vvm, 200 rpm 30 . Table 15  
 15 7.0 x10<sup>9</sup>/ ml .

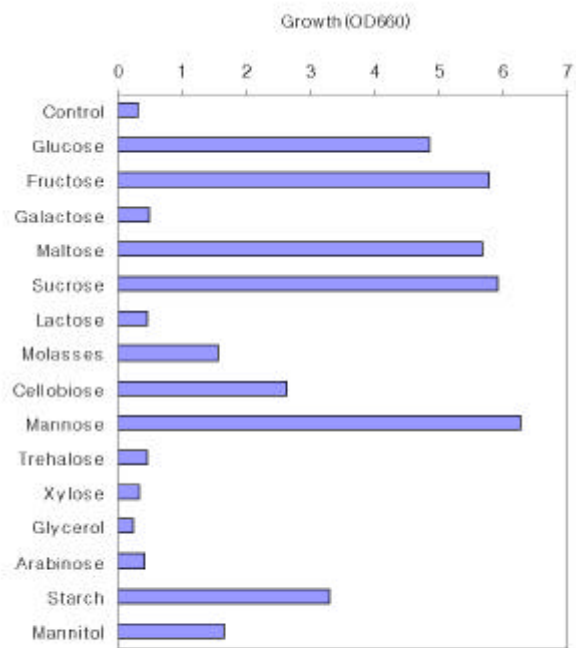


Fig. 3. Effect of carbon source on growth of *Lactobacillus* sp. HL-48

Fig. 4. Effect of nitrogen source of *Lactobacillus* sp. HL-48

Fig. 5. Effect of minerals on growth of *Lactobacillus* sp. HL-48

Table 15. Fermentation profiles of *Lactobacillus* sp. HL-48

Time(hr)	O.D(600nm)	c.f.u./ml
0	0.000	2.2x 10 <sup>7</sup>
2	0.012	2.0x 10 <sup>7</sup>
4	0.340	1.4x 10 <sup>8</sup>
6	1.147	3.0x 10 <sup>8</sup>
10	1.788	8.0x 10 <sup>8</sup>
12	1.862	2.4x 10 <sup>9</sup>
14	1.867	7.8x 10 <sup>9</sup>
48	2.020	< 10 <sup>5</sup>

2. *Bifidobacterium* sp.

가

*Bifidobacterium*

*Bifidobacterium*

*Bifidobacterium*

가

가

*Bifidobacterium*

pH,

,

*Bifidobacterium*

,

가

*Bifidobacterium*

2 : 3

L(+) lactic acid

pH

가

, *Bifidobacterium*

가

amine,

phenol, ammonia, steroid metabolites, bacterial toxin, indole, skatole

가

*Bifidobacterium*

. *Bifidobacterium*

(, ),

(, ),

. *Bifidobacterium*

가

glycopeptide

macrophage

B

가

. *Bifidobacterium*

B1, B2, B6, B12

K

$\alpha$ -galactosidase



가.

6 6 ,  
Enterobacteriaceae가 12 100% 105-108  
. *Bacillus* 58% , *Staphylococcus* 33%, *Yeast* 17%  
. *Bacteroidaceae*, *Eubacterium* 12 109-1010  
, *Clostridium* 67%, *Peptococcaceae* 42%, *Veillonella*  
33%, *Megasphaera* 25% . 12  
(total counts) 10K-1011 . *Streptococcus*, *Lactobacillus*, *Bifidobacterium*  
, (selective medium) TATAC, LBS, BS  
medium, (non-selective medium) BL, EG medium  
, *Streptococcus* 72 , *Lactobacillus* 98 , *Bifidobacterium* 84  
. *Bifidobacterium* 12 11 ( 92%), 107-101C  
. *Lactobacillus* 12 100% ,  
104-105 . *Streptococcus* 12 10  
( 83%), 105-108 .  
*Bifidobacterium* . 12  
pH 4.0 가 15 Bifidobacteria  
pH 3.0 가 BK-11 가  
가 .(Table 16)  
*Bifidobacterium* BK-11 (identification) ,  
23 (Carbohydrate fermentation) (species)  
. PYF broth 23 (Carbohydrate) 0.5% , EGF  
*Bifidobacterium* , 37°C 7 ,  
(Table 17). *Bifidobacterium* BK-11  
GC content 60.2% *Bifidobacterium longum*

. *Bifidobacterium longum*

가

Screening , *Streptococcus* 72 , *Lactobacillus* 98  
 , *Bifidobacterium* 84 EGF broth , pH 3.0  
 pH 4.0 Diluent A , 37 (C 3 hr ,  
 Table 18 *Streptococcus* 72 32 (44.5%)가,  
*Lactobacillus* 98 25 (25.5%), *Bifidobacterium* 84 15 (17.9%)가  
*Streptococcus* 32 ,  
*Lactobacillus* 25 , *Bifidobacterium* 15 Screening  
 , EGF broth , 0.15% oxgall(pH  
 7.0) 가 Diluent A , 37 (C 3 hr ,  
 Table 18 *Streptococcus* 32 12 (37.5%), *Lactobacillus* 25  
 11 (44.0%), *Bifidobacterium* 15 7 (46.7%) .

Bifidobacteria fermenter 가

107 / ml

109-IC

100-1000 , skim milk 8%, yeast  
 extract 0.3%, soytone 0.3%, dextrose 3%, L-cystein HCL 0.05%, MgSO<sub>4</sub>7H<sub>2</sub>O 0.01%,  
 Tween 80 0.1% , 18 9.7X 10<sup>9</sup> .  
 7가 , 0.01M phosphate buffer 100ml lactose 5g,  
 peptone 1g, MSG 0.1% 93.1% ,

Bifidobacteria (MRS broth 16hrs) ⇒ (fermenter, 1% , N<sub>2</sub> gas ,  
 (28%) pH 7.4 ) 37.C , 17hrs⇒ (30.C,4hrs) ⇒ 12,580g/ 10 min

⇒ 가 ( 2%) ⇒ 10min ⇒ -70.℃ ⇒  
30 hrs (-40.℃, 10 mHg ) ⇒ , 1g 3.0 X 10<sup>11</sup>

Table 16. Characteristics of Bifidobacterium strains isolated from the intestine of human s.

No.	Isolated medium	Sample No.	Cella	Gram	AEb	Identification	pHc resistance	Biled resistance
BK-1	BL	1	Y	+	-	<b>Bifidobacterium</b>	+	+
BK-2	BS	1	Y	+	-	"	++	++
BK-3	BS	2	Y	+	-	"	+	+
BK-4	"	3	Y	+	-	"	+	+
BK-5	"	3	Y	+	-	"	+	+
BK-6	BL	3	Y	+	-	"	++	++
BK-7	"	6	Y	+	-	"	+	++
BK-8	BS	7	Y	+	-	"	+	+
BK-9	"	7	Y	+	-	"	+	+
BK-10	BL	8	Y	+	-	"	+	+
<b>BK-11</b>	"	10	Y	+	-	"	+++	++
BK-12	"	10	Y	+	-	"	+	+
BK-13	BS	11	Y	+	-	"	+	+
BK-14	BL	12	Y	+	-	"	+	+
BK-15	"	12	Y	+	-	"	+	+

a Cell morphology(Y: y-shaped rods).

b Aerobic growth.

c pH 3.0 resistance.

d 0.3%Oxgall resistance.

Table. 17. Carbon utilization of isolated *Bifidobacterium* sp. BK- 11

Factors	Results	Factors	Results
Arabinose	Positive	Melibiose	Positive
Xylose	Positive	Raffinose	Positive
Ribose	Positive	Melitriotose	Positive
Glucose	Positive	Soluble starch	Negative
Mannose	Positive	Inulin	Negative
Fructose	Positive	Manitol	Negative
Galactose	Positive	Sorbitol	Negative
Sucrose	Positive	Inositol	Negative
Maltose	Positive	Esculin	Negative
Cellobiose	Negative	Salicin	Negative
Lactose	Positive	Amygdalin	Negative
Trehalose	Negative	Gluconate	Negative

Table 18. Number of resistant strains for pH 3.0 and bile(0.15% oxgall) treatment

No. isolated strains		pH 3.0	0.15% bile
Streptococcus	72	32/ 72 (44.5%)	12/ 32 (37.5%)
Lactobacillus	98	25/ 98 (25.5%)	11/ 25 (44.0%)
Bifidobacterium	84	15/ 84 (17.9%)	7/ 15 (46.7%)
Total	254	72/ 254 (28.4%)	30/ 72 (41.7%)

Bifidobacterium Trypticase-phytone-glucose medium,  
 Bifidobacterium medium, Oatmeal medium, BL(Glucose-blood-liver) medium,  
 GAM medium BL medium .  
 Bifidobacterium . Bifidobacterium  
 N2 gas 0.1 vvm flow rate .  
 Bifidobacterium (H2O2)  
 , 가 fructose-6-phosphate phosphoketolase  
 가 . BL medium 5  
 6 , , ,  
 fructose lactose가 가 . Bifidobacterium  
 growth factor 가 Bifidobacterium ,  
 가 가  
 . BL medium  
 multifactorial screening method  
 Plackett-Burman design Bifidobacterium  
 yeast extract tween 80 positive effect . 가  
 positive surface response methodology contour plotting  
 . modified BL medium Table 22  
 . Table 22 5  
 15-20 3.0×10<sup>8</sup> .

Table 19. Plackett- Burman design

Bifidobacterium

Run	A	B	C	D	E	F	G	Growth (A <sub>610</sub> )	Cell (no./ml)
1	+	+	+	-	+	-	-	3.99	3.6E+08
2	-	+	+	+	-	+	-	4.23	3.8E+08
3	-	-	+	+	+	-	+	3.22	2.9E+08
4	+	-	-	+	+	+	-	4.22	3.8E+08
5	-	+	-	-	+	+	+	3.54	3.2E+08
6	+	-	+	-	-	+	+	3.39	3.1E+08
7	+	+	-	+	-	-	+	3.05	2.8E+08
8	-	-	-	-	-	-	-	3.97	3.6E+08
Effect	- 0.08	0.06	0.01	- 0.04	0.08	0.29	- 0.80		
MS	0.01	0.00	0.00	0.00	0.01	0.17	1.29		

Factors ; A:Lab- lamco powder, B:Proteose peptone No.3 (Difco 0122), C:Trypticase p eptone (BBL 11920), D:Phytone (BBL 11906), E:Yeast extract, F:Tween 80, G:5% Cy steine



Table 20. Bifidobacterium

Carbohydrate	Growth (A <sub>610</sub> )	Cell (no./ ml)	Medium pH
Inulin	0.886	8.0E+07	5.02
Starch	1.962	1.8E+08	5.76
Dextrin	0.751	6.8E+07	5.39
Arabinose	0.809	7.3E+07	4.37
Xylose	0.940	8.5E+07	4.05
Ribose	1.339	1.2E+08	4.22
Glucose	0.955	8.6E+07	4.22
Galactose	0.629	5.7E+07	4.40
Mannose	0.553	5.0E+07	5.66
Fructose	3.225	2.9E+08	3.93
Maltose	1.280	1.2E+08	4.07
Sucrose	1.438	1.3E+08	4.27
Lactose	2.550	2.3E+08	3.73
Cellobiose	0.457	4.1E+07	5.70
Trehalose	1.484	1.3E+08	4.88
Mannitol	0.997	9.0E+07	4.16
myo- Inositol	0.457	4.1E+07	5.87
Sorbitol	1.604	1.4E+08	4.33
Arabitol	0.403	3.6E+07	5.92
Xylitol	0.509	4.6E+07	6.01

Table 21. Bifidobacterium growth factor

	<b>Growth factor</b>	<b>Concentration</b>	<b>Growth (A610)</b>	<b>Cell (no./ ml)</b>	<b>Medium pH</b>
1	Nicotinic acid	16.246mM	0.478	4.3E+07	6.01
2		1.6246mM	0.533	4.8E+07	5.92
3		97.5nM	0.575	5.2E+07	5.83
4	Pantothenic acid	100mM	-	-	-
5		20mM	0.538	4.9E+07	5.79
6		100nM	0.520	4.7E+07	5.87
7	Biotin	164nM	0.647	5.8E+07	5.83
8		16.4nM	0.590	5.3E+07	5.87
9		1.64nM	0.520	4.7E+07	5.81
10	Casamino acid	1%	0.480	4.3E+07	5.83
11		0.10%	0.714	6.5E+07	5.88
12		0.01%	0.458	4.1E+07	5.85
13	Menadione	11.62mM	-	-	-
14		116.2nM	0.529	4.8E+07	5.83
15		11.62nM	0.533	4.8E+07	5.87

Table 22. Bifidobacterium

Lab- lamco powder	2.4	g/L
Proteose peptone No.3 (Difco 0122)	10.0	g/L
Trypticase peptone (BBL 11920)	5.0	g/L
Phytone (BBL 11906)	3.0	g/L
Yeast extract	30.0	g/L
Liver extract	150.0	ml/L
Lactose	10.0	g/L
Soluble starch	0.5	g/L
K <sub>2</sub> HPO <sub>4</sub>	1.0	g/L
KH <sub>2</sub> PO <sub>4</sub>	1.0	g/L
MgSO <sub>4</sub> 7H <sub>2</sub> O	0.2	g/L
NaCl	0.01	g/L
FeSO <sub>3</sub> 7H <sub>2</sub> O	0.01	g/L
MnSO <sub>4</sub> xH <sub>2</sub> O	0.006	g/L
Tween 80	3.0	g/L
5% L- Cysteine HCl H <sub>2</sub> O	10.0	ml/L

Fig. 7. Bifidobacterium

2 가

1.

가 (幹) ,

가 .

,

( )

(乾鹽法)

(習染法)

( )

pH

가

가

30

5

55 1

5

가

pH

가

6.98

7.52,

1.84

1.78

pH

pH

pH

pH가

가

test

Table 23

Fig. 8

가

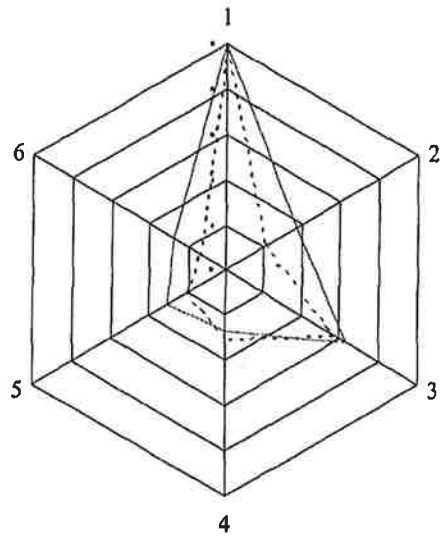


Fig. 8. A spider-web digram of difference between two salts

1, 짠맛; 2, 신맛; 3, 부드러운 맛; 4, 상쾌한 맛; 5, 신내; 6, 불쾌취

실선: salt treated lactic acid      점선: salt untreated lactic acid



2.

가.

pH

, , ,  
 $6 \times 10^5$ ,  $8 \times 10^5$ ,  $8 \times 10^6$ ,  $1.5 \times 10^6$ ,  
 $2 \times 10^5$ ,  $5 \times 10^4$ ,  
, 105-106 가  $1.5 \times 10^6$ ,  $6 \times 10^5$ ,  $5.4 \times$   
10 $\epsilon$ ,  $6.5 \times 10^6$ ,  $1 \times 10^6$  .

가

15%, 20%, 25%

4 , 30 $\text{C}$

pH

Table

24

NaCl

가

pH

가

가

5

4 ,

30 $\text{C}$

가

가

가

20%

10

4 $\text{C}$

10

Table 25

NaCl

가

가

가

가



Table 24. Effect of NaCl concentration on pH and microbial population of chinese cabbage.

NaCl concentration	pH			Microbial population (logN)		
	0 day	3 day	5 day	0 days	3 days	5 days
15%	5.7	5.0	5.0	6.30	7.20	7.98
20%	5.8	5.5	5.3	6.20	7.10	7.80
25%	5.9	5.8	5.4	6.20	6.80	7.10

\* 4°C

Table 25. Effect of preservation temperature on pH, texture, and microbial population of chinese cabbage treated 20% NaCl digestion.

Temperature	pH		texture*		Lactobacillus	
	0 days	10 days	0 days	10 days	0 days	10 days
4.C	5.87	5.51	5	4	1.92×10 <sup>7</sup>	2.65×10 <sup>8</sup>
Room Temperature	5.87	5.44	5	3	1.92×10 <sup>7</sup>	3.62×10 <sup>8</sup>

\* Texture was determined by pannel test ( 5 is highest score )



Table 26. Change of microbial populations after salt-acid treatment of Chinese cabbage.

unit : log N/ g

Items	Coliform bacteria				Total bacteria				Yeast			
	Non- treat*		Treat		Non- treat		Treat		Non- treat		Treat	
	leaf	stem	leaf	stem	leaf	stem	leaf	stem	leaf	stem	leaf	stem
0 hr	6.26	5.78	6.84	5.51	6.78	6.00	6.87	6.75	4.46	4.26	4.99	4.76
4 hr	5.60	5.45	- **	-	6.15	5.79	2.84	2.70	3.59	3.45	1.95	1.95
8 hr	5.40	5.38	-	-	5.90	5.42	2.48	2.70	3.48	3.45	1.77	1.85
12 hr	5.26	5.26	-	-	5.60	5.62	2.22	2.30	3.48	3.92	1.48	1.60
16 hr	5.08	5.00	-	-	5.40	5.38	2.00	2.00	3.14	3.09	1.25	1.60

pH ,

Fig. 9

가 가

, , 15% 가 . Fig. 10

가 . Fig. 11

2-3% 3-5 가 ,

가 .

pH가 2.5가 가 15% 20-25 3-5

가 가 가 ,

, 2-3% 가

*Lactobacillus*

sp. HL-48 *B.fidobacterium longum* BK-11

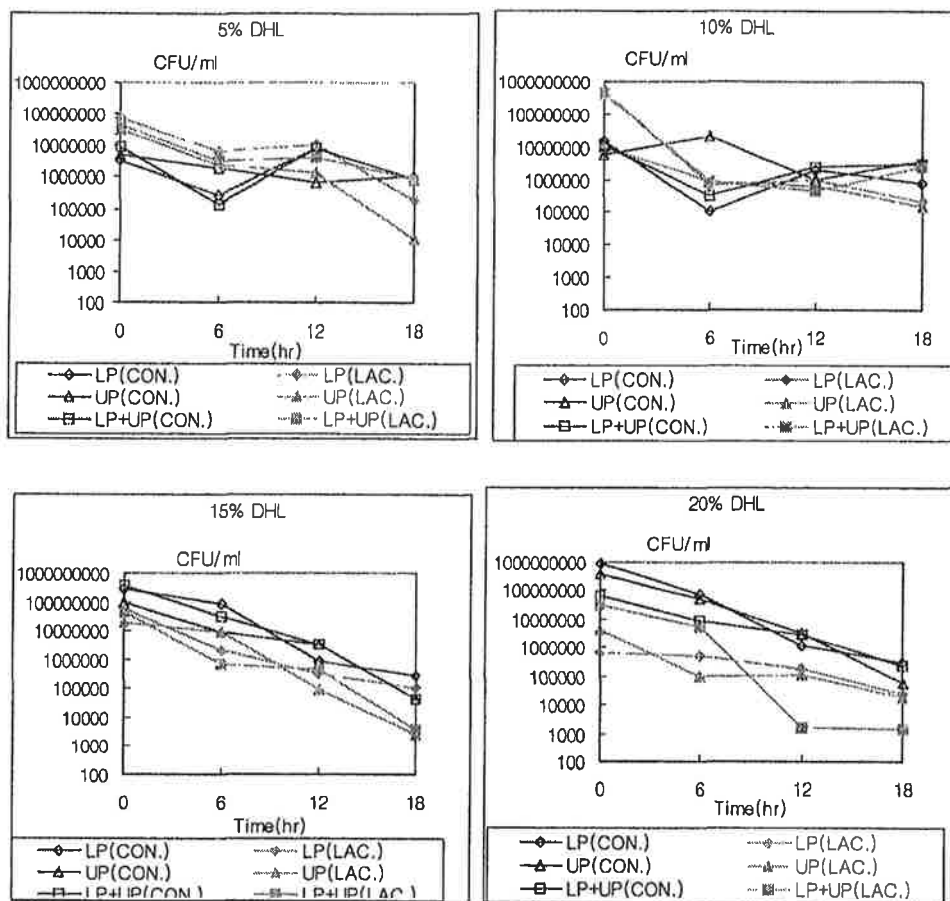


Fig. 9. Time profiles of coli-form bacteria in cabbage after treatment with lactated NaCl (LAC.) and NaCl only (CON.) according to NaCl concentration ; LP : Stem of cabbage, UP : Leaf of cabbage, LP+UP : Total cabbage

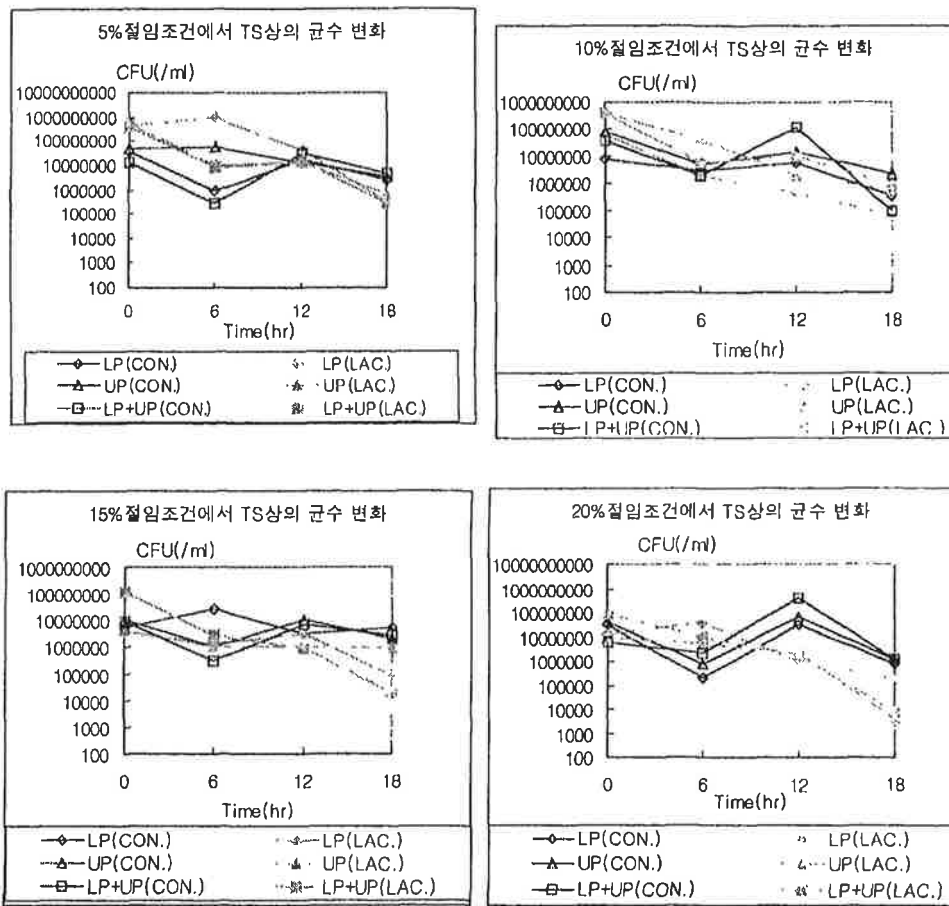


Fig. 10. Time profiles of total bacteria in cabbage after treatment with lactated NaCl (LAC.) and NaCl only (CON.) according to NaCl concentration ; LP : Stem of cabbage, UP : Leaf of cabbage, LP+UP : Total cabbage

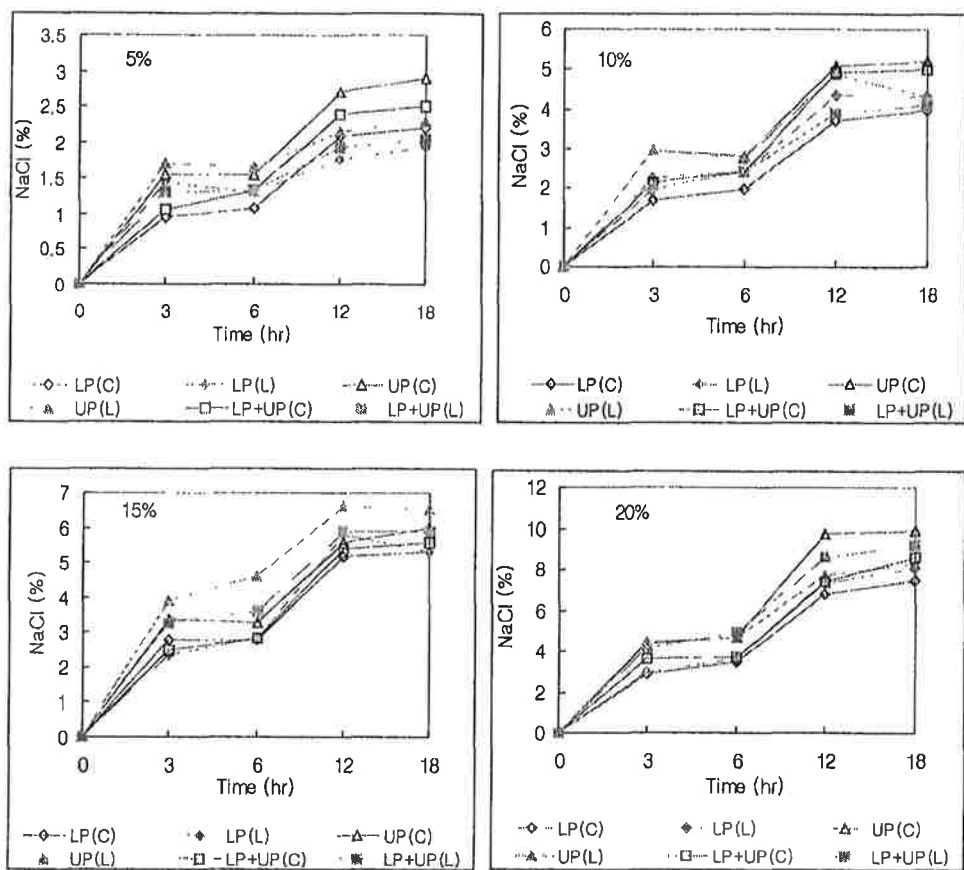


Fig. 11. Time profiles of NaCl concentration in cabbage after treatment with lactated NaCl (LAC.) and NaCl only (CON.) according to NaCl concentration ; LP : Stem of cabbage, UP : Leaf of cabbage, LP+UP : Total cabbage



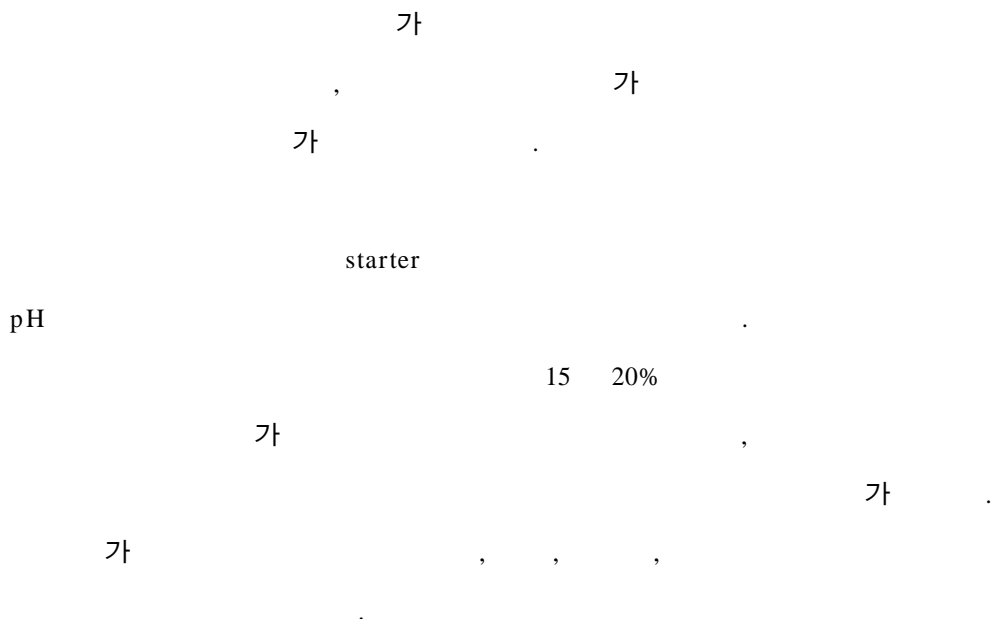


Fig. 12. 가

가 , 가

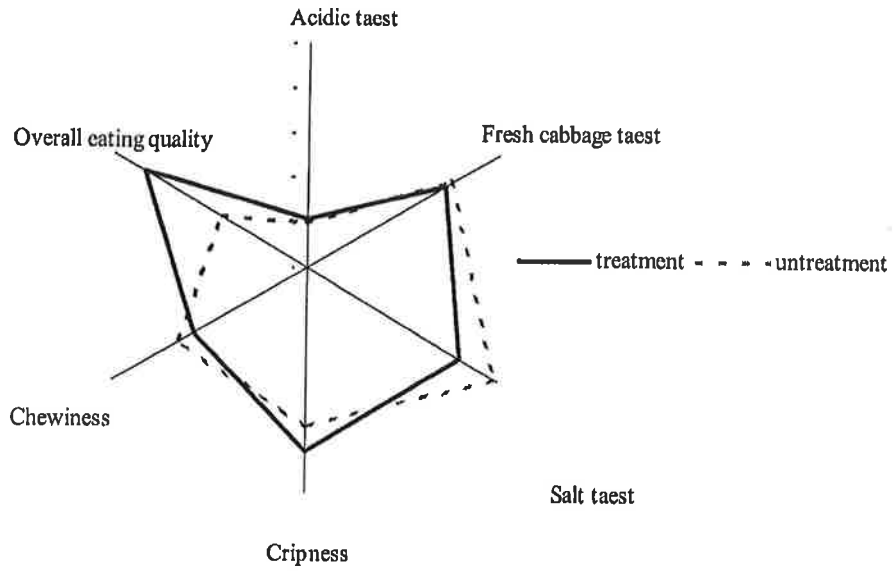


Fig. 12. Comparisons of the sensory evaluation of treated with lactic acid and untreated that after salting time on Chinese cabbage.

3.

, pH Fig. 13  
 가 pH가  
 , 가 가 가 pH 4.5 pH 3.7  
 42  
 140 ,  
 DHL , MRS

Fig. 14

, ,  
 가  
 8-10 , 가

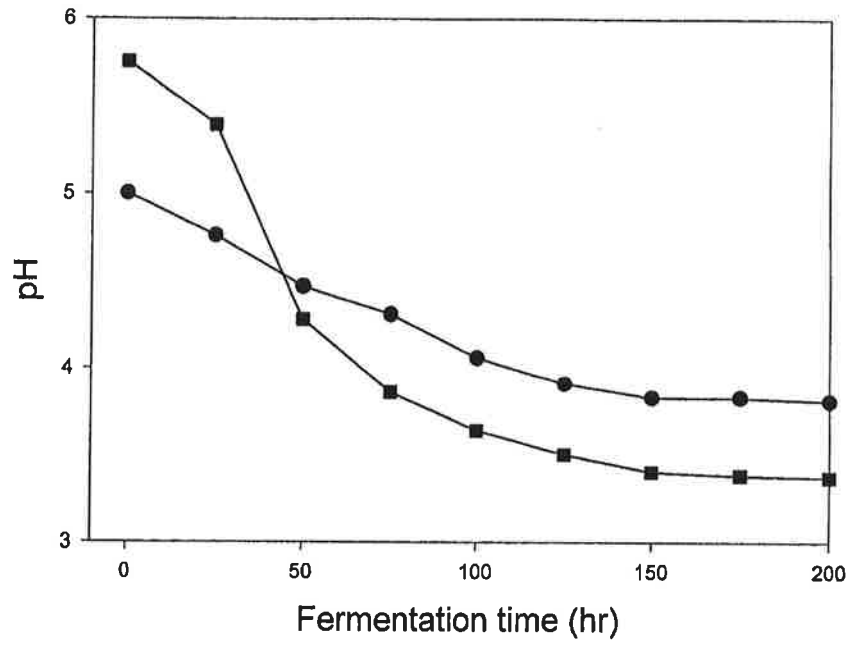


Fig. 13. Comparison of pH profile between control(■-■) and development (●-●) during Kimchi fermentation.

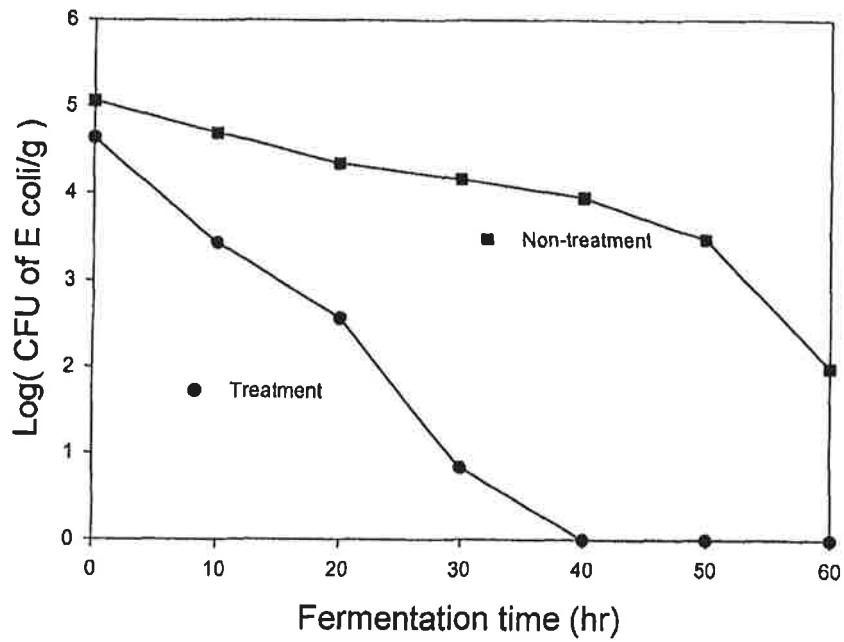


Fig. 14. Comparison of coli-form bacteria profile between control(■-■) and development (●-●) during Kimchi fermentation.

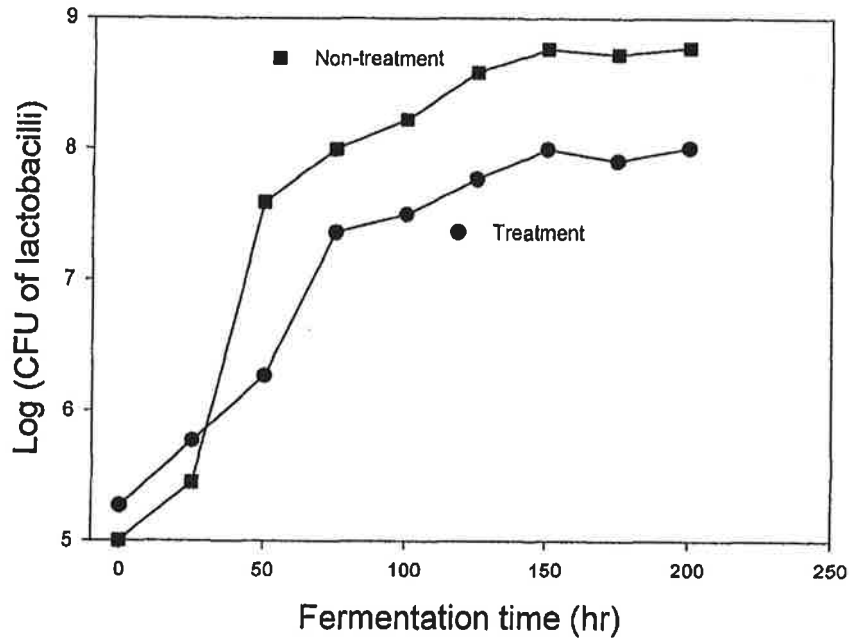


Fig. 15. Comparison of lactobacilli profile between control(■-■) and development (●) during Kimchi fermentation.

4.

가.

가 Bifidobacterium BK-11

, MRS broth pH 3.0, 4.0, 5.0, 6.0

, 4 1 , 3 , 5 , 7

. 4 , pH Log

, Table 27 . 8.4 , pH

3.0 1 5.2, 5.3 , 3

. pH 4.0 5 2.0, 2.6 , 7

. pH 5.0 pH 6.0 7 6.8-7.6

.

Table 27. Changes of Bifidobacterium counts by storage period and pH in Kimchi solution at 4°C

pH \ Days	Days									
	0	1	2	3	4	5	6	7	8	
3.0	8.41)	5.2	5.3	0	0	0	0	0	0	
4.0	8.4	6.8	6.4	4.9	5.6	2.0	2.6	0	0	
5.0	8.4	7.6	7.9	8.3	8.1	6.9	7.2	6.9	6.8	
6.0	8.4	8.0	8.0	8.3	8.6	7.9	7.3	7.6	7.6	

1) Log<sub>10</sub> counts of Bifidobacterium / ml of Kimchi solution.



가

1)

*Bifidobacterium longum*

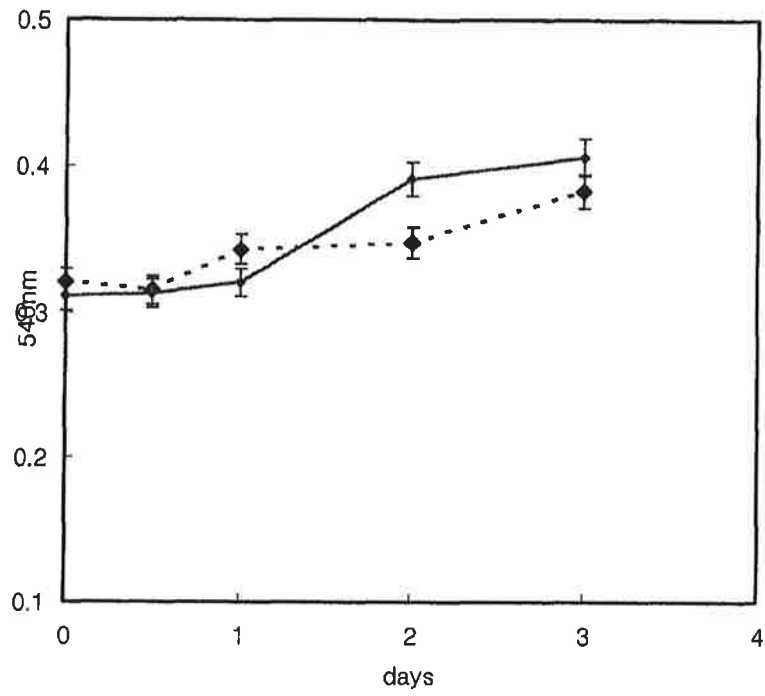


Fig. 16. Viability of *Bifidobacterium* in kimchi broth.

—◆— the growth of *Bifidobacterium* at 37°C  
 ...◇... the growth of *Bifidobacterium* at 15°C

2)

*Bifidobacterium* 가 ( 가 ) 가

. 101 cfu/ g 가 가 , 20

3 × 10<sup>10</sup> cfu/ g 가 .

*Lactobacillus* , 가 가 4×10<sup>3</sup> cfu/ g ,

15 . 가 가

가 *Bifidobacterium*

*Lactobacillus* .

*Leuconostoc* ,

가 가 가 .

, .

, 가 , 가

가 . *Bifidobacterium*

*Bifidobacterium* BL-blood F6PPK .

F6PPK *Bifidobacterium*

*Bifidobacterium* .

*Bifidobacterium* *Bifidobacterium* .

*Bifidobacterium* ,

(ex. cysteine)

pH 4.9 , *Lactobacillus*

pH 4.0 .

가

*Bifidobacterium* pH 4.0 가 .  
*Bifidobacterium* cot  
 , *Bifidobacterium* 105 cfu/ g 가 .  
*Bifidobacterium* 106 cfu/ g 가 . *Bifidobacterium*  
 가가 ,  
*Bifidobacterium* 가 , pH 가  
 . *Bifidobacterium*  
 가 가 .

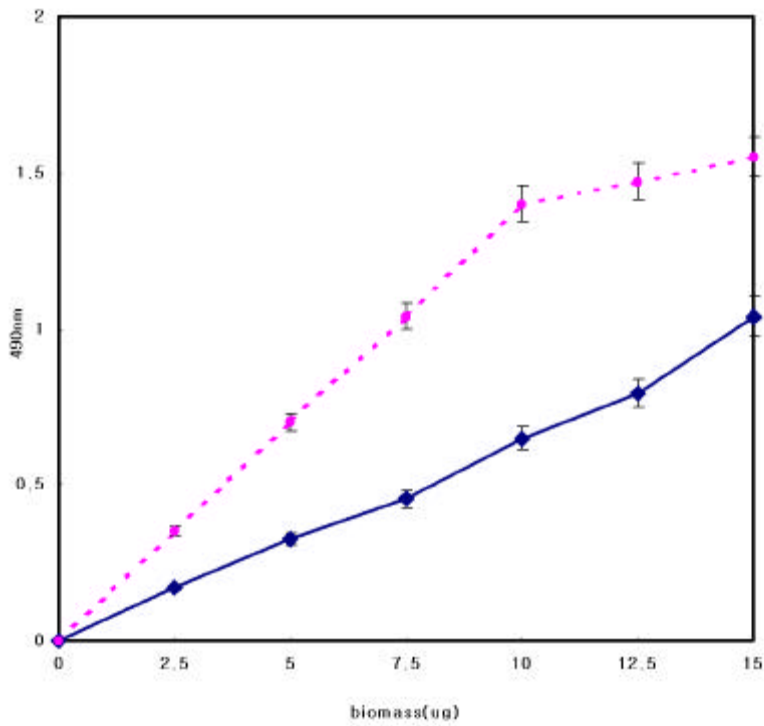


Fig. 17. A Standard curve for *B. longum* counts using Fructose-6-phosphate phosphoketolase.

- ... : The Fructose-6-phosphate phosphoketolase of *B. longum* in kimchi juice
- ◆— : The Fructose-6-phosphate phosphoketolase of *B. longum*

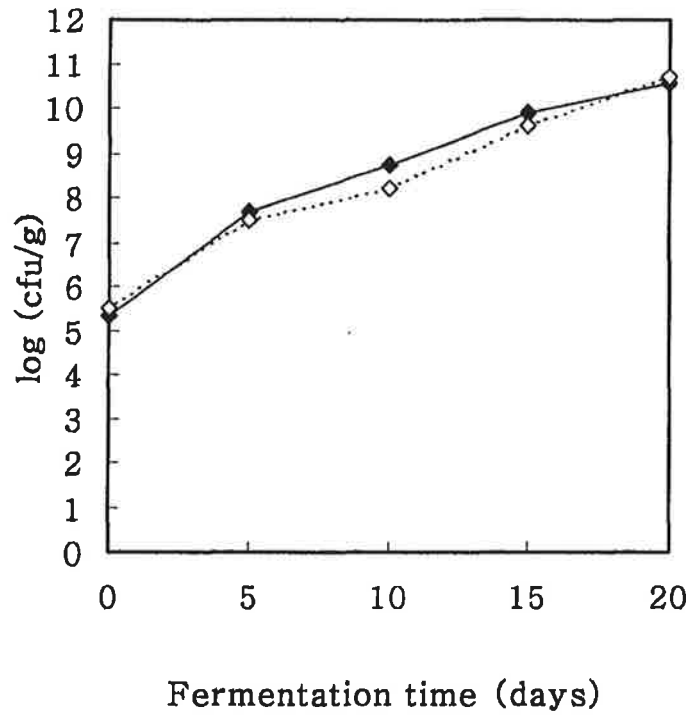


Fig. 18. Changes of total aerobes in kimchi during fermentation at 4°C

—◆— kimchi with Bifidobacterium  
 ...◇... kimchi without Bifidobacterium

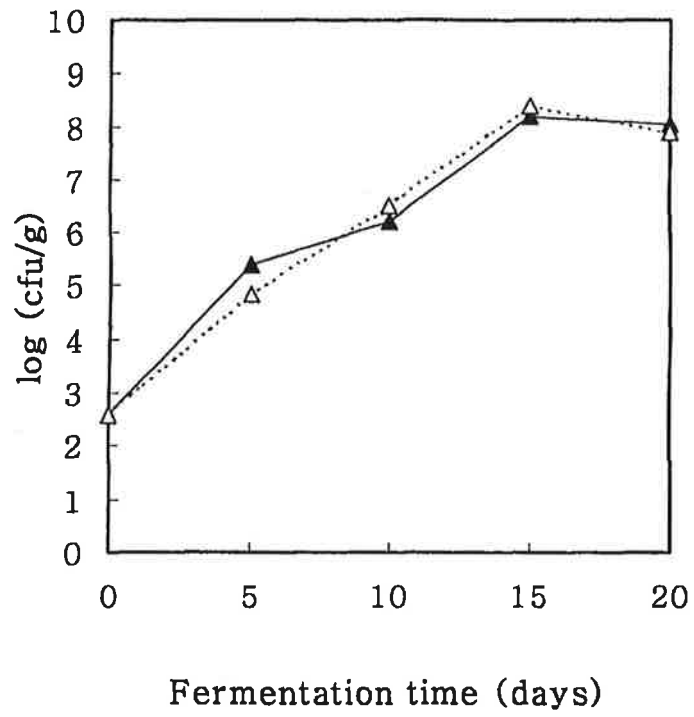


Fig. 19. Changes of *Lactobacillus* spp. in kimchi during fermentation at 4°C  
 —▲— kimchi with *Bifidobacterium*  
 ...△... kimchi without *Bifidobacterium*

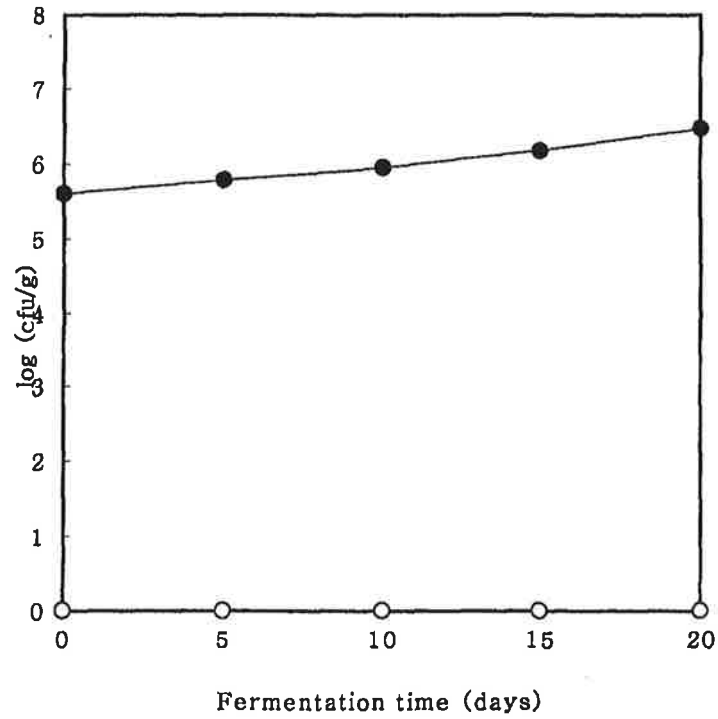


Fig. 20. Changes of Bifidobacterium in kimchi during fermentation at 4°C

—●— kimchi with Bifidobacterium  
 ...○... kimchi without Bifidobacterium



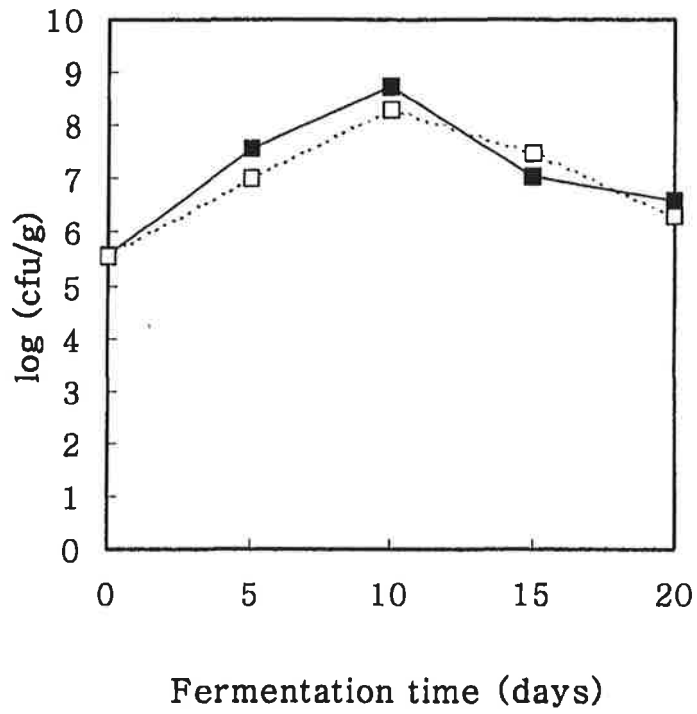


Fig. 21. Changes of *Leuconostoc* spp. in kimchi during fermentation at 4°C  
 —■— kimchi with *Bifidobacterium*  
 ...□... kimchi without *Bifidobacterium*

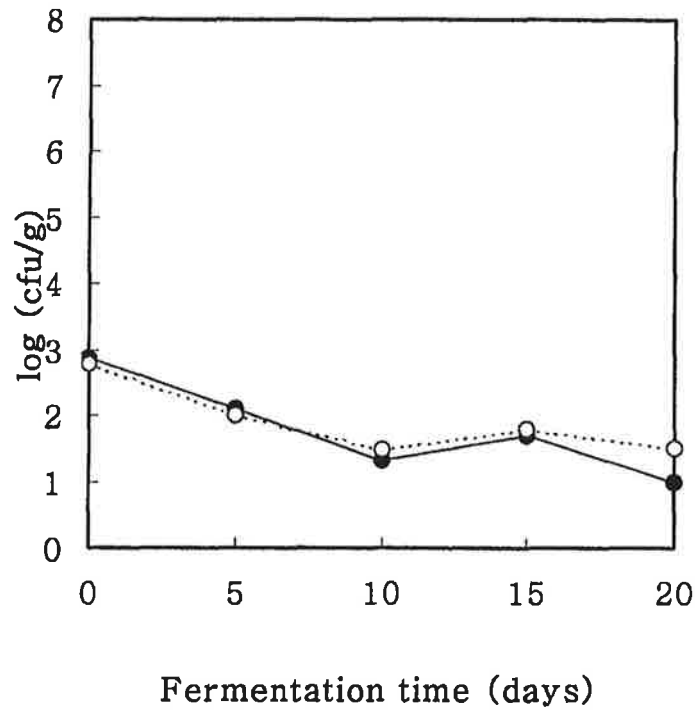


Fig. 22. Changes of yeasts in kimchi during fermentation at 4°C  
 —●— kimchi with Bifidobacterium  
 ...○... kimchi without Bifidobacterium

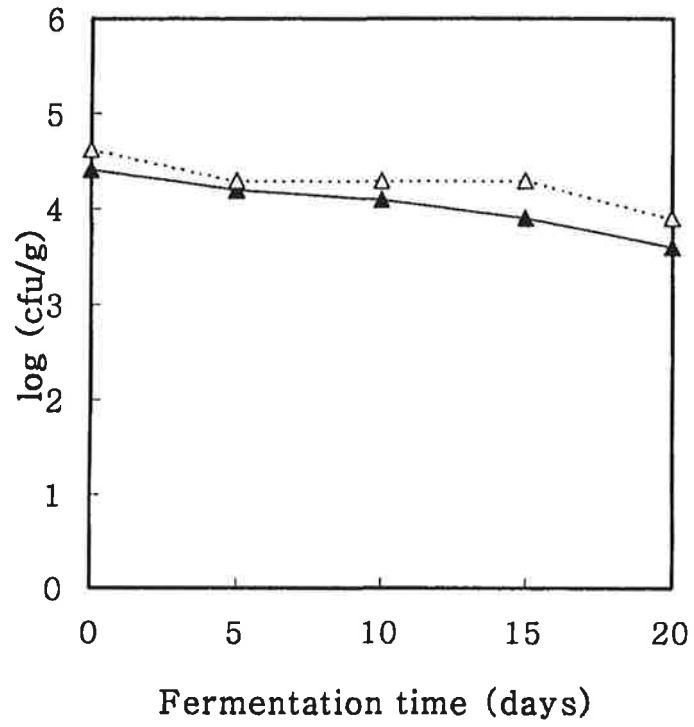


Fig. 23. Changes of coliforms in kimchi during fermentation at 4°C  
 —▲— kimchi with Bifidobacterium  
 ...△... kimchi without Bifidobacterium

3)

pH가 4.7 가 .  
 5 , 가  
 . pH , 가  
 , pH -  
 . , 가  
 . 가 *B.fidobac- terium*  
 .  
 가 가 lactic acid, acetic acid  
 . *B.fidobacterium* (A/ L) 1.5 : 1  
 1 : 0 lactic acid  
 . *B.fidobacterium*  
 . *Lactobacillus*가 .

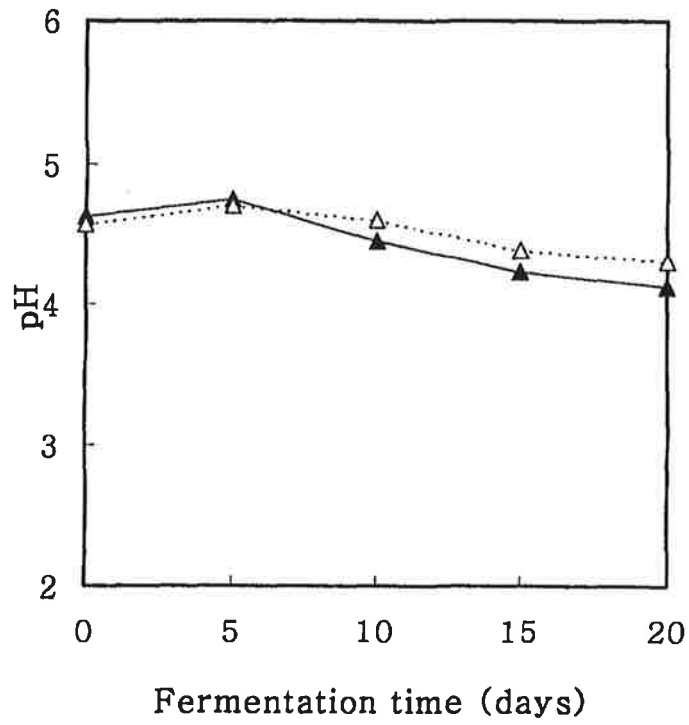


Fig. 24. pH changes of kimchi during fermentation at 4°C  
 —▲— kimchi with Bifidobacterium  
 ...△... kimchi without Bifidobacterium

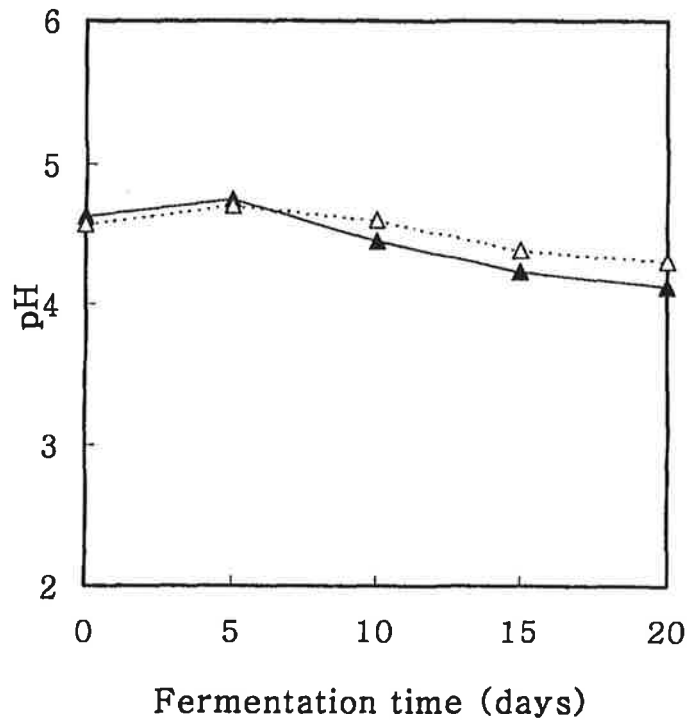


Fig. 24. pH changes of kimchi during fermentation at 4°C  
 —▲— kimchi with Bifidobacterium  
 ...△... kimchi without Bifidobacterium

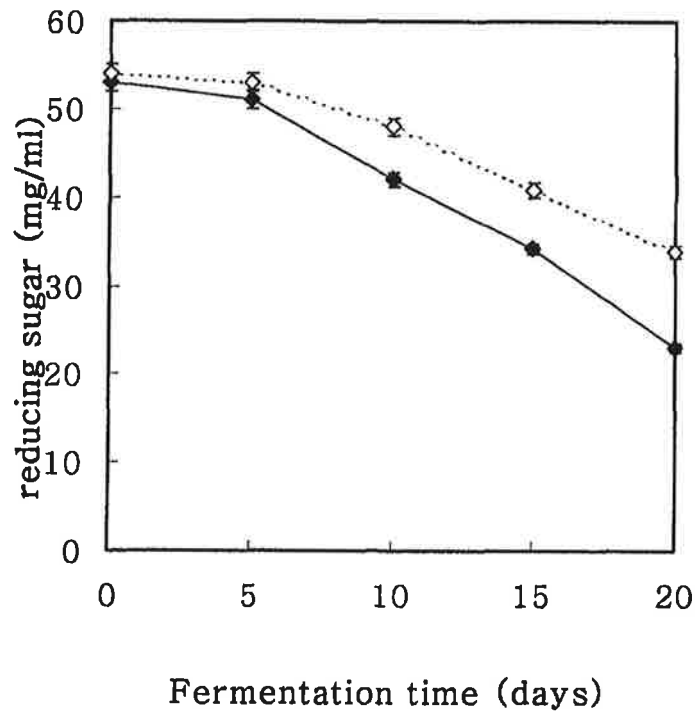


Fig. 25. Changes in reducing sugar content of kimchi during fermentation at 4°C

—◆— kimchi with Bifidobacterium  
 ...◇... kimchi without Bifidobacterium

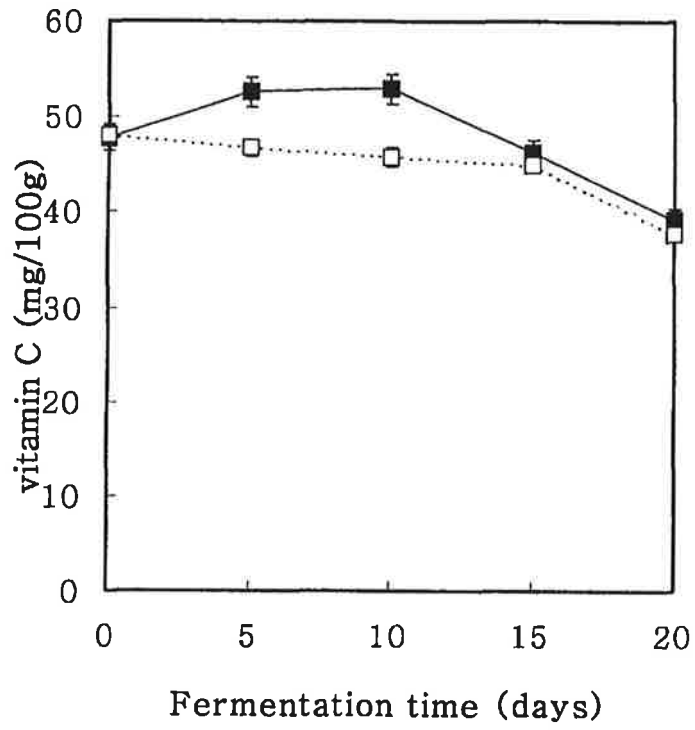


Fig. 26. Changes in vitamin C content of kimchi during fermentation at 4°C

—■— kimchi with Bifidobacterium  
 ...□... kimchi without Bifidobacterium



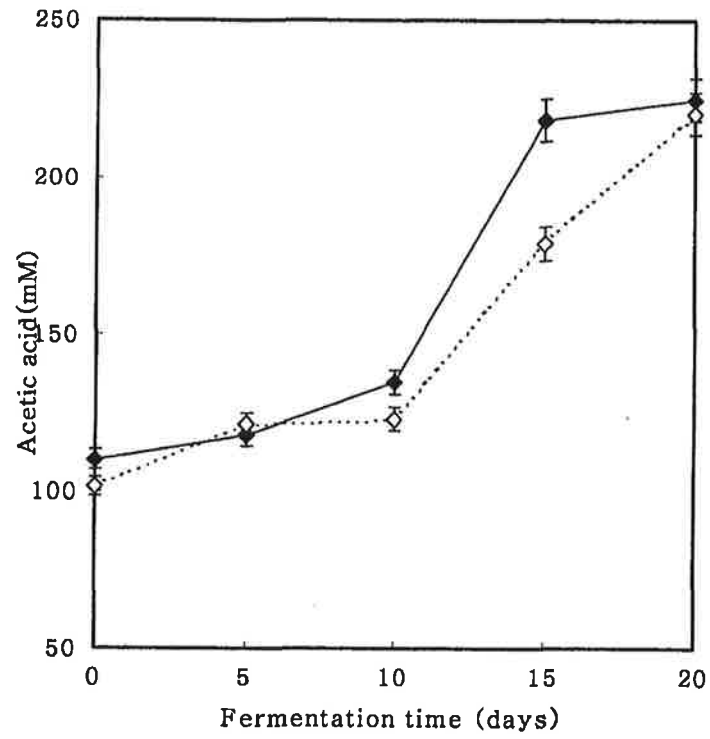


Fig. 27. Acetic acid content in kimchi during fermentation at 4°C  
 —◆— kimchi with Bifidobacterium  
 ...◇... kimchi without Bifidobacterium

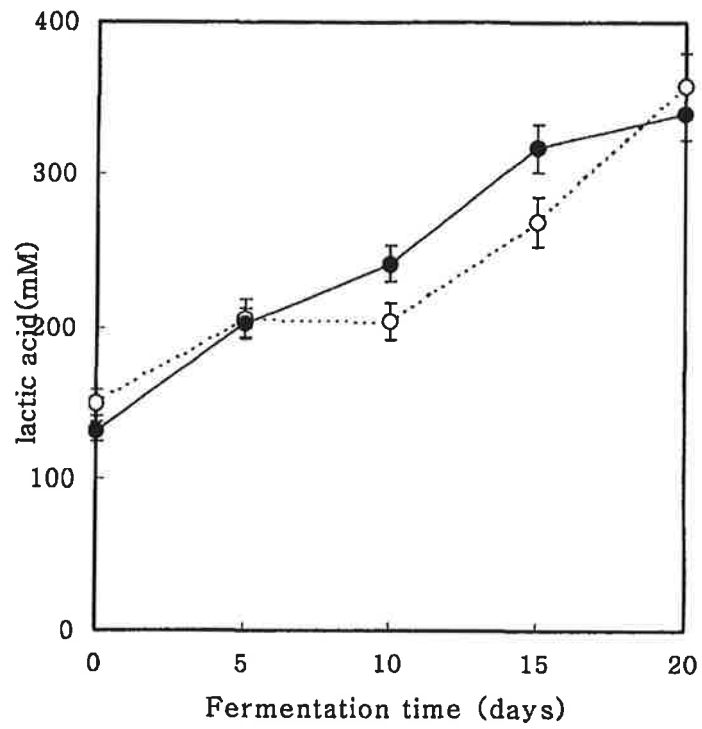


Fig. 28. Lactic acid content in kimchi during fermentation at 4°C  
 —◆— kimchi with Bifidobacterium  
 ...◇... kimchi without Bifidobacterium

4)

20 pH가 4.2가 , , , 9  
가 가 .  
, *B.fidobacterium* 가 가 ,  
가 .  
*B.fidobacterium* 가 가 . 가 *B.fidobacterium*  
acetic acid

Table 28. Sensory sheet of score test of kimchi

가

\_\_\_\_\_ : \_\_\_\_\_ : \_\_\_\_\_ :  
 \_\_\_\_\_ , \_\_\_\_\_ , \_\_\_\_\_ , \_\_\_\_\_ , \_\_\_\_\_ 가

1 2 3 4 5 6 7 8 9  
 -----+-----+-----+-----+-----+-----+-----+-----+-----+-----

1 2 3 4 5 6 7 8 9  
 -----+-----+-----+-----+-----+-----+-----+-----+-----+-----

-----+-----+-----+-----+-----+-----+-----+-----+-----

1 2 3 4 5 6 7 8 9  
 -----+-----+-----+-----+-----+-----+-----+-----+-----+-----

-----+-----+-----+-----+-----+-----+-----+-----+-----

-----+-----+-----+-----+-----+-----+-----+-----+-----

1 2 3 4 5 6 7 8 9  
 -----+-----+-----+-----+-----+-----+-----+-----+-----+-----

-----+-----+-----+-----+-----+-----+-----+-----+-----

-----+-----+-----+-----+-----+-----+-----+-----+-----

1 2 3 4 5 6 7 8 9  
 -----+-----+-----+-----+-----+-----+-----+-----+-----+-----

Table 29. Sensory score of kimchi with *Bifidobacterium* during fermentation at 4

<b>Attributes</b>	<b>A</b>	<b>B</b>
<b>Color</b>	5.38±2.13 <sup>1)</sup>	6.00±1.00
<b>Odor</b>		
Acetic odor	3.25±1.03 <sup>2)</sup>	6.71±1.38 <sup>a</sup>
Fresh cabbage odor	4.75±1.39 <sup>a</sup>	3.00±2.16 <sup>b</sup>
Fresh acidic odor	3.63±1.69 <sup>b</sup>	6.57±1.52 <sup>a</sup>
<b>Taste</b>		
Hot taste	3.63±1.19	4.71±1.25
Acidic taste	3.12±0.83 <sup>b</sup>	6.00±1.73 <sup>a</sup>
Fresh cabbage taste	4.50±2.33	4.86±2.79
Fresh sourness taste	2.88±0.99 <sup>b</sup>	5.86±1.95 <sup>a</sup>
<b>Texture</b>		
Softness	4.75±1.58	4.29±1.70
Fracturability	5.38±1.51	6.00±1.00
Chewiness	5.00±2.00	5.14±1.86
<b>Overall acceptability</b>	4.25±1.28 <sup>b</sup>	5.71±0.49 <sup>a</sup>

1) Mean±SD, Data were analyzed by independent t-test.

2) Different superscripts within a column indicate significant difference (p<0.05).

A : kimchi without *B. bifidobacterium*

B : kimchi with *B. bifidobacterium*

5)

가) *Bifidobacterium* 105 cfu/ g 106 cfu/ g ,  
*Bifidobacterium* 가 가 *Lactobacillus* .

) *Bifidobacterium* 가 .

) *Bifidobacterium* 가 , 15

) *Bifidobacterium* 가 pH가 , 가  
pH 가 .

) *Bifidobacterium* 가 가  
가 .

5.

가 pH

가 .

, *Lactobacillus*

가.

pH 0 pH 6 7  
 pH 4 7 pH 5  
 가 가 (Fig. 29). HPLC

lactic acid oxalic acid가

(data not shown).

. (%) 가 가 , pH 가  
 7 3%가 (Fig. 30).  
 : (TS medium)  
 (MRS medium), (DHL medium) pouring culture method

가 ,  
 5 가 가 (Fig. 31, 32).

log 4 5 ,  
 5 log 8 가 ,  
 7 log 3 가 (Fig. 33).

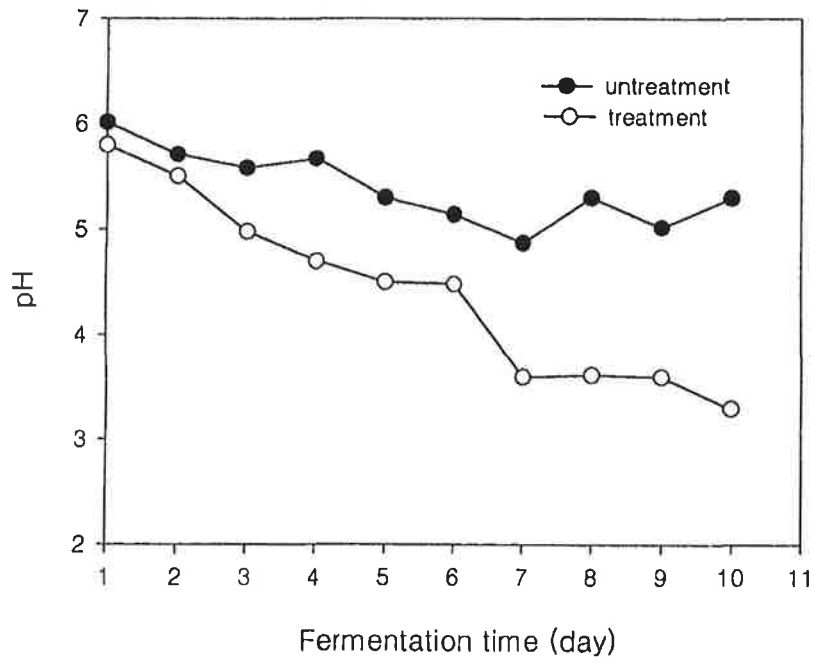


Fig. 29. The change of pH in cabbage during fermentation period.



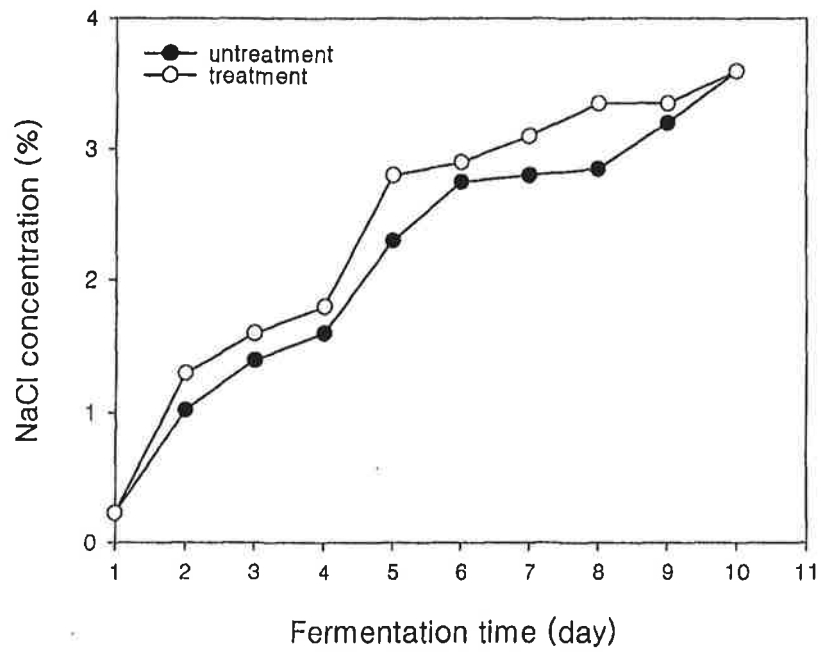


Fig. 30. The change of salt concentration in cabbage during fermentation period.

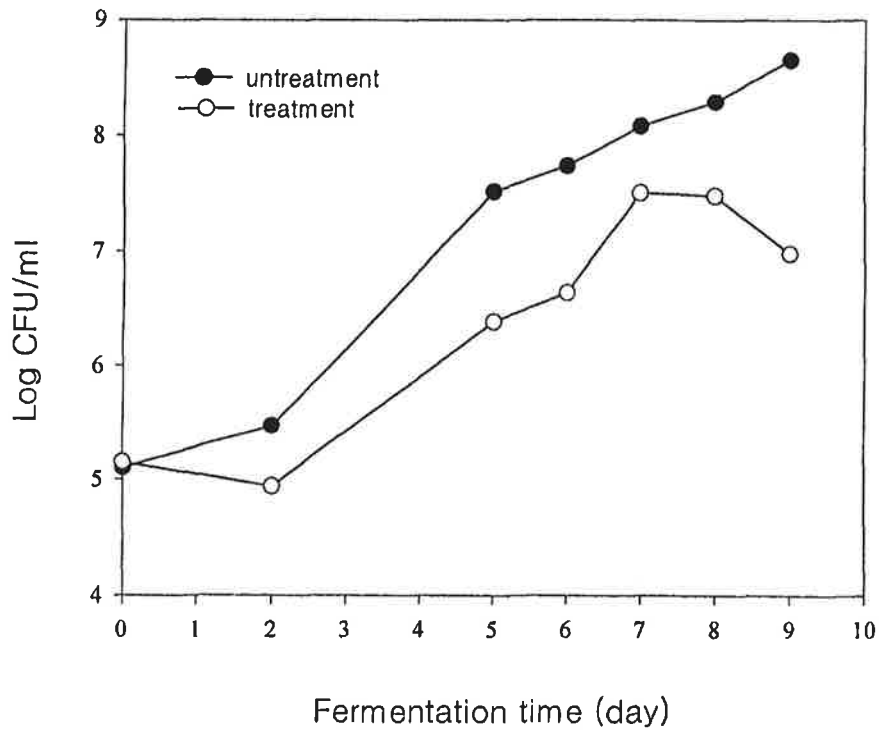


Fig. 31. Comparisons of total microbial profile between untreated and treatment during cabbage fermentation

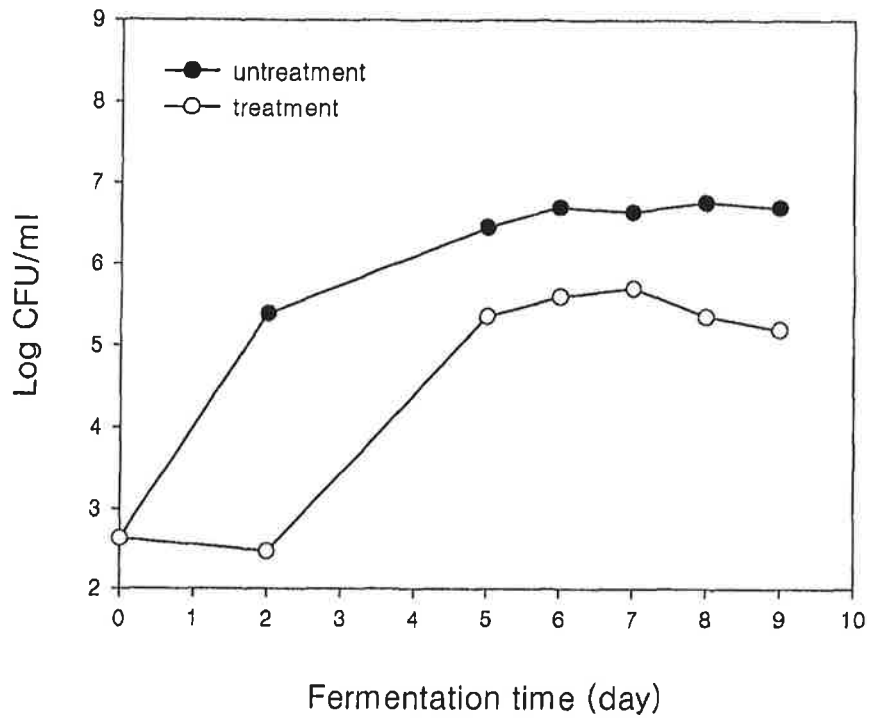


Fig. 32. Comparisons of *Lactobacillus* profile between untreated and treatment during cabbage fermentation

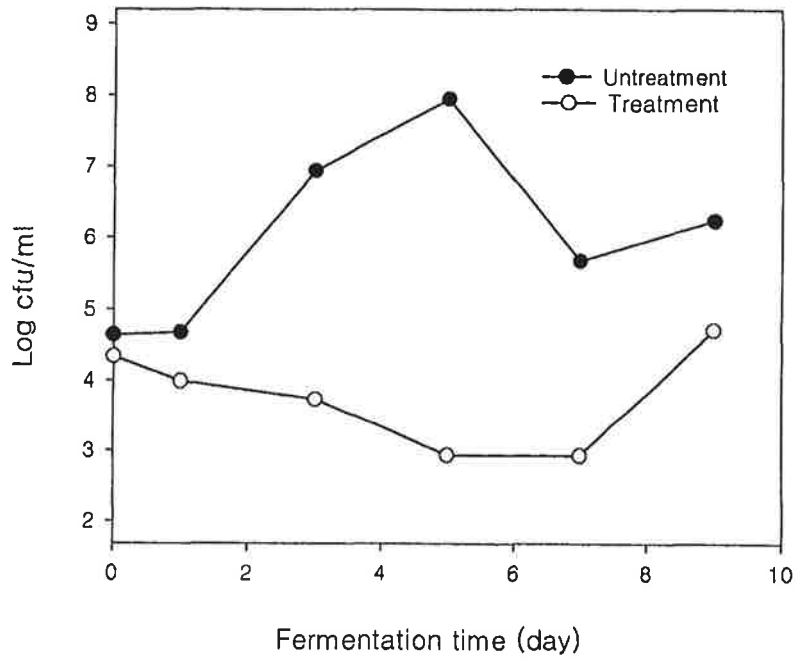


Fig. 33. Comparisons of *E. coli* profile between cabbage treated lactic acid and untreated during cabbage fermentation.

:

, 7

가

가 Photo. 1

(Spectro colorimeter JS555 Tokyo, Japan)

Fig. 35, 36

, 가 untreatment  
treatment

Fig. 35 untreatment treatment . 10

E untreatment 30 treatment 10

가

'L' 10 untreatment 30 가 , treatment  
-10

treatment 가 ,

가 가 가 .

'a' , 'b' untreatment가 10 5

treatment -5 가 가

. Fig. 36 untreatment treatment .

untreatment가 40 treatment 45 가

, 'b' untreatment가 20 가 treatment

10 'a' untreatment treatment

가 . Fig. 35 36

가 pH 가

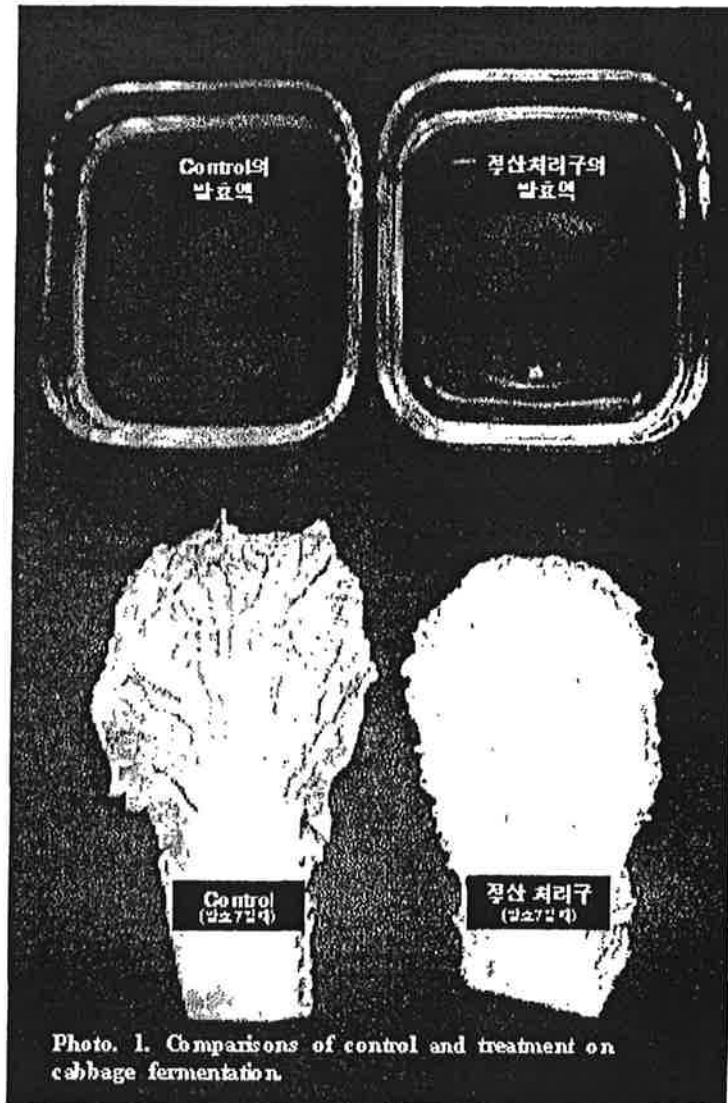


Fig. 34. Comparisons of control and treatment on cabbage fermentation

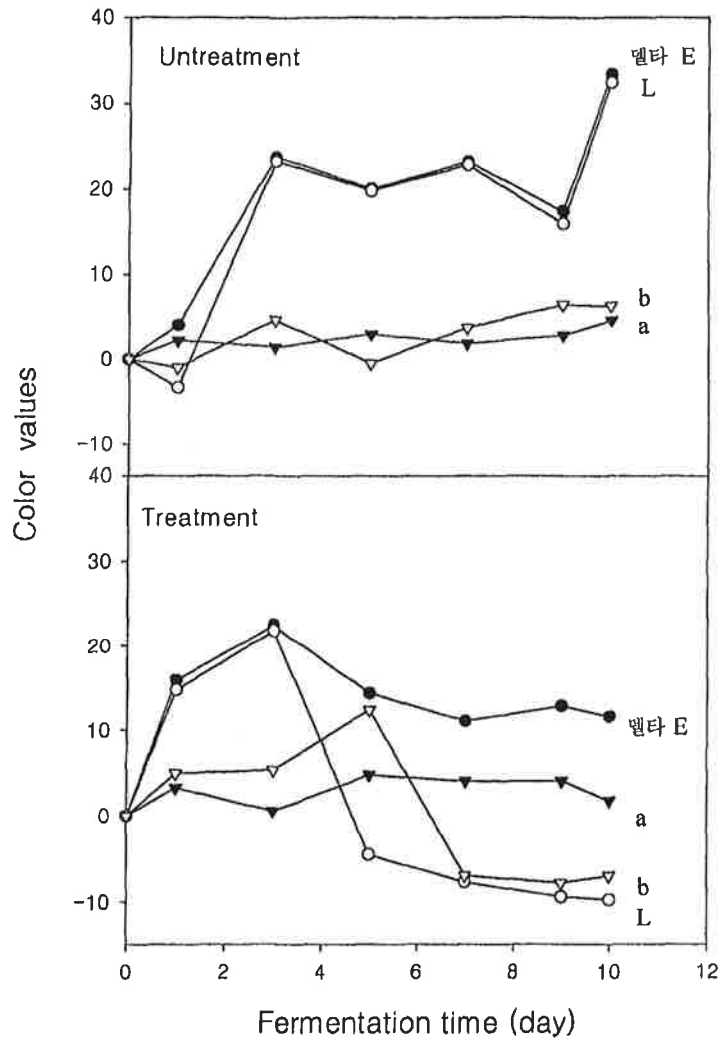


Fig. 35. Changes in color values of cabbage fermentation  
 델타 E: 변화율, 'L': 밝기, 'a': 적색-녹색, 'b': 황색-청색

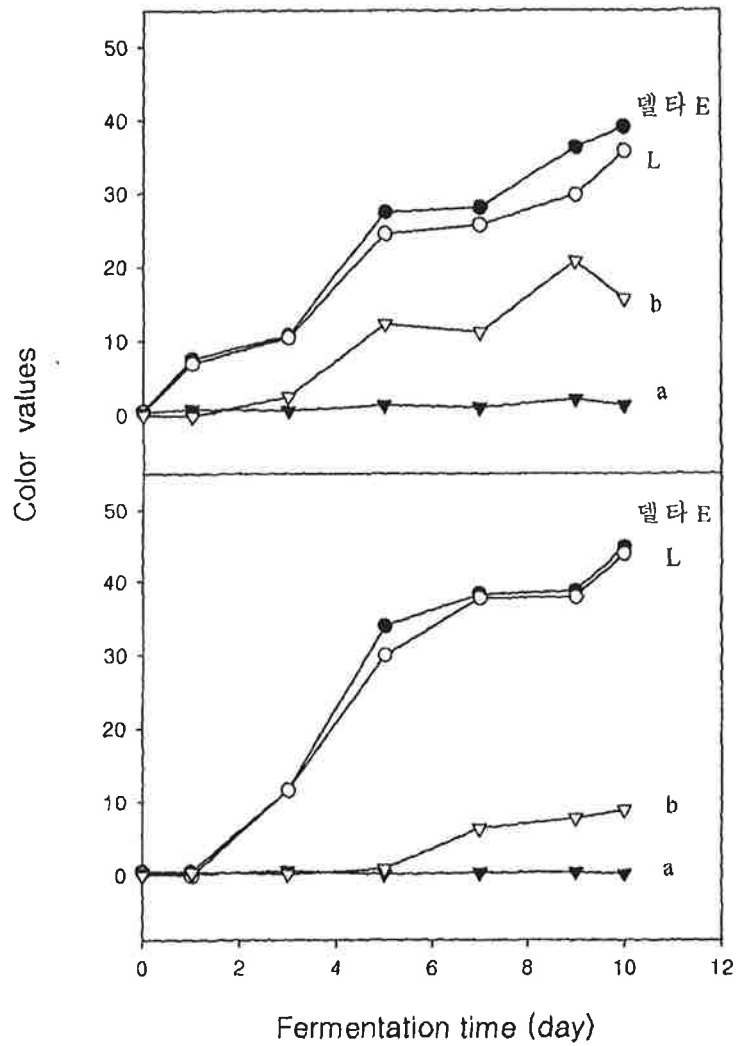


Fig. 36. Changes in color values of fermentation solutions on cabbage fermentation. 델타 E: 변화율, 'L': 밝기  
'a': 적 색 - 녹색, 'b': 황 색 - 청 색



: (pectin)  
 (lignin) methyl ester ,  
 , (未熟)  
 가 protopectin  
 (成熟) protopectinase protopectin 가  
 pectin 가 가  
 β- (transelimination) glycoside  
 가  
 가 ,  
 가 pH 7.1 pH 6.3  
 pH 4.1

TAXT2i Texture Analyser  
 (MHK Trading Co., English) Compression test ( ) TPA  
 (Texture Profile Analysis) Fractuability ( ), Hardness  
 ( ), Chewiness ( ), Guminess( ) 4가  
 5% (untreatment) 5% 0.25%(pH 2.5) 가  
 (treatment)  
 20 2  
 sampling TPA Fig. 37  
 pH  
 2 가  
 1.5 5 Kg 가 가 1,5 2 Kg

가 .

,

.

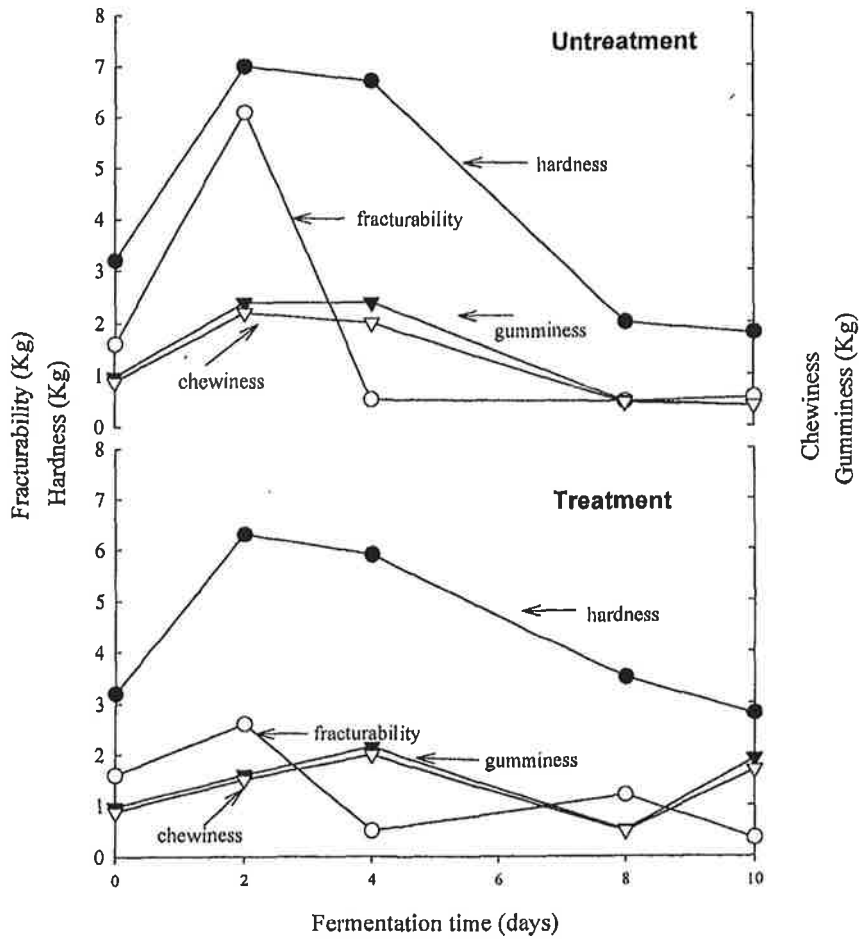


Fig. 37. Texture profile parameter of Chinese cabbage during fermentation in salt solution and lactic acid added solution fermentation solution is added 5% NaCl and adjusted pH 2.5 as lactic acid.

: Pectin      pectinic acid  
 가 methyl ester      polygalacturonic acid      ,  
 가      .  
 가      polygalacturonase(poly- $\alpha$ -  
 -1,4-galacturonide glycanohydrolase)      pectinesterase(pectin-pectyl hydrolase)가      .  
 Pectinesterase      methylester group      가  
 (deesterification)      .      가  
     . Polygalacturonase       $\alpha$ -1,4      가  
     , polygalacturonic acid      .      가  
     가      가      ,      protopectin      pectin  
 (depolymerization)가      가      .

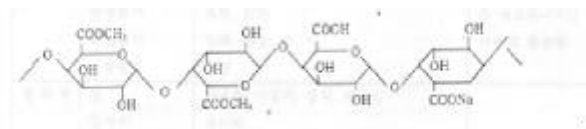


Fig. 38. Polygalacturonic acid

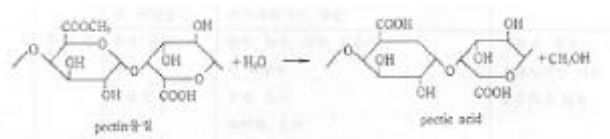


Fig. 39.

PG      PE      . PG      1M  
 NaCl      24      75%      ,

PE 0.1M phosphate (pH 8.0) 24 ,  
 . PG dinitrosalicylic acid (DNS)  
 , PE titrimetric assay .

Fig. 40. standard curve

PG 가

Fig. 41.

PG 가가 가 .

가

Fig. 42, 43

10

2

PE PG 가

. PE PG 가

가

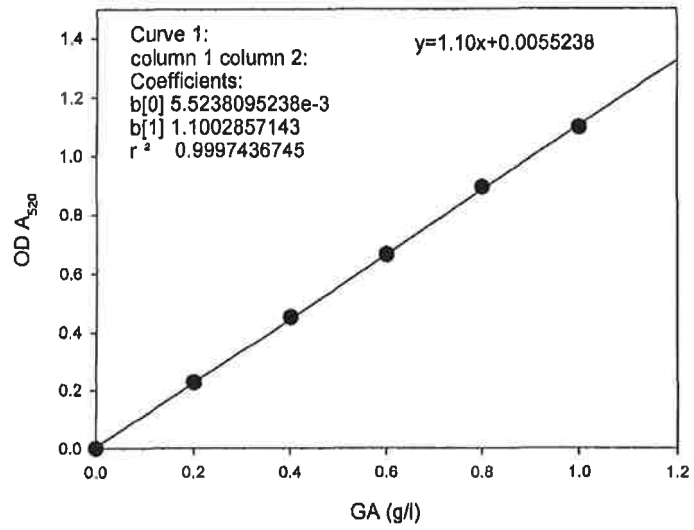
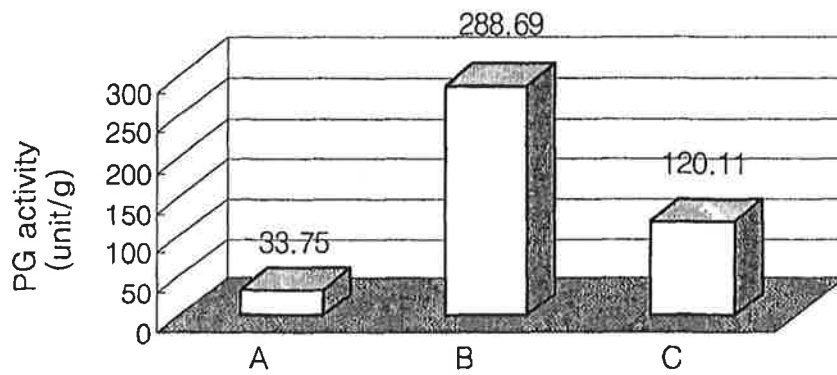


Fig. 40. Galacturonic acid standard curve



A: Law cabbage  
 B: Untreatment(10일발효)  
 C: Treatment(10일발효)

Fig. 41. 배추·단독 발효 후의 polygalacturonase의 역가측정.

가

가

. Table 30

4

가

Table 31

Table 32

4

BA

. flavor

color

가 *Lactobacillus* sp. HL-48

3

BA-8

(film yeast)

8

가

(Fig.44, Fig. 45, Fig. 46)

가

BA-8

가

8

(film yeast)

bacteriocin

Gram

(13)

10

20

BA-8

8

가 가

(flavor),

가

BA-8

가

bacteriocin

가

가



Table. 30 Sensory evaluation of salted cabbage without inoculation of Lactic Acid Bacteria by hedonic scale.(1)

parameter \ day	0	1	2	3	4	5	6	7	8
taste	3.00	2.80	3.33	2.78	2.38	2.40	2.60	2.48	2.23
flavor	3.00	2.55	2.90	3.08	3.15	3.18	3.30	2.98	2.73
color	3.00	2.88	2.75	2.72	2.70	2.60	2.35	2.31	2.28

1) Each value represented the mean of 10 observation using on hedonic scale of 1 (dislike very much) to 5 (like very much).

Table. 31 Sensory evaluation of salted cabbage with *Lactobacillus* sp. HL-48 by hedonic scale<sup>1)</sup>.

parameter \ day	0	1	2	3	4	5	6	7	8
taste	3.00	3.15	3.78	3.40	2.90	2.70	3.60	3.29	2.90
flavor	3.00	3.20	3.20	3.34	3.48	3.65	3.73	3.42	3.10
color	3.00	3.00	3.20	2.76	2.38	2.18	2.18	2.09	2.03

1) Each value represented the mean of 10 observation using on hedonic scale of 1 (dislike very much) to 5 (like very much).

Table 32. Sensory evaluation of salted cabbage with Lactobacillus strain BA-8 by hedonic scale.(1)

day parameter	0	1	2	3	4	5	6	7	8
taste	3.00	3.03	3.43	3.54	3.63	3.70	3.88	3.47	3.25
flavor	3.00	3.25	3.38	3.43	3.55	4.05	3.98	3.32	3.18
color	3.00	3.20	3.53	3.50	3.48	3.45	3.50	3.22	2.95

1) Each value represented the mean of 10 observation using on hedonic scale of 1 (dislike very much) to 5 (like very much).

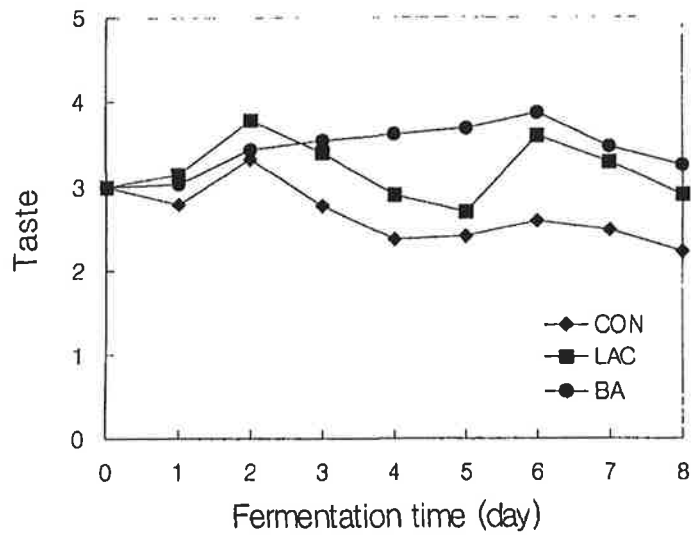


Fig. 44. Changes of taste of salted cabbage on the addition of lactic acid bacteria. (-◆- control ; -■- Lactobacillus sp. HL-1 ; -●- Lactobacillus strain BA)

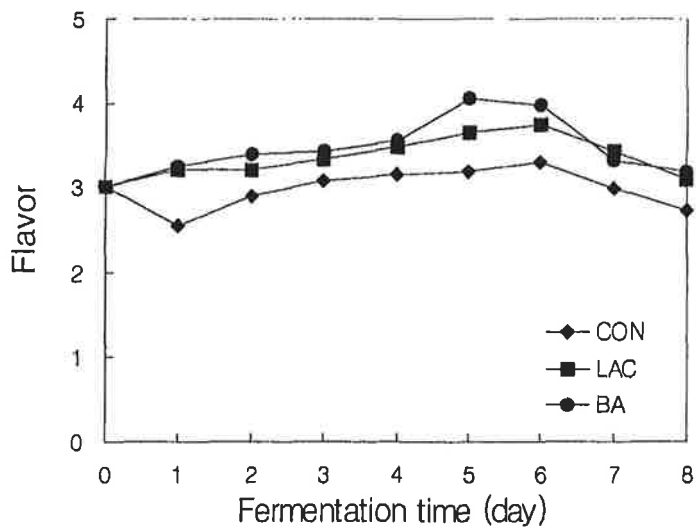


Fig. 45. Changes of flavor of salted cabbage on the addition of lactic acid bacteria  
 (-◆- control ; -■- Lactobacillus sp. HL-1 ; -●- Lactobacillus strain BA)

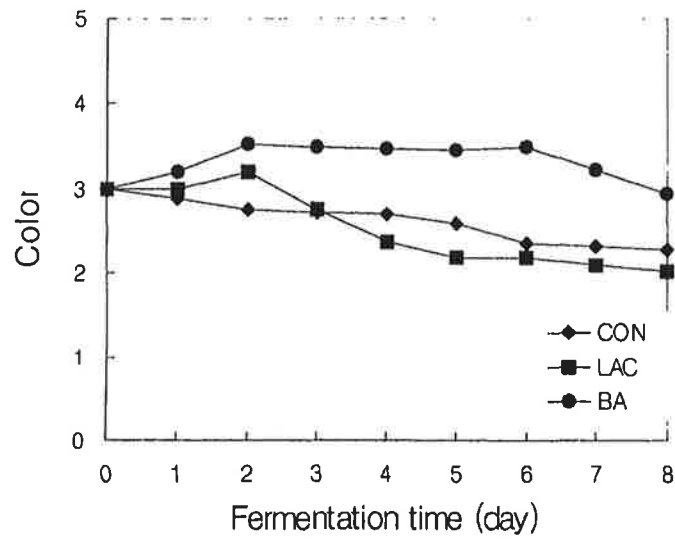


Fig. 46. Changes of color of salted cabbage on the addition of lactic acid bacteria. (-◆- control ; -■- Lactobacillus sp. HL-1 ; -●- Lactobacillus strain BA)

#### 나. 부원료의 각개발효 기법 연구

부원료들의 영향: 각각의 부원료를 따로 발효시키면서 비교해본 결과 Fig. 47에서 보여 주듯이 대조구에 비해서 새우젓과 당, 스타터 첨가구에서는 양념내 유산균의 증식이 높은 데 반해 소금 첨가구에서는 증식이 낮았다. 이에 따라 소금 첨가구의 pH 변화는 다른 모든 양념구에 비해 서서히 pH도 낮아졌다. 유기산 변화는 HPLC로 조사하였으며, 전처리는 solid phase extraction cartridge로 처리하여 양념의 색소를 제거한 후 0.2 pore disk-type filter로 여과하여 분석 시료로 사용하였다. 그 결과 김치 양념은 lactic acid, malic acid, citric acid, succinic acid, oxalic acid를 포함하고 있었으며, citric acid는 발효 전반에 있어서 감소하였고, 그 외의 유기산들은 증가하는 경향을 보였다(Table 33).

Table 33. The change of other organic acids concentration during Kimchi-seasoning fermentation. (n/n)%

day sam ple	Lactic acid						Acetic acid						Malic acid					
	0	3	6	9	12	15	0	3	6	9	12	15	0	3	6	9	12	15
1	39.15	38.33	280.5	455.2	524.6	861.7	-*	-.	6.176	13.52	18.29	40.83	16.56	2.596	14.93	27.11	32.17	37.39
2	123.2	68.52	68.45	135.9	161.4	314.4	-	-*	-.	-.	-.	-.	10.49	2.500	18.33	23.47	21.55	19.33
3	156.8	197.0	478.7	447.4	587.7	587.3	-	9.26	18.15	13.24	21.81	34.04	11.12	6.666	26.84	18.66	25.18	23.45
4	29.52	60.13	305.4	163.7	402.7	598.3	-	-*	8.16	1.86	3.89	19.93	2.388	6.795	31.11	9.418	16.80	23.08
5	251.4	165.2	322.3	328.1	549.8	723.0	-	14.03	13.67	15.12	16.88	18.38	13.12	3.436	16.76	14.53	18.49	26.69
6	261.7	519.1	264.6	426.9	428.4	411.4	-	-*	-*	-*	-.	-.	21.68	29.92	21.44	24.86	32.52	23.26
7	782.8	172.1	260.3	301.9	406.4	327.6	-	-*	-*	-*	-.	-*	24.73	9.039	25.14	29.00	19.63	20.51

day sam ple	Citric acid						Succinic acid						Oxalic acid					
	0	3	6	9	12	15	0	3	6	9	12	15	0	3	6	9	12	15
1	-	-*	-*	-*	-.	-*	22.08	1.969	32.88	50.49	41.36	-.	17.63	4.47	34.18	40.11	37.15	34.53
2	10.54	4.663	11.39	8.867	3.924	4.627	-.	-.	30.04	38.52	34.91	27.64	14.27	18.53	37.69	40.49	33.68	40.35
3	-	-*	-*	-*	-.	-*	-.	32.19	47.29	36.97	35.92	35.39	14.72	27.39	48.71	32.17	35.15	42.14
4	4.824	5.313	6.575	3.068	0.518	0.531	10.78	14.17	54.86	21.87	32.96	17.38	9.631	8.419	32.20	27.04	32.96	46.13
5	16.64	0.346	0.381	0.321	0.321	0.949	-.	12.30	51.92	46.35	43.65	37.37	31.41	11.34	28.43	25.65	43.58	46.83
6	8.791	10.82	8.994	7.350	10.86	5.136	23.62	36.84	25.17	35.21	22.80	30.51	44.17	29.98	45.99	37.80	38.20	39.89
7	1.691	10.26	18.93	17.31	11.74	11.24	22.66	18.62	25.43	34.31	30.48	19.50	47.36	40.82	56.06	41.67	50.45	48.15



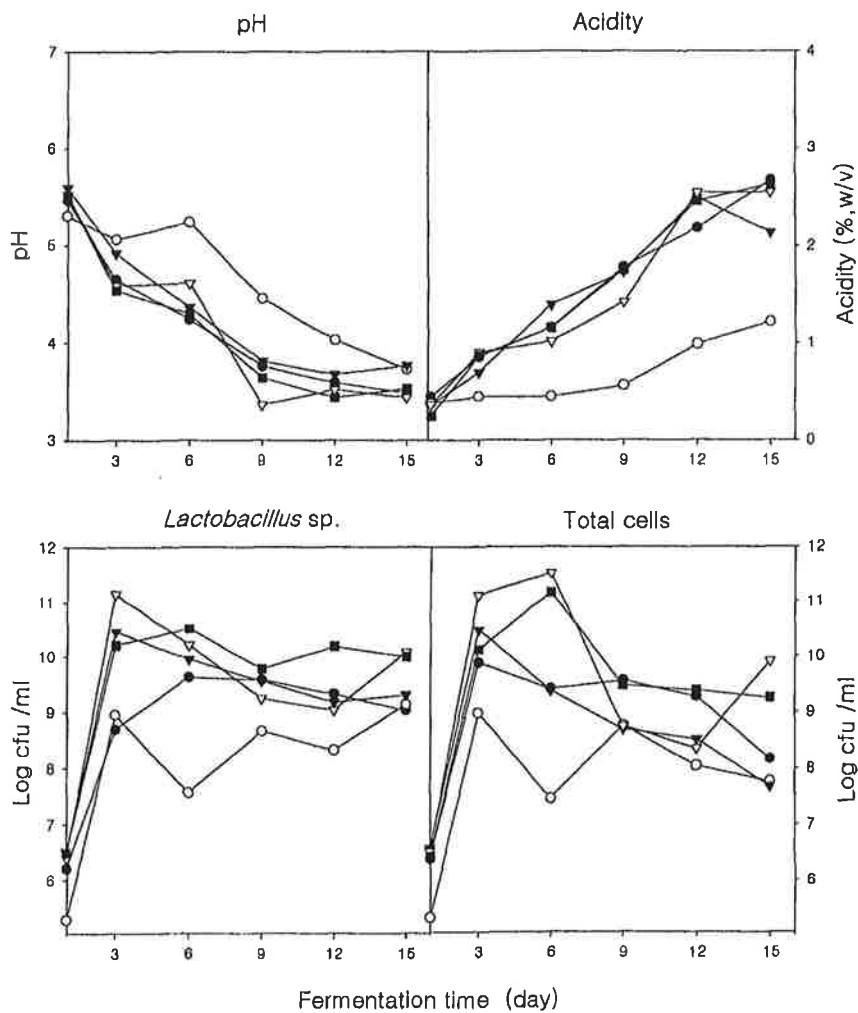


Fig. 47. The change of pH, acidity and microbials during kimchi seasoning fermentation. ● basal seasoning  
 ▼ added S. shrimp    ▽ added sugar    ■ added starter    ○ added salt

가 :

가

Table 34

가

E 0 6

color 'b'

. pH

6

40

3.6

,

0.4%

가

4.8

40 4.3

가

pH

(Fig. 48).

Fig. 49

log

9

가

가

,

0.4% 가

가

가

,

가

가

0.4%

가

가

Table 34. Changes in color values of Kimchi-seasoning added various lactic acid concentration during fermentation at 20.techno system

Fermentation time (days)	0	3	6	9	12	16	24	30	36	42	
Control	∇E	0.05	2.19	2.53	2.43	0.92	1.43	4.75	4.33	3.74	6.69
	"L"	0.02	1.06	1.26	1.23	0.11	0.73	2.40	2.17	1.92	3.38
	"a"	-0.02	0.74	0.62	0.43	-0.90	0.14	0.92	1.00	-0.46	1.24
	"b"	0.03	1.77	2.10	2.05	0.16	1.22	4.00	3.61	3.18	5.63
0.4%	∇E	0.09	1.71	1.85	2.34	0.75	1.67	1.36	0.72	1.48	1.43
	"L"	-0.04	0.87	0.93	1.19	0.38	0.86	-0.52	0.34	0.74	0.71
	"a"	-0.04	0.24	0.36	0.39	-0.17	0.12	-0.90	-0.29	-0.38	-0.42
	"b"	-0.07	1.45	1.55	1.97	0.62	1.43	-0.88	0.56	1.22	1.17
0.6%	∇E	0.03	0.82	0.97	0.73	0.81	0.88	0.91	0.80	1.27	0.91
	"L"	-0.01	0.42	0.49	0.38	0.39	0.45	0.44	0.39	-0.15	0.31
	"a"	-0.03	0.15	0.17	-0.09	0.26	-0.17	-0.33	-0.27	-1.24	-0.71
	"b"	-0.01	0.69	0.82	0.62	0.66	0.74	0.73	0.64	-0.27	0.49
0.8%	∇E	0.06	0.72	0.84	0.59	1.16	0.35	0.55	0.46	1.29	0.87
	"L"	-0.03	0.37	0.43	0.30	-0.49	0.06	-0.13	-0.03	-0.41	0.03
	"a"	-0.03	0.08	0.13	-0.09	-0.64	-0.33	-0.48	-0.46	-1.01	-0.87
	"b"	-0.05	0.61	0.71	0.50	-0.83	0.09	-0.23	-0.05	-0.69	0.03

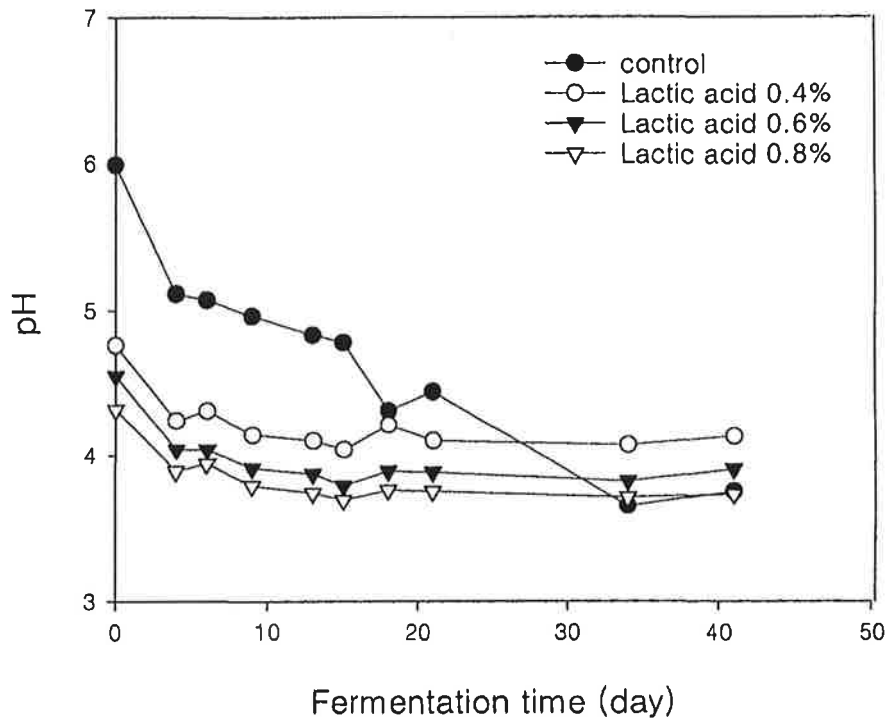


Fig. 48. Changes of pH on Kimchi-seasoning.

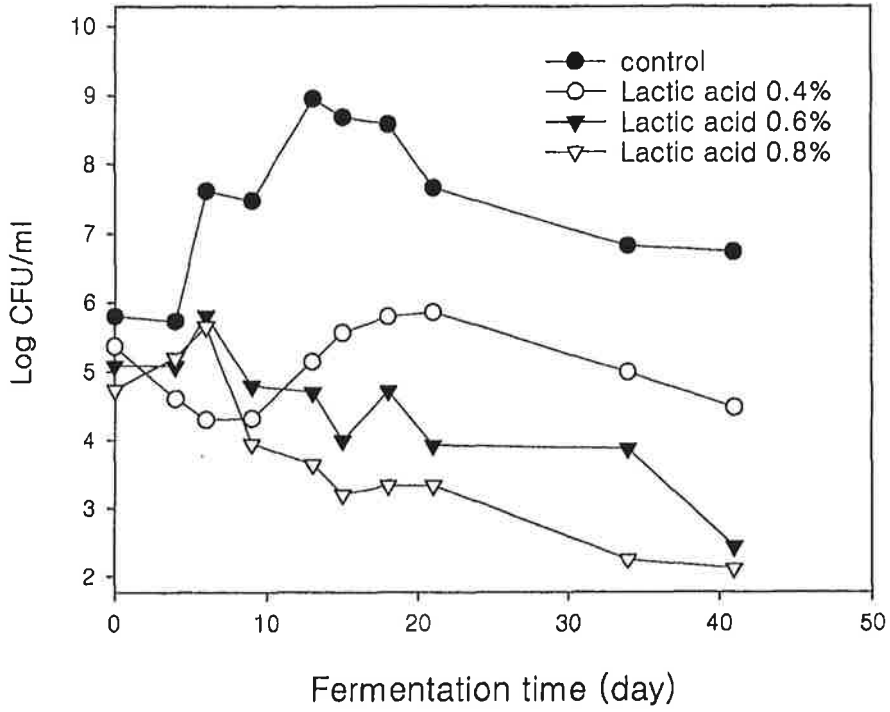


Fig. 49. Profiles of lactic acid bacteria on Kimchi-seasoning

1.

가 , pH, , ,

가 ,

: 가 3

가 . 5-6 가 9

가 , 11 가 . 12

(Fig. 50). 가

가 가

PET-JDSF 가 ,

12 (Fig. 51).

**pH** : 가 pH

.(Fig. 52) 10 가 가 pH가

가 . pH 가

가 11 pH

PET-JDSF

pH가 .

: 가

. 0 , 가

2% . 3 가 가

가 2.8 2.6

가 4

(Fig. 53).

:

, ,

2 hedonic scale

1 ( ) 5 ( )

. 4

, 6

. LDPE HDPE

6

, PET-JDSF

. 6 HDPE LDPE

가 , 가

가 . 7

가 , HDPE LDPE

PET-JDSF

. HDPE가 가 . PET-JDSF

가 , 6

. PET-JDSF

: 8 12 , 8

LDPE PET-JDSF가 가 (Fig. 56).

HDPE . 12

PET-JDSF . PET-JDSF

가 . LDPE 가 가

가 PET-JDSF

, 가 .

HDPE 가 ,

PET-JDSF 가

가 pH 10 가 가

가 HDPE LDPE 가

PET-JDSF pH .

PET-JDSF 가 가 , 가

가 . PET-JDSF 가 가

가

가 ,

PET-JDSF 가 HDPE LDPE .

12 .

4 , 6 .

6 , PET-JDSF

LDPE . 6 HDPE

가 ,

가 가 7

가 , HDPE LDPE

PET-JDSF . HDPE가 가 .

PET-JDSF 가 , 6

. PET-JDSF



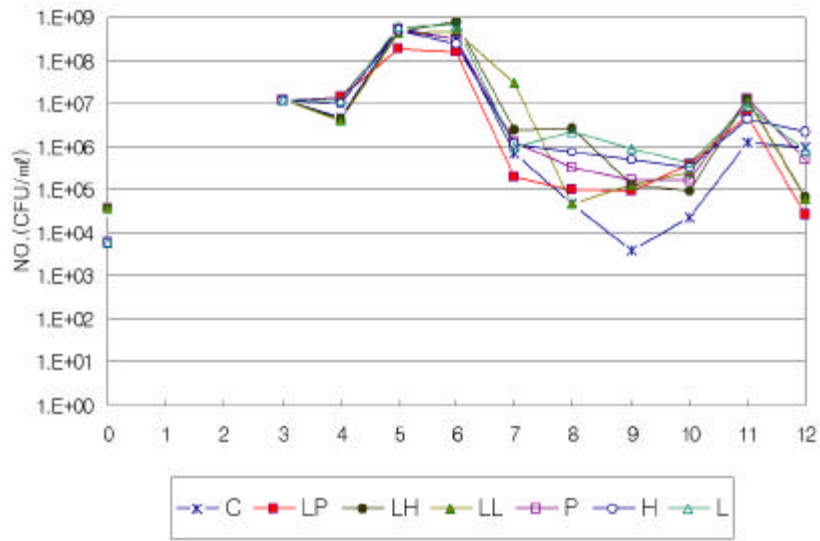


Fig. 50 Changes of *Lactobacillus* number of cabbage preservation in the various packaging materials.

C: control with *Lactobacillus* inoculation, LP: PET-JDSF with *Lactobacillus* inoculation, LH: HDPE with *Lactobacillus* inoculation, LL: LDPE with *Lactobacillus* inoculation, P: PET-JDSF, H: HDPE, L: LDPE

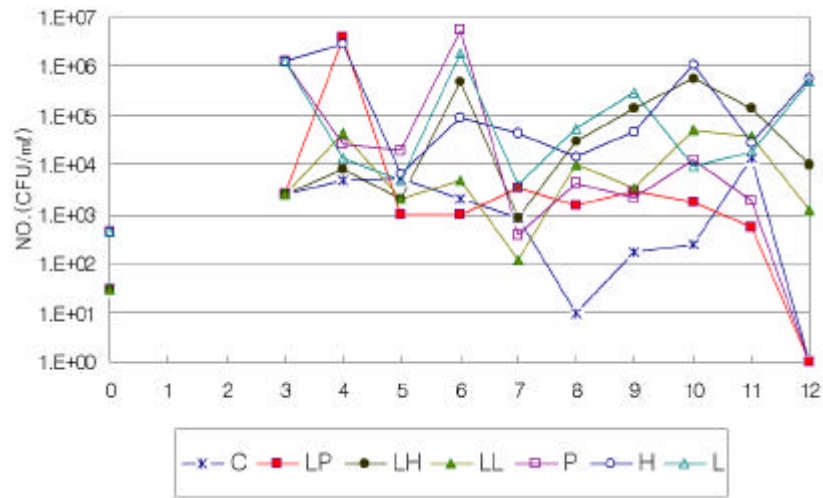


Fig. 51. Change of *E. coli* populations of cabbage during preservation in the various packaging materials.

C: control with *Lactobacillus* inoculation, LP: PET-JDSF with *Lactobacillus* inoculation, LH: HDPE with *Lactobacillus* inoculation, LL: LDPE with *Lactobacillus* inoculation, P: PET-JDSF, H: HDPE, L: LDPE

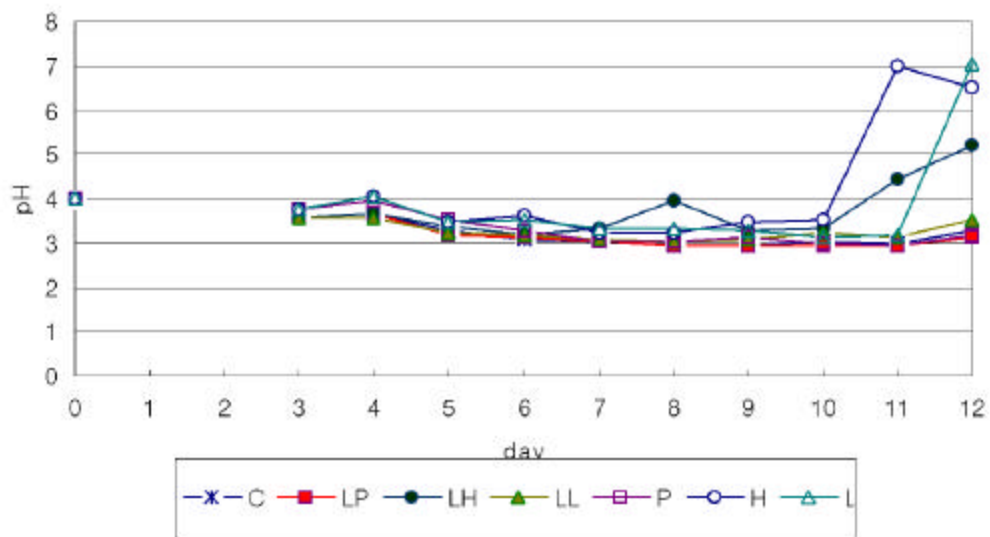


Fig. 52. Changes of pH of cabbage during preservation in the various packaging materials.

C: control with *Lactobacillus* inoculation, LP: PET-JDSF with *Lactobacillus* inoculation, LH: HDPE with *Lactobacillus* inoculation, LL: LDPE with *Lactobacillus* inoculation, P: PET-JDSF, H: HDPE, L: LDPE

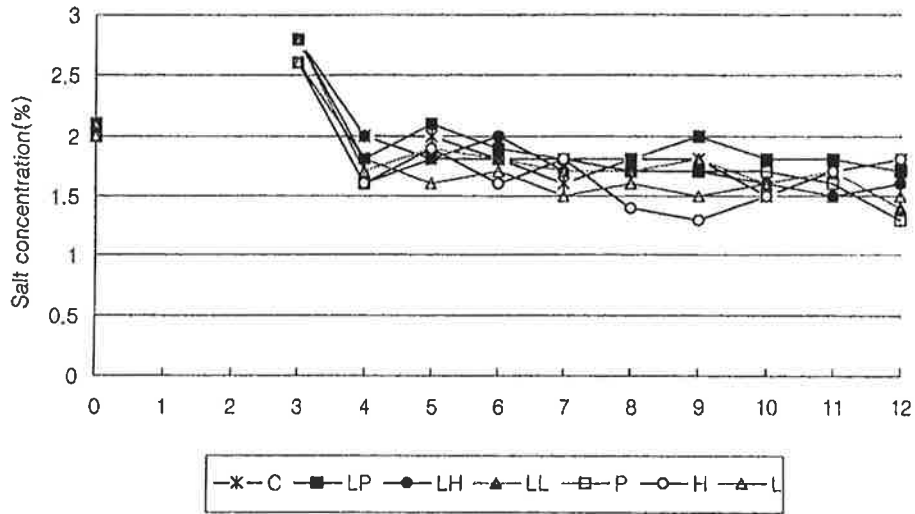


Fig. 53. Changes of salt concentrations of cabbage during preservation in the various packaging materials.

C: control with *Lactobacillus* inoculation, LP: PET-JDSF with *Lactobacillus* inoculation, LH: HDPE with *Lactobacillus* inoculation, LL: LDPE with *Lactobacillus* inoculation, P: PET-JDSF, H: HDPE, L: LDPE

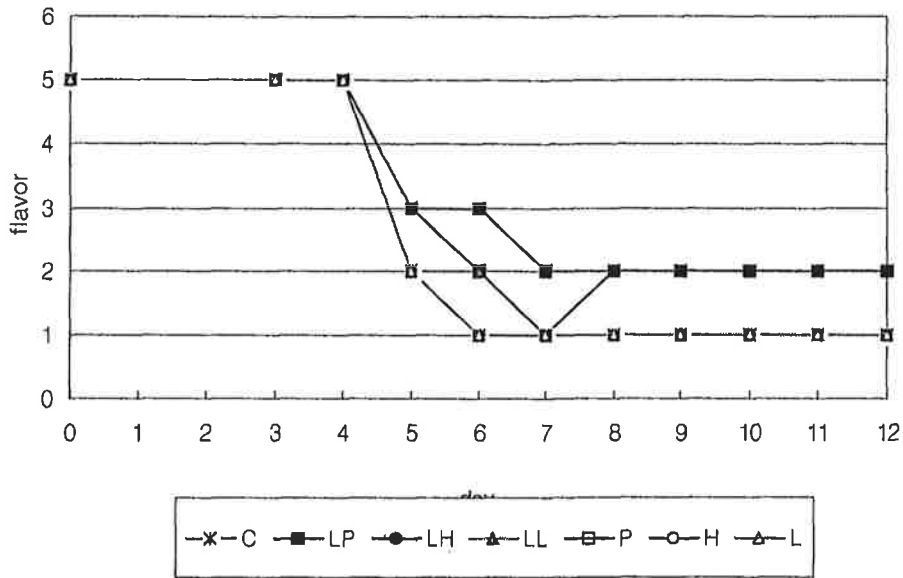


Fig. 54. Changes of tastes of cabbage during preservation in the various packaging materials.

C: control with *Lactobacillus* inoculation, LP: PET-JDSF with *Lactobacillus* inoculation, LH: HDPE with *Lactobacillus* inoculation, LL: LDPE with *Lactobacillus* inoculation, P: PET-JDSF, H: HDPE, L: LDPE

Table 35. Sensory evaluation of taste of salted cabbage with/ without inoculation of Lactic acid bacteria to the various packaging materials by hedonic scale

Sample \ day	0	3	4	5	6	7	8	9	10	11	12
control with <i>Lactobacillus</i> inoculation	5	5	4	2	1	1	1	1	1	1	1
PEP+JDSF with <i>Lactobacillus</i> inoculation	5	5	4	3	3	2	2	2	2	2	2
HDPE with <i>Lactobacillus</i> inoculation	5	5	4	2	2	1	1	1	1	1	1
LDPE with <i>Lactobacillus</i> inoculation	5	5	4	2	2	1	1	1	1	1	1
PEP+JDSF	5	5	4	3	2	2	2	2	2	2	2
HDPE	5	5	4	2	1	1	1	1	1	1	1
LDPE	5	5	4	2	1	1	1	1	1	1	1

\* Indicated number is the most frequent number is chosen in the score made by panel.

Table 36. Sensory evaluation of flavor of salted cabbage with/ without inoculation of Lactic acid bacteria to the various packaging materials by hedonic scale

Sample \ day	0	3	4	5	6	7	8	9	10	11	12
control with <i>Lactobacillus</i> inoculation	5	5	5	3	1	1	1	1	1	1	1
PEP+JDSF with <i>Lactobacillus</i> inoculation	5	5	5	4	3	2	2	2	2	2	2
HDPE with <i>Lactobacillus</i> inoculation	5	5	5	3	2	1	1	1	1	1	1
LDPE with <i>Lactobacillus</i> inoculation	5	5	5	3	2	1	1	1	1	1	1
PEP+JDSF	5	5	5	3	2	2	2	2	2	2	2
HDPE	5	5	5	3	1	1	1	1	1	1	1
LDPE	5	5	5	3	1	1	1	1	1	1	1

\*Indicated number is the most frequent number is chosen in the score made by panel.

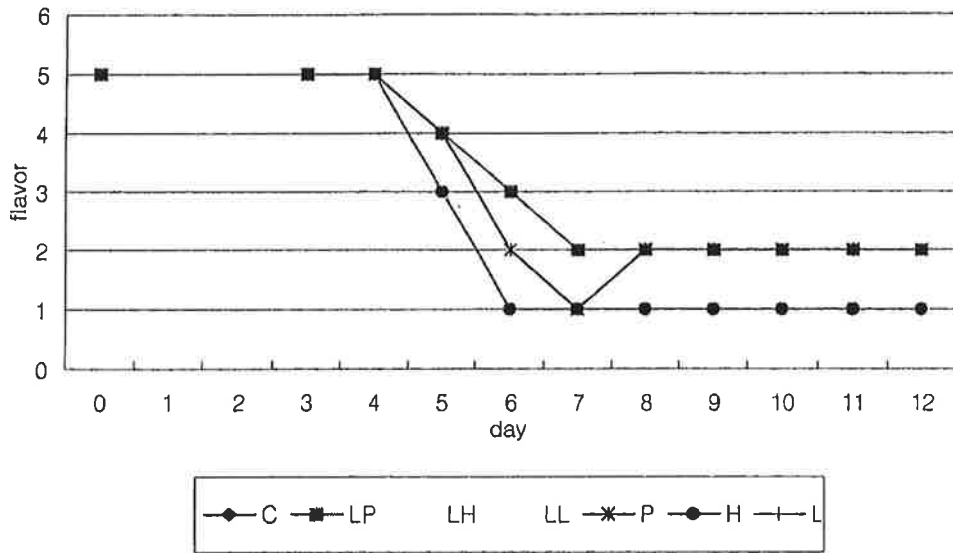


Fig. 55. Changes of flavor of cabbage during preservation in the various packaging materials.

C: control with *Lactobacillus* inoculation, LP: PET-JDSF with *Lactobacillus* inoculation, LH: HDPE with *Lactobacillus* inoculation, LL: LDPE with *Lactobacillus* inoculation, P: PET-JDSF, H: HDPE, L: LDPE



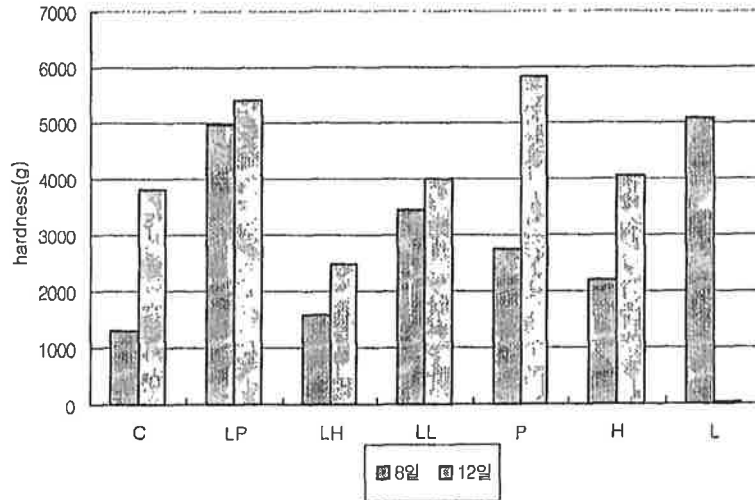


Fig. 56. Hardness of tastes of cabbage during preservation in the various packaging materials.

C: control with *Lactobacillus* inoculation, LP: PET-JDSF with *Lactobacillus* inoculation, LH: HDPE with *Lactobacillus* inoculation, LL: LDPE with *Lactobacillus* inoculation, P: PET-JDSF, H: HDPE, L: LDPE

2. 가 가

가

가

가.

	가	5%가	pH 2.5
	(salt taste),	(acidic taste),	(fresh cabbage taste),
	(crispness),	(chewiness),	(yeast moldy taste),
	(overall eating quality) 7가	1013	

가 20-30

, Fig. 57

가

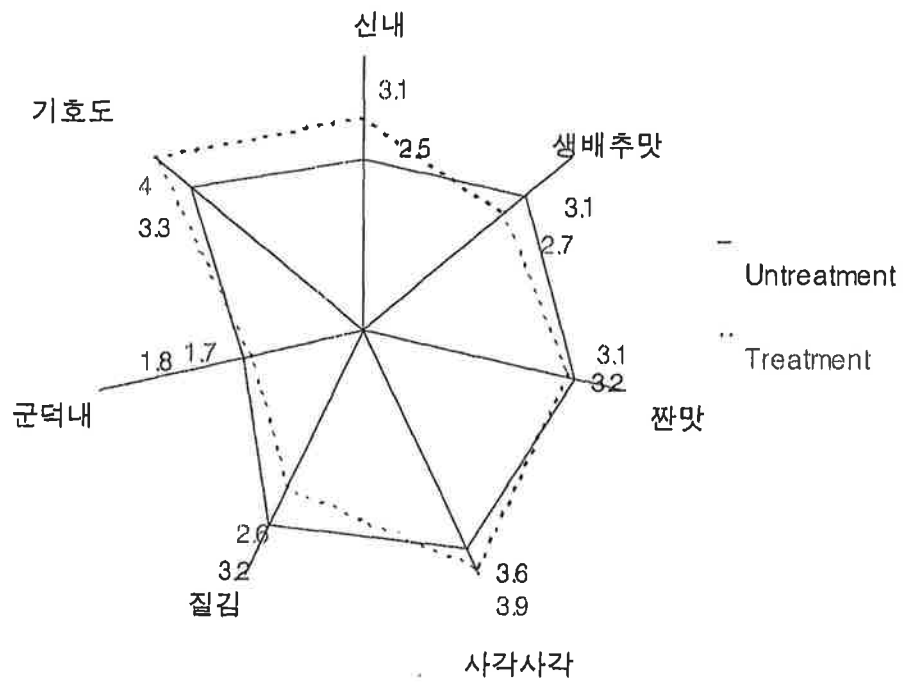


Fig. 57. A spider-web diagram of difference between treatment and untreatment with lactic acid on

가 가 . Table  
8 가 ,

가 , ,  
Table 8 가  
(flavor), (taste), (texture), (overall acceptability) 4가  
, 5 (scoring test) (Table 6).

sample No. 9 (  
) sample No. 16 3.5 가 ,  
sample No. 16 3.3 가 .  
sample No. 16 가 3.5  
가 (Table 37).

가 가

가 ,

Table 37. Palatability test score.

Sample No.					Total score
1	3.1	3.7	3.2	3.2	13.2
2	2.9	3.2	2.9	3.2	12.2
3	3.0	3.2	3.1	2.9	12.2
4	3.1	3.4	3.2	3.2	12.9
5	3.2	3.3	2.8	3.4	12.7
6	2.8	3.0	2.8	2.8	11.4
7	2.9	3.4	2.8	2.9	12.0
8	2.4	3.2	2.8	2.6	11.0
9	3.5	3.5	3.0	3.0	13.0
10	3.0	3.7	3.3	3.3	13.3
11	2.8	3.7	2.7	2.7	11.9
12	3.3	3.7	2.7	3.0	12.7
13	3.0	3.5	2.7	2.8	12.0
14	3.1	3.7	2.7	3.2	12.7
15	3.3	3.8	3.0	3.3	13.4
16	3.5	3.7	3.3	3.5	14.0

## 4 .

1. , , , : 가  
 . , 28, 888 (1996)
2. , : Na-acetate Na-malate K-sorbate가 pH,  
 . , 20, 40 (1988)
3. , , , : 가.  
 , 23, 188 (1991)
4. , , , : 가 가  
 . 23, 183 (1991)
5. , : . , 305 (1967)
6. : Sorbic acid가 ascorbic acid .  
 , 1, 18 (1985)
7. : .  
 , 4, 39 (1988)
8. , : pH sodium malate buffer 가가  
 . , 17, 358 (1988)
9. : pH . , 14, 259 (1985)
10. , : 가가 . ,  
 21, 331 (1981)
11. , , , : .  
 , 27, 257 (1995)
12. , : Chitosan  
 가 . , 11, 309 (1995)

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(1987).

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