



Epidemiological survey of calf diseases, and
development of immunotherapy and diagnostic
method to calves with failure of immunoglobulin
passive transfer

Epidemiological survey of calf diseases and mortality

Diagnostic method for measuring colostrum transfer

Development of artificial colostrum and immunotherapy

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2000. 1. 25.

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1997	1999	3		676
			가 419 (62.0%)	, (257) .
			67.9% (459)	,
69.9%	(64.6%)		가	.
가	69.8%	가		, (22.6%)

(4.4%) . 가 46.0%

, (14.7%) (4.3%) .

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52.5% 가 ,

(35.2%) (2.5%) .

가 40.0% , 18.3% .

48.4% 35.6% ,

76.3% 55.7% .

59.1% 가 , (34.6%)

75.7% , 46.1% . 가

(46.5%) 73.9%

. 23.9% 11.8%

77.6% 61.9% 가 .

14.4% (3.4%) .

24.7% 가 ,

(21.4%), (17.2%),

(15.1%), (13.0%), (5.5%), (2.5%)

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가

. 가 66.7%

가 , (43.9%), (35.5%),

(33.9%) .

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1 52.5%

, 1-3 (25.1%) 4-6 (22.4%) 1

37.9% 가 , 1-3 18.1%, 4-6

13.8% 가 가 가

1-2 (41.1%)

가 1 44.3% 가

1-3 (26.1%) 4-6 (29.6%) 가

21.0- 29.0% 가

13.2- 18.3%

(31.5%) 가 (20.2%) 가

(p<0.05) , (26.8%) 가 (21.4%)

가 (19.5%) 가

(56.5%) 가

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(174) 가

가 (p<0.05).

, MCV, MPV 가 가

가 가

, 가

(241)

가 가

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147 98 (66.7%) 가

33.3% 가 , (16.7%), 가 (10.2%),

(0.7%)

(5.4%) 가 1-3

66.7- 73.1% , 4 100% 가 4

62.0%

(44.9%) 가

)

117 1-2

10 16 (13.7%)가 2 92 (78.6%)가

(43.78%) 가

(35.9%) , (16.2%)

59% 가

(52.1%)

65.8% (77/117)

Pasteurella spp.가 44.4% (52/117) 가 ,

29.9% BVDV(19), IBRV(18), PI-3V(14)

coronavirus(14)

Pasteurella spp. PI-3V IBRV 가 11

9 , *Pasteurella* spp. BRSV 7

(3)

(32) 가 . BVD(19), BCV (14),

(5), (5), (4) .

(4) (2) .

92 BCV 100%

64 가 78.3%

. BVDV 87% 50% 가 16 .

BEV 71.7% 8 가 89.4%

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) 가

(125) IBRV가

(84.8%) 32 가 72%

. PI- 3V BVDV 82.4%

PI- 3V 32 가 60.8%

BVDV 32 67.2% 가 . BEFV

57.6% 64 16- 64

.

) 가

9 305 가 .

가 40 가 53% 가

40 가 29%, 50 60 18% .

가 21- 40 가 30% 가 , 41- 60

가 25%, 61- 100 가 26%, 100 6% .

가 71%
 (9%) (5%)
 (43%)가 가 (35%) (11.53%)
 , , , .
 (13%) 가 , 10% ,
 (9.5%), (5.5%), (5.4%)
 2% . 가
 (44%) (35%) (10%), (5%),
 (4%) .

2.

97 9 99 9 6 258

. 258
 51 (19.8%) 13 (5.0%)
 가 가
 BCV
 가 96.1% 가 , BVDV(73.6%), IBRV(73.6%)
 PI- 3V(43.8%) 4
 FPT ZSTT 가 가 (P<0.01).
 FPT IgG 가
 TP, Alb, Glob . sRID IgG(< 1,000 mg/dl)
 FPT 20.5%(53)

45.3% 15.1% . TP(< 4.0 g/dl) FPT 29.8% (77) Glob(<2.0 g/dl) FPT 30.2% (78) . SSPT(<1+) FPT 13.1% (34) ZSTT(<1,000 mg/dl) 27.1% (70) . 6 FPT Alb Glob 2.0 g/dl , TP 4.0 g/dl , sRID IgG 1,000 mg/dl , SSPT +1 , ZSTT 1,000 mg/dl 가 . Alb FPT FPT 2.63 7.38 . FPT SSPT가 0.79 0.89 가 (P<0.01).

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, casein- N

. Holstein

. IgG

가 120

Ig 가 ,

Ig

6

가

Ig

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1 FPT

FPT

TP 5.8 ± 0.9 g/dl,

Alb 2.2 ± 0.5 g/dl, Glob 3.6 ± 0.5 g/dl, IgG 2.2 ± 0.5 g/dl .

가 BVDV 8.1 ± 1.0 log₂, BCV 6.4 ± 2.3 log₂, PI-3V 1.9 ± 1.7 log₂, IBRV 1.9 ± 1.7 log₂ . 1-7

23 , TP 4.5 g/dl , IgG 0.8 g/dl , SSPT가 1+ FPT , 가

가 11 (48%), 2가 가 6 (26%), 3가

6 (26%) . FPT 23 (7)

, (6)

5 3 . ,

2

. Kg 0.28 g , 0.16 g

1-7 1 .

13 2 (25%)

가 10 12 ,

2 3 15 .

FPT transfer factor [TF; mg/dl(IgG

) / Kg] 11.17 ± 1.27 , $5.46 \pm$

2.74 , 1.40 ± 0.21 . FPT

(P<0.01).

24 Ig , BVDV, BCV

가 (P<0.05).

FPT

7 FPT가 , 14 30 FPT

. FPT

CD2, CD4 가 (P<0.05).

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SUMMARY

Cattle industry in Korea has been confronting by new environment of world trade on beef and milk products which should be forced to overcome the low productivity and competition of the domestic management system comparing to those of some exporting countries. For the persistent establishment of cattle in the coming era, there may be considering many contributing factors associated with management, nutrition, breeding and diseases. Among them, the stable growth and supply of calves is most likely to be one of the high priorities for strengthening the productivity and performance in cattle industry. In this study, there was aimed at the understanding of the basic epizootiological information of calf diseases and mortality, the establishment of the diagnostic techniques to measure the status of failure of passive immunoglobulin transfer(FPT) from dams to calves under the field condition, the evaluation of artificial colostrum and immunotherapy using bovine immune sera.

1. Epizootiological survey of calf diseases and mortality

From Sep. 1996 to Sep. 1999, there was examined 676 calves with disease and carcasses including 419 Korean-native and 257 Holstein calves by microbiological and pathological means. Carcasses were consisted of 69.9% of Korean-native calves and 64.6% of Holstein calves among the calves to be required for diagnosis. Calf diseases and mortality were most frequently associated with alimentary disorders (69.8%), and followed by respiratory

(22.6%) and reproductive (4.3%) problems. Also, enteric diseases (46.0%) were most frequently involved to cause death, and the following orders were taken by respiratory (22.6%) and reproductive (4.4%) disorders. In addition, calf mortality was occurred by abomasal impaction, bacterial septicemia, enterotoxemia, Akabane disease, Chuzan disease, abortion, malformation and toxicosis.

Calf diseases were classified by bacterial (52.5%), viral (35.2%), and parasitic (2.5%) origin. There was up to 40.0% mortality of bacterial diseases, 35.2% of viral diseases, and 2.5% of parasitic diseases in calf disorders to be diagnosed. In Korean-native calf diseases, 48.4% was related to bacteria and 35.6% to viral agents. Mortality in Korean-native calves showed 76.3% by bacterial diseases and 55.7% by viral diseases. In Holstein calf diseases, 59.1% was involved with bacteria and 34.6% with some viruses. Mortality in Holstein calves represented up to 75.7% by bacterial infection and 46.1% by viral infection. Frequent bacterial disorders were colibacillosis(46.5%), pasteurellosis(23.9%) and salmonellosis(11.8%). Also, there were confirmed the outbreaks of bacterial septicemia(16.9%), enterotoxemia(6.2%) and campylobacteriosis(3.4%) in bacterial calf diseases. Frequent viral diseases were composed by rotavirus infection(24.7%), bovine viral diarrhea(21.4%), coronavirus infection(17.2%), bovine respiratory syncytial virus infection(15.1%), infectious bovine rhinotracheitis(13.0%), Akabane disease(5.5%), and Chuzan diseases(2.5%). Interesting results through this studies were obtained the first isolate to Chuzan virus and Inovirus in this country which could be promised the development for diagnostic method and vaccines. Mortality among viral diseases was 66.7%

by bovine viral diarrhea, 43.9% by BRSV, 35.5% by IBR, and 33.9% by rotavirus infection. Main parasitic diseases were coccidiosis and babesiosis. Malformations were consisted of anasarca, derodidymus, polymelia, humerus hypoplasia, and tracheal collapse. Calf diseases were mostly occurred in one-month-old(52.5%). Prevalence of it was 25.1% in two to three-month-old and 22.4% in four to six-month-old. In mortality, there were 37.9% in one-month-old, 18.1% in two and three-month-old, and 13.8% in four to six-month-old, respectively. The older the age of calf, the less the prevalence of calf enteric diseases. Respiratory diseases in Korean-native calves were more frequently occurred in one to two-month-old(41.4%). Seasonal prevalence and mortality of calf diseases were not a significant difference. In Korean-native calves, prevalence of diseases in summer(31.5%) was frequently occurred to compare that in winter(20.2%). Abortion and malformation in Korean-native calves were frequently occurred in spring.

Hematological examination to 174 calves with clinical signs showed mild to marked leukocytosis ; the tendency of it was appeared more significant in Korean-native calves than that in Holstein ones($p < 0.05$). In addition, there were slight increased in hematocrit, platelet, mean corpuscular volume and mean plasma volume, but all of those were included the higher level of normal ranges. Korean-native calves with respiratory signs were slight erythrocytosis. Two hundred forty one calves without clinical signs were not significant difference between breed and age in hematological values, but platelets were higher normal range in Korean-native calves than that in Holstein ones.

2. Viral-particle detection in feces of Korean-native calves with diarrhea

Presently, viral isolation in the diarrheal feces can be reached by many tools such as fluorescent antibody test, negative contrast electron microscopy(NCEM), neutralization, cell culture, and so on. The purpose of the study was to aimed at the establishment of simplified NCEM technique which can be efficiently applied for diarrheal feces and also the understanding on prevalence of viral-induced diarrhea in calves. One hundred forty-seven Korean native calves with diarrhea were examined to their feces by the modified NCEM. Among them, 98 cases(66.7%) were confirmed to have one or more viruses in their feces. The viruses detected were identified as rotavirus(33.3%), coronavirus(16.3%), togavirus(10.2%) and herpesvirus(0.7%). Ten cases of combined viral infection were consisted of 8 with rotavirus+coronavirus, one with rotavirus+togavirus and one with rotavirus+herpesvirus. Diarrheal types could classified by yellowish watery(44.9%), blood-tinged(19.7%), white watery(17.7%), brownish watery(14.3%), greenish watery(3.4%) diarrhea, respectively. Yellowish watery diarrhea(66cases) was frequently included rotavirus(31.8%), coronavirus(15.2%), and togavirus(13.6%), respectively. Consequently, these results suggest that the modified NCEM is reliable and efficient diagnostic tool for detection of viruses in the diarrheal feces and many calves rearing in Chonnam province have been exposed to some enteric viral agents mainly including rotavirus and coronavirus.

3. Diseases and mortality of Holstein calves with clinical signs

One seventy four Holstein calves with clinical signs were moved the experimental farm operated with the intensive management system in winter season. Among them, 92 cases(78.6%) died in the two months after the introduction. Main causes of their outbreaks were involved with respiratory(43.8%) and alimentary(35.9%) problems; most of the mortality was caused by respiratory(59.9%) and alimentary(52.1%) pathogens, and also showed more frequent prevalence(65.8%) related to combined infection. Principle pathogens to cause mortality were associated with *Pasteurella* spp.(44.4%), *E. coli*(29.9%), BVDV(16.2%), IBRV(15.4%), PI-3V(12.0%) and bovine coronavirus(12.0%). Mortality due to *Pasteurella* spp. was combined with IBRV(11 cases), PI-3V(9 cases), and BRSV(7 cases). In enteric diseases, colibacillosis was simultaneously occurred with BVD(19 cases), bovine coronavirus infection(14 cases), salmonellosis(5 cases), coccidiosis(5 cases) and clostridial infection(4 cases). Ninety two cases to death were appeared 100% with the neutralizing antibodies including 78.3% with it more than \log_{26} against bovine coronavirus, 50% with it higher than \log_{25} to BVDV, and 71.7% with it including 89.4% cases having it lower than \log_{23} to bovine ephemeral fever virus

4. Prevalence of neutralizing antibodies to some viruses in Korean-native calves with diseases

One hundred twenty five Korean-native calves were 84.8% cases with the antibody to IBRV including 72% cases with the antibody titer lower than \log_{25} . The antibodies higher than \log_{25} to PI-3V and BVDV were 60.8% and

67.2% cases, respectively. There was 57.6% cases having the antibody level lower than log₂5 to BEFV.

5. General survey on diseases and management in Korean- native cattle farms

The survey was included 350 farms located in 9 provinces in Korea during the middle of 1997. Owners to the farms mostly belonged to the age of 40's(53%) and these rest was classified as 29% younger than 40's, 18% in 50's and 18% in 60's, respectively. The farms to be examined were mostly held less than 60 cattle(55%) and 6% of them only had more than 100 cattle. Feedstuff as raw materials was mostly occupied by straw(71%) and also supplied by silage(99%) and corn(5%) as well. Main diseases in the field were enteric disorders(43%) with diarrhea, respiratory problem(35%) and skin diseases(11.5%). In addition, cattle was frequently suffering by tympany, reproductive problems, urolithiasis and laminitis. Provincial prevalence of enteric problem a farm was appeared the highest in Kyongpuk(13%) and the rest were less than 10%. Provincial prevalence of respiratory diseases was also the highest in Kyongpuk(9.5%) and then followed by Kyongnam(5.5%) and Chungnam(5.4%). Chonbuk and Cheju of it were less than 2%. Owners had recognized that enteric(44%) and respiratory(35%) diseases were most frequently involved in minimizing the performance and productivity of cattle. There was correlated between facility pattern of premise and the outbreak of respiratory diseases ; to make the cement bottom in a shed was to decrease the prevalence of respiratory distress.

6. Establishment of diagnostic methods on FPT to Korean-native calves

For making the criteria and selection of diagnostic techniques to failure of passive immunoglobulin transfer (FPT) in Korean-native calves younger than 6-week-old, 258 sera were examined by spectrophotometry for total protein (TP), albumin (Alb) and Globulin (Glob), sodium sulfite precipitation test (SSPT), zinc sulfate turbidity test (ZSTT), and single radial immunodiffusion test (sRID). During 8 weeks after birth all calves were clinically monitored at the daily level and showed various morbidity(19.8%) as well as mortality(5.0%) due to many diseases mainly including enteric and respiratory origin. Prevalence of seropositives to bovine viral diarrhea virus, bovine coronavirus, bovine infectious rhinotracheitis virus, and parainfluenza type-3 virus in Korean-native calves was examined, showed the highest rate(96.1%) to BCV, and followed those to BVDV(73.6%), IBRV(73.6%) and PI-3V(43.8%) in decreasing orders. There was significant relationship($P < 0.01$) between the seronegatives to viral agents tested and the ZSTT or sRID test. Risk factors to be able to expose the diseases in the calves with FPT were increased up to 2.63-7.38 times than the control. Calves with FPT were classified as 29.8%(78/258) when the cutoff point of TP was determined less than 4.0 g/dl and among them the morbidity and mortality during the experiment was 35.1% and 10.4%, respectively. it was 30.2%(78/258) when the cutoff point of Glob was made less than 2.0g/dl and among them the morbidity and mortality was 24.6% and 10.3%, respectively. There was 13.1% (34/258) when the cutoff point of SSPT was under the degree of 1+ and among them the morbidity and mortality was 79.4% and

29.4%, respectively. it was 20.5% (53/258) when the cutoff point of IgG with sRID was 1,000 mg/dl and among them the morbidity and mortality was 45.3% and 15.1%, respectively. Also, mean concentration of IgG in calves tested by sRID test was 2,165 mg/dl at 9-day-old as the highest level, but dropped down to 1,140 mg/dl at 15-day-old. Based on sensitivity and specificity of each test to be evaluated, the cutoff point of FPT for Korean-native calves was made as 1,000 mg/dl by sRID and ZSTT, less than 4.0 g/ dl by TP, less than 2.0 g/dl by Alb and Glob, below 1+ by SSPT. SSPT was the highest sensitivity(0.79) and specificity(0.89) compared to the others. Consequently, the results of the study were suggested that SSPT can be useful as the diagnostic tool with relatively reliable efficacy and convenience for detecting FPT cases of Korean-native calves under the field condition.

7. Development of artificial colostrum

By general analysis, colostrum to be tested was higher concentrations in protein, total solid components, casein-N than those in milk. The colostrum produced from Korean-native dams showed higher in all main components than that in Holstein ones. The levels of immunoglobulin and protein in colostrum were paralleled to increase the parity of dams. However, milk taken at 120 hours after parturition was similar the levels of them to the normal one. To make a artificial colostrum is strongly recommended to add immunoglobulin and anti-diarrheal components. The formula obtained in the study was described. Within 6 hours after feeding of artificial colostrum, concentrations of immunoglobulins in the recipient's blood had been markedly

decreased. Consequently, the results were suggested that the supply of the enough and qualified colostrum to neonatal calves as suggested the formula is critical to obtain the positive effect immunologically for establishing the resistant ability to pathogens after birth.

8. Studies on immunotherapy to calves with failure of immunoglobulin passive transfer using bovine immune sera

The efficacy of bovine immune sera to correct the calves with failure of passive transfer (FPT) was evaluated. Immune sera used were produced from 14 one-year-old Holstein cattle which were inoculated commercial combined viral vaccine, administered by the challenge of some main enteric or respiratory viruses, aseptically filtered and stored at 4 °C before used. Serum IgG concentrations of donor calves were calculated by single radial immunodiffusion assay and were ranged from 1750 mg/dl to 2700 mg/dl during the experiment. The serum total protein (TP) concentrations of donor calves were ranged from 4.9 g/dl to 6.7 g/dl and the serum neutralizing antibody titers specific to bovine viral diarrhea virus and bovine coronavirus were $8.1 \pm 1.0 \log_2$ and $6.4 \pm 2.3 \log_2$, respectively. FPT calves were selected in the case of at least one of the three options ; serum IgG concentration was less than 0.8 g/dl, TP concentration was less than 4.5 g/dl, and sodium sulphate precipitation test was less than +1. Twenty-three calves with FPT younger than 7 days of age were used as the recipient and monitored two months after the administration of bovine immune sera to be produced. Hyperimmune bovine sera used were administered into seven Korean-native FPT calves (0.16 ± 0.06 g of Ig/kg of body weight) by intraperitoneal

route(K-IP group), six Holstein FPT calves(0.28 ± 0.17 g of Ig /kg of body weight) by intravenous route(H-IV group). Controls were allocated as five Korean-native FPT calves to K-IP group and three Holstein FPT calves to H-IV group without any treatment of immune sera. In addition, two Korean-native FPT calves with recumbency(K-IV group) was intravenously treated with the bovine immune sera. After the treatment of bovine immune sera, Mean transfer factor(mg/dl of IgG administered/kg of body weight) was 5.46 ± 2.74 in K-IP group, 11.17 ± 1.27 in H-IP group, and 1.40 ± 0.21 in K-IV group. The corrective effect of bovine immune sera to FPT calf without any clinical signs showed that intravenous route was more effective than intraperitoneal administration($P < 0.01$). FPT calves with severe signs were not effective response to the immunotherapy used and consequently died within 10 days after the treatment. Ten percentage of controls appeared the clinical signs including diarrhea. On contrary, there were not any clinical signs in K-IP and H-IV group. There was significant increase of the neutralizing titer against bovine viral diarrhea virus and bovine coronavirus as well as of cell population including CD2, CD4, and monocytes in K-IP and H-IV group after the immunotherapy($P < 0.05$). Also, K-IP and H-IV group showed the successful correction to FPT within one week after the immunotherapy, but controls had kept the FPT two four weeks even after the same treatment. Consequently, the results were suggested that the bovine immune sera could be used the corrective and efficient material to calves with FPT.

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(Curtis,).

(Bunger , 1979 ; Martin , 1990), (Simensen, 1983 ; Correa

, 1988), (Britney , 1984 ; Correa , 1988 ; Curtis ,

1989), (Simensen, 1983) .

가 , 가 (Lovell Hill 1940 ; Debnath , 1990), (Speicher Hepp, 1973) . 가 , 가 (Martin , 1990). 가 가 , 가 가 (Webster , 1985), 4 가 (Schumann , 1990), (Martin Meek, 1986). 가 (Speicher Hepp, 1973; Marin , 1975b ; Simensen Norheim, 1983b). 가 .

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(Sanford, 1984). ,

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(Webster , 1985; , 1999). 6

(Martin Meek, 1986; , 1999).

bovine rotavirus(BRV), bovine coronavirus(BCV), bovine viral diarrhea virus(BVDV) *E. coli*, *Salmonella* spp, *Clostridium* spp, *Campylobacter* , *Eimeria* spp, *Isospora* spp, *Cryptosporidium* spp (Britney , 1984; Himura , 1985; Holland, 1990; Barker , 1993).

bovine respiratory syncytial virus(BRSV), infectious bovine rhinotracheitis virus(IBRV), parainfluenza type 3 virus(PI-3V), bovine adenovirus, *Pasteurella multocida hemolytica*, *Mycoplasma* spp. (, 1991; Dungworth, 1993; Graham , 1998).

(Oxender , 1973; Speicher Hepp, 1973; Tsuda, 1999).

(Simensen, 1983c; Schumann, 1990; Smith , 1998).

4) :

5) Negative

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가 2 3 -70 , 10

1 2 g

PBS(pH 7.4) 1 ml 3,000 rpm 10

800 $\mu\ell$

600 $\mu\ell$

(70Ti) PBS(pH 7.4) 20 M ℓ 27,500 rpm 4

(900- XL, Beckman)

grid(400 mesh) amyl acetate 2% silicagel

PBS(pH 7.4) 1 M ℓ

grid 10 $\mu\ell$ 3

grid 2%

phosphotungstic acid(PTA) 10 $\mu\ell$ 13 20

grid 37 1

grid

(JEM- 2000 FX II, Jeol) 가 80 KV

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6) :

Giemsa

7) :
 10ml
 (EDTA)가
 (Medonic CA530, Oden, Sweden),
 Vacutainer™(Becton dickinson)

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Hematoxylin & Eosin (HE)

1) : 1998 10 1 117
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3) : 117 82 bovine ephemeral virus(BEV),
 bovine viral diarrhea virus(BVDV), bovine coronavirus(BCV)

1) 1997 1999 125 1998

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가) :
. BVDV NADL , BEFV Tong-re (), IBRV PQ7 ,
PI-3V NADL Chuzan, Akabane virus OBE-1 ,
Ibaraki virus Imaizumai .
) : Marden-Darby bovine
kindey(MDBK) .
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2 (200 TCID₅₀/ml) 37cC
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Student's t test .

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1997- 1999	676
Table 2- 1	
가 62.0% (419)	(257)
(64.6%)	가 (69.9%)가
67.9%	69.8%
	22.6%
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가 46.0%	
(14.7%)	
(4.3%) (2.8%)	72.8% 가
	(17.4%)
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4.5- 4.8%	
71.7%	15.4%
6.5%	
	65.0%
	14.7% , 4.3%
	39.3% 가

21.8% .
 60.8% 가 , 가 33.7%
 , 5.4% .
 60.3% 68.9%
 95.1%
 가 79.0% 가 , 60.4%

Table 2-1. Prevalence of calf diseases and mortality by body system from 1996 to 1999

System	Disease	Korean- navive calves	Holstein calves	Mortality (%)
Enteric	Collibacillosis+complication	79/107*	43/58	122/165(73.9)
	Rotavirus infection+ complication	11/31	9/28	20/59 (33.9)
	Bovine viral diarrhea	25/34	9/17	34/51 (66.7)
	Salmonellosis	19/28	7/14	26/42 (61.9)
	Coronavirus infection + complication	11/25	7/16	18/41 (43.9)
	Abomasal impaction	24/28	8/8	32/36 (100)
	Unclassified septicemia	18/18	11/11	29/29 (100)
	Unclassified enterotoxemia	16/16	6/6	22/22 (100)
	Coccidiosis with complication	2/10	0/5	2/15 (13.3)
	Campylobacteriosis	4/8	1/4	5/12 (41.7)
	<i>Subtotal</i>	210/305	101/167	311/472(65.9)
Respiratory	Pasteurellosis	19/26	47/59	66/85 (77.6)
	Bovine respiratory syncycial pneumonia	15/23	7/13	22/36 (61.1)
	Infectious bovine rhinotracheitis	9/23	2/8	11/31 (35.5)
	Tracheal collapse	1/1	0/0	1/1 (100)
	<i>Subtotal</i>	44/73	56/80	100/153(65.4)
Reproductive	Akabane disease	7/7	6/6	13/13 (100)
	Abortion & malformation	8/8	2/2	10/10 (100)
	Chuzan disease	5/5	1/1	6/6 (100)
	Aino virus infection	0/1	0/0	0/1 (0.0)
	<i>Subtotal</i>	20/21	9/9	29/30(96.7)
Others	Toxicosis	19/19	0/0	19/19 (100)
	Endoparasitic infection with complication	0/1	0/1	0/2 (0.0)
	<i>Subtotal</i>	19/20	0/1	19/21(90.5)
Total		293/419(69.9)	166/257(64.6)	459/676(67.9)

*No. of death/No. of diagnosed.

Table 2-2

52.5% 가 , 35.2%
2.5% (Figure 2-1).
40.0% ,
18.3% .

35.6% 48.4% ,
76.3% 55.7%
(Figure 2- 2). 59.1% 가
, (34.6%) , 2.3%
(Figure 2- 3). 75.7% ,
46.1% .
(Figure 2- 4 6)
46.5% , 73.9%
. (74.1%) (52.7%)
73.9% .
23.9% 11.8% ,
77.6% 61.9% .
(79.7%) (73.1%) (67.9%)가
(50%) . 가
14.4% (21.6%)가
(11.2%) .
3.4% 41.7% 가 . (Figure 2- 7,8)
24.7% 가 ,
(21.4%), (17.2%),
(15.1%), (13.0%), (5.5%), (2.5%)
1 가 .
가 . 가
66.7% ,
43.9% 35.5% ,

33.9% . 100%

가 .

13.3%

4 (5.3%)가 (3.1%)

(6.7%)

(4.5%)

Table 2-2. Prevalence of calf diseases and mortality by pathogen from 1996 to 1999

Pathogen	Causative agent	Korean-native calves	Holstein calves	Mortality (%)
Bacteria	<i>E. coli</i>	79/107*	43/58	122/165 (73.9)
	<i>Pasteurella</i> spp.	19/26	47/59	66/85 (77.6)
	<i>Salmonella</i> spp.	19/28	7/14	26/42 (61.9)
	Unclassified septicemia	18/18	11/11	29/29 (100)
	Unclassified enterotoxemia	16/16	6/6	22/22 (100)
	<i>Campylobacter</i> spp.	4/8	1/4	5/12 (41.6)
	Subtotal	155/203	115/152	270/355 (76.1)
Virus	Rotavirus	11/31	9/28	20/59 (33.9)
	BVDVa	25/34	9/17	34/51 (66.7)
	Coronavirus	11/25	7/16	18/41 (43.9)
	BRSVb	15/23	7/13	22/36 (61.1)
	IBRVc	9/23	2/8	11/31 (35.5)
	Akabane virus	7/7	6/6	13/13 (100)
	Chuzan virus	5/5	1/1	6/6 (100)
	Ainovirus	0/1	0/0	0/1 (0.0)
	Subtotal	83/149	41/89	124/238 (52.1)
Parasite	<i>Eimeria</i> spp.	2/10	0/5	2/15 (13.3)
	<i>Babesia</i> spp.	0/1	0/1	0/2 (0.0)
	Subtotal	2/11	0/6	2/17 (11.8)
Other	Abomasal impaction	25/28	8/8	33/32 (91.7)
	Toxicosis	19/19	0/0	16/16 (100)
	Abortion & malformation	8/8	2/2	10/10 (100)
	Tracheal collapse	1/1	0/0	1/1 (100)
	Subtotal	53/56	10/11	63/66 (91.5)
Total		293/419	166/257	459/676 (67.9)

*No. of death / No. of diagnosed.

aBovine viral diarrhea virus.

tBovine respiratory syncytial virus.

cnfectious bovine rhinotracheitis virus.

(1.6%) (2.1%)가 (0.8%)
(100%) .
, , .

Table 2-3 (Figure 2-1
9) (Figure
2-10 12) 1 가 52.5% ,
1-3 (25.1%) 4-6 (22.4%) (p<0.05).
(Figure 2-13) 1 37.9% 가 , 1-3 가
18.1%, 4-6 13.8% 가 가
(p<0.05). 가
1-3 41.1% 가 . 1
가 80.0% 가 (p<0.05),
9.5% 8.2% . 1-3 4-6
가 65.7% 63.8% , 가 28.6%
26.6% .
1 44.3% 가
(p<0.05), 4-6 29.6%, 1-3 26.1% 가
(Figure 2-14) 61.1- 65.8%
가 . 1 1-3 79.8%
65.7% 가 4-6
(61.8%)가 (38.2%) (p<0.05).
1 7.9% 가 .

Table 2-3. Prevalence of calf diseases and mortality by age during 1997- 1999

Breed	System	Age(months)			
		<1	1- 3	4- 6	Total
Ka	Enteric	121/176c	50/69	39/60	210/305
	Respiratory	14/18	20/30	10/25	44/73
	Reproductive	20/21	0/0	0/0	20/21
	Others	4/5	6/6	9/9	19/20
	Subtotal	159/220	76/105	58/94	293/419
Hb	Enteric	59/94	25/44	17/29	101/167
	Respiratory	7/11	16/22	33/47	56/80
	Reproductive	9/9	0/0	0/0	9/9
	Others	0/0	0/1	0/0	0/1
	Subtotal	75/114	41/67	50/76	166/257
Total		234/334	117/172	108/170	459/676

Table 2- 4

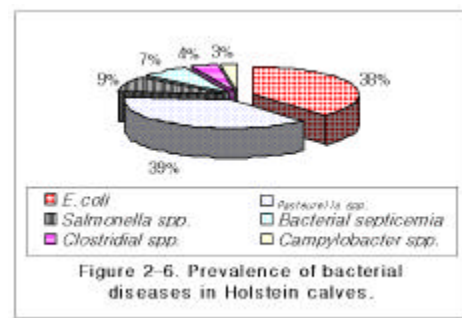
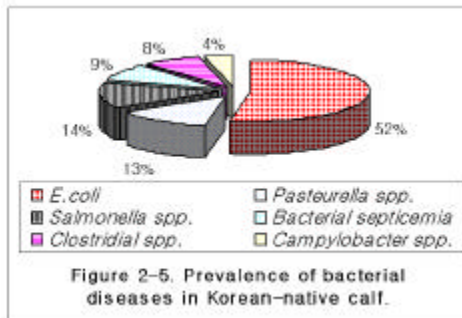
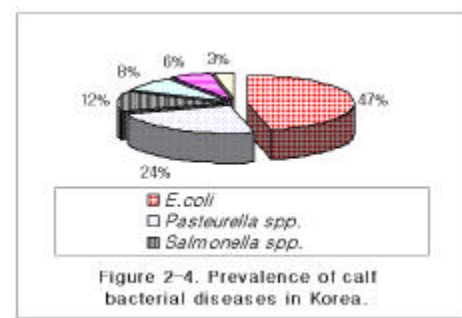
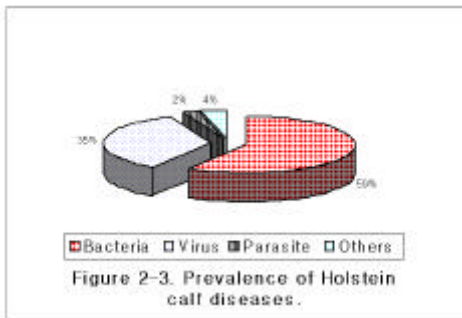
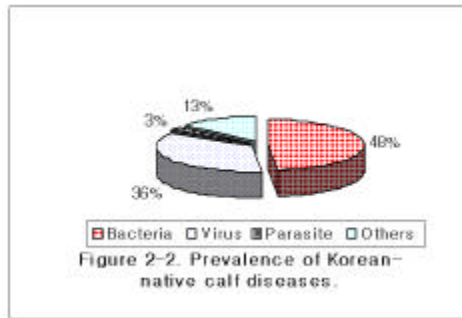
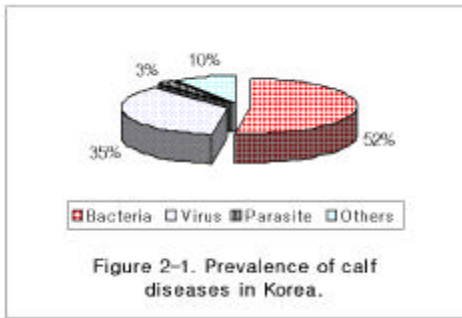
(29.0%), (27.2%), (22.8%), 가 (21.0%)
 , (21.9%), (18.3%), (14.5%), 가 (13.2%)
 . (31.5%) , (27.4%), 가
 (20.8%), (20.2%) (25.1%), (18.9%),
 (13.4%), 가 (12.6%) (p<0.05). 가
 79.5% 78.2%
 가 (19.5%) (7.6%)
 (11.3%)
 5.2% 9.8%
 (26.8%), (26.8%), (24.9%),
 가 (21.4%) , (17.5%), (16.7%), (16.3%),
 가 (14.0%) 가 ..
 (85.9%) 가 , 가

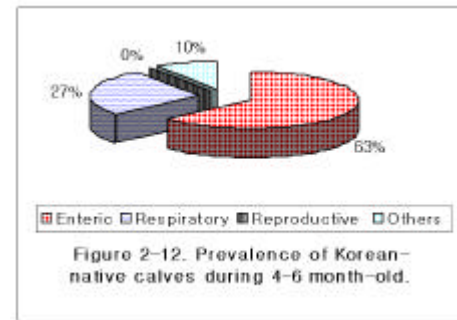
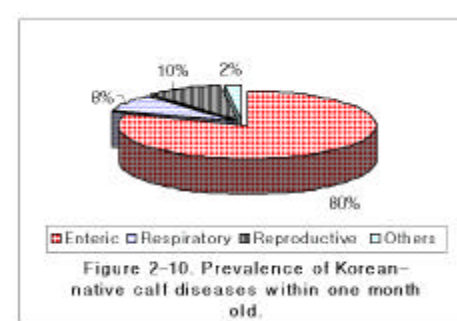
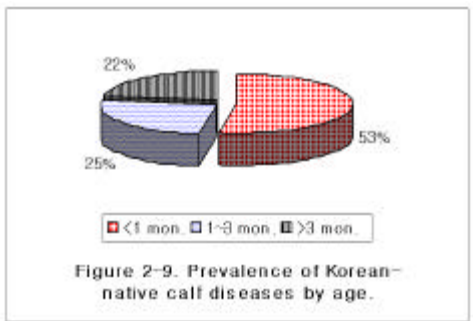
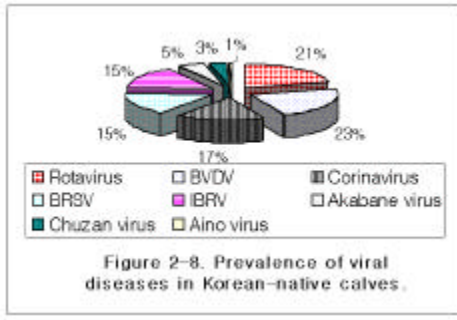
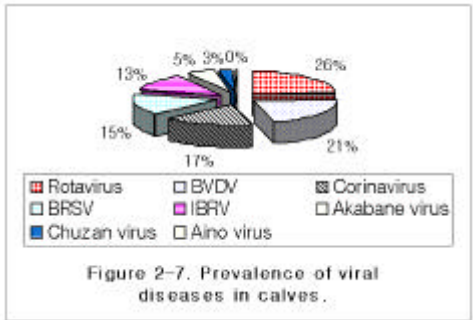
(67.3%) (66.7%) (42.0%)
 (57.8%) 가 , (40.5%), 가 (38.2%),
 (21.7%) (56.5%) 가
 (32.7%), (26.1%), (7.8%) , (37.7%), 가 (27.3%),
 (17.4%), (4.7%) (7.2%) (4.7%)

Table 2-4. Seasonal prevalence of calf diseases and mortality during 1997- 1999

Breed	System	Season				
		Spring	Summer	Autumn	Winter	Total
Korean native	Enteric	48/76*	84/105	42/68	36/56	210/305
	Respiratory	12/20	4/10	11/18	18/26	45/74
	Reproductive	13/13	4/4	1/1	2/3	19/20
	Others	6/6	13/13	0/1	0/0	19/20
	Subtotal	79/115	105/132	53/87	56/85	293/419
Holstein	Enteric	28/46	37/55	21/37	15/29	101/167
	Respiratory	12/18	3/5	15/18	26/39	56/80
	Reproductive	5/5	3/3	0/0	1/1	9/9
	Others	0/0	0/1	0/0	0/0	0/1
	Subtotal	45/69	43/64	36/55	42/69	166/257
Total		124/184	148/196	89/142	98/154	459/676

*No. of death/No. of diagnosed.





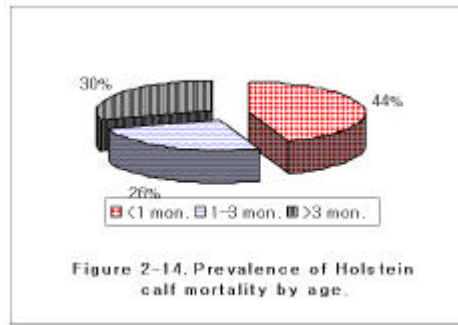


Figure 2-14. Prevalence of Holstein calf mortality by age.

174 (Table 2-5)

231 (Table 2-6)

가

가

($p < 0.05$).

, MCV MPV

가

가

가 ($p < 0.05$)

, MCHC

Table 2- 5. Hematological profile of calves with clinical signs*

Breed	Disease	RBC ^a (10 ⁶ /mm ³)	WBC ^b (10 ³ /mm ³)	HCT ^c (%)	HGB ^d (g/dℓ)	PLT ^e (10 ³ /mm ³)	MCV ^f (fl)	MCH ^g (pg)	MCHC ^h (g/dℓ)	MPV ⁱ (μm ³)
K ^{**}	Respiratory (n=29)	10.24 ^j ± 1.01	20.27 ± 8.80	30.18 ± 11.18	11.47 ± 1.47	182.17 ± 74.34	33.88 ± 3.33	12.18 ± 2.39	33.12 ± 1.15	4.08 ± 0.08
	Enteric (n=55)	8.30 ± 3.18	20.77 ± 7.13	23.27 ± 17.25	9.90 ± 3.76	376.00 ± 217.73	35.67 ± 2.01	12.20 ± 0.36	33.63 ± 2.07	5.07 ± 1.76
	Subtotal (n=84)	8.87 ± 3.18	20.36 ± 7.82	26.63 ± 12.89	10.13 ± 3.56	225.23 ± 219.22	34.59 ± 3.74	12.20 ± 3.41	33.35 ± 9.77	4.59 ± 1.58
	Respiratory (n=24)	8.12 ± 1.22	8.95 ± 3.46	29.35 ± 4.94	8.86 ± 1.13	762.90 ± 339.86	36.18 ± 4.46	10.99 ± 0.66	30.64 ± 3.52	9.08 ± 2.94
H ^{***}	Enteric (n=66)	7.32 ± 1.34	16.62 ± 5.88	16.62 ± 5.88	19.54 ± 8.75	530.61 ± 430.08	35.81 ± 6.91	11.47 ± 1.99	30.54 ± 4.23	11.15 ± 5.07
	Subtotal (n=90)	7.62 ± 1.35	13.78 ± 6.34	27.81 ± 5.06	15.59 ± 7.28	616.64 ± 413.91	35.94 ± 6.09	11.29 ± 1.63	30.57 ± 3.97	10.38 ± 4.50
Total (n=174)		8.11 ± 1.75	15.43 ± 7.21	27.87 ± 7.72	14.43 ± 4.36	524.18 ± 208.11	35.58 ± 5.47	11.52 ± 1.72	30.57 ± 3.69	10.89 ± 4.70

^aDiarrhea, salivation, depression, emaciation, coughing, decreased appetite, and so on.
^{**}Korean-native calves. ^{***}Holstein calves. ^aRed blood cell. ^ℓWhite blood cell. ^cHematocrit.
^dHemoglobin. ^ePlatelets. ^fMean corpuscular volume. ^gMean corpuscular hemoglobin. ^hMean
corpuscular hemoglobin concentration. ⁱMean plasma volume. ^jMean ± SD.

가 ,

가

(Table 2- 6).

Table 2- 6. Hematological profile of normal calves

Breed	Sex	RBCa (10 ⁶ /mm ³)	WBCb (10 ³ /mm ³)	HCTc (%)	HGBd (g/dl)	PLTe (10 ³ /mm ³)	MCVf (fl)	MCHg (pg)	MCHCh (g/dl)	MPVi (μ m ³)
K*	(n=121)	8.84j ± 2.0	8.99 ± 2.96	27.89 ± 7.22	12.18 ± 2.96	156.90 ± 76.60	31.90 ± 3.41	11.03 ± 2.18	33.68 ± 2.48	4.18 ± 0.74
	(n=52)	8.60 ± 2.12	9.70 ± 4.50	28.43 ± 7.76	10.46 ± 3.03	133.19 ± 61.40	32.76 ± 5.09	13.51 ± 2.65	33.47 ± 5.70	4.02 ± 0.51
	Subtotal (n=173)	8.87 ± 1.88	9.34 ± 3.43	28.45 ± 6.84	11.7 ± 4.9	154.21 ± 89.53	32.31 ± 4.0	11.8 ± 9.9	33.6 ± 3.7	4.1 ± 0.7
H**	(n=46)	8.06 ± 1.24	10.75 ± 3.17	28.34 ± 4.26	8.65 ± 1.38	719.13 ± 321.39	34.80 ± 6.33	10.78 ± 1.96	30.84 ± 4.01	9.76 ± 4.43
	(n=22)	7.81 ± 2.55	8.39 ± 4.56	25.63 ± 9.14	8.19 ± 2.83	698.68 ± 563.28	32.50 ± 3.05	10.37 ± 0.66	32.26 ± 4.24	8.92 ± 4.78
	Subtotal (n=68)	7.98 ± 1.75	9.99 ± 3.81	27.46 ± 6.33	8.50 ± 1.96	712.51 ± 411.00	34.06 ± 5.57	10.65 ± 1.66	31.30 ± 4.11	9.49 ± 4.53
Total (n=241)		8.62 ± 1.89	9.53 ± 3.54	28.2 ± 6.7	10.77 ± 3.98	308.67 ± 239.62	32.83 ± 4.20	11.5 ± 8.67	33.1 ± 3.52	5.67 ± 3.44

*Korean- native calves. **Holstein calves. aRed blood cell. bWhite blood cell. cHematocrit. dHemoglobin. ePlatelets. fMean corpuscular volume. gMean corpuscular hemoglobin. hMean corpuscular hemoglobin concentration. iMean plasma volume. jMean ±SD.

1)

147 가 66.7% (98
) , (33.3%) (16.3%)가
, 가 10.2% (15) .
가 5.5% (Table 2- 7).

Table 2- 7. Detection of viral-like particles in the feces collected from 147 Korean-native calves with diarrhea

Virus	No. detected(%)
Rotavirus	49(33.3)
Coronavirus	24(16.3)
Togavirus	15(10.2)
Rotavirus + Coronavirus	8(5.4)
Rotavirus + Togavirus	1(0.7)
Rotavirus + Herpesvirus	1(0.7)
Not detected	49(33.3)
Total	147(100.0)

2)

65- 75 nm RNA genomic
 RNA가 가
 가 60- 70 nm

(Figure 2- 15).

80- 160 nm RNA 가
 110- 130 nm
 가 (Figure 2- 16).
 120- 200 nm DNA
 가 130 nm

양이 관통된 전형적인 형태를 띠고있고 외피주변에 털과 같은 특징적인 구조물이 관찰되었다(Figure 2-17).

단일나선 RNA바이러스인 토가바이러스는 40-70 nm 정도의 크기로 전자현미경상에서 원형 혹은 불규칙한 형태로 주로 비어있는 형태로 관찰되었다. 이 실험에서 관찰된 토가바이러스는 약 50 nm 정도의 크기로 난원형의 공복형과 불규칙형 등 여러 가지 형태로 관찰되었다(Figure 2-18).

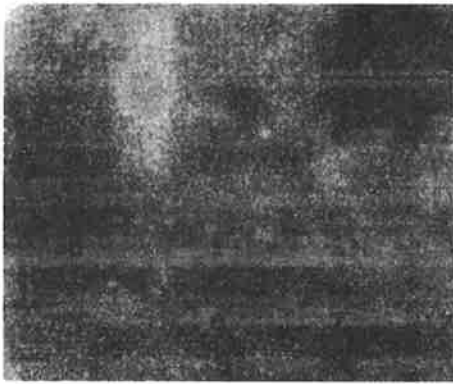


Figure 2-15. Electron photomicrograph of rotavirus-like particles in feces of Korean-native calf with diarrhea. Phosphotungstic acid(PTA) stain. Bar = 50 nm.

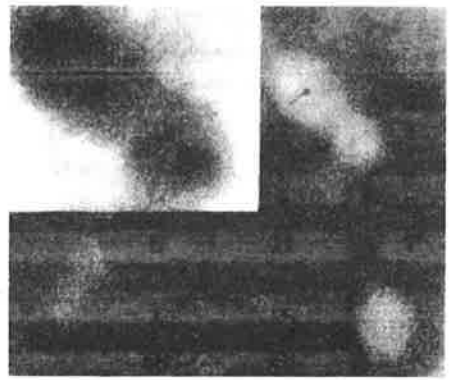


Figure 2-16. Electron photomicrograph of coronavirus-like particles in feces of Korean-native calf with diarrhea. PTA stain. Bar = 100 nm.

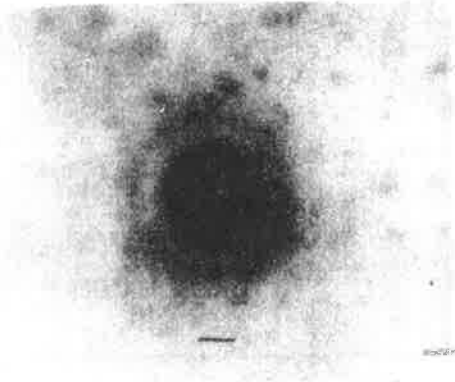


Figure 2-17. Electron photomicrograph of herpesvirus-like particles in feces of Korean-native calf with diarrhea. PTA stain. Bar = 50 nm.

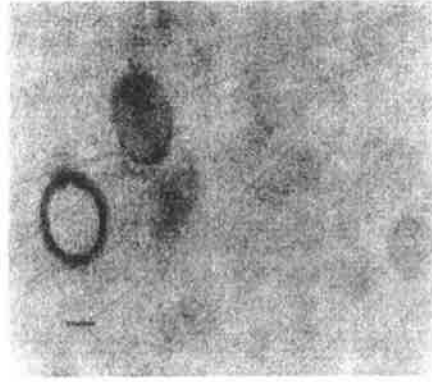


Figure 2-18. Electron Photomicrograph of togavirus-like particles in feces of Korean-native calf with diarrhea. PTA stain. Bar = 100 nm.

3) 설사 송아지 일령별 분변내 바이러스 동정

설사 송아지의 일령별 바이러스 검색률을 알아본 결과(Table 2-8), 1주일령의 경우 19마리의 분변에서 바이러스가 분리되었고(73.1%) 이들은 주로 로타바이러스와 코로나바이러스였다. 2주일령과 3주일령 이상의 경우 각각 10두(66.7%)와 16두(69.6%)의 송아지에서 바이러스가 분리되었으며 1주일령과 비슷한 바이러스 분포를 보이고 있었다.

Table 2-8. Prevalence of viral-like particles in the feces of 147 Korean native calves by age

Age(weeks)	No. examined	No. detected(%)
1	26	19(73.1)
2	15	10(66.7)
3	23	16(69.6)
4	4	4(100.0)
>4	79	49(62.0)

*(No. detected/No. tested) × 100

4)

Table

2-9 가 44.9%(66) 가
, (31.8%)
(15.2%), 가 (13.6%)
가 가 +
(7) + (1)
가 17.0%
,
(p<0.01). + 가
(1) +
(1)가 .

Table 2-9. Correlation between the kinds of diarrhea and the viral-like particles in the feces of 147 Korean native calves with diarrhea

Diarrhea	No. detected(%)	Viruses detected
yellow	44.9	rotavirus coronavirus togavirus rotavirus+coronavirus rotavirus+herpesvirus
bloody	19.7	rotavirus coronavirus togavirus rotavirus+togavirus
white	17.0	rotavirus coronavirus togavirus
brown	14.3	rotavirus coronavirus togavirus rota+togavirus
green	3.4	rotavirus coronavirus

117 가 (43.78%) 가
 (35.9%) ,
 16.2% . 2 92
 (78.6%)가 10 가 16 (13.7%)
Pasteurella spp.
 가 가
 가
 59% 52.1% 95% 가

(Table 2- 11).

65.8% (77/117)	<i>Pasteurella</i>
spp.가 44.4% (52/117) 가	
29.9% (35/117) .	BVDV(19), IBRV(18), PI-3V(14)
coronavirus(14) .	
	가
	PI-3V 가 11 가
<i>Pasteurella</i> spp.	<i>Pasteurella</i> spp. IBRV
가 9 .	, <i>Pasteurella</i> spp. BRSV
7 .	<i>Pasteurella</i> spp.
(9)	(43) . <i>Pasteurella</i> spp.
	IBRV (4)
(14)	가 . 61
	가 35 가 (3)
	(32) 가 . BVD(19), BCV
(14),	(5), (5), (4)
.	(4) (2)
.	

Table 2-11. Classification of Holstein calf diseases in intensively rearing system

Infection	Pathogen	No. of occurred	%
Enteric	Bovine viral diarrhea virus(BVDV)	2	1.7
	Bovine coronavirus(BCV)	3	2.5
	Bovine rotavirus(BRV)	2	1.7
	<i>E.coli</i>	3	2.5
	<i>Clostridial</i> spp.	4	3.4
	<i>Salmonella</i> spp.	2	1.7
	<i>Eimeria</i> spp.	1	0.9
	Subtotal	17	14.4
Enteric complex	BVDV + BCV	2	1.7
	BVDV + BRV	1	0.9
	<i>E.coli</i> + BCV	3	2.5
	<i>E.coli</i> + BVDV	4	3.4
	<i>E.coli</i> + BRV	3	2.5
	<i>E.coli</i> + <i>Eimeria</i> spp.	1	0.9
	BVDV + <i>Eimeria</i> spp.	1	0.9
	<i>E.coli</i> + <i>Salmonella</i> spp.	2	1.7
	BVDV + BRV + BCV	2	1.7
	BVDV + BCV + <i>Eimeria</i> spp.	1	0.9
	<i>E.coli</i> + BVDV + BCV	2	1.7
	<i>E.coli</i> + BVDV + BRV	1	0.9
	<i>E.coli</i> + BCV + BRV	1	0.9
	<i>E.coli</i> + BCV + BRV + <i>Eimeria</i> spp.	1	0.9
Subtotal	25	21.5	
Respiratory	<i>Pasteurella</i> spp.	9	7.7
	Infectious bovine rhinotracheitis virus	4	3.4
	Bovine respiaroty syncytial virus	2	1.7
	<i>Mycoplasma</i> spp.	2	1.7
	Subtotal	17	14.5
Respiratory complex	<i>Pasteurella</i> spp + Parainfluenza-3 virus	11	9.4
	<i>Pasteurella</i> spp. + IBRV	9	7.7
	<i>Pasteurella</i> spp. + BRSV	7	6.0
	<i>Pasteurella</i> spp. + IBRV + BRSV + PI-3V	1	0.9
	<i>Mycoplasma</i> spp. + <i>Pasteurella</i> spp.	2	1.7
	<i>Mycoplasma</i> spp. + PI-3V	2	1.7
	<i>Mycoplasma</i> spp. + IBRV	1	0.9
	Subtotal	33	28.3

(continued)

42 (35.9%) .
 16.2% 가
 6 (5.1%)가 . 가
 117 *Pasteurella* spp.가 52 가
Pasteurella spp.
 가 39 .
 25 ,
 33 , 19 117
 77 가 .
 PI-3V, IBRV, BRSV
 PI-3V
Pasteurella spp.

Table 2-12. Serum neutralization antibody titers to some viruses in 92 Holstein calves with clinical signs

Antibody titer	BEVa (%)	BVDVb (%)	BCVc (%)
<2	27d(29.3)	12 (13.0)	0 (0.0)
2	31 (33.7)	11 (12.0)	1 (1.1)
4	16 (17.4)	14 (15.2)	0 (0.0)
8	10 (10.9)	9 (9.8)	1 (1.1)
16	7 (7.6)	13 (14.1)	8 (8.7)
32	0 (0.0)	11 (12.0)	10 (10.9)
>64	1 (1.1)	22 (23.9)	72 (78.3)
Total	92 (100)	92 (100)	92 (100)

aBovine ephemeral virus. bBovine viral diarrhea virus. cBovine coronavirus.
 dNo. of calf.

92 BCV 100%
 64 가 78.3%
 . BVDV 87% 50% 가 16
 . BEV 71.7% 8 가 89.4%

Table 2-13. Serum neutralization antibody titers to some viruses in Holstein calves with clinical signs by sex

Antibody titer						
	BEFVa	BVD ^b	BCV ^c	BEFV	BVD	BCV
<2	10d	2	0	17	10	0
2	8	4	0	23	7	1
4	1	5	0	15	9	0
8	4	1	1	6	8	0
16	1	2	1	6	11	7
32	0	3	1	0	8	9
>64	0	7	21	1	15	51
Total	24	24	24	68	68	68

aBovine ephemeral fever virus. bBovine viral diarrhea virus. cBovine coronavirus. dNo. of calf.

가 가 (Table 2-14).

BVDV

(Table 2-14, 15).

Table 2-14. Serum neutralization titer against bovine viral diarrhoea virus in female Holstein calves with clinical signs by age

Weeks	Serum neutralization titer (log ₂)							Total
	<2	2	4	8	16	32	64	
2.5	0	0	1	0	0	0	1	2
3	0	1	1	1	1	1	1	6
4	1	3	2	0	1	1	3	11
5	1	0	0	0	0	1	0	2
6	0	0	1	0	0	0	2	3
Total	2	4	5	1	2	3	7	24

Table 2-15. Serum neutralization titer against bovine viral diarrhoea virus in male Holstein calves with clinical signs by age

Weeks	Serum neutralization titer (log ₂)							Total
	<2	2	4	8	16	32	64	
2.5	2	1	0	1	3	1	4	12
3	3	1	2	4	2	1	1	14
4	3	4	7	2	3	6	7	32
5	0	1	0	0	4	0	2	7
6	2	0	1	1	2	0	1	7
Total	10	7	10	8	14	8	15	62

가
 (125) Table
 2-16 IBRV가 (84.8%)

32 가 72% .
 PI-3V BVDV 82.4%
 PI-3V 32 가 60.8% BVDV 32
 67.2% 가 . BEFV 57.6%
 64 16-64 .

Table 2- 16. Prevalence of neutralizing antibody to some viruses in Korean- native calves with clinical signs

Antibody titer	No. of calves to			
	IBRV ^a	PI-3V ^b	BVDV ^c	BEFV ^d
< 2	19	22	22	53
2	8	11	2	4
4	19	13	3	8
8	33	33	7	9
16	20	19	7	11
32	10	13	22	10
64	7	10	28	28
128	5	3	19	2
> 256	4	1	15	-
Total	125	125	125	125

^aInfectious bovine rhinotracheitis virus. ^bParainfluenza-3 virus.
^cBovine viral diarrhea virus. ^dBovine ephemeral fever virus.

(Table 1- 17)

IBRV PI-3 가 90
 1 55.6% 가
 Chuzan, Akabane, Ibaraki virus
 63.3%, 54.4% 54.4% ,
 BVDV 90 84.4% 가

Table 2-17. Prevalence of neutralizing antibody to some viruses in slaughtered Korean-native cattle

Antibody titer	No. of cattle to						
	Chuzan virus	Akabane virus	Ibaraki virus	IBRVa	PI-3Vb	BVDVc	BEFVd
< 2	33	41	41	89	89	14	40
2	2	0	1	0	0	0	0
4	4	1	2	1	1	1	1
8	9	0	4	0	0	4	1
16	16	14	7	0	0	3	2
32	16	14	11	0	0	3	10
64	9	8	10	0	0	2	12
128	1	9	4	0	0	5	14
> 256	0	3	10	0	0	49	10
Total	90	90	90	90	90	90	90

aInfectious bovine rhinotracheitis virus. bParainfluenza-3 virus.

cBovine viral diarrhea virus. dBovine ephemeral fever virus.

가
 가 9
 (45), (39), (30),
 (20), (30), (19), (35), (79), (8 가) 305

가

가

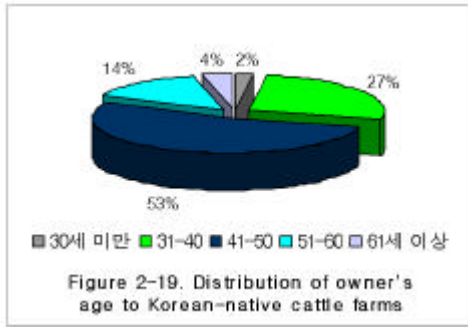
Figure 1-19

40 가 53% 가

40 가 29%, 50 , 60 18% 30-40

가 21- 40 가 30% 가
 41- 60 가 25% , 61- 100 가 26%
 100 가 6% 가
 가

(Figure 2- 20).



(Figure 2- 21) 가 71%
 9% , 5%

(Figure 2- 22) 가 가
 가 가
 , , ,

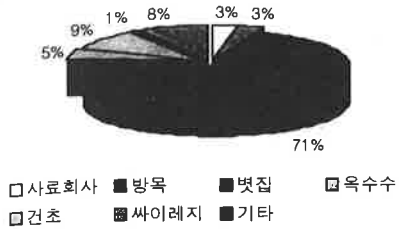


Fig 2-21. Distribution of kinds of feedstuff used.

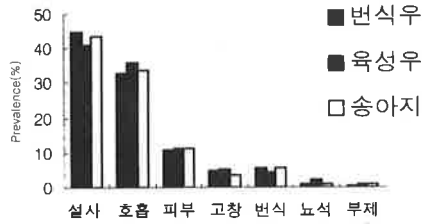


Fig 2-22. Prevalence of frequent diseases occurring in Korean-native cattle by age.

지역별 설사 발생률은 경북과 경기지역이 다른 지역에 비하여 보다 많이 발생하고 있는 것으로 조사되었고(Figure 2-23) 호흡기 질환의 발생률은 경북, 경남, 충남 등이 비교적 다발하였으며, 전북과 제주지역이 2%이하로 낮은 발생률을 보였다(Figure 2-24).

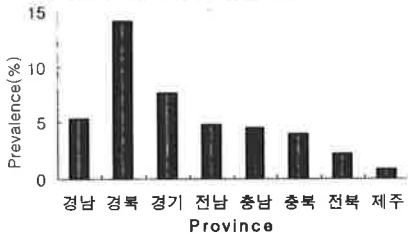


Fig 2-23. Provincial prevalence of enteric diseases of Korean-native cattle.

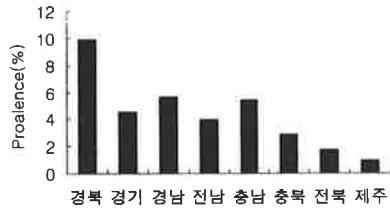
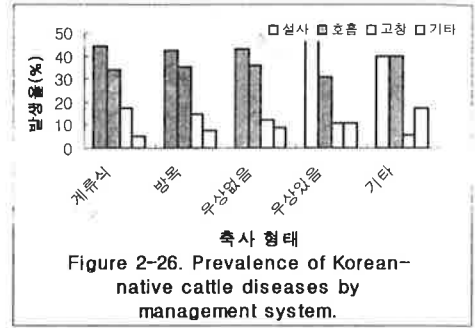
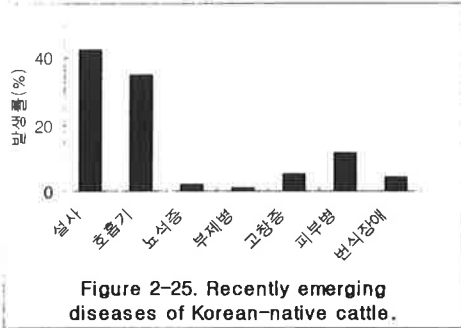


Fig 2-24. Provincial prevalence of respiratory diseases in Korean-native cattle.

한편, 축산인들이 가장 문제시되는 질병으로 생각하는 것으로는 설사증과 호흡기 질환이 가장 문제가 되고 있다고 답변하였고 피부병, 고창증, 번식장애가 그 다음으로 문제가 되는 질병으로 나타났다(Figure 2-25).

축사의 형태별 질병발생상황은 Figure 2-26에서 보는 바와 같이 축사의 형태와 사육방식이 질병의 발생여부에 크게 영향을 미치지 않았으나 우상이 있는 축사의 경우가 다른 축사에 비해 호흡기 질환의 발생률이 낮게 나타나는

것으로 조사되어 한우사육에 있어서 우상의 설치가 호흡기질환의 발병 방지에 도움이 되는 것으로 나타났다.



가 65.7% (67.9%)

가

. Santra

Pachalag(1996)

(58.3%) 가

(20.8%)

72.8% 65.0%
17.4% 14.7%

가

10

4

가

가

(, 1996)

가

(Barker, 1993).
 55.6%
 BRSV(23.5%) IBR(20.3%)
 (, 1998).
 가 (, 1998;
 1998).
 가 (Tsuda, 1999).
 4.5%가
 가 10
 , 4
 (, 1996).
 가 52.5% , 35.2%
 가 87.7%
 (가) (1996- 1998)
 (44.7%) 가 , (17.6%)
 62.3%가 (, 1996,
 1997 ; , 1998).

가

IMF가

20

가

가 가

419

203

(48.48%)

37.0%

257

115 (44.7%)

(39.3%)

가 가

1989; , , 1991).

(Bachmann, 1983; , 1984; Crouch, 1984; Peter ,

, BVDV,

가

가

가

가

가

90 100% 가 20 30% 가
 가 . 80 가
 가 41% 가 가 6
 (K- 88) 21% (Allen , 1985).
 가 46.5% 가 .
 가 52.7%, 38.2%
 가 73.9%
 가

(Bachmann, 1983; Hall , 1988).

1

가 , 가
 . ,
 가

(Petrie, 1987).

, (lipopolysaccharide)
 (0.2- 20 ug/kg, IV) 가
 , , , , , ,
 , 가 , , , ,
 , (Gerros , 1995).

(Pospischil, 1987; Berg, 1985).

가

가

B

vero

(Allen, 1985; Barker

, 1993).

60

(40%)

86.6% 가

(11.8)

44 (73.3%)

(

, ,

,

, BVD,)

(Janke, 1990).

가

(Hall, 1984, 1988).

21

19 (90.5%)가 가

가 ,

6 ,

가 4 , *S. typhimurium* 2 ,

K99+

1

(Hall ,

1988).

,

1 x 10¹⁰

가

1 x 10⁷

가

Munich K28, V1158/79,

V1124/81

가

가

가

가

(Bachmann, 1983).

(Bachmann, 1983).

7-40

가

가

, K-99

50- 60% 15- 20% , 10% 0.6%

(Bachmann, 1983; Allen , 1985).

Clostridium perfringens

가 가
가

(Bachmann, 1983)

가

가

가가

.

676

59 (8.7%)가

1969

Nebraska

1988

가

.

2

1

가

가

80%

30%

2

(, 1988),

가

8.3%

1.2%

가

6

36%,

4.1%

100%, 20 45% (, 1991).
33.9% (1991)

BVD (676)
51 (7.5%)가 (472) 10.8%

가

(Baker , 1993).

(Evermann Faris,
1981).

,

,

가 ,

(Potgieter , 1997; Graham , 1998)

가

. BVD

2

(Baker , 1993).

BCV

82.0 93.5%

(, 1991)

BCV

6.0%

43.9%

BCV 1972

(16.4%),

(59%),

(3- 31%)

(Tsunemitsu , 1991).

가

,

BCV

2

4

,

BCV

(, 1991).

4

BCV

4

BVDV

20- 50%

(, 1999).

BCV

BCV

가

가

(, 1998). BCV

43.9%

가 .

, D- xylose (Hall ,

1984).

가 (42)

Salmonella typhimurium

(Petrie, 1987) *S. dublin*

*Salmonella muenster*가 ,

(Sanford, 1984)가

가 .

(1996)

Clostridium perfringens A

가

가 .

*Clostridium perfringens*가

4

2

100%

가

가 .

1

가

,

, 1 2 , ,
 , ,
 ,
 (Randhawa , 1981; Petrie, 1987).

Eimeria spp 4 15 가

2

Chlamydia psittaci

24

(Doughri , 1974)

(12.6%),

BRSV (5.3%), IBR(1.2%)

77.6%, 61.1%, 35.5%

Pasteurella multocida

Pasteurella hemolytica 가

가 2 9 , ,

, , 가 , . 1

3 4% 25% , 30%

(Panciera Corstvet, 1984; Martin

, 1990).

(Reynold , 1985). 12.8%

38.8%

(Dungworth, 1993). BRSV, PI- 3V, BVDV ,

가

(Turk , 1985; Dungworth, 1993).

P. hemolytica *P. multocida*가 가

. IBRV *P. hemolytica*

P.

hemolytica 가 myeloperoxidase 가

가 myeloperoxidase

가 가 (Turk , 1985).

BRSV

3.3%

(Elvander , 1998). 1970

(Ames, 1993),

(Dungworth, 1993).

(Sanford, 1984; Ames, 1993)

(61.1%)

BRSV 가 (Baker, 1985)가
(, 1997)
15% 가
가

Pasteurella spp., IBRV, *Mycoplasma* spp.

IBRV 13.0%

35.5% 1 , ,

가

(Briges, 1988; van Donkersgoed, Babiuk, 1991). 2

10 , ,

IBRV *P. multocida*가

가 (, 1998)

. IBR 20 30% ,

1 10% (Briggs, 1988)

가

가

. IBRV 가 가

(van Donkersgoed Babiuk, 1991; Dungworth, 1993).

가
PI-3V가

(Table 2- 11)

(Briggs , 1988).

(, 1988).

2
40- 45%가
가

(Sanford, 1984).

4

가

(13)

(6),

(1)

bunyaviridae

bunyavirus

1959

1964

Aedes vexans, *Cules tritaeniorhynchus*

(Tsuda, 1999)

1999 .
가 ,
(Tsuda, 1999). Reoviridae
orbivirus 1985 *Culicoides oxystoma*
1956 Kasba

가
, , , ,
(Tsuda, 1999). ,
, 가
1998 .
, , , ,
가 10-30
(Tsuda, 1999),
가 가

10 가
가 1 .

1 ,
follow - up

10 가

가 가 (, 1999).

(, 1997). (,
1997, , 1999) , (, 1998)
(, 1998) .

Neospora canimun

1998). , , , (,
가 (, 1998).

(19)

가 1 2

가

(, 1998).

가 52.5% 가 1
 가 22.4 25.1% . 1
 가 가 (p<0.05), 1 (37.9%)
 가 6 2 . 가가

(, 1998).

가 가
 1 1-3
 ,
 90 7%, 15.4%
 Jensen (1976)
 . 1 가
 (44.3%) , 4-6 (29.6%) 1-3 (26.1%)
 . 4-6
 (61.8%) (38.2%)

가 . 25.0% , , , 가
 (29.0%) 가 (21.0%) 가 .
 가

(Jensen , 1976)

가
 가 가

가

(Mitruka Rawnsley, 1981)

가

(, 1982), 가

가

가 가

가 ,

(, 1982).

가

가

(England , 1998).

가

가

가

가

가

가 가

가

가

가

, , , , ELISA, 가 , , in situ . (Hiroshi , 1984; Dirk Deregts , 1992; Baszler , 1995; Dar , 1998).

가 가 (Janke , 1989; Ronald , 1989) 가 (Marsolais , 1978)

가 (England , 1976).

(Langpap , 1979; Hiroshi , 1991) 가

가

가 ,

가

(Acres , 1975; Ronald , 1989).

(Reynolds

, 1985; Peter , 1989).

(Reynolds , 1985; Marsolais , 1987).

PTA uranyl acetate 가

(, 1996).

가

가 106/ml

가

(Reynolds , 1985; Marsolais , 1987).

가 가 (Reynolds , 1985; Peter , 1989) 3 7

(England , 1976)

1 2
 1% PTA grid formvar

(Horne , 1974; England , 1976).

(Horne , 1974)

가 .

. , 1 2

grid

1% PTA 2% PTA

10 20

amyl acetate 2% colloidion

80 kV 가

grid

147

가

66.7% (98) ,

가 33.3% 가

가

16.3% 10.2% 가

1 가 97.6% 2)

4 가 72.1%

가

가

80% 가

(30.8%) (14.0%)가 (Dirk , 1992).
 , , , ,
 (Weldon , 1979; Janke , 1989; , 1990). (35.3%),
 (35.3%), (5.9%) (
 , 1994). (33.3), (16.3%), 가
 (10.2%) .
 가

(, 1991)

가

1969
 RNA (, 1984; , 1990; , 1996) 65 75 nm
 가 6 (, 1988).
 1 2
 (, 1996; Janke , 1989).
 23% (, 1984), 5- 14 70%
 (Janke , 1989) 가 . 1980
 (, 1990) 가 40%

1972

80 160

nm 가 RNA

(Hiroshi , 1991) , ,

16.4%, 59%, 3 31% (Langpap , 1979; Janke , 1989)

가

(Reynolds , 1985)

(Hiroshi , 1991)

16.3%

가

가

가

1946

(,

1986; Ronald , 1989).

가

RNA

90 150 nm

5 7 nm

가

38.4%가

가

(, 1988;

Ronald , 1989; Baszler , 1995),

10.2%가

가

120 200 nm

DNA

negative

nucleocapside가

(Marsolais , 1978).

가

+

가

가 2

가

44.9%

72.7%

61.3%

가

10

E. coli *Salmonella* spp.가 7 3

가 (Chasey Lucas, 1977; Langpap
, 1979; Reynolds , 1985; , 1988).

가 가

가 가

4 117 2

14.4% 14.5% 2
 21.5% 28.3%
 , 16.2%
 66.0%

가

가

가

BCV, BVDV, BRV, *Eimeria* spp., *Salmonella* spp., *Pasteurella* spp.,

IBRV, BRSV 8

4

117

92

BEFV,

BVDV, BCV

가

BCV

100%

64

78.3%

. BVDV 8

59.8% ,

125

78.4%

가

BEFV

29.3% 가

125

45.6% 가

. 3

가

. , 125
 가 2 가 IBRV 21.6% PI-3V
 26.4% %
 .
 325 (van
 Donkersgoed , 1993) PI-3V BRSV가 90% IBRV 70%
 1995
 BEFV가 26.3%, IBRV가 10.5%, PI-3V가
 7.9%, BCV가 87.5%, BVDV가 49.0%

가 .
 .
 가
 IMF가 가 가
 30-40 가 (305
 가) 81% .
 가
 가 21
 - 60 가 55%
 20 가가 13% .
 가

(15.1%), (13.0%), (5.5%), (2.5%)

가

가 66.7%

가 , (43.9%), (35.5%), (33.9%)

(, , ,)

1 52.5%

, 1-3 (25.1%) 4-6 (22.4%) 1

37.9% 가 , 1-3 18.1%, 4-6

13.8% 가 가 가

1-2 (41.1%)

가 1 44.3% 가

1-3 (26.1%) 4-6 (29.6%) 가

21.0- 29.0% 가

13.2- 18.3%

(31.5%) 가 (20.2%) 가

(p<0.05) , (26.8%) 가 (21.4%)

가 (19.5%) 가

(56.5%) 가

2)

(174) 가
 가 (p<0.05). ,
 , MCV, MPV 가 가
 . 가 가
 , 가 .
 (241)
 가 가

3)

147 98 (66.7%) 가
 33.3% 가 , (16.7%), 가 (10.2%),
 (0.7%) .
 (5.4%) 가 1-3
 66.7- 73.1% , 4 100% 가 4
 62.0% .
 (44.9%) 가

4)

117 1-2 .
 10 16 (13.7%)가 2 92 (78.6%)가
 . (43.78%) 가
 (35.9%) , (16.2%)

59%

가 (52.1%)

65.8% (77/117)

Pasteurella spp.가 44.4% (52/117) 가 ,

29.9% BVDV(19), IBRV(18), PI-3V(14)

coronavirus(14)

Pasteurella spp. PI-3V IBRV 가 11

9 , *Pasteurella* spp. BRSV 7

(3)

(32) 가 BVD(19), BCV (14),

(5), (5), (4)

(4) (2)

92 BCV 100%

64 가 78.3%

BVDV 87% 50%가 16

BEV 71.7% 8 가 89.4%

5) 가

(125) IBRV가

(84.8%) 32 가 72%

PI-3V BVDV 82.4%

PI-3V 32 가 60.8%

BVDV 32 67.2% 가 BEFV 57.6%

64 16-64

6) 가

9 305 가

가 40 가 53% 가 40

가 29%, 50 60 18%

가 21- 40 가 30% 가 , 41- 60 가

25%, 61- 100 가 26%, 100 6%

71%

(9%) (5%) (43%)가

가 (35%) (11.53%) ,

, , (13%)

가 , 10% , (9.5%),

(5.5%), (5.4%) 2%

가

(44%) (35%) (10%), (5%), (4%)

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가 . 1997, p.55- 58.

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Kasba(Chuzan) virus .
(), 1998, 38:71.

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parainfluenza- 3 respiratory syncytical virus .
(가) 1988,30: 39- 35.

. 1982.

3

1

(syndesmochorial)

가

(Osburn et al 1982).

(John 1997).

(Failure of passive immunoglobulin transfer: FPT)

6.4

5.4 Whittum (1995)

(1989; Saif and Smith

1985).

10 30% 가

(1989;

Logan and Gibson 1975; McBeath et al 1974).

FPT 가

(Rajala and Castren 1995; 1989; Stott and

Fellah 1983; Naylor et al 1977; McGuire et al 1976; Selmen et al 1971).

(Julie et al 1993).

		Banks (1982)	IgG	100
IgM, IgA가	24	14	97%	
	()		
		가	가	

(Hudson and Payne 1976).

가

FPT

(TP), glutaraldehyde test, sodium sulfite precipitation test (SSPT), zinc sulfate turbidity test (ZSTT), single radial immunodiffusion test (sRID)

FPT (Tyler et al 1996; Rea et al 1996, Julie et al 1993; Hopkins et al 1984; Naylor et al 1977; Nancy and Travis 1977; McEwan et al 1970).

FPT

FPT

가

IgG

FPT

, bovine viral diarrhoea virus (BVDV), parainfluenza-3 virus (PI-3V), infectious bovine rhinotracheitis virus (IBRV), bovine coronavirus (BCV)

가

2

가.

97 9 99 9 6 258

2

2 2

10ml

vacutainer

(Beckton Dickson™)

2 3

4°C 3,000 rpm 15

- 70°C

. sRID

sRID IgG 12 well Kit (VMRD™)

IgG

well 3 μ l

20 24

IgG 400, 800, 1,600,

3,200 mg/dℓ

가

. TP, Alb, Glob

Vitros DT 60 , DTSC Chemistry System (Johnson & Johnson™)
(total protein;TP) (albumin;Alb)
(globulin;Glob)

. SSPT

SSPT Nancy Travis (1977)
14%, 16%, 18% Na₂SO₄ (anhydrous)
1.9 Mℓ 0.1 Mℓ
1
, 1+, 2+, 3+

. ZSTT

McEwan (1970)
ZnSO₄ · 7H₂O 가 208 mg/L 가
10 15
1 6 Mℓ 0.1 Mℓ
가 1
(spectrophotometry, DUTM, USA)

490 nm 400, 800,
1,600, 3,200 mg/dl IgG

10% fetal bovine serum (FBS)
- minimum essential medium (MEM gibcoTM, USA) MDBK
80%
(cytopathic effect: CPE) 가 90%
- 70
strain . BVDV, BCDV, IBRV PI3- V
BVDV NADL strain, Kagegawa strain, PQ7 strain (), PI-3V
NADL strain . 96 well microplate
가 56 30 50 µl
96 well microplate 가 37 1
10% FBS가 - MEM MDBK
96 well microplate 24 0.1 Ml
. 37 CO2 가 200 TCID₅₀/Ml
MDBK 가 96 well microplate 4 CPE
. CPE가 10%
가 , 가가 2

Student's
T - Anova (analysis of variance: SAS)

3

가.

258 51 (19.8%)
 13 (5.0%)
 33 가 11
 (9) 가 , (2)

(Table 3- 1).

Table 3- 1. Biochemical values of Korean- native calves used

Clinical signs	No. occurred	Biochemical value ^a			
		TP (g/dl)	Alb (g/dl)	Glob (g/dl)	IgG (g/dl)
Respiratory	11(2 ^b)	4.1 ± 0.4 ^c	2.0 ± 0.3	2.0 ± 0.3	1.2 ± 0.5
Enteric	33(9)	4.1 ± 0.4	2.2 ± 0.3	1.9 ± 0.4	1.2 ± 0.4
Weakness	5(0)	4.8 ± 0.4	2.3 ± 0.3	2.4 ± 0.3	1.6 ± 0.4
Others	2(2)	3.4 ± 0.3	1.8 ± 0.2	1.7 ± 0.1	0.8 ± 0.1
<i>Subtotal</i>	51(13)	4.1 ± 0.4	2.2 ± 0.3	2.0 ± 1.0	1.2 ± 0.5
No	207	4.9 ± 0.5	2.2 ± 0.3	2.6 ± 0.3	1.6 ± 0.6
Total	258	4.8 ± 0.5	2.2 ± 0.3	2.5 ± 0.8	1.5 ± 0.5

^aTP, total protein; Alb, albumin; Glob, globulin. ^b No. of death. ^c Mean ± SD.

질병이 발생한 송아지들에 대한 TP, Alb, Glob, IgG 검사결과 임상증상이 나타나지 않았던 송아지들보다 TP, Glob, IgG의 혈청내 농도가 낮게 나타났으나 Alb은 더 높았다.

나. sRID를 이용한 IgG 수준

sRID를 이용한 IgG 검사에서 전체 258 두의 IgG 평균값은 $1,530 \pm 410$ mg/dl 였으며 일령별 평균값은 Figure 2-1에서와 같으며 초유 섭취 후 9 일령에서 $2,160 \pm 460$ mg/dl 로 가장 높은 값을 나타냈으며 이후 15일령에서 $1,140 \pm 380$ mg/dl로 낮아졌으며 다시 22 일령에서 $2,040$ mg/dl 까지 상승하였다(Figure 1).

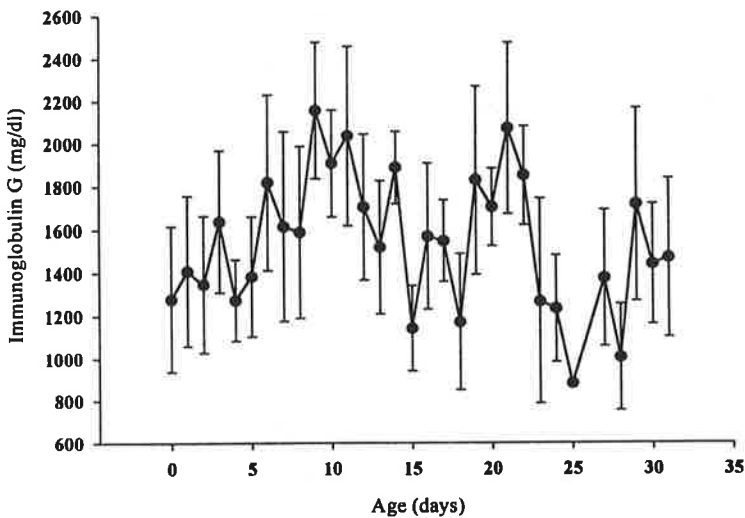


Figure 1. Mean IgG concentrations of Korean-native calves using single radial immunodiffusion test.

Error bar was standard deviation. The value at 31 days was indicated the mean level between day 31 and 42.

Ig 수준을 800, 1,000, 1,300 mg/dl로 기준을 삼아 FPT를 측정 한 결과 FPT 진단 은 각각 34두, 53두 및 93두였으며 이들에 대한 민감도는 1,000 mg/dl를 기준으

0.45 가 1,300 mg/dℓ 0.88 가
 , 1,000 mg/dℓ 0.15
 가 (Table 3-2).

Table 3-2. Sensitivity and specificity of the cutoff point to Korean-native calves with FPT by the concentration of immunoglobulin G

Cutoff point (mg/dℓ)	No. of FPT	No. diseased	No. died	Sensitivity	Specificity
800	34	14(10a)	5(2)	0.41	0.83
1,000	53	24(16)	8(4)	0.45	0.87
1,300	93	31(21)	9(5)	0.33	0.88
Mean	60	23(16)	7(4)	0.40	0.86

aNo of calves with enteric disorders.

. TP

TP 258 TP 4.76 ± 0.6 g/dℓ
 IgG (sRID) 1 가
 TP (5.4 ± 0.6 g/dℓ) 15 가 (3.2 ± 0.5
 g/dℓ) 19 TP (5.8 ± 0.5 g/dℓ)

(Figure 3-2).

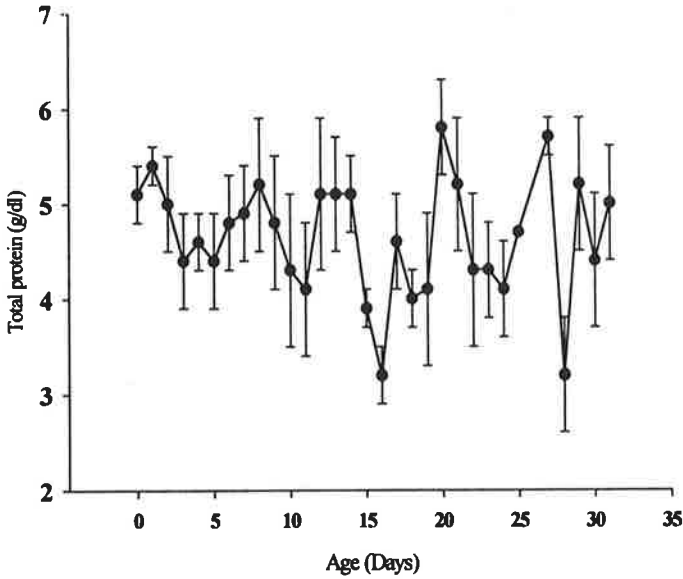


Figure 2. Mean serum total protein levels of Korean-native calves by age.

Error bar was standard deviation. The value at 31 days was indicated the mean one between day 31 and 42.

TP수준을 3.5 g/dl, 4.0 g/dl, 4.5 g/dl이하로 각각 FPT를 구분했을 경우 FPT 발생은 61두, 77두 및 96두 이었고 이들에 대한 민감도는 4.0 g/dl 이하를 FPT 기준점으로 했을 경우 0.35로 가장 높았으며 특이도는 3.5 g/dl이하를 FPT로 잡았을 경우 0.89로 가장 높았다. 또한 폐사율에 대한 민감도는 4.0 g/dl이하를 기준으로 했을 경우 0.10으로 가장 높았다(Table 3).

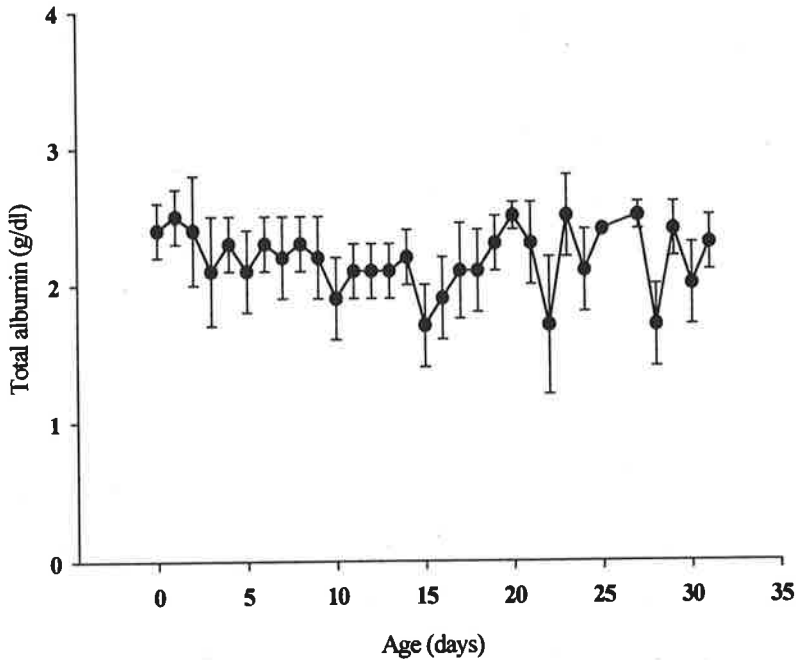


Figure 3. Mean serum albumin levels of Korean-native calves by age.

Error bar was standard deviation. The value at 31 day was the mean levels between day 31 and 42.

혈청 Alb수준을 각각 1.5, 2.0, 2.5 g/dl로 FPT를 구분했을 경우 각각 검사대상 송아지 중 FPT 해당두수는 31두, 70두 및 172두이었다. 각각의 기준점에 따른 민감도와 특이도 검사에서 2.5 g/dl 이하를 기준으로 했을 경우 가장 높은 민감도(0.22)와 특이도(0.85)를 나타내었으며 FPT 진단율(66.7%)이 가장 높았다 (Table 4).

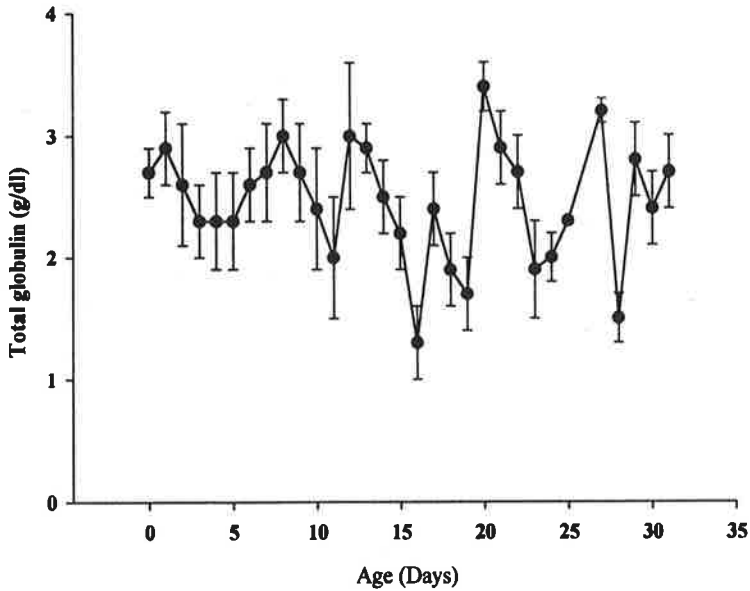


Figure 4. Mean serum globulin levels of Korean-native calves by age.

Error bar was standard deviation. The value at the day 31 was indicated the mean level between the day 31 to 42.

혈청내 Glob수준을 각각 1.5, 2.0, 2.5 g/dl 이하로 잡고 FPT로 구분했을 경우 각각의 해당두수는 45두, 78두, 110두씩이었다. 이들에 대한 민감도는 1.5 g/dl 이하를 FPT 기준점으로 했을 경우 0.38로 가장 높은 민감도를 나타냈으며 특이도는 2.5 g/dl 이하를 FPT 기준으로 했을 경우 0.89로 가장 높았다. 또한 폐사율에 대한 민감도 측정에서 2.5 g/dl 이하를 FPT 기준점으로 했을 경우 0.1로 가장 높았다 (Table 5).

Table 3-5. Sensitivity and specificity of the cutoff point of FPT to 258 Korean-native calves by total globulin

Cutoff point (g/dl)	No. of FPT	No. diseased	No. died	Sensitivity	Specificity
1.5	45	17(12a)	4(3)	0.38	0.84
2.0	78	27(17)	8(5)	0.35	0.87
2.5	110	34(23)	11(7)	0.31	0.89
Mean	78	26(18)	8(5)	0.34	0.87

aNo of calves with enteric diseases.

. SSPT

가 14%, 16%, 18%

19% 34 ,

51 . 16% 18% 71 가

186 . 1+ FPT

34 가 FPT 27 가 10

가 . 2+ 51 36 가 12 가

. 3+ FPT 37 가 12 가

(Table 3-6).

Table 3- 6. Sensitivity and specificity of the cutoff point of FPT to 258 Korean- native calves by sodium sulfite precipitation test

Cutoff point	No. of FPT (%)	No. diseased	No died	Sensitivity	Specificity
<1+	34(13.2)	27(20a)	10(7)	0.79	0.89
<2+	51	36(25)	12(9)	0.71	0.93
<3+	71	37(25)	12(9)	0.52	0.93
Mean	52	33(23)	11(8)	0.67	0.92

aNo. of calves with enteric diseases.

1+ FPT 0.79 가
 2+ 3+ FPT
 0.93 가
 가 가

. ZSTT

490 nm IgG 1,440
 ± 420 mg/dℓ sRID
 IgG 9 가 IgG 가 2,300 mg/dℓ
 . 18 1,150 mg/dℓ 25
 IgG 가 1,000 mg/dℓ (Figure 3- 5).

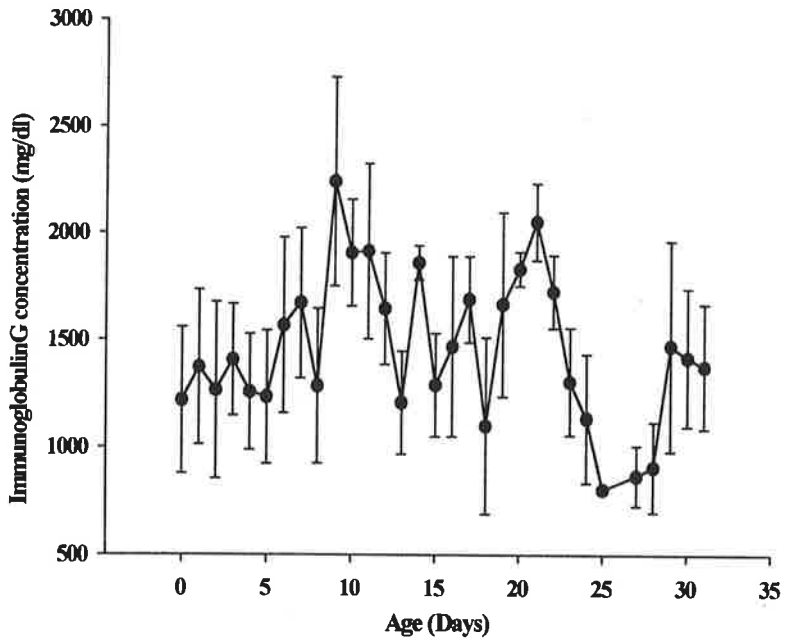


Figure 5. Immunoglobulin G concentrations of Korean-native calves with zinc sulfate turbidity test.

Error bar was standard deviation. The value at the day 31 was indicated the mean level between the day 31 to 42.

ZSTT를 통한 혈청 IgG농도를 800 mg/dl, 1,000 mg/dl 및 1,300 mg/dl이하로 FPT 기준을 각각 잡았을 경우 FPT로 진단된 송아지는 각각 59두, 70두 및 111두이었다. 민감도와 특이도 검사에서 민감도는 800 mg/dl 이하를 FPT 기준점으로 했을 경우 가장 높은 0.42를 나타냈으며 특이도는 1,300 mg/dl 이하를 FPT 기준점으로 했을 경우 가장 높은 민감도(0.88)를 나타냈다. 또한 폐사율에 측정에 대한 FPT 검사에서 800 mg/dl 이하를 FPT 기준으로 했을 경우 0.12로 가장 높은 민감도를 보였다 (Table 7).

Table 3-7. Sensitivity and specificity of the cutoff point of FPT to Korean-native calves by zinc sulfate turbidity test

Cutoff point (mg/dℓ)	No. of FPT	No. diseased	No. died	Sensitivity	Specificity
800	59	25(16a)	7(3)	0.42	0.87
1,000	70	27(18)	8(4)	0.39	0.87
1,300	111	34(23)	9(5)	0.31	0.88
Mean	80	29(19)	8(4)	0.37	0.87

aNo of calves with enteric diseases.

SSPT가 가 0.79 0.89

0.80 TP 0.35 0.87

Alb 0.19 0.80 Glob 0.35 0.87 sRID

IgG 1,000 mg/dℓ cutoff point 가 0.41,

가 0.87 . ZSTT 1,000 mg/dℓ cutoff point

가 0.39 0.87 (Figure 3- 6).

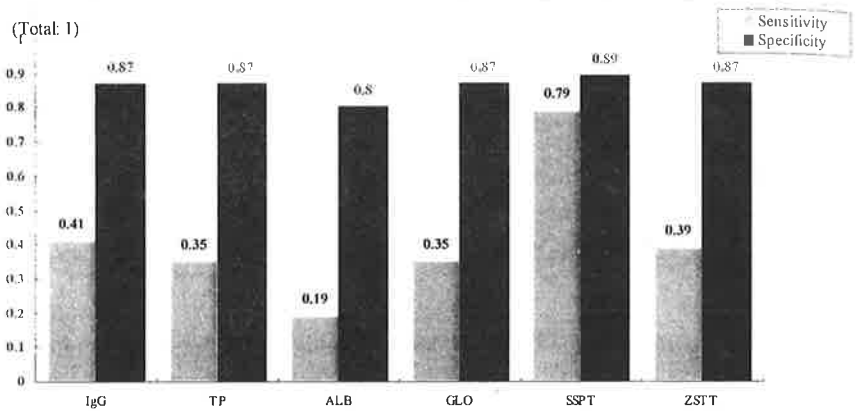


Fig 6. Sensitivity and specificity of Korean-native calves with failure of passive transfer by each test.

*The cutoff point of each test was made the following; sRID and ZSTT were less than 1,000 mg/dl, TP(total protein) less than 4.0 g/dl, Alb (albumin) less than 2.0 g/d, Glob (globulin) less than 2.0 g/d, SSPT (sodium sulfite precipitation test) less than <1+.

또한 이들에 대한 비교 위험도 검사에서 SSPT는 1+ 이하인 한우 송아지들은 그 이상의 송아지들보다 7.38배이상 질병에 이환될 확률이 높았다. IgG는 1,000 mg/dl 이하일 경우 그 이상인 개체보다 질병에 이환될 확률이 3.45배가 높았다. 또한, Alb을 이용한 검사에서는 2.0 g/dl 이하인 개체가 질병에 이환될 확률이 더 높게 나타났다(Table 8).

Table 3-8. Sensitivity, specificity and relative risk to each test for determining FPT to Korean-native calves

Test	IgG	TP	Alb	Glob	SSPT	ZSTT
Sensitivity	0.45	0.35	0.19	0.35	0.79	0.39
Specificity	0.87	0.87	0.80	0.87	0.89	0.87
Relative risk	3.45	2.65	0.09	2.63	7.38	3.07

The cutoff point was based on the following: IgG (sRID) was less than 1,000 mg/dl, total protein less than 4.0 g/dl, albumin less than 2.0 g/dl, globulin less than 2.0 g/dl, sodium sulfite precipitation test less than 1+, and zinc sulfate turbidity test less than 1,000 mg/dl.

258 BVDV, BCV, IBRV, PI-3V

Table 9 BCV BVD 가

96.1% 73.6% PI-3V IBRV 37.9%, 43.8%

(P > 0.05).

Table 3-9. Prevalence of the antibodies to main viruses in Korean-native calves by sex

Sex	No. of calves with antibody to			
	BVDV	BCV	PI-3V	IBRV
Female	91/123a	116/123	43/123	51/123
Male	99/135	132/135	55/135	62/135
Total	190/258	248/258	98/258	113/258

aNo. of positive / No. of sera tested. All positives were more than 1 : 2 or higher of neutralization test to each virus.

4

가 Table 10

8 log₂

(< 2) 8 log₂

가

Table 3-10. Distribution of antibody titer to some viruses in Korean-native calves

Virus ^a	Antibody titer ^b								
	<2	2	4	8	16	32	64	128	256
BVDV	73 ^c	3	1	3	5	12	16	28	117
BCV	10	1	4	2	4	4	9	26	298
PI-3V	164	14	16	8	12	5	9	9	20
IBRV	151	20	22	12	8	11	9	8	17

^aBVDV, bovine viral diarrhea virus; BCV, bovine coronavirus; PI-3V, parainfluenza-3 virus; IBRV, infectious bovine rhinotracheitis virus. ^bReciprocal of final serum dilution inhibiting viral cytopathic effects. ^cNo of calf.

가

Figure 7

6 4 가

가 14

가 BCV, BVDV, IBRV, PI-3V

BVDV, BCV, PI-3V, IBRV

가 4.2, 6.4, 1.2, 1.3 log₂

가 Table 3-11

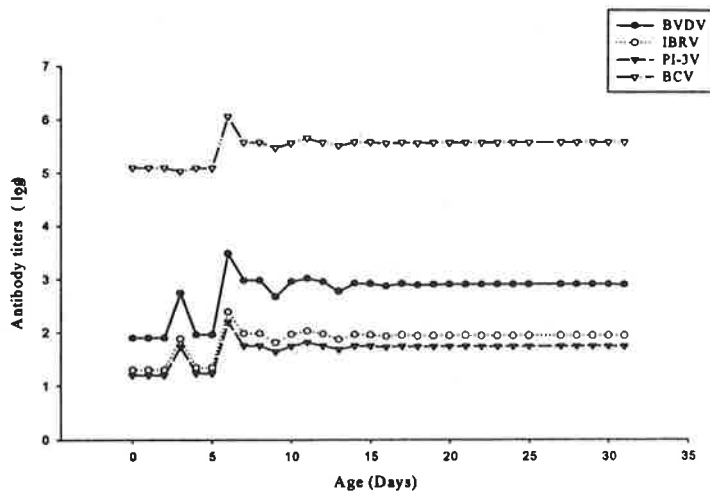


Figure 7. Mean antibody titers to main viruses in Korean-native calves by age.

Table 11. Mean neutralizing antibody titers to main viruses in Korean-native calves with clinical signs

Clinical signs	Antibody titers to			
	BVDV ^a	BCV	PI-3V	IBRV
Respiratory	4.5 ^b	6.6	1.4	1.4
Enteric	4.5	6.4	1.0	1.2
Weakness	2.0	7.2	0	0
Others	4.0	4.0	4.0	5.0
<i>Subtotal</i>	4.2	6.4	1.2	1.3
No	5.4	7.4	1.7	1.8
Total	5.12	7.15	1.6	1.7

^aBVDV, bovine viral diarrhea virus; BCV, bovine coronavirus; PI-3V, parainfluenza-3 virus; IBRV, infectious bovine rhinotracheitis virus. ^bAntibody titer(log₂)

FPT		Table 2-12		BVDV 가	
FPT		IgG, 1,000 mg/dl		가 FPT	
. BVDV		IgG 1,520 mg/dl (ZSTT)		.	
BVDV	PI-3V	IBRV		BCV	
6.5 log ₂	가	. BVDV			
FPT	IgG (sRID)				
FPT	. BVDV				
	가 18	10	가		
7	4	가	.		
BCV	10	FPT	IgG		
FPT	30.0%	가	FPT		
	30.0%				
	2		. BCV		
PI-3V	가	.			
PI-3V	165	가	. 165		
FPT	IgG (sRID)		FPT	20.6%	
가	FPT	(ZSTT)			
30.9%		FPT			
			36		
24	가		6	가	. ,
11			8		
2	가	.			
IBRV	151	FPT	IgG (sRID)		
FPT	22.5%	FPT			

32.5% 가 . 21.2% 6.6%

(Table 12).

가 (20)

(7) , 10 (7)가

가 (2) .

Table 3- 12. Relationships among antibodies to viruses, different test to FPT, and morbidity and mortality of diseases in 258 Korean- native calves

Antibody	Virus	No of calves	% of calves with FPT ^a by					SSP T	Morbi- dity(%)	Mor- tality(%)
			TP	Alb	Glob	sRID	ZSTT			
Serone- gatives to	BVDV	73	17.8 (5.0) ^b	11.0 (2.4)	23.3 (2.5)	30.1 (1,380)	42.5 (1,520)	17.8	24.7	9.6
	BCV	10	30.0 (4.5)	20.0 (2.0)	20.0 (2.5)	30.0 (1,640)	30.0 (1,569)	10.0	20.0	0
	PI- 3V	165	20.6 (4.9)	18.8 (2.3)	25.5 (2.6)	20.6 (1,460)	30.9 (1,370)	15.2	21.8	6.7
	IBRV	151	17.9 (4.9)	15.9 (2.3)	23.8 (2.6)	22.5 (1,450)	32.5 (1,340)	13.9	21.2	6.6
Sero- positives to	BVDV	185	34.6 (4.7)	33.5 (2.1)	33.0 (2.5)	16.8 (1,590)	21.1 (1,520)	11.4	17.8	3.2
	BCV	248	30.0 (5.4)	27.4 (2.4)	30.6 (3.0)	19.4 (1,670)	27.1 (1,460)	13.3	19.8	5.2
	PI- 3V	93	46.2 (4.9)	62.0 (2.3)	38.7 (2.6)	20.4 (1,640)	20.4 (1,570)	9.7	16.1	2.2
	IBRV	107	46.7 (4.4)	43.0 (2.0)	39.3 (2.4)	17.8 (1,640)	19.6 (1,590)	12.1	17.8	2.8

^aThe cutoff point to failure of passive transfer was <4.0 g/dl of TP, <2.0 g/dl of albumin, <2.0g/dl of globulin, <1,000 mg/dl of IgG, <1,000 mg/dl of ZSTT, and <1+ of SSPT. ^bMean serum concentration by each cutoff point.

(FPT)

19.7% 5.0%

가

Salman (1991)
11,767

1987 1988
가

5.0%

(Rea et

al 1996).

Glob IgG , 2.0 ± 0.4 g/dl IgG

1,220 ± 340 mg/dl

Glob IgG

Glob IgG 가 1.9 ±

0.4 g/dl, 1,170 ± 430 mg/dl

51

(1989)

IgG IgG 2940 ± 790

mg/dl, 1720 ± 760 mg/dl

가

IgG

IgG

(Tizard 1996; Besser et al 1991).

Ig
glucocorticoid , PI-3V
2 가 *Salmonella dublin* 24 ,
BVDV 28 100 가 (Hudson and
Payne 1976; Eberhart and Patt 1971). FPT 1

6 IgG
1,530 mg/dl IgG
9 가 IgG 15 가
(1989)

가 , IgG 가 1,483mg/d, 1,860mg/d, 2,245mg/d
Julie (1993) IgG , ,

IgG FPT cutoff point Virtala (1997)
800mg/dl, 1,000mg/dl, 1,300 mg/dl FPT FPT
13.2%, 20.5%, 36.0% 1,000
mg/dl 가 , 1,300 mg/dl
가 1,000 mg/dl
IgG FPT 1,000 mg/dl

TP Donovan (1998)
TP TP 4.0 5.0 g/dl 5.0
6.0 g/dl 6.0 g/dl

, Julie (1993)

가 TP가
 0.5 g/dl 4.67
 g/dl TP (6.57 7.07 g/dl)
 (1989) TP(6.8 ± 0.9 g/dl)

가 97

99 IMF

가 TP
 FPT Rea (1996) FPT 4.5g/dl
 4.0g/dl 3.5g/dl 23.6%, 29.8%,
 37.2% FPT 0.34
 가 4.0g/dl 0.35 가
 3.5g/dl 0.89 가
 TP FPT 4.0 g/dl가

Alb FPT Alb
 2.2 g/dl , 6
 Alb FPT 1.5g/dl, 2.0g/dl 2.5 g/dl ,
 12.0% , 27.1% , 66.7% 0.82 가
 0.20 Alb
 FPT 가 가
 2.0g/dl
 6 Glob 2.54 g/dl
 1 가 6 가 7

20 가 Glob(3.4 g/dl)

Glob 2 3

Glob 가 2.55 g/dl (DeNise ,1989), 2.57

g/dl(Robinson ,1988)

(,1989) Glob (3.59 g/dl)

FPT Glo 1.5 g/dl, 2.0 g/dl 2.5 g/dl

17.4%, 30.2%, 42.6% 가 FPT 1.5 g/dl

가 (0.38) 2.0 g/dl

가 (0.89) Glob

FPT 2.0 g/dl

SSPT FPT Nancy Travis (1977)

1+ FPT 가 0.79 0.89

가

SSPT 29.4% 가

sRID IgG

. Nancy Travis (1977) 18%

Ig 가 5 mg/ml

18% 16%

5 15 mg/ml, 18%, 16%, 14% 15

mg/ml 258

(+1) Ig가 5 mg/ml

51 sRID FPT

(1,000 mg/dl) 53 SSPT

가 .

ZSTT FPT

(McEwan et al 1970; Selmen et al 1971; McGuire et al 1976).

IgG가 9 가 가 21

25 가 sRID

IgG (P > 0.01). FPT 800

mg/dl 가 (0.42) , 1,300 mg/dl

(0.88)가 가 , 1,000 mg/dl

가 가 .

가 BCV

(96.1%) 가 6 가가

BVDV가 73.6% PI-3V IBRV 40.9%

BVDV, BCV, PI-3V, IBRV 가

1.1- 1.5 IgG

FPT

ZSTT가 가 FPT SSPT

가

FPT FPT IgG 가

TP, Alb, Glob 가 .

BCV, BVDV, IBRV PI-3V가

96.1%, 73.6%, 43.8% 37.9% (1995)

87.1%, 44.2%, 8.8%

5.6%

가

5

97 9 99 9 6 258

258 51 (19.8%) 13
(5.0%) 가

가

BCV 가 96.1% 가 , BVDV(73.6%),
IBRV(73.6%) PI-3V(43.8%) 4
FPT ZSTT가 가 (P<0.01).

FPT

IgG 가 TP, Alb, Glob

sRID IgG(<1,000 mg/dl) FPT 20.5% (53)

45.3% 15.1% TP(<

4.0 g/dl) FPT 29.8% (77) Glob(<2.0 g/dl)

FPT 30.2% (78) SSPT(<1+) FPT

13.1% (34) ZSTT (<1,000 mg/dl) 27.1% (70)

6 FPT Alb Glob
 2.0 g/dl , TP 4.0 g/dl , sRID IgG 1,000 mg/dl , SSPT
 +1 , ZSTT 1,000 mg/dl 가 . Alb
 FPT FPT

2.63 7.38 .

FPT SSPT가 0.79
 0.89 가

(P<0.01).

6

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가 . 12 . 29; 153-170, 1989.

,
, 29; 155- 162, 1989.

,
, 29; 171- 178, 1989.

,
, 35; 615- 624, 1995.

4

1

1. IgG

가.

, whey
 immunoglobulin 가
 (Hemmings, 1976). Williams and Miller (1978) IgG1
 IgG2 가 . Devery-Pocius and Larson
 (1983) immunoglobulin 가 가
 , IgG 가
 (Oyeniya and Hunter 1978). IgG
 IgG

1)

Holstein .
 5
 - 20
 IgG whey 6,000 rpm
 30 2 pH 4.6
 casein (Fey et al., 1976) whey pH 6.6
 - 20

2)

casein (1987),
A.O.A.C. (1984).

3. ELISA immunoglobulin G

IgG single radial immunodiffusion (VMRD, Inc., Pullman, WA). Kit
3ul wells,
well 3ul. Plate cover 24
ring. Standard curve
sample IgG.

pH

(Table 4-1).

가 pH가

(Quigley et al., 1994)

12.8%

Holstein

5.1%

5.8%

3.3%

3.6%

가

58%

Holstein

가

Im et al. (1991)

4.2 가

immunoglobulin

가

(Perino et

al., 1995; Singh et al., 1993).

가 Holstein

2

Table 4-1. Comparison of milk and colostrum in Korean-native and Holstein cattle

Cattle Item(unit)	Holstein		Korean-native	
	Colostrum	Milk	Colostrum	Milk
pH	6.64	6.72	6.68	6.74
Sp. gravity	1.072	1.068	1.076	1.071
H2O(%)	76.4	88.4	73.4	86.7
Total Solid(%)	23.6	11.6	26.6	13.3
Fat (%)	5.1	3.3	5.8	3.6
SNF*(%)	18.5	8.3	20.8	9.7
Protein (%)	14.1	3.1	17.3	3.7
Lactose (%)	2.23	4.61	2.15	4.52

*solid non- fat.

Table 4-2

Holstein		Korean-native	
Casein	23.16g/l	28.59 g/l	3
Casein-N	5.48g/l	6.35g/l	3
Casein-N		가	
Casein-N		60%	2.5
Casein-N			
Holstein			

Table 4-2. Analysis of protein components of colostrum and milk in Korean-native and Holstein calves

Cattle Item (unit)	Holstein		Korean-native	
	Colostrum	Milk	Colostrum	Milk
Total N (g/l)	23.16	5.48	28.59	6.35
NPN*	0.82	0.52	0.91	0.34
Protein- N	22.34	4.96	27.68	5.92
Casein- N	7.51	2.31	7.74	2.40
Non- casein- N	15.65	3.17	20.85	3.95

*non-protein- N ()

IgG Table 4-3 .
 IgG 6 220.1 ± 30.6 mg/ml 가
 , 2 가 4-6 316.2 ± 24.4
 mg/ml 가 .
 120 IgG .
 50% Scott et al., (1981) , 12
 24 3 IgG Oyeniyi and
 Hunter (1978) . Ig
 가

Table 4-3. Immunoglobulin concentration in Korean-native cattle colostrum and milk

Time after calving	Lactation (mg/dl)			Mean (mg/dl)
	1	2	4-6	
0-6hr	220.1 ± 30.6a	240.1 ± 21.6a	316.2 ± 24.4b	258.8 ± 25.5
24hr	18.2 ± 5.3a	32.6 ± 3.7b	38.2 ± 6.3b	29.7 ± 5.1
48hr	5.6 ± 1.2a	21.8 ± 2.3c	15.3 ± 2.7b	14.2 ± 2.1
120hr	0.8 ± 0.2a	2.1 ± 0.2b	3.4 ± 0.3c	2.1 ± 0.2
Milk	0.4 ± 0.1a	0.8 ± 0.1b	0.7 ± 0.1b	0.6 ± 0.1

* Mean in the same row with different superscripts differ (P<0.05)

Table 4-4. Protein content in Korean-native cattle colostrum and milk(%)

Time after calving	Lactation			Mean
	1	2	4-6	
0-6hr	14.2 ± 0.6a	15.3 ± 0.5a	17.2 ± 0.3b	15.6 ± 0.5
24hr	5.0 ± 0.3a	6.2 ± 0.3b	4.9 ± 0.2a	5.4 ± 0.3
48hr	4.2 ± 0.2a	3.8 ± 0.2a	4.8 ± 0.4b	4.3 ± 0.3
120hr	3.7 ± 0.1b	3.2 ± 0.1a	3.6 ± 0.1b	3.5 ± 0.1
Milk	3.6 ± 0.1a	3.1 ± 0.1a	3.3 ± 0.1a	3.4 ± 0.1

*Mean in the same row with different superscripts differ (P<0.05)

Table 4-4

가 14.2 ± 0.6% 4-6 17.2 ± 0.3% 120

24 $5.4 \pm 0.3\%$ 1/3 $15.6 \pm 0.5\%$
 Parrish et al., (1950) 60% 36%
 Oyeniyi and Hunter (1978) 84% 64%

가 .

.

, casein-N

. Holstein

IgG

가

120

2.

가.

1)

Holstein
 6 , 24 36
 - 20 . IgG
 whey 6,000 rpm 30 2
 pH 4.6 casein
 (Fey et al., 1976) whey pH 6.6
 - 20 .

2)

casein

(1987)

3) ELISA immunoglobulin G
 IgG single radial immunodiffusion (VMRD, Inc., Pullman, WA). Kit
 3ul wells ,
 well 3ul . Plate cover 24
 . ring . Standard curve
 sample IgG .

Holstein
 (Table 4-5).

Table 4-5. Concentration of colostrum and milk ingredients in Korean-native cattle

Time after birth	Spec. Gravity	TS*	Ash	Protein	Fat	Lactose
6hr	1.044	27.45	1.22	11.66	10.74	3.84
24hr	1.030	13.98	0.87	3.99	4.88	4.24
36hr	1.030		0.86	3.85	4.08	4.75

*total solid

Ig Holstein
 (Table 6). IgG1 Holstein
 59% Ig .

Holstein

2

가

Ig

Ig

Table 6. Concentration and relative percentage of immunoglobulins in the serum and mammary secretion

Cattle		Concentration (mg/ml)			Relative Ratio (%)		
		serum	colostrum	milk	serum	colostrum	milk
Holstein	IgG1	10.8	45.4	0.62	49.3	81.9	42.5
	IgG2	8.1	2.7	0.31	37.0	4.9	21.2
	IgA	0.6	3.4	0.42	2.7	6.1	28.8
	IgM	2.4	3.9	0.11	11.0	7.0	7.5
Korean-native	IgG1	9.3	26.8	0.47	49.2	77.2	40.9
	IgG2	7.6	1.7	0.26	40.2	4.9	22.6
	IgA	0.4	3.2	0.32	2.1	9.2	27.8
	IgM	1.6	3.0	0.10	8.5	8.6	8.7

Ig

가

가

Ig

Ig

Ig

가

IgG1

Table 4-7

Table 4-7. The formulation of immunoglobulin enforced artificial colostrum

Feeding	Col- 200	Col- 300	Col- 400
DCP	200mg Ig/ml	300mg Ig/ml	400mg Ig/ml
D- Electrolyte (%)	4.0	4.0	4.0
Lactoferrin (mg)	0.2	0.2	0.2

DCP (Dried Colostrum Powder)

Ig , 24
 Ig
 Ig
 , Ig , Ig
 Ig , , Ig
 가 , Ig
 . IgG 가 . IgG
 Ig
 12 100g Ig Ig
 Ig
 1811 가
 26% 가 2L
 74% 가 <3.84L 23% 가
 <8.5kg IgG1 <35mg/ml
 Ig
 40% IgG ml 10mg

Table 4- 8. Commercial products of enhance immune system

Products	Application	Manufacturer
DCW concentrate™	Dried colostrum powder	DC Smarte Int. Inc.
D- Electrolyte™	Anti- Diarrhea	Walco Int. Inc., USA
Kaolin- Pectin Anti- Diarrheal Liquid™	Anti- Diarrhea	Durvet Inc., USA
DCP	Dried colostrum powder	Milk Specialties Co.,

Ig

(Table 8). DCW concentrate™ DCP

Ig

Anti- Diarrheal

D- Electrolyte™ Kaolin- Pectin Anti- Diarrheal Liquid™

가

Ig Anti- Diarrheal

Ig Anti- Diarrheal

3.

가.

1)

2 3
500ml 2 1kg

는 즉시 냉동하였다. 수집된 초유를 냉동상태로 실험실로 운반하여 면역항체 분석과 냉동건조에 이용하였다.

냉동된 초유는 해동하여 여러 어미소로부터 수집된 초유를 섞어서 균일한 batch를 만들었다. 1000ml 크기의 냉동건조용 튜브에 냉동 coating하고 freeze dryer에 연결하여 완전히 냉동건조 하였다. 건조된 초유를 수거하여 분말로 분쇄하고 냉동실에 보관하였다(그림 1).

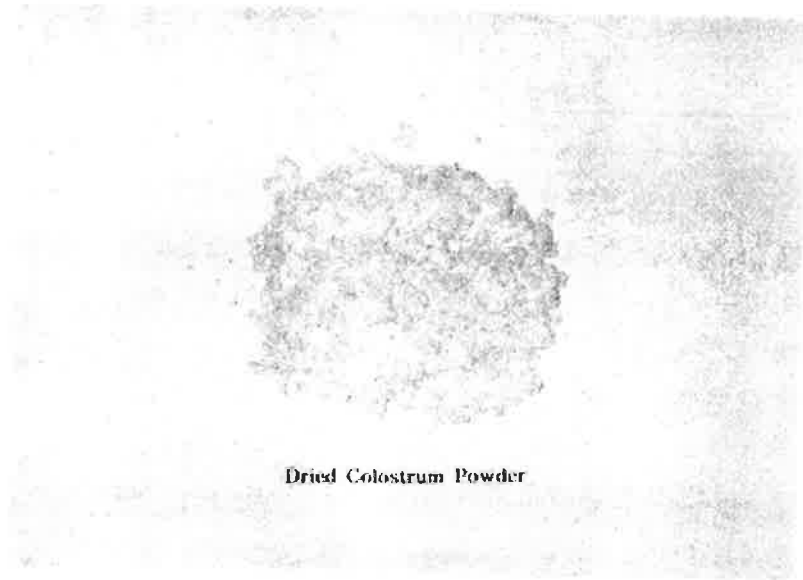


Figure 1. Dried colostrum powder collected from local dairy farm.

2) 냉동건조 초유의 면역항체 수준측정

냉동건조된 초유를 보통 우유에 희석하였다. 희석수준은 100ml 우유에 0, 1, 5, 10, 20g을 완전히 용해하여 상기 기술한 방법에 따라 면역항체 수준을 측정하였다(Table 4-9), 면역 항체수준은 신선한 초유에 비하여 냉동건조한 초유에서 별다른 감소현상은 없었다. 또한 정상우유에 대한 희석 수준에 따라 면역항체의 수준은 커다란 변화를 보이지 않았다. 이러한 결과로 볼 때 냉동건조한 초유를 면역증강제를 위한 인공초유제조에 적합한 것으로 보인다. 그러나, 많은 량을 냉동건조하기에는 비용이 소요될 것이다.

Table 4- 9. Immunoglobulin level in dried colostrum powder(DCP)

DCP(g/dl) (mg/ml)	0	1	5	10	20	Fresh Colostrum
IgG1	1.6	2.8	11.3	23.6	43.2	45.4
IgG2	0.07	0.11	0.67	1.52	2.71	2.7
IgA	0.12	0.17	0.83	1.62	3.49	3.4
IgM	0.15	0.19	0.92	1.82	3.81	3.9

3) 가

가 Anti- diarrheal D- Electrolyte .

4)

2

가

가

Immunoglobulin 가 200mg/ml(Col- 200),
300mg/ml(Col- 300), 400mg/ml(Col- 400)

Col- 200, Col- 300, Col- 400 500ml 3

immunoglobulin

(Table 4- 10).

Table 4-10. Serum immunoglobulin after artificial colostrum feeding(mg/ml)

Feeding	colostrum	Col- 200	Col- 300	Col- 400
Before	1.3	2.3	1.7	2.0
6hr	6.2	37.3	49.5	53.1
12hr	3.6	8.6	7.2	4.8
24hr	1.7	4.3	2.0	2.7

500ml
 6, 12,24
 total immunoglobulin
 6
 Col- 200 6 37.3 mg/ml
 , Col- 300 49.5mg/ml , Col- 400 mg/ml
 53.1 mg/ml 12 24
 가 가
 가

1)
 , casein- N . Holstein
 IgG
 가 120

2) Ig Anti- Diarrheal
 Ig Anti- Diarrheal

3)

6

가

가

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J dairy Sci. 76:831- 836.

, , . 1994. ELISA
. 36(5):453- 458.

IgG

, , , , . 1991.
. 13(4):225

Immunoglobulin G alc

2

1.

(Ig)
(Ivanoff , 1975).

(failure of passive transfer; FPT) Ig
(MacGuire , 1976), FPT
(Tizzard , 1996).

10 30%가 (Logan , 1974).
Ig
(Gay , 1965).
Ig 가 (McEwan , 1970)
(Logan , 1974), (RV)
(Klingenberg , 1999) (Boyd , 1972)
(Rea , 1995; MacGuire , 1976)
(Williams
, 1975). , Wittum (1995) (Total Protein)
Ig 가 12
5
8 9 (, 1989).
, FPT (Tizzard , 1996; Reburn ,
1995), IgG (Nollet , 1999;
Fernandez , 1998), (Quigley , 1996;
Selim , 1995) , Ig
(Quigley , 1996) (Quigley , 1996;
Crowford , 1995) , FPT
가가

가 Ig
 (Kirk , 1988) (Fernandez ,
 1998) 가 FPT 가
 . Nollet (1999)
 (Sanchez , 1993) *E. coli* F17+, F5+
 fimbria 가

. Selim (1995) *E. coli* O111:B4(J5)
 2 FPT
 가 .

bovine
 viral diarrhea virus (BVDV), bovine corona virus (BCV), infectious bovine
 rhinotracheitis virus (IBRV), parainfluenza-3 virus (PI-3), bovine respiratory
 syncytial virus (BRSV)

, 8 FPT
 TP Ig FPT 가

2.

가.

1)

3 14 1998 8

7

. 1 Bar Vac[®]Elite4- HSTM (Bovine Rhinotracheitis
 virus, Diarrhea-Parainfluenza²-Respiratory syncytial virus killed vaccine +
Haemophilus somnus bacterin; Boehringer Ingelheim) ScourGuard 3[®](K)
 (Bovine rota-coronavirus killed vaccine + *Escherichia coli* bacterin; Pfizer)
 2 1 가 . (R)

2 ml infectious bovine rhinotracheitis virus (PQ , 10⁵ TCID₅₀/ml),
 bovine respiratory syncytial virus (A51908 , 10⁵ TCID₅₀/ml),
 parainfluenza-3 virus (NADL , 10⁶ TCID₅₀/ml) , (D) 5
 ml bovine viral diarrhoea virus (KD-26-1 , 10⁵ TCID₅₀/ml) bovine
 corona virus (5 , 10⁵ TCID₅₀/ml) 2 2

2)

Quigley (1996)

4°C 24 3,000 rpm 20 3
 Wattman (No. 2) 3.0
 μm (Osmonics, USA), 0.8 μm , 0.45 μm membrane filter (Gelman, USA)
 -70°C
 IgG, IgM, IgA VMRDIM (Pullman,
 USA) (single radial diffusion tests; sRID)

(3 μl) 가

18-24

semilogarithmic Ig logarithm ,
 ring arithmetic

ring

Ig

(total protein; TP), (albumin; Alb)

(Johnson&Johnson clinical Diagnostics Inc. U.S.A)

(Johnson&Johnson vitros DT60 , DTSC)

, (globulin; Glob) TP Alb

가

14

BVDV, BCV, IBRV, PI-3V

가

Baker

(1985)

BVDV NADL , BCV Kagegawa , IBRV PQ7 , PI-3V
 NADL Marden-Darby
 bovine kindey (MDBK) 96 microplate
 . 56cC 30 -MEM 2
 (200 TCID₅₀/ml) 37cC 1
 . MDBK 가 2 5×10⁵/ml
 100 μl/well 5% humidified CO₂
 37cC 3 가

1)

(A,B) 23
 . A (9) 2 CattleMaster4
 (Bovine rhinotracheitis, Viral diarrhea, Parainfluenza3, Respiratory syncytial
 virus modified live and killed vaccine; Pfizer) Scourguard 3[®]
 , B (14)
 3 . 1999 6 8
 . 1
 , 80cC 100cC 1 2
 1 3

2)

FPT 1-7 23
 . (7) (IP) (6
) (IV) . (IP 5 , IV 3)
 . 2
 (IV)
 Kg 0.28 g , 0.16 g 1

(Table 4- 11).

Table 4- 11. Experimental design

Breed	Route	No. tested	BW(Kg) at start	Mean dose(g) of IgG / Kg of BW
Korean native cattle	Control	5	25 ± 4	-
	IP	7	21 ± 3	0.28 ± 0.17
	I- IV	2	25 ± 3	0.23 ± 0.05
Holstein	Control	3	36 ± 5	-
	IV	6	39 ± 3	0.16 ± 0.06

IP ; Intreperitoneous, I-IV ; Group with clinical signs like recumbency, IV ; Intravenous BW ; Body weight.

3)

(- 1), 1 (1), 7 (7),
 14 (14), 30 (30) .
 4cC 24 3,000 rpm 10
 3
 - 70cC .

4)

EDTA가
 (Medonic CA 530, Oden, Sweden)
 (RBC), (WBC), (Hb), (Hct),
 (MCV), (MCH), (MCHC)

5)

가)

(Sodium sulfite precipitation test; SSPT) Stone
 (1969) . 14 g, 16 g, 18 g
 anhydrous Na₂SO₃ 100 ml
 , 1.9 ml 0.1 ml
 (22°C) 1 .

(Table 4-12).

Table 4-12. Estimation of serum immunoglobulin concentration by sodium sulfite precipitation test

Concentration of Ig (mg/ml)	Sodium sulfite		
	14%	16%	18%
< 5	-	-	+
5 to 15	-	+	+
> 15	+	+	+

- ; No precipitation 1 hour after the reaction.

+ ; Flocculent precipitation 1 hour after the reaction.

) , , , 가
 , , 가

1)

Davis (1990)
 acid citrate dextrose (ACD ; sodium citrate 22.0 g,
 citric acid 7.3 g, dextrose 24.5 g, D.W 1,000 ml) 3:1
 , 1,500 rpm 30 . Buffy coat

36°C 가 0.87% tris-buffered ammonium chloride (tris
 -NH₄CL, 0.01M tris, pH 7.2) 37°C 5
 가 . 1,500 rpm 10
 (PBS; sodium chloride 7.6 g, disodium
 phosphate 1.2 g, monosodium phosphate 0.1 g, monopotassium phosphate 0.2
 g, pH 7.2) ACD 9:1 PBS-ACD buffer 3
 . 1.0 ml PBS tryphan
 blue 가 1 × 10⁷/ml

2)

MHC
 class , T cell , BoCD2 cell, BoCD4 cell, BoCD8 cell, N cell, sIgM,
 Monocyte 8 (Table 4- 13).

Table 4- 13. Monoclonal antibodies specifically reactive with bovine leukocyte differentiation antigens

Specificity	Monoclonal antibodies	Cell type
MHC class	H58A	All nucleated cell
MHC class	H42A (DP)	Antigen presenting cell
BoCD2	BAQ95A	T cell
BoCD4	CACT 138A	T helper/inducer cell
BoCD8	CACT 80C	T cytotoxic /suppressor cell
TCR1- N12	CACT 61A	Non T/B cell
sIgM	PIG45A	B cell
G+M	CH59B	Monocyte

Specificity ; Bovine leukocyte differentiation molecules.

Monoclonal antibodies ; Monoclonal antibodies that specifically react with leukocyte differentiation antigen. Cell type ; Cells expressing molecules.

3)

Davis (1990)

50 $\mu\ell$ (15 $\mu\text{g}/\text{ml}$) $1 \times 10^7/\text{ml}$ 가
4cC 30 , 4cC first washing buffer (PBS 450 ml,
ACD 50 ml, 20% NaN3 5 ml, gamma globulin free horse serum 10 ml, 250
mM EDTA 20 ml, 0.5% phenol red 1 ml) 3 (2,000 rpm, 3 , 4cC)

Vortex

200 2 (FITC-conjugated goat anti-mouse
IgG + IgM antibody , Caltag Lab, INC, USA) 100 $\mu\ell$ 가
4cC 30 4cC 2 (1
) 3 2% PBS-formalin (38%
formalin 20 ml, PBS 980 ml) 200 $\mu\ell/\text{well}$
(4cC)
flowcytometry 2,000

Dickinson (USA) Conson 32 Lysys Becton

SAS (Correlation anylisis) Student t'

Wilkins (1994)

$$\frac{\nabla \text{IgG}}{\sum \text{IgG}/\text{BW}} = \text{transferfactor (TF)}$$

IgG = final serum IgG (mg/ml) - initial serum IgG (mg/ml).

IgG = total administered IgG (mg).

BW = initial body weighct (kg). P < 0.05.

3.

가.

14

TP, Alb, Glob

Table 14

TP, Alb, Glob

(IgG, IgM)

2.2 g/dℓ 110 mg/dℓ

TP

5.8 ± 0.9 g/dℓ, Alb 2.2 ± 0.5 g/dℓ, Glob 3.6 ± 0.5 g/dℓ, IgG 2.2 ± 0.5 g/dℓ

Table 4-14. Mean values of total protein, albumin and globulin in bovine immune sera used

Group	TP (g/dℓ)	Alb (g/dℓ)	Glob (g/dℓ)	IgG (g/dℓ)	IgM (mg/dℓ)	IgA (mg/dℓ)
R (N=7)	5.5 ± 0.9	2.3 ± 0.5	3.2 ± 0.5	2.2 ± 0.5	110.3 ± 18.5	10.1 ± 4.2
D (N=7)	6.0 ± 1.0	2.0 ± 0.5	4.0 ± 0.5	2.2 ± 0.5	110.7 ± 18.1	8.9 ± 4.2

R group was intramuscularly injected with infectious bovine rhinotracheitis, parainfluenza 3 virus, and bovine respiratory syncytial virus after commercially respiratory and enteric pathogen combined vaccination.

D group was intramuscularly injected with bovine viral diarrhea virus and bovine corona virus after commercially respiratory and enteric pathogen combined vaccination.

Values were Mean ± standard deviation.

가

가 Table 4-15

가

가

BVDV ($8.1 \pm 1.0 \log_2$)가 가 (P<0.01) BCV ($6.4 \pm 2.3 \log_2$)
 IBRV ($1.7 \pm 1.9 \log_2$) PI-3V ($1.7 \pm 1.9 \log_2$)
 가 BVDV
 1.5 1.4 가 ,
 1 IBRV PI-3V 가
 가 2 8 \log_2 ,
 BVDV BCV 가가 8 \log_2
 가 BVDV $8.1 \pm 1.0 \log_2$, BCV $6.4 \pm 2.3 \log_2$, IBRV 1.7 ± 1.9
 \log_2 , PI-3V $1.7 \pm 1.9 \log_2$.

Table 4-15. Mean neutralization antibody titers against some viruses of cattle for production of immune sera before and after commercially combined viral vaccine and then viral challenge

Group		Antibody titer ^b (\log_2) against			
		BVDV	BCV	IBRV	PI-3V
R (n=7)	before	5.4 ± 1.7	4.7 ± 2.6	- c	-
	after	8.1 ± 0.9	5.9 ± 2.9	2.4 ± 2.6	2.4 ± 2.6
D (n=7)	before	5.7 ± 1.4	6.3 ± 0.8	-	-
	after	8.0 ± 1.2	7.0 ± 1.5	-	-

R group was intramuscularly injected with infectious bovine rhinotracheitis, parainfluenza 3 virus, and bovine respiratory syncytial virus after commercially respiratory and enteric pathogen combined vaccination.

D group was intramuscularly injected with bovine viral diarrhea virus and bovine corona virus after commercially respiratory and enteric pathogen combined vaccination.

a ; Bar Vac[®]Elite4- HSTM (Bovine Rhinotracheitis virus, Diarrhea-Parainfluenza3- Respiratory syncytial virus killed vaccine + *Haemophilus somnus* bacterin; Boehringer Ingelheim), ScourGuard 3[®](K)(Bovine rota-coronavirus killed vaccine + *Escherichia coli* bacterin;Pfizer)

b ; Reciprocal of final serum dilution inhibiting viral cytopathic effects.

c ; Negative for viral neutralization at 1:2 dilution. Values were mean \pm standard deviation.

FPT

1) FPT

FPT 가 TP <4.5 g/dℓ, IgG <0.8 g/dℓ, SSPT <1+ 가 3 (17%), 3가 6 (26%), 13 (57%), 2가 4 (Table 4-16)

Table 4-16. Criteria of failure of passive transfer for calves to be used

Breed	Route	Criteria to be applied (n)					
		TP+IgG	TP+SSPT	TP+IgG+SSPT	IgG	TP	SSPT
Korean native cattle	Con (n=5)	1	1	1	1	1	-
	IP (n=7)	2	-	1	-	4	-
	I-IV(n=2)	-	-	2	-	-	-
Holstein	Con (n=3)	-	-	-	2	1	-
	IV (n=6)	-	-	2	2	2	-

Cutoff Point ; TP <4.5 g/dℓ, IgG < 800 mg/dℓ, SSPT <1+.

TP ; Serum total protein , IgG : Serum immunoglobulin G, SSPT : Sodium sulfate precipitation test.

IP ; Intraperitoneous, I-IV ; Group with clinical signs like recumbency, Con ; No treatment. IV; Intravenous.

23 가 TP, Alb, Glob, IgG 가 4.0- 4.4 g/dℓ 3.1- 3.6 g/dℓ , Alb 1.9- 2.1 g/dℓ 1.6- 1.7 g/dℓ , Glob 2.0- 2.1 g/dℓ 1.5- 2.0 g/dℓ 가 (Table 4-17).

Table 4-17. Total protein, globulin, albumin and immunoglobulin levels of neonatal calves with failure of passive transfer before administration of bovine immune sera

Breed	Route	Age (days)	TP (g/dℓ)	Alb (g/dℓ)	Glob (g/dℓ)	IgG (g/dℓ)	IgM (mg/dℓ)	IgA (mg/dℓ)
Korean native cattle	Con (n=5)	3 ± 1	3.5 ± 0.8	1.7 ± 0.4	1.7 ± 0.9	0.8 ± 0.4	89.4 ± 27.6	17.0 ± 10.1
	IP (n=7)	3 ± 1	3.1 ± 0.9	1.6 ± 0.4	1.5 ± 0.6	1.3 ± 0.9	62.8 ± 8.4	17.8 ± 15.8
	I-IV (n=2)	2 ± 1	3.6 ± 0.8	1.7 ± 0.4	2.0 ± 0.4	0.6 ± 0.1	104.6 ± 1.7	10.0 ± 5.0
Holstein	Con (n=6)	3 ± 1	4.3 ± 1.9	2.1 ± 0.5	2.1 ± 1.8	1.2 ± 1.0	84.3 ± 35.4	27.1 ± 17.2
	IV (n=3)	3 ± 2	4.0 ± 0.9	1.9 ± 0.4	2.0 ± 0.7	1.0 ± 0.8	88.0 ± 30.2	27.1 ± 17.2

TP ; Total protein, Alb ; Albumin, Glob ; Globulin.

IP ; Intraperitoneous, I-IV ; Group with clinical signs like recumbency, Con ; No treatment.

IV ; Intravenous, Values are Mean ± standard deviation.

2)

FPT , , ,
 , 7 13
 , 30 (2)
 10 12 . 1
 2 가 3 15
 (<IgG 770mg/dℓ)

3)

1 , 1 , 8 , 14 , 30 (RBC),
 (Hct), (WBC), (Hb), (MCV),
 (MCH), (MCHC) (Figure 2).

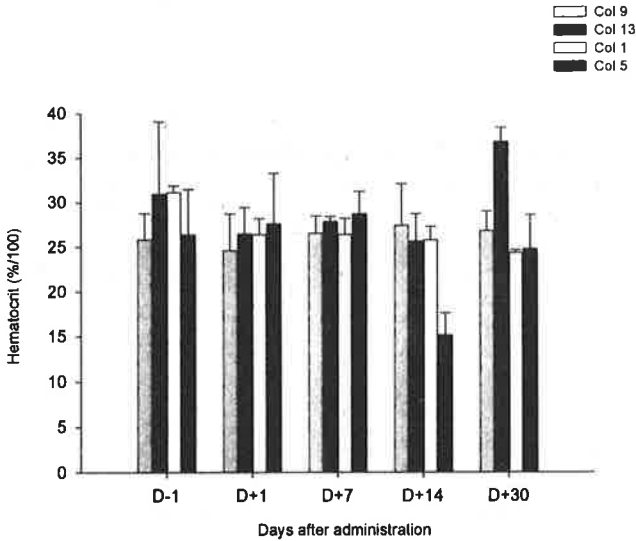


Figure 4-2. Changes of haematocrit neonatal calves with failure of passive transfer administered with bovine immune sera.

K- IP: Intraperitoneous, H- IV: Intravenous, Con : No treatment.

모든 면역혈청 투여군에서는 혈액학적 수치가 전 실험기간동안 정상범위에서 두렷한 변화는 없었다. 다만, 투여 후 30일째 한우 IP 대조군의 Hct ($36.8 \pm 2.26\%$)가 상승하였고 홀스타인 IV 대조군은 투여 후 14일 제에 크게 떨어지는 것을 확인할 수 있었다.

4) 면역혈청의 FPT 보정효과

면역혈청 투여 전후 FPT 보정 효과를 조사한 결과는 Table 18과 같다. 한우와 홀스타인 송아지의 대조군에서는 투여전의 FPT 선정우 5두와 3두가 투여 후 14일부터 점차적으로 FPT 보정효과가 인정되었으나 일부 송아지는 투여 후 30일째에도 보정 효과가 인정되지 않았다. 이와는 반대로 복강투여군에 있

가

Table 4-18. Corrective effect of calves with failure of passive transfer after administration of bovine immune sera

Breed	Route	Day(s)	Criteria to be applied (n)					NFPT
			TP+IgG	TP+SSPT	TP+IgG+SSPT	IgG	TP	
Korean native cattle	Con (n=5)	-1	1	1	1	1	1	-
		1	-	1	1	1	2	-
		7	-	1	1	1	2	-
		14	1	-	-	-	3	1
		30	-	-	-	-	-	5
	IP (n=7)	-1	2	-	1	-	4	-
		1	-	-	-	5	-	2
		7	-	-	-	4	1	2
		14	-	-	-	-	-	7
		30	-	-	-	-	-	7
Holstein	Con (n=3)	-1	-	-	-	2	1	-
		1	1	-	-	1	1	-
		7	-	-	-	-	2	1
		14	-	-	-	-	1	2
		30	-	-	-	-	1	2
	IV (n=6)	-1	-	-	2	2	2	-
		1	2	-	2	-	-	2
		7	2	-	-	1	1	2
		14	-	-	-	-	-	6
		30	-	-	-	-	-	6

Cutoff Point ; TP <4.5 g/dl, IgG < 800 mg/dl, SSPT <1+. TP ; Total protein, SSPT ; Sodium sulfate precipitation test. NFPT ; No. of corrected cases from failure of passive transfer. -1 ; one day before inoculation, 1 ; one day after inoculation. IP ; Intraperitoneous, IV ; Intravenous, Con ; No treatment.

5) FPT

Ig

Ig
(TF; mg/dℓ/IgG administered/Kg BW) IV 가 ,
IP I-IV . IP I-IV
IgG IV 1.75 1.38 (P<0.05)
(P<0.01)가 (Table 4- 19).

Table 4- 19. Transfer factor of immune sera in calves with failure of passive transfer

Breed	Route	Clinical signs	Total administered IgG (g/kg)	Transfer factor (mg/dℓ/G administered/kgBW)
Korean	IP	No	0.28 ± 0.17	5.46 ± 2.74
Holstein	IV	No	0.16 ± 0.06	11.17 ± 1.27
Korean	IV	Recumbency	0.23 ± 0.05	1.31 ± 0.36

Korean ; Korean native cattle. IP ; Intraperitoneous, IV ; Intravenous. Values were Mean ± standard deviation. (P<0.05).

1 1, 7, 14 30 TP, Alb, Glob, IgG
Table 10 . TP (3.5 ± 0.8 g/dℓ)
dℓ) 7 (4.8 ± 0.2 g/dℓ) ,
(3.1 ± 0.8 g/dℓ) 1 가
7 14 3.9 ± 1.4 g/dℓ 4.8 ± 0.7 g/dℓ
가
(3.8 ± 0.8 g/dℓ) 1 7
4.4 ± 1.1 g/dℓ 4.6 ± 0.9 g/dℓ 가 .
Alb Glob (Table 20),
가 Alb (1.6 ± 0.4 g/dℓ 1.9 ± 0.4
g/dℓ) 7 (1.9 ± 0.5 g/dℓ 2.0 ± 0.3 g/dℓ) 가
Glob (1.5 ± 0.6 g/dℓ 1.9 ± 0.7 g/dℓ)

Table 4-20. Total protein, globulin and immunoglobulin G levels in calves with failure of passive transfer before and after the administration of bovine immune sera

Breed	Route	Day(s)	TP (g/dℓ)	Alb (g/dℓ)	Glob (g/dℓ)	IgG (g/dℓ)
Korean native	Con (n=5)	- 1	3.5 ± 0.8	1.7 ± 0.4	1.7 ± 0.9	1.2 ± 1.0
		1	3.8 ± 1.0	1.9 ± 0.5	2.0 ± 0.5	1.0 ± 0.4
		7	4.8 ± 0.2	2.1 ± 0.1	2.6 ± 0.1	1.5 ± 0.2
		14	2.5 ± 3.3	1.1 ± 1.5	1.4 ± 1.8	1.3 ± 0.4
		30	5.8 ± 1.0	2.3 ± 0.3	3.5 ± 0.7	1.0 ± 0.4
	IP (n=7)	- 1	3.1 ± 0.8	1.6 ± 0.4	1.5 ± 0.6	1.3 ± 0.9
		1	3.6 ± 1.5	1.7 ± 0.6	1.8 ± 0.9	1.7 ± 0.6
		7	3.9 ± 1.4	1.9 ± 0.5	2.0 ± 1.0	1.7 ± 0.8
		14	4.8 ± 0.7	2.2 ± 0.5	2.6 ± 0.5	1.7 ± 0.1
		30	4.9 ± 0.9	2.1 ± 0.5	2.8 ± 1.3	1.8 ± 0.8
Holstein	Con (n=3)	- 1	4.8 ± 1.9	2.2 ± 0.4	2.6 ± 1.8	0.8 ± 0.4
		1	4.3 ± 2.3	2.0 ± 0.3	2.2 ± 2.0	1.8 ± 0.8
		7	4.3 ± 1.4	1.6 ± 0.5	2.7 ± 1.0	1.8 ± 0.4
		14	3.8 ± 1.3	2.4 ± 0.2	1.5 ± 1.5	1.7 ± 0.5
		30	4.8 ± 0.5	2.0 ± 0.1	2.8 ± 0.6	1.4 ± 0.4
	IV (n=6)	- 1	3.8 ± 0.8	1.9 ± 0.4	1.9 ± 0.7	0.9 ± 0.7
		1	4.4 ± 1.1	2.0 ± 0.3	2.4 ± 0.8	1.5 ± 0.9
		7	4.6 ± 0.9	2.0 ± 0.3	2.5 ± 0.7	1.2 ± 0.5
		14	4.4 ± 0.8	2.1 ± 0.2	2.6 ± 2.5	1.1 ± 0.3
		30	4.5 ± 0.9	2.2 ± 0.3	2.3 ± 0.6	1.1 ± 0.2

TP ; Total protein, Alb ; Albumin, Glob ; Globulin. IP ; Intraperitoneous, Con; No treatment. IV ; Intravenous. - 1 ; one day before inoculation, 1 ; one day after inoculation. Values were Mean ± standard deviation.

6) 가
 BVDV, BCV 가
 BVDV 가
 (P<0.05),
 1
 5 log₂ 가가 30
 , 8 log₂ 가가 30
 .
 가가 30 1 가
 6 log₂ 가가
 (Figure 4-3).
 , BCV 가
 (P<0.01) 5 log₂ 30 7
 log₂ , 가가
 (Fig3).
 8 log₂ 가 14 5
 log₂ 가가
 BVDV 가 3.5 log₂ 5 log₂ BCV 5.0 log 6.5 log
 (Figure 4-4).

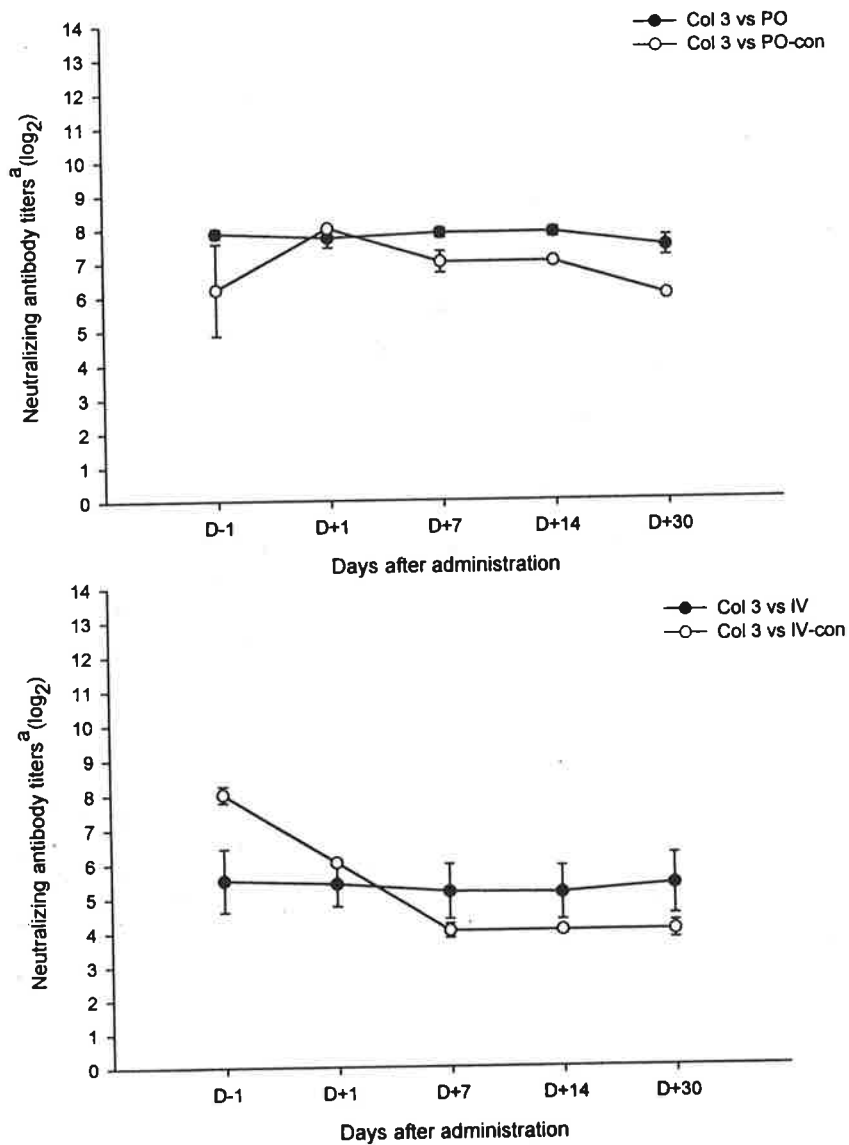


Figure 4-3. Changes of antibody titers against bovine viral diarrhea virus in neonatal calves with failure of passive transfer which were administered the bovine immune sera

a: Reciprocal of final serum dilution inhibiting viral cytopathic effects.
 IP: Intraperitoneous, Con: Not treatment, IV: Intravenous.

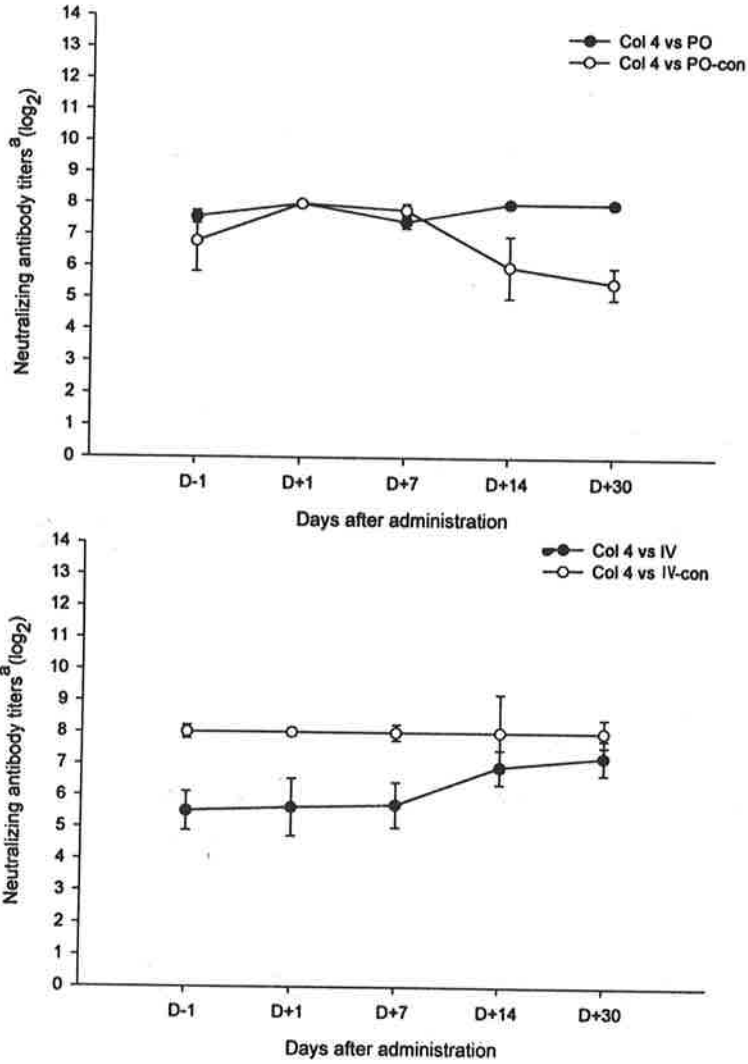


Figure 4-4. Changes of antibody titers against bovine coronavirus of neonatal calves with failure of passive transfer which were administered with the immune sera.

a: Reciprocal of final serum dilution inhibiting viral cytopathic effects.
 IP: Intraperitoneous, Con: No treatment. IV: Intravenous.

7)

가

7 , 14 , 21

(Table

21,22; Figure 5).

T, B,

N 33.0- 47.0% , 7.6- 9.7% , 42.7- 53.4%

. T

CD4/CD8 1.1- 1.4

가 MHC class

12.2- 16.1% , Monocyte 9.3- 11.8%

CD2, CD4, CD8

가 (P<0.05), CD2 (33.0%)

가 21 44.6% 가 ,

CD4 . CD4/CD8

가 (2) 14 (0.9 ± 0.2)

1.0 .

sIgM B

가 ,

(9.3 ±

4.1%) 7 14 13.7 ± 8.8% 15.7 ± 6.0%

가 .

, 10- 20

CD4 , CD8

가, sIgM 가, N 가 .

Table 4- 21. Percentage of lymphocyte subsets in calves with failure of passive transfer after the administration of bovine immune sera

Breed	Route	Day(s)	Monoclonal antibodies(%)				
			ClassI	ClassII	sIgM	N cell	M
Korean native	Con (n=5)	- 1	86.3 ± 7.4	12.3 ± 5.3	9.7 ± 2.6	53.4 ± 18.9	11.5 ± 0.5
		7	97.1 ± 0.5	25.3 ± 2.2	9.1 ± 1.0	45.3 ± 12.3	8.5 ± 2.7
		14	96.2 ± 3.7	20.5 ± 11.3	7.8 ± 3.0	38.1 ± 18.1	8.7 ± 3.6
		21	97.0 ± 3.7	21.5 ± 12.0	7.6 ± 2.9	35.0 ± 16.0	8.7 ± 3.6
	IP (n=7)	- 1	91.3 ± 2.1	14.6 ± 3.8	9.0 ± 3.5	42.7 ± 11.5	11.8 ± 6.9
		7	93.4 ± 4.3	21.3 ± 5.5	12.4 ± 2.7	43.3 ± 5.4	9.8 ± 2.7
		14	94.0 ± 3.6	16.5 ± 4.6	8.1 ± 3.5	43.9 ± 8.1	14.2 ± 6.8
		21	94.0 ± 3.7	16.5 ± 4.6	8.9 ± 3.5	43.0 ± 9.0	14.3 ± 6.9
Holstein	Con (n=3)	- 1	96.3 ± 3.5	12.2 ± 5.9	7.6 ± 3.1	48.3 ± 8.6	10.5 ± 2.1
		7	91.3 ± 2.8	21.6 ± 0.8	5.0 ± 1.4	46.0 ± 1.1	11.1 ± 1.6
		14	91.3 ± 0.1	21.6 ± 0.4	10.6 ± 0.9	40.6 ± 0.8	11.1 ± 3.7
		21	95.6 ± 4.4	25.1 ± 0.6	9.1 ± 0.1	45.9 ± 2.1	7.6 ± 0.6
	IV (n=6)	- 1	91.9 ± 6.7	16.1 ± 8.8	8.9 ± 5.6	46.9 ± 10.2	9.3 ± 4.1
		7	93.5 ± 6.7	15.9 ± 6.7	9.4 ± 4.3	44.7 ± 12.2	13.7 ± 8.8
		14	94.6 ± 6.4	15.5 ± 4.7	11.0 ± 4.6	44.1 ± 10.2	15.7 ± 6.0
		21	91.0 ± 8.9	17.5 ± 6.3	12.0 ± 4.8	50.2 ± 10.8	13.1 ± 6.4

- 1 ; one day before inoculation, 7 ; 7 day after inoculation. IP ; Intraperitoneous, Con ; No treatment, IV ; Intravenous. Values were mean ± standard deviation.

Table 4- 22. Percentage of lymphocyte subsets in calves with failure of passive transfer after the administration of bovine immune sera

Breed	Route	Day(s)	Monoclonal antibodies(%)			
			CD2	CD4	CD8	CD4/ CD8
Korean native	Con (n=5)	- 1	41.3 ± 12.4	21.3 ± 8.7	17.1 ± 2.6	1.2 ± 0.3
		7	37.4 ± 21.2	21.8 ± 4.1	14.9 ± 5.0	1.5 ± 0.2
		14	37.1 ± 12.4	23.0 ± 0.2	17.9 ± 0.3	1.3 ± 0.1
		21	36.0 ± 12.1	21.9 ± 0.2	17.0 ± 0.3	1.2 ± 0.1
	IP (n=7)	- 1	47.1 ± 13.8	13.5 ± 2.2	11.1 ± 3.9	1.4 ± 0.6
		7	50.8 ± 14.8	18.2 ± 3.5	12.7 ± 2.0	1.5 ± 0.5
		14	48.0 ± 16.1	19.7 ± 6.8	13.1 ± 4.0	1.5 ± 0.4
		21	48.0 ± 14.0	19.8 ± 4.0	13.1 ± 3.1	1.5 ± 0.4
Holstein	Con (n=3)	- 1	33.1 ± 5.3	17.5 ± 7.8	15.7 ± 3.5	1.1 ± 0.3
		7	41.5 ± 0.7	18.2 ± 0.6	12.3 ± 0.4	1.4 ± 0.2
		14	41.5 ± 7.1	10.1 ± 0.1	16.5 ± 3.0	0.9 ± 0.2
		21	46.5 ± 2.1	18.0 ± 1.1	9.6 ± 0.6	1.5 ± 0.6
	IV (n=6)	- 1	33.0 ± 8.9	16.8 ± 8.2	14.4 ± 4.8	1.2 ± 0.4
		7	37.4 ± 17.4	18.6 ± 7.2	13.8 ± 6.0	1.5 ± 0.7
		14	39.5 ± 10.4	18.1 ± 4.6	15.3 ± 4.9	1.3 ± 0.7
		21	44.6 ± 5.4	24.5 ± 11.3	16.7 ± 8.7	1.5 ± 0.6

- 1 ; one day before inoculation, 7 ; 7 day after inoculation. IP ; Intraperitoneous, Con ; No treatment, IV ; Intravenous. Values are Mean ± standard deviation.

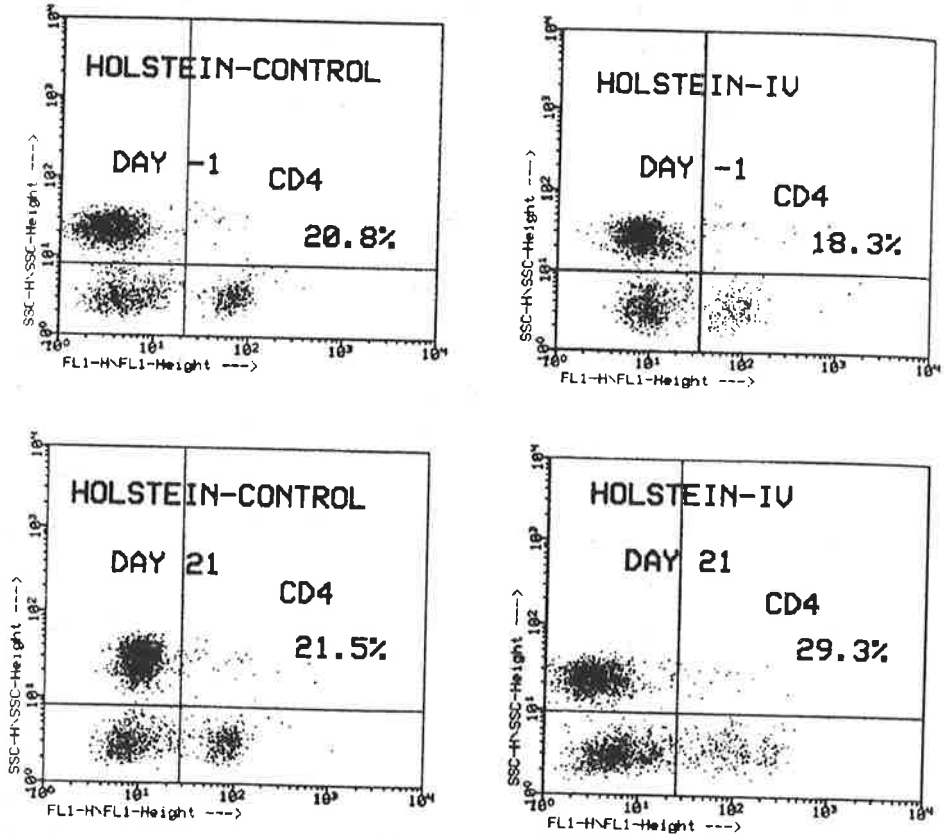


Fig 4-5. Change of percentage of CD4 before and after the administration of bovine immune serum to calves with failure of passive transfer.

4. 고찰

초유의 수동전이 부전의 치료를 위하여 초유를 직접먹이거나 IgG의 경구투여 하는 것이 야외상황에서 많이 적용되고 있지만 IgG의 경구투여는 송아지의 장관내 흡수능력을 고려하면 생후 12시간 이내에 이루어져야 한다. 하지만 현실적으로 FPT는 생후 12시간 이후에 관찰이 가능하며, 장의 흡수 폐쇄 시간이 출생 후 24-48시간 이내에 이루어지기 때문에 (Scott 등, 1979) IgG를 다른 경로로 투여하는 것이 필요하다.

한편, FPT 보정을 위한 전혈의 사용은 1-2L의 양을 송아지에게 투여 (Reburn 등 1995)가 권장되지만 시행시간과 용량의 부담감이 있으며, 채취한

1 2 , Ig
(Quigley , 1996).
가 1
가 (Jeffcott , 1974)
FPT
가 Grahan (1998) (1995),
(1989)
IBRV, BRSV, PI-3V, BVDV BCV
bovine rota virus (BRV) 가
가
(Table 5) (D) 가
(R)
1
가
가가 2 log2
가 가
가 가 , FPT
가가
IgG 1.7 g/dℓ 3.2 g/dℓ
FPT IgG
1.9 g/dℓ 3.3 g/dℓ (Wilkins , 1993). IgM, IgA
Tizzard (1996) TP Alb Mirtuka (1981)
1
TP 가 가

FPT

가 ,

가

(Boyd , 1972)

FPT

Perino

(1995)

Stone

(1969)

TP

<4.5 g/dℓ

, IgG

<0.8 g/dℓ

, SSPT

1

FPT

가

2

7

(Rea

, 1996)

(1989)

2

TP 6.7 ± 0.9 g/dℓ, Glob 3.5 ± 9.7 g/dℓ, IgG 2.9 ± 0.8

g/dℓ, IgM 49.1 ± 29.9 mg/dℓ, IgA 19.4 ± 9.9 mg/dℓ

TP,

Glob, IgG가 2

, IgM IgA

MaBeath (1971)

2

Holstein

TP 4.8 ± 0.2 g/dℓ, IgG 0.6 ± 0.1 g/

dℓ, IgM 19.8 ± 22 mg/dℓ

가

가

1

1

FPT

1

TP 3.2 g/dℓ

IgG

0.91 g/dℓ

가

가

1

10 12

Ig

(Clement, 1995; Julie, 1996; Virtala, 1996).

Hct가 30
Hct가 14
(P<0.05). Hct 가
(Fisher, 1972; Tennant, 1972)

, FPT
7 (Table 8),
Wilkins (1994)

IgG IgG
(TF; mg/dl/IgG administered/kgBW)
H- IV 11.17 ± 1.27, K- IP 5.46 ± 2.74, I- IV 1.40
± 0.21 1.75 ,
1.38 가 4- 10
가 . Wilkins (1994)

가 (P<0.01).
TP (Donovan ; 1998) Ig 가 (Boyd, 1972) Alb Glob
(Besser ;1993)

TP, Alb, Glob, IgG
Alb . TP Glob K- Con
(r=0.49) H- Con (r=0.47) 14 (7) 가 가
21 40 (r=0.47)
(r=0.53) 30 .
Logan (1975) IgG 가 21.0 , 4
(1989) IgG 가
21.1 10
Ig 8 Ig

가 , 40 Ig Ig

가 Ig

가 가가 BVDV, BCV

가 BVDV BCV 가가

가

가

가

(Davis , 1990; Wyatt , 1994; Sordillo , 1997).

Sordillo (1997) Major histocompatibility complex (MHC) Class I

T Class II (Antigen presenting cell; APC)

B , T . B

CD2 T

, CD4 (helper T) (cytokine) B

APC-MHC complex

, CD8 cytotoxic T cell

helper T cell IL-2 MHC class I , Class

I target cell , N

(NonT/NonB)가 T cell receptor (TCR2)

가 30

B 20% 20

(Wyatt , 1994). Wyatt (1994)

N cell (15- 30%), CD2 (51.8- 45.7%), CD4 (29.8- 21.7%),

CD8 (16- 17%), MHC class I (90-95%), MHC class II (16- 9%), G+M

(16- 8%) sIgM , Wilson (1996) 10

4 30 CD2 34% 30% ,
 CD4 20% 16% , CD8 9% 8% , N cell 35% 25%
 Menge (1999) 3-9 10
 CD2 ($42.8 \pm 5.9\%$), CD4 ($20.2 \pm 4.0\%$), CD8 ($20.8 \pm 3.8\%$), N cell ($20.2 \pm 7.6\%$),
 B cell ($10.0 \pm 3.9\%$), MHC class II ($33.6 \pm 5.5\%$), Monocyte ($18.9 \pm 6.8\%$)

(Menge , 1999). 3
 T, B, N 33.0- 47.0% , 7.6- 9.7% ,
 42.7- 53.4% , CD4/CD8 1.1- 1.4 , MHC class II 12.2- 16.1% ,
 Monocyte 9.3- 11.8%

CD2, CD4, CD8
 가 (P<0.05), CD2
 (33.0%) 가 21 44.6% 가
 CD4
 Monocyte

Monocyte
 ($9.3 \pm 4.1\%$) 7 14 $13.7 \pm 8.8\%$
 $15.7 \pm 6.0\%$ 가 T Monocyte
 Menge (1999)
 4

MHC class II Monocyte 가
 가 T
 Menge (1999)
 T

T 가 가
 10- 20

CD4
 , CD8 가, sIgM 가, N 가
 Sordillo (1997) CD4/CD8
 1 .

가

FPT
 FPT Ig 가 가
 가 가
 BVDV, BCV 가
 CD2, CD4 Monocyte 가
 FPT Ig
 가 .

5.

1 FPT
 FPT
 TP $5.8 \pm$
 0.9 g/dl , Alb $2.2 \pm 0.5 \text{ g/dl}$, Glob $3.6 \pm 0.5 \text{ g/dl}$, IgG $2.2 \pm 0.5 \text{ g/dl}$
 가 BVDV $8.1 \pm 1.0 \log_2$, BCV $6.4 \pm 2.3 \log_2$, PI-3V $1.9 \pm$
 $1.7 \log_2$, IBRV $1.9 \pm 1.7 \log_2$.
 1-7 23 , TP 4.5 g/dl ,
 IgG 0.8 g/dl , SSPT가 1+ FPT ,
 가 가 11 (48%), 2가 가 6
 (26%), 3가 6 (26%) . FPT 23
 (7) , (6)

5 3

2

Kg 0.28g

0.16g 1-7 1

13

2 (25%)가 10 12

2 3 15

FPT transfer factor

[TF; mg/dl(IgG) / Kg] 11.17

$\pm 1.27,$ $5.46 \pm 2.74,$ 1.40 ± 0.21

FPT

(P<0.01). 24 Ig

, BVDV, BCV 가

(P<0.05). FPT

7 FPT가

14 30 FPT . FPT

CD2, CD4 가

(P<0.05).

6.

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29; 163-170, 1989.

, :
29; 171-178, 1989.

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29; 155-162, 1989.

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35: 615-623, 1995.

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1) 가 2) 3)

4) 5) ()

가 ?

1) 2)

3) 가 4) ()

(IBR)			
(BRSV)			
IBR+ (BVD-MD)			
IBR+BVD-MD+ (PI3)			
IBR+BVD-MD+PI3+BRSV			
+			
+			

8.

	()			
		(6)	(6-24)	(24)
		: (%) : (%)	: (%) : (%)	: (%) : (%)
		: (%) : (%)	: (%) : (%)	: (%) : (%)
		: (%) : (%)	: (%) : (%)	: (%) : (%)
		: (%) : (%)	: (%) : (%)	: (%) : (%)
		: (%) : (%)	: (%) : (%)	: (%) : (%)
		: (%) : (%)	: (%) : (%)	: (%) : (%)
		: (%) : (%)	: (%) : (%)	: (%) : (%)

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10. (, , ,)

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	1993	1994	1995	1996	1997

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- 1) 2) 3) 가 4)
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- 1) 2) 3) 4) :

11.

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- 가.
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- 가 ?
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4) 5) ()

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- 1) 2) 가 3)
4) 5) ()

(500Kg) 가 가?

- 1) 200 2) 220 3) 250
4) 270 5) 300 6) 350

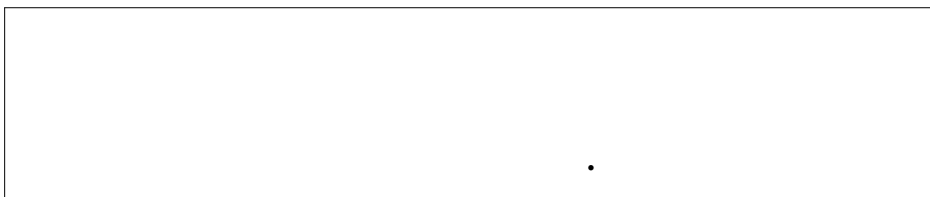
2001 (500Kg) 가

가 가?

- 1) 200 2) 220 3) 250
4) 270 5) 300 6) 350

16.

가 .



2.

가.

(polymelia) 6
 8 가 (heterotropic polymelia) 1
 2 가
 (notomelia), (cephalomelia), 가 (thoracomelia),
 (pygomelia) (Hattacharya, 1964; Hofmann, 1969; Petersen
 de Boom, 1970; Leipold Dennis, 1972; Leipold Huston, 1983; Hiraga ,
 1989).
 Abe et al.(1978)
 Leipold et
 al(1983) 6 가 4
 accessory hind leg .
 平賀 (1987) 5 가
 (1993)
 가 가 .
 1998 7 가 2 가
 . 480kg

38kg

가 1

1

(Akabane virus, Ibaraki virus)

가 2 가

(Fig 1,2).

72cm

48cm

(Fig 3-5)

가

가

/

BVD

가

2 가

612cm

1/2

(Fig 6).

8mm

가 가

(Fig

7,8).

平賀 (1987)

(1993)

가

Leipold et al.(1983)

가

5

가

1

38kg

가

2

가

72cm

48cm

가

가

BVD 가

2 가

6 × 12cm 1/2

(6).

8mm

가 가

Abe M, Hiraga T, Iwasa K, et al. A case of bovine thoracopagus. *J Coll Dairying* 1978; 7: 331- 336

Hattacharya MM. Six-legged monster in calf. *Ind Vet J* 1964;41:739.

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金鐘涉, 安東元, 鄭順熙. 五足 臀部 過剩後肢 過剩尾 가 二臀體.
大韓獸醫學會誌 1990;30(4):401- 406.

金鐘涉, 許贊權, 鄭憲植 等. 韓牛 胸部 前肢가 附着
多肢畸形. 大韓獸醫學會誌 1993;33(1):1- 5.

二頭二顔體

가 (Gilmore Fechheimer, 1969; Roberts 1986).

(Roberts, 1986). Arey (1960) 가

가 (diprosopus)

. , Witt (1963) Leipold

Dinnis(1972) 가

Denis(1972) Angus . Leipold
 體(Dicephalus dipus dibrachius) 二頭二足二腕
 Holstein Friesian
 二頭四足二腕體(Dicephalus tetrapus dibrachius)
 . Wacker Glaser (1988) 가 2 3 4
 가 二臀畸形 Arthur (1956) 가
 가
 (1990) 가
 . Leipold Dennis (1972)
 (derodidymus)

3 가

2

380kg

35kg

2

가 가 (Fig. 1,2)

(Fig. 3).

(Fig 4). 3 가 2 가 1 가

1 가 1 13

(Fig. 5). 2 2 가 1 가

(Fig. 6). 3 3 가 1 1 5cm 가

(Jenkins Hardy, 1968; Leipold , 1972; Deore, 1984),
(Thuringer, 1919; Partlow , 1981), (Dennis, 1975), (Bissonette
1933), (Misk Hifny, 1988)

McGirr (1985) 2 2
가 가
가

2
, (1987) 가

1/3

, McGirr (1985)

2 ,
가
2 , 1
1 가 1 가

(1990)

Partlow (1981) 2 2 4 , 3
가 가 , 가

8

1 2

Leipold (1972)

가

Leipold (1972)

Misk Hifny (1988)가 Buffalo

Leipold (1972)

, Ricardi Bergaman(1977)

2

가

가

2

1

1

1 가
1 가
1 가
2 , 1 가
가
가 1
5cm

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(Deore 1984; Leipold 1982; Roberts 1986).

, , , ,

(Fechheimer 1968, Roberts 1986). Arey (1960) Kurogi (1977)

(congenital dropsy)

(anasarca) 가 (Roberts, 1986; Young , 1986).

Akabane (Miura, 1974); Kurogi , 1975)

(Kahrs , 1970), (Kniazeff ,

1967), 3 (Kniazeff, 1967) Ibaraki (Roberts, 1986),

Bluetongue (Mckercher , 1970), Chuzan (Cottral, 1978;

, 1998) , Aino , Peaton

Tinaroo (Coverdal , 1978)

4

가 . 3

400 kg

가

가

2

3

, Akabane

, Ibaraki

, Chuzan

, Bluetongue

가

2/3

82cm,

25 kg

가

20 × 17cm

가

(Fig. 1)

가

(Fig. 2).

(Fig. 3).

가

1/2
가 32 × 49 mm (56
× 45 mm)
(Fig. 4, 6).

가
(Fig. 5).

15 × 21cm 13 × 18cm
13 × 9cm
가

가 Akabane
64 , Ibaraki Chuzan ,
, Bluetongue 2

가
(Arey , 1960; Kurogi ,
1977; Roberts 1986).
, Bulldog ,

1984; Roberts, 1986). (Donaldson Mason, Roberts (1986)가

Bulldog

가

(1970)

(1990)

(1990)

가 가

平賀 (1987)

가

가

가

가

Akabane

, Ibaraki

, Bluetongue

, Chuzan

(Russel ,

1985),

(Hall , 1989),

A (Markusfeld,

1989)

(Hartley Haughey, 1974)

가

Akabane

(Kurogi et al, 1975, 1976). Whitten (1957)

Akabane

, , -
. Miura (1974)

Akabane

가

Kurogi (1975)

-

가

가

Akabane

가 (1994)

206

가 2

가

가가

가

(, 1994).

Akabane

, ,
.

2/3

(82cm, 25kg)

.

,

가

가

가 32 × 49 mm (56 × 45 mm)

(15 × 21cm, 13

× 18cm) 1 (13 × 9cm)

가

64

가 Akabane 가

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麥角中毒

가 *Claviceps purpurea*

, bromes, bluegrasses, ryegrasses

1-4.

0.6%

ergotamine ergonovine

가

가

7

10

3 6

가

5.

2

7

1 가 1997 2

1996 11

3

1997 12

, 41

12

1997 1

가
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 20
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 3-4
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 가

(Fig 1).

“L”

(Fig 2).

가

1/3 ,

, 1/3 ,

가 .
가 .

dermatophyte (*Microsporum gysium*) .

tall fescue

, *Salmonella dublin* , .

fescue foot tall fescue

가

가 ,

€.

가

Microsporum

gysium

2

가 58.

. 가

17.

가

가

가

ergotamine ergotoxine

가

prolactin

가

가

134.

7

2 가 1997 2

X-ray

Microsporium gysium

1. Burfening PJ. Ergotism. JAVMA 1973; 163: 1288- 1290.
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•

5.15, 5.14 7, 2.12

,
14.11.16. 가 ,

9-11.

9

12 , 臀位
가 ,

4.8.11.

가 , ,

11,13,15,16 .

x-ray

4.11.

x-ray

140 kg 3 holstein 가

1 ,

,

1 ,

가

,

가

가 1 가

,

67 /min.

70 /min.

40.8

握雪音

(crackles) 喘鳴(wheezes)

가

가

(Spiramycin 400mg,

Streptomycin sulfate 800 mg, Procain HCl 16mg)

Pasteurella spp. 가 ,

(WBC 14.3×10^3 , neutrophils 71%, lymphocytes 13% ,

monocyte 16%) , gas

2

x - ray

(left and right lateral radiographs) (Fig 1 Fig 2). x - ray

bronchial cuffing

1 가

(radio- density)가 가

2 8

2 3

가 5 2

(thoracic inlet) 가

x - ray

(Fig 3).

(tracheal collapse) ,

19 41 (Fig 4), 1 4

가

*Setaria digitata*가

glycosaminoglycans

9-11.

6

4&11,13.

(Table 1).

13

10

1

2

가

가

3

10

Table 1. Summary of clinical data of the calves with tracheal collapse

Breed	Age (weeks)	Sex	Location of Collapse	Rib Trauma	Dystocia at birth	History of trauma	Reference
Holstein	12	female	C5- T2	Yes	No	No	
Holstein	4	female	Entire length of trachea	No	No	No	Ashworth <i>et al</i> 1
Charolais	2	male	Thoracic inlet	Yes	Yes (bp)	No	Jelinski <i>et al</i> 8
Guernsey	10	female	C5- T2	Yes	Yes (bp)	No	Fingland <i>et al</i> 4
Holstein	20	male	C6- T1	Yes	not witnessed	No	Fingland <i>et al</i> 4
Simmental	8	male	C5- T2	Yes	Yes (bp)	No	Fingland <i>et al</i> 4
Holstein	6	male	C6- T1	Yes	NA	No	Fingland <i>et al</i> 4
Holstein	4	female	C5- T1	Yes	No	No	Scarratt <i>et al</i> 13
Angus	3	male	C5- T2	Yes	NA	NA	Scarratt <i>et al</i> 13
Simmental	2	NA	C6- T1	Yes	NA	NA	Vestweber <i>et al</i> 15
Limousin	14	male	C6- T2	No	NA	No	Vestweber <i>et al</i> 15, Watt16
Shorthorn	3	male	C4- T5	Yes	Yes (bp)	No	Vestweber <i>et al</i> 15, Watt16
Holstein	20	female	C2- C4	NA	NA	NA	Hopney6

NA : Not available.

(bp) : breech presentation.

가 4&12.

가

4,11,15

13

가

4

4

3 가

가

(chondrodysplasia)

1,

(tracheal

stenosis)

가 3.

19.

Ashworth 1

(cartilage matrix)

가

가

. 가

“D”

가

4,8,13.

가 가

4,11,13.

11.

(tracheal prothesis)

가

4.11.

30%

11.

가

,

가

8,11,13.

,

.

3

holstein

가

1

.

握雪音(crackles) 喘鳴(wheezes)

.

가

.

Pasteurella spp. 가

,

.

x-ray

bronchial

cuffing

, .

1

.

가

,

(radio-density)가 가

, .

2

8

.

5

2

.

(tracheal collapse)

19 41

, . 1 4

가

,

,

.

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3. Legends for Figures

가. Polymelia in Korean- native calf

Fig 1. Polymelia appearing at dorsal pelvic region occurring in left hindlimb of 3-month old Korean-native calf.

Fig 2. Close-up of Fig 1.

Fig 3. Skeleton of pelvic hindlimb in Korean-native calf with pygomelia.

Fig 4. Dorsal view of pelvic girdle in Korean-native calf with pygomelia.

Noted incomplete formation of acetabulum(arrow) attached by extra hindleg.

Fig 5. Cranial view of extra hindlimb. Compared to two legs occurring pygomelia; left one(L) might be true extra hindleg.

Fig 6. Three kindeys taken from two left side occurring pygomelia and one right side as shown in normal size. Note multicystic Kidney(center) had multicystic form.

Fig 7. Urinary system taken from a Korean-native calf with pygomelia.

Noted markedly enlarged ureter(U) with cyst-like blind end(E) containing lots of creamy fluid toward urinary bladder. Uterus(T), Ovary(O)

Fig 8. Opening view of ureter and its cyst-like blind end of Fig 7.

. Diprososus in A Korean- native Calf

Fig 1. Dorsal view of a Korean-native calf with congenital derodidymus.

Noted marked distortion of both heads.

Fig 2. Ventral view of a Korean-native calf with congenital derodidymus.

Fig 3. Dorsal view of the left (L) and right (R) skulls. Note the right skull with congenital malformation of the right maxilla and right parietal bone.

Fig 4. Photograph of the two pairs of cervical and thoracic vertebrae. Note the fused and distorted vertebrae.

Fig 5. Normal thirteen pairs of ribs of a Korean-native calf with congenital derodidymus.

Fig 6. Skeletal bone of the three pairs of forelimbs. A pair of limbs was normal and the others were hypoplastic.

. Anasarca of a Korean-native calf

Fig 1. Dorsal view of a Korean-native calf with congenital dropsy and anasarca. Noted marked systemic hypoplasia of limbs and edema.

Fig 2. Ventral view of a Korean-native calf with congenital dropsy and anasarca. Noted marked change of cranial and cervical area due to congenital dropsy and anasarca. U = umbilical cord.

Fig 3. Cervical view of a deformed Korean-native calf with anasarca. Noted a large amount of serous fluid and marked edema in subcutaneous tissue. E = epiglottis.

Fig 4. Cut surface of cervical and thoracic area of deformed Korean-native calf. Noted marked hypoplastic lung but grossly normal trachea, esophagus and heart. H = heart; L = lungs; T = trachea.

Fig 5. Longitudinal section of the right head of a Korean-native calf with congenital dropsy and anasarca. Noted marked edema in subcutaneous tissue at the area of parietal bone and mandible.

Fig 6. Hypoplastic lungs (4x6 cm) of a Korean-native calf with anasarca.

. Ergot poisoning of Korean-native calves

Fig 1. Seven-month-old Korean native beef female calve appeared uncomfortable and showed typical gangreous changes of lower hindlimbs and tail.

Fig 2. Calf of Fig 1 was clearly indented lines formed between viable and necrotic tissue at just above the fetlock joint and below the hock joint. The skin on the lower metatarsal regions was cold, dry, insensitive on palpation as a bark of tree and sloughed.

. Tracheal collapse of a Holstein calf

Fig 1. Lateral inspiratory radiograph of cervical and thoracic area. The trachea is narrow at the thoracic inlet, multiple old rib fractures are apparent, and there is an exuberent non-bridging callus formation on the first rib.

Fig 2. Lateral thoracic radiograph of the left caudodorsal lung field in which there are very mild bronchial thickening.

Fig 3. Expiratory thoracic radiograph. Compare to inspiratory film, there is a minor change on the diameter of the collapsed trachea.

Fig 4. Exposed trachea from the ventral mid-line approach at necropsy. The collapsed segment is located at the thoracic inlet.

바. Hypoplasia of right forelimb in a Holstein calf

- Fig 1. Front view of forelimb with hypoplasia of right side.
- Fig 2. Lateral view of forelimb. Noted marked curvature of carpal joint due to hypoplastic right forelimb.
- Fig 3. Humerus and scapula of forelimb. Noted the marked hypoplasia of right humerus compared to the normal left one(upper).
- Fig 4. Close-up of right humerus with hypoplasia.
- Fig 5. Comparison of carpal joints. Noted marked hypoplasia of right side.
- Fig 6. Comparison of bones of forelimbs. Noted no formation of the bone under right humerus.

사. Others

- Fig 1. Scoliosis of vertebra of Korean-native calf with Akabane virus.
- Fig 2. Hydranencephaly of Korean-native calf with Chuzan virus.

가. Polymelia in Korean-native calf



Fig.1



Fig.2

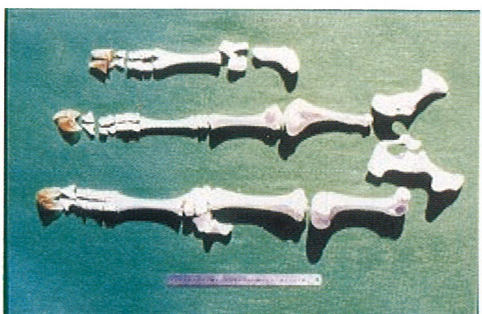


Fig.3

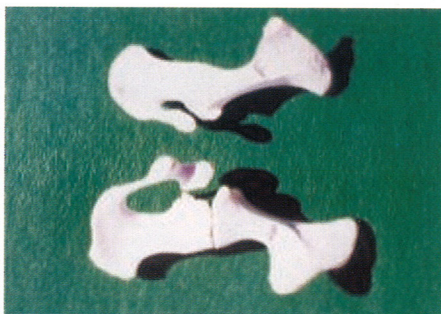


Fig.4



Fig.5

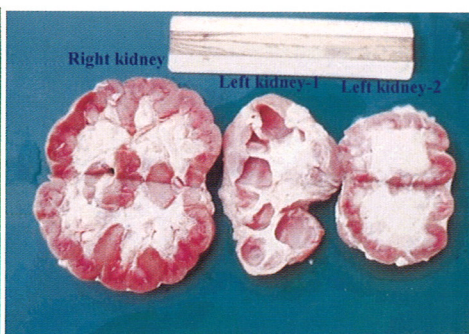


Fig.6

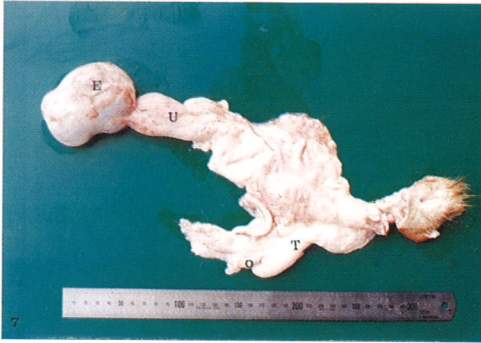


Fig.7

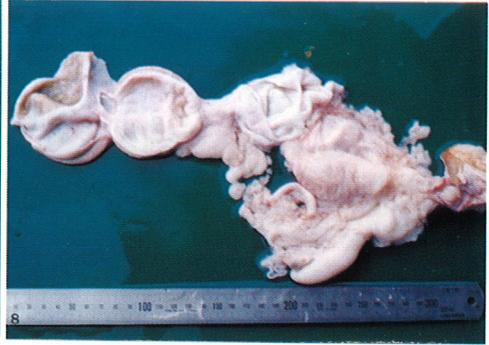


Fig.8

4. Diprososis in A Korean-native calf



Fig.1



Fig.2

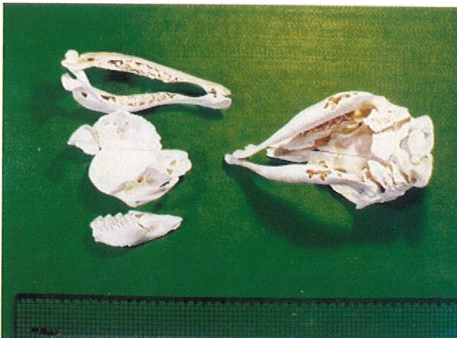


Fig.3

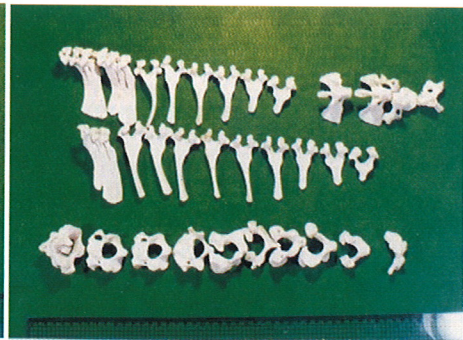


Fig.4

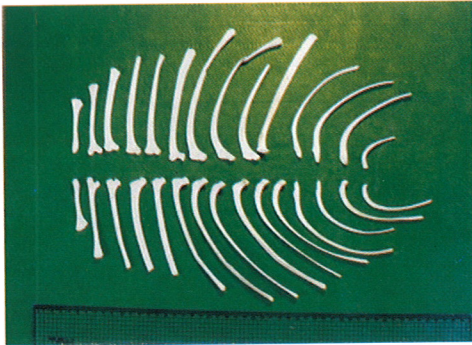


Fig.5

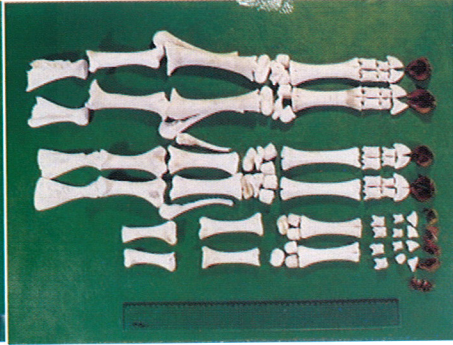


Fig.6

다. Anasarca of a Korean-native calf

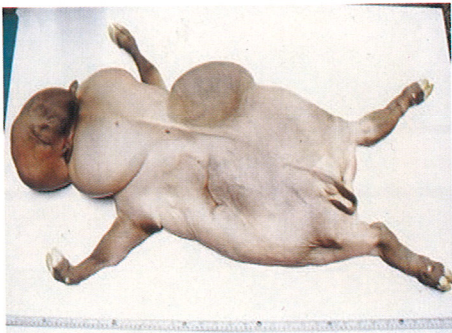


Fig.1

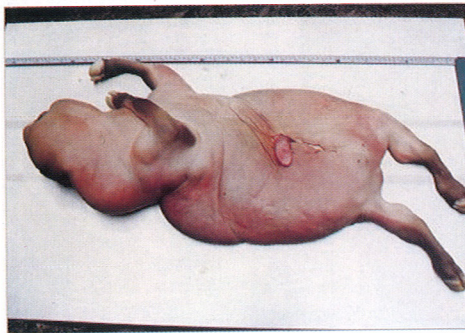


Fig.2

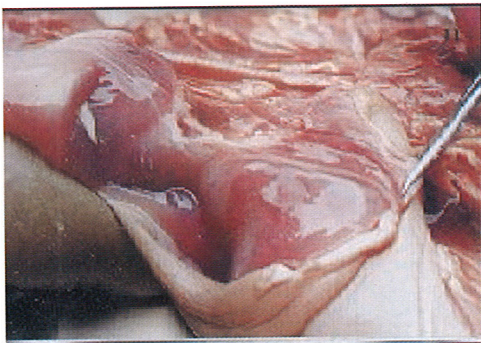


Fig.3

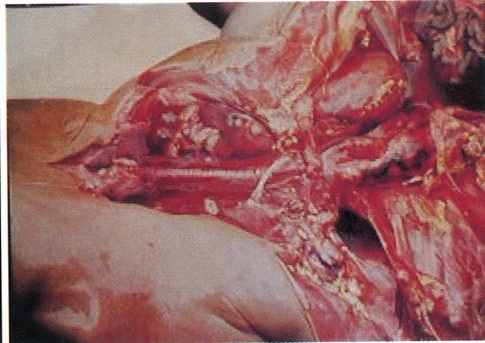


Fig.4



Fig.5

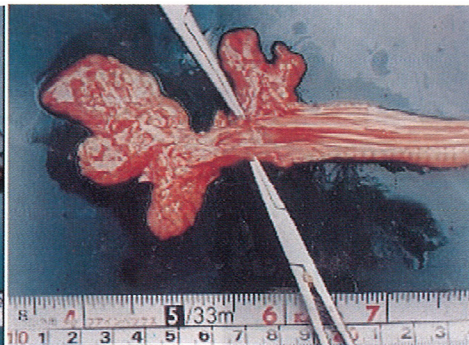


Fig.6

라. Ergot poisoning of Korean-native calves

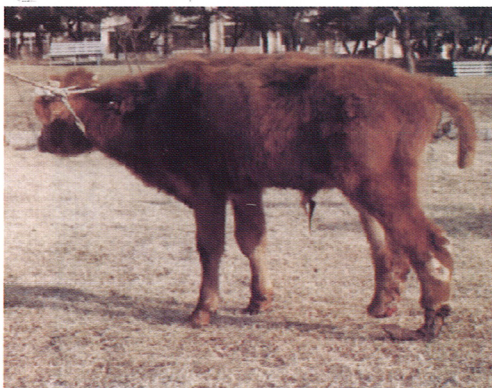


Fig.1



Fig.2

마. Tracheal Collapse



Fig.1



Fig.2

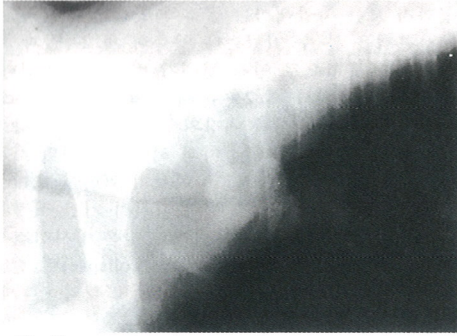


Fig.3

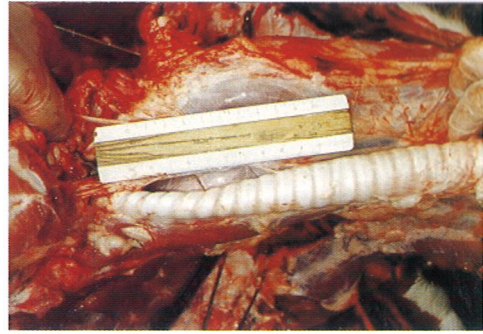


Fig.4

바. Hypoplasia of right forelimb in a Holstein calf



Fig.1

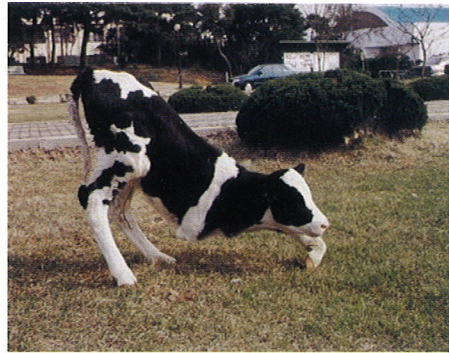


Fig.2



Fig.3

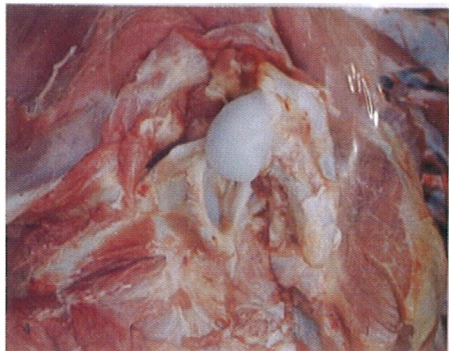


Fig.4

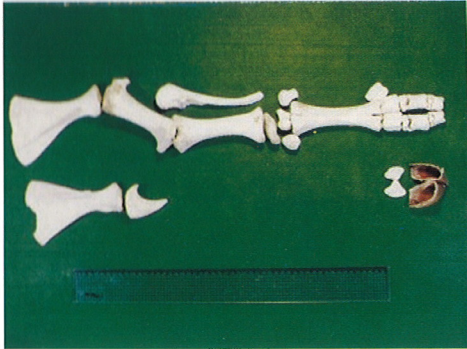


Fig.5

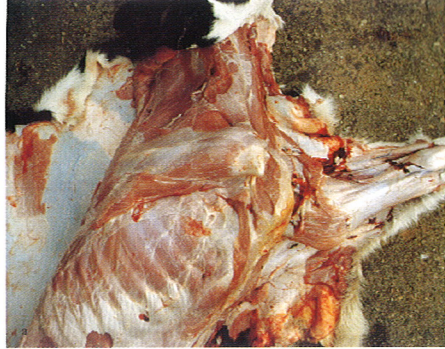


Fig.6

사. Others



Fig.1

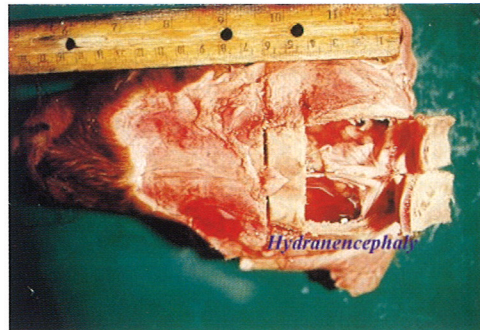


Fig.2