

GA0117-9904

**Research on Development of the New Liquor  
Product with Green Tea and Its  
Industrialization**

“ ”

1999. 10. 30.

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I.

II.

- 281 ('96 )  
가
- 가 가 가가
- , ,
- 가 가 가가 가
- 가 , 가 , , ,
- 가

III.

1.

2.

, ,

3.

가

4.

5.

6.

40

,

7.

,

가

8.

가

,

,

IV.

1.

tannin 16.65%, catechins 101.56  
mg/g, caffeine 32.47 mg/g, free sugar 10.49 mg/g, free amino acid  
58.86 mg/g tannin caffeine  
roasted 1-ethyl-2-  
formylpyrrol grassy cis-3-hexanol 36

2.

1 가  
170 7  
가 ,  
가 .

3.

95%  
가 .  
95%  
가 . 3

4. 가 0.5 g/100 mL  
 가 .  
 가 95%
5. 가 , 가  
 가 . , -  
 C, C  
 가 . stevioside, vitamin C,  
 disodium succinate, alanine, oligosaccharide .
6. 가 가  
 . 가  
 가 PVPP  
 가 ,  
 가 .  
 PVPP 가 0.05%, 10 가 .
7. 95% PVPP 0 120

20%

가 가

40

가

, ,

8.

,

가

가

가

가

9.

가

가

가

가

가

가

1 (360 mL)

( , ,

) 212

가

# SUMMARY

## . Title

Research on Development of the New Liquor Product with Green Tea and Its Industrialization

## . Purpose and Significance of the Study

*Posong-Koon* in *Chonnam* province is one of major regions producing green tea, where its production reaches 281 ton per year and tea cultivation is increasing continuously every year. This circumstance made tea farmers establish *Posong* green tea Farmer's Association to promote the consumption of their produces by selling them in the types of green tea leaves, instant tea (tea bag) and canned tea drink, etc.

Recently, *Posong* green tea Farmer's Association also decided to develop a new type of processed product with green tea to create a new demand and requested the development of a commercial green tea liquor product to Korea Food Research Institute (KFRI). One of the most important things to be considered in the development and commercialization of green tea liquor would be to develop the product maintaining the unique flavor and taste of green tea, and having the suitable shelf life for distribution in the market. Plant design was also required for the industrial production



## . Scope and Content of the Study

### 1. Final objective at the end of the study

Development of green tea liquor from green tea cultivated at *Posong-Koon* in *Chonnam* province

### 2. Scope and content

1) Analysis of functional components and flavor compounds of green tea

2) Determination of the pre-treatment condition to improve the flavor of green tea leaves, and the optimal extraction method, alcohol content and extraction time

3) Determination of the optimal amount of green tea leaves added for the liquor preparation and the alcohol content showing preference through a sensory evaluation

4) Determination of the contents of additives to improve the liquor quality

5) Methods to remove the precipitate formed possibly during the aging and storage period of green tea liquor

6) Storage test at 40 through the analysis of color, turbidity and

microorganism of products to estimate the quality changes during distribution

7) Diversification of products by adding apple and/or pear, and improvement of taste by adding high grade soju

8) Plant design for the industrial production of green tea liquor by estimating production cost, drawing flow sheet, layout and listing machinery and facilities

## **. Results and Recommendation**

### 1. Chemical analysis of green tea leaves

The composition of green tea leaves of *Posong-koon* was 16.65% tannin, 101.56mg/g catechins, 32.47mg/g caffeine, 10.49mg/g free sugar, and 58.86mg/g free amino acid, with the tendency of higher tannin and caffeine contents compared to tea leaves growing naturally. Thirty six flavor components including 1-ethyl-2-formylpyrrol of roasted fragrance and cis-3-hexanol of grassy fragrance were identified.

### 2. Pre-treatment condition of green tea leaves

To improve the flavor of green tea leaves, dry green tea leaves primarily process by the method of steamed tea were heat-treated differently. Sample heated at 170 for 7min showed the best

results in tastes, flavor and overall preference. However, it was a little worse in color of solution after extraction.

### 3. Optimization of extracting condition

The extraction of green tea leaves with 95% alcohol followed by diluting it with purified water was the most effective in maintaining the typical green color of leaves during extraction process. Whereas total polyphenol content was higher in a sample prepared by extracting leaves with solvent of lower alcohol content, sensory score was higher in sample extracted with 95% alcohol. The optimal extraction time was 3 hr.

### 4. Preparation of green tea liquor using extracts

For the production of green tea liquor, green tea leaves added at the level of 0.5mg/100mL showed the optimal sensory quality. Considering original cost including overall preference, storage and aging container, we have chosen the method of extraction with 95% alcohol, aging at 5 and then, dilution to the final alcohol concentration at the step of bottling.

### 5. Selection and combination of additives

For the stabilization of green tea color in product, acids such as citric acid, iso-vitamin C and vitamin C were revealed to be effective, while sugar had no effect regardless of its kind and concentration. Stevioside, vitamin C, disodium succinate, alanine, and

oligosaccharide were used for the formulation of the liquor.

#### 6. Clarification of green tea liquor

The formation of precipitate can be fatal in the quality of final product as it affect the quality index such as flavor during aging process and distribution in the market. To remove precipitates, filtration was selected when the product quality and the processing cost were considered. Among filtration methods, PVPP treatment was the most effective for clarification, maintenance of green color and sensory quality although the total polyphenol content was lowered. The treatment with 0.05% PVPP for 10 minutes gave the best results.

#### 7. Storage experiments of green tea liquor

Green tea leaves were extracted with 95% alcohol, treated with PVPP, stored at 0 °C for 120 days, diluted to alcohol content of 20%, seasoned with several food additives, and stored at 40 °C.

The results were that no large variation was found in turbidity and color, and it was microbiologically safe without contamination by bacteria, yeast and mold.

#### 8. Diversification of green tea extracts

The flesh and extract of fruits such as apple and pear added to diversify green tea extracts caused the rapid browning reaction in extract, resultin in that fruit juices were not desirable for the liquor

in the respect of maintaining the native color of green tea.

In order to improve the preference of green tea liquor, marketed distilled liquor and high grade diluted liquor were blended.

#### 9. Manufacturing process and unit cost of products

The unit cost of products was estimated. The price of green tea was based on that of *Posong's*, and other food additives were based the offer of providers. The cost(including material, production and packaging) for 1 bottle was calculated with 212 won. In addition, the name of machine required for the construction of processing factory was listed, and then flow sheet and layout was drawn for the factory design.

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2.	.....	39
3.	.....	40
1)	.....	40
2)	.....	40
(1)	.....	41
(2) 95%	.....	41
(3)	.....	44
(4)	.....	45
(5)	.....	50
4.	.....	50
1)	가 .....	50
2)	.....	54
3)	.....	55
5.	.....	56
1)	가 .....	56
2)	가 .....	56
3)	.....	57
6.	.....	61

1) 가	.....	61
2) PVPP 가	.....	72
(1) PVPP 가	.....	72
(2) PVPP	.....	74
(3) PVPP	.....	74
3)	.....	81
7.	.....	83
8.	.....	84
1) 가	.....	84
2)	.....	84
9.	.....	85
1) 가	.....	86
2)	.....	87
3)	.....	88
4	.....	90





## 2

1.

1)

4 5 8 1, 2 가

2)

(-)-epicatechin(EC), (-)-epigallocatechin (EGC), (-)-epigallocatechingallate(EGCg), (-)-epicatechingallate(ECg) caffeine, glutamic acid, fructose, glucose, sucrose, maltose Sigma , acetonitrile(CH<sub>3</sub>CN), ethyl acetate(EtOAc) HPLC , chloroform(CHCl<sub>3</sub>) 가 ( )

2.

1)

Ethanol 100 mL 70 mL가 가 100 mL가

15

2) Tannin

0.1 g 80 70 mL messflask  
 80 30 가 100 mL  
 20 mL 20 mL  
 5 mL 5 mL 25 mL  
 volumetric flask pH 7.5 Sorensen's phosphate  
 buffer solution spectrophotometer  
 (Backman, DU-7) 540 nm ethyl-gallate  
 (Wako chemical Industries, Japan)  
 tannin

$$\text{Tannin(\%)} = G \times 1.5 \times 100/W$$

G = ethyl gallate

W = 100 mL (mg)

3) Catechins

0.1 g 80 70 mL messflask  
 80 30 가 100 mL  
 1 Matsuzaki &  
 Hara counter current chromatography crude  
 catechin HPLC , 10 mL  
 chloroform 10mL 3

ethyl acetate 10 mL 3  
 (Büchi rotavapa R-124,  
 Switzerland) HPLC  
 {CH<sub>3</sub>CN-EtOAc-0.05% H<sub>3</sub>PO<sub>4</sub> (12:2:86, v/v/v)} 0.45 μm  
 membrane filter HPLC  
 1 .

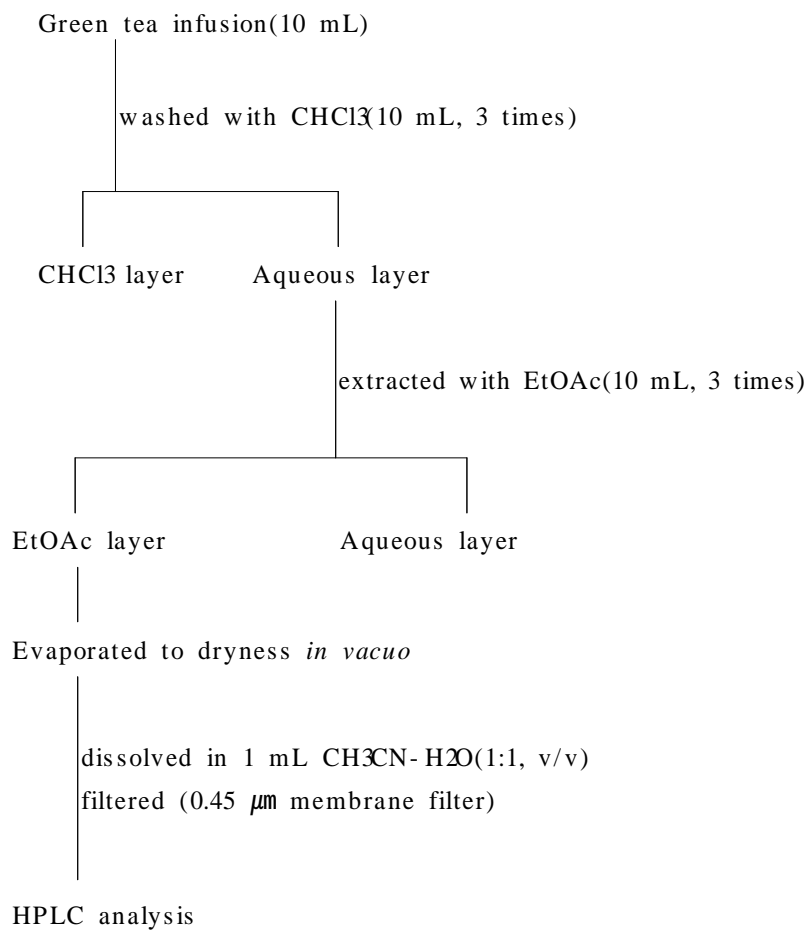
1. HPLC

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Instrument	Jasco LC-960 (Japan)
Column	CentriTMμ Bondapak C18 guard column (125 , 3.5 × 20mm, Waters) μ-Bondapak C18 (3.9 mm I.D. × 300 mm)
Mobile phase	CH <sub>3</sub> CN-EtOAc-0.05% H <sub>3</sub> PO <sub>4</sub> (12:2:86, v/v/v)
Detection wavelength	280 nm
Flow rate	1.0 mL/min
Column Temp.	35

---





## 1. HPLC

4) Caffeine

0.1 g 80 70 mL messflask  
80 30 가 100 mL  
polyvinylpolypyrrolidone(PVPP) 200 mg 가  
30 PVPP  
0.45  $\mu$ m membrane filter HPLC  
HPLC catechin

5) Free amino acids

PVPP 가 , 30  
1 mL vial  
phenylisothiocyanate A  
200  $\mu$ L 0.45  $\mu$ m membrane filter 2  
HPLC chromatogram  
peak area peak area

6) Free sugars

2  
JASCO HPLC system Waters  
Carbohydrate analysis column(3.9  $\times$  300 mm, USA) CH<sub>3</sub>CN- H<sub>2</sub>O  
(75:25, v/v) 30 1.4 mL/min RI

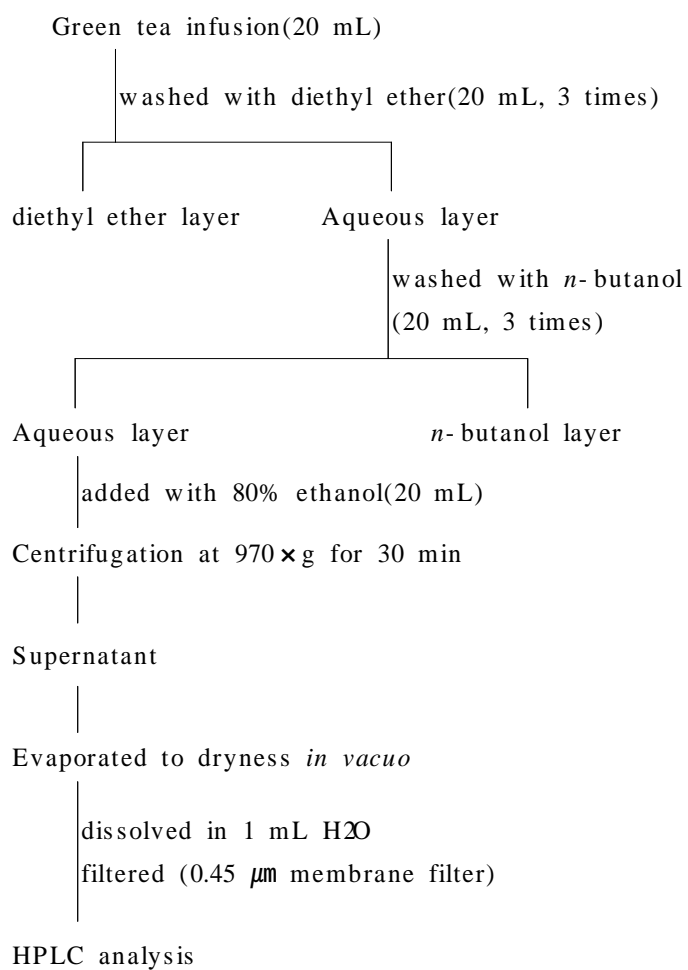
2.

HPLC

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Instrument	Waters PicoTag system
Column	Picotag 8.5 × 300 mm
Pump	Waters 510
Injector	Waters 712 WISP
Detector	Photodiode array detector Waters 990 at 254 nm
Solvent	A) 1.4 mM NaHAc, 0.1% TEA, pH 6.3 : CH <sub>3</sub> CN (94 : 6, v/v) B) 60% CH <sub>3</sub> CN
Elution	Linear gradient of solvent B (0 - 100%)
Flow rate	1.0 mL/min

---



## 2. HPLC

7)

Plate count agar(PCA) Potato dextrose agar(PDA)

8)

(ColorQuest , Hunter Lab, USA) L(lightness),  
a(redness/ greenness), b(yellowness/blueness)

Hunter L, a, b (1) chroma  
(2) total color difference(TCD) pH가

$$\text{Chroma value} = \sqrt{a^2 + b^2} \quad (1)$$

$$\text{TCD value} = \sqrt{(L_1 - L_0)^2 + (a_1 - a_0)^2 + (b_1 - b_0)^2} \quad (2)$$

9) UV- Visible Spectrometry

Spectrophotometer(Backman, DU- 650, USA) scanning

10)

Folin- Denis  
5 mL Folin 5 mL 가 3

10% Na<sub>2</sub>CO<sub>3</sub> 5 mL 가 1 760 nm  
tannic acid .

11)

dynamic  
headspace . Dynamic headspace  
purge-trap system , Tekmar (Cincinnati,  
U.S.A.) purge-trap LSC 3000 .  
10 mL (55 mm × 120 mm) 50  
(50 ~ 60 mL/ , 20 ) purging 3 dry  
purging . Tenax-GC (polymer based on the  
2,6-diphenyl-p-phenylene oxide, 60/80 mesh, Alltech, U.S.A.)가  
(1/8" x 12" stainless steel) .

Purge-trap system desorb preheat 180 ,  
valve, mount line 120 , desorb 220 (3 ) , bake 250 (20 )

(Innowax 30m  
× 0.32mm, 0.25 μm in film thickness, Hewlett Packard., U.S.A.)  
, 35 (2 ) 160 2.5  
180 , 280  
, 1.2 mL/  
split ratio 1 : 1 .

gas chromatograph-mass  
spectrometric detector(MSD5972, Hewlett Packard, USA)

Wiley NBS 138

library spectrum .

12) pH

pH meter(HANNA HI9321, Singapore)

13)

Spectrophotometer 665 nm

14)

5 가 , , ,  
가 .

### 3

1.

3 .

3.

---

Tannin(%)	16.65
Catechins(mg/g)	101.56
Caffeine(mg/g)	32.47
Free sugar(mg/g)	10.49
Amino acid(mg/g)	58.86

---

1) Tannin

16.95%  
14.42%, 14.25%, 15.46%

가

2) Catechins

HPLC 10.16%  
10.04 10.85%



10.35 14.58%      煎茶 13.46 14.14%

가

standard

chromatogram      3      epigallocatechin(EGC),  
epicatechin(EC), epigallocatechingallate (EGCg), epicatechingallate  
(ECg)      EGC 1.17%, EC 0.84%,  
EGCg 6.28%      ECg 1.87%

가

가      가      15%

가

EC, EGC

ECg, EGCg      가

3) Caffeine

32.47 mg/g      1 g

22.1 31.1 mg

100%

가

4) Free amino acids

58.86 mg/g

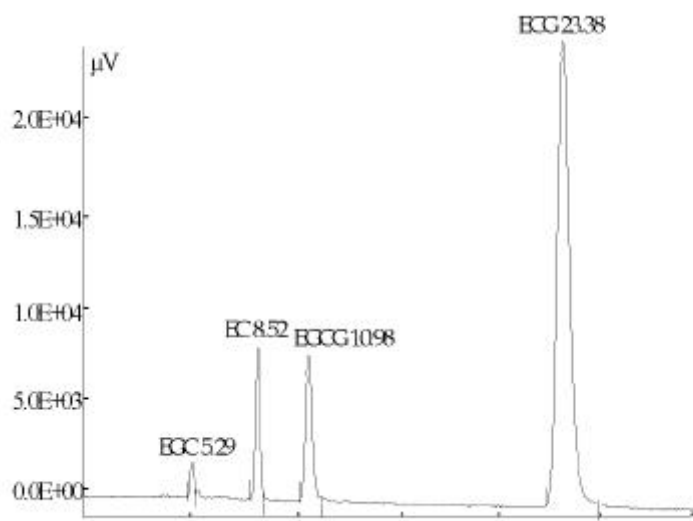
40.06 mg/g

chromatogram 4

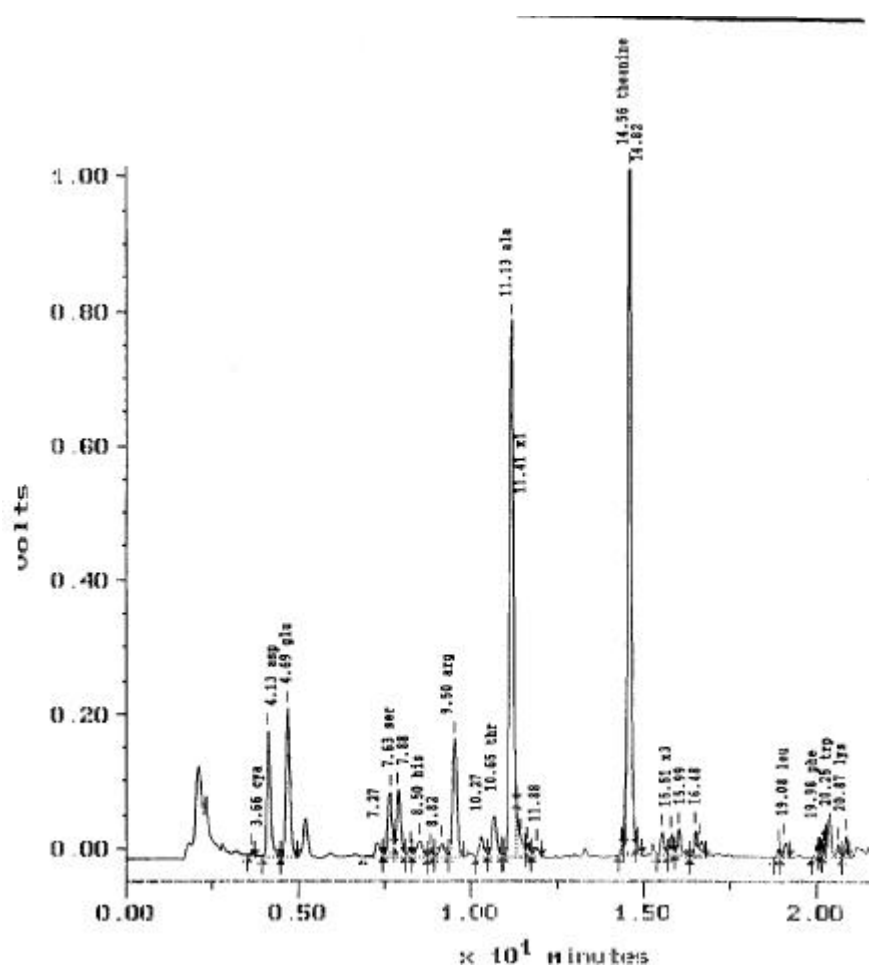
5) Free Sugars

10.49 mg/g

가



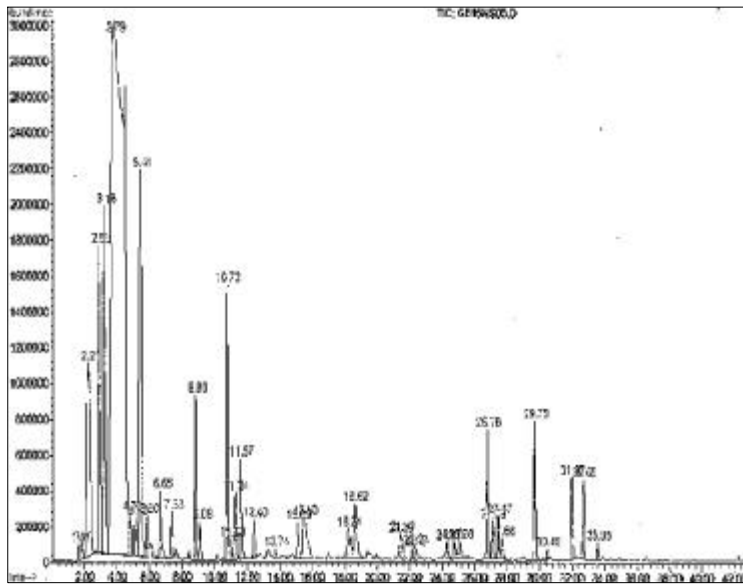
3. HPLC chromatogram



4.

6)

5 chromatogram  
, GC/MS library  
4 . Library peak  
가 quality  
rose, sweet, roasted,  
grassy, ligneous, bitter  
roasted alkyl prazine 1-ethyl-2-formyl pyrrol  
grassy cis-3-hexenol  
가 rose  
geraniol .



5. chromatogram

4.

Peak No.	Retention Time (min)	Compounds
1	1.62	propane, 2- isocyanato-
2	1.81	methane, thiobis
3	2.21	buthane
4	2.85	ethyl acetate
5	3.19	butanol, 2- methyl
6	3.79	ethanol
7	4.75	2- heptanol
8	5.13	2- pentanone, 4- methyl
9	6.65	2,3- penanedione
10	9.08	3- penten- 2- one
11	10.73	1H- pyrrol, 1- ethyl
12	11.00	2- heptanone
13	11.24	1- penten- 3- one
14	11.57	2- pentano, 4- methyl-
15	12.40	oxazole, trimethyl-
16	13.74	fran,2- (methoymethyl)-
17	15.09	1- pentanol
18	15.51	pyrazine, methyl-
19	18.21	pyrazine, 2,5- dimethyl-
20	18.61	pyrazine, ethyl-
21	21.39	cis- 3- hexenol
22	21.49	pyrazine, 2- ethyl- 5- methyl
23	22.16	pyrazine, 2- ethyl- 3- methyl
24	22.34	pyrazine, trimethyl-
25	24.31	pyrazine, 3- ethyl- 2,5- dimethyl
26	24.72	acetic acid
27	25.08	fufural
28	26.78	1- hexanol- 2- ethyl
29	27.08	ethanone, 1- (2- furanyl)
30	27.46	1H- pyrrole
31	27.68	benzaldehyde
32	29.70	delta- 3- carene
33	30.48	2- furancarboxaldehyde
34	31.97	1- ethyl- 2- formyl pyrrol
35	32.69	hotrienol
36	33.58	benzeneamine,2- methoxy- 5- methylhotrienol

2.

1 (4 ), 2 (6 ), 3 (8  
 ), 4 (10 ) 가  
 C  
 가 .  
 가 ,  
 1  
 5 .  
 6  
 . 가 ,  
 a b  
 ,  
 170 7 , 5 가  
 가 .

5.

No.			
1			
2	120	30	(1203)
3	140	8	(14008)
4	140	20	(14020)
5	170	7	(17007)



6. 가

No.	Hunter value			a)
	L	a	b	
1	60.86	- 18.86	38.17	3.0
2	58.98	- 16.84	38.09	2.8
3	59.68	- 16.57	38.49	2.5
4	65.92	- 15.83	42.30	2.2
5	72.39	- 12.57	46.20	1.5

a) values are mean : 1;very good, 2;good, 3;fair, 4;poor, 5;very poor

3.

1)

45, 60, 75, 95%

750 ppm      가      4      72

750 ppm      가

2)

,  
 ,  
 ,  
 . , 20%  
 1 mL 0.005 g  
 0.2  $\mu\text{m}$  filter pad  
 50 mL cap vial head space  
 30 incubator ,

(1)

20% 40%  
 7 . 3  
 a 가 .  
 , .  
 가 가

(2) 95%

95% 20% 40%  
 8 . a  
 . 가

7.

			Color	
			20%	40%
0	Hunter	L	73.47	75.93
		a	- 5.39	- 6.55
		b	36.33	36.13
3	Hunter	L	69.29	73.11
		a	- 0.04	- 2.90
		b	36.45	35.34
6	Hunter	L	66.23	67.25
		a	4.69	3.07
		b	37.89	36.62
12	Hunter	L	59.25	65.47
		a	12.57	9.04
		b	37.63	39.42
15	Hunter	L	62.95	61.80
		a	12.65	11.33
		b	39.74	38.38

8. 95%

			Color	
			20%	40%
0	Hunter	L	89.43	83.74
		a	- 5.36	- 6.84
		b	18.45	29.49
7	Hunter	L	87.57	80.90
		a	- 3.66	- 4.91
		b	19.70	27.57
14	Hunter	L	85.98	79.75
		a	- 3.16	- 4.00
		b	20.62	27.80
21	Hunter	L	86.03	78.11
		a	- 3.06	- 3.50
		b	20.40	27.89
28	Hunter	L	85.44	79.96
		a	- 2.71	- 3.53
		b	20.74	27.03
35	Hunter	L	85.04	77.45
		a	- 2.48	- 2.61
		b	21.68	28.74

(3)

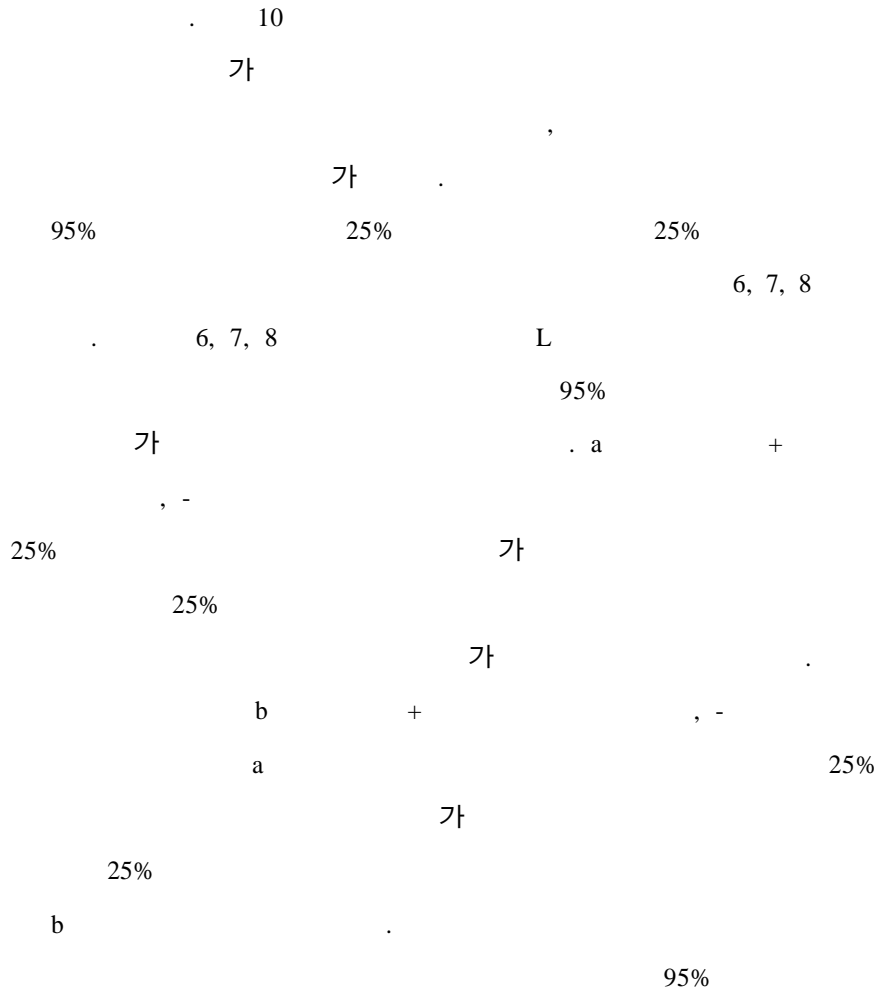
20% 40%  
9 . 9  
a 20%, 40%  
a 가  
.

9.

Color			20%	40%
0	Hunter	L	83.07	73.33
		a	- 3.23	- 5.22
		b	21.90	29.73
7	Hunter	L	78.14	73.73
		a	- 1.05	- 2.29
		b	23.26	29.62
14	Hunter	L	76.36	72.57
		a	1.12	- 0.62
		b	27.49	33.23
21	Hunter	L	74.64	68.19
		a	3.67	0.81
		b	30.57	34.98
28	Hunter	L	71.49	70.52
		a	4.13	2.51
		b	31.86	36.35
35	Hunter	L	71.02	68.40
		a	5.28	4.13
		b	33.80	36.52

(4)

10



10.

---

a)

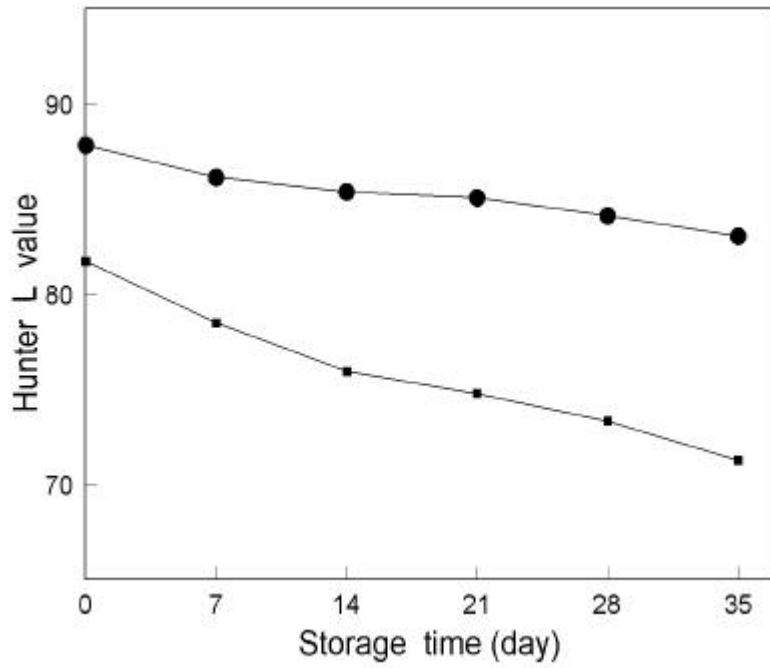
(mg/100mL)

---

	25%	153.16	4.0
25%		150.88	3.0
95%	25%	142.72	2.5

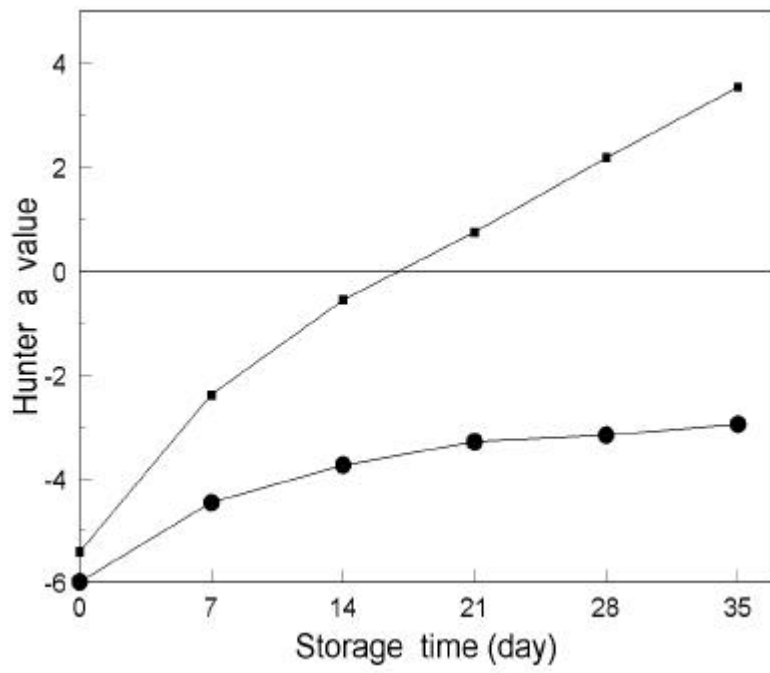
---

a) values are mean : 1;very good, 2;good, 3;fair, 4;poor, 5;very poor

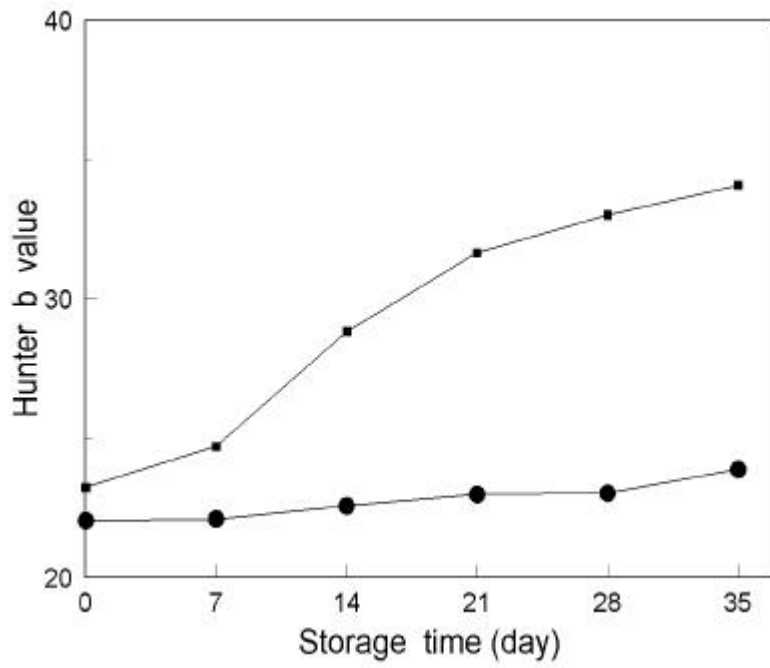


6. Hunter L  
●: 25%  
■: 25%





7. Hunter a  
●: 25%  
■: 25%



8. Hunter b  
●: 25%  
■: 25%

(5)

95%

9, 10, 11

9 11

3

665nm

10

color intensity 가

3

4.

1) 가

가

95%

20%

100 mL

0.1, 0.5, 1, 1.5, 2 g

11 . 0.5 g/100 mL 가 가 , ,

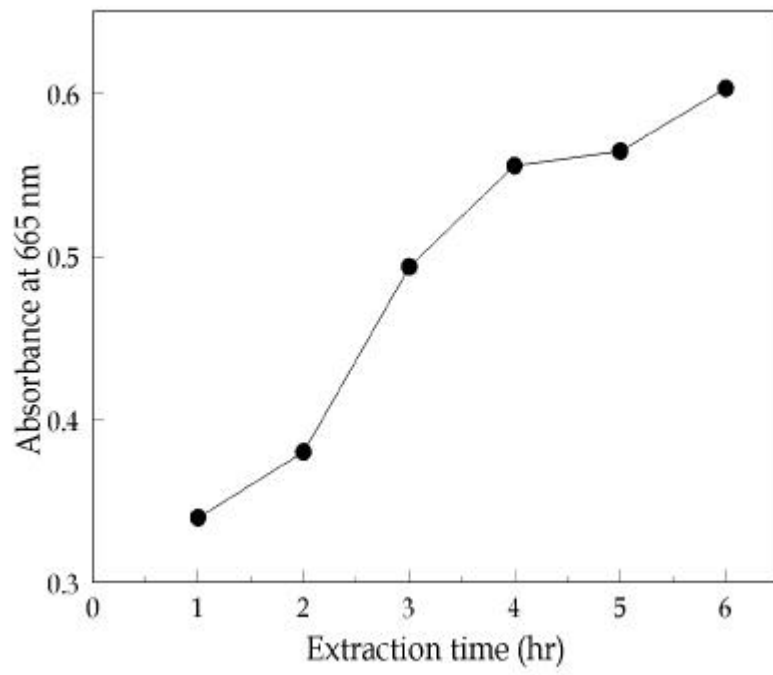
가

가

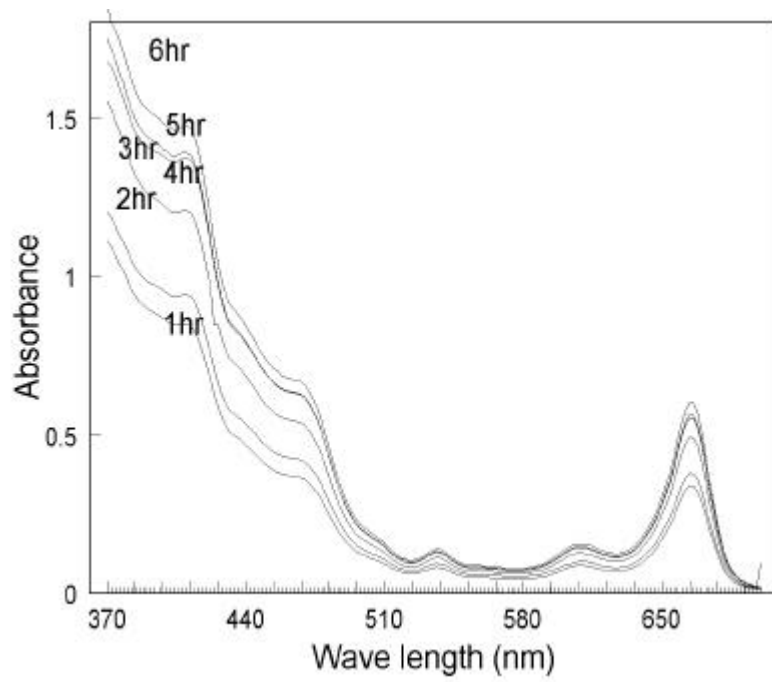
가

가

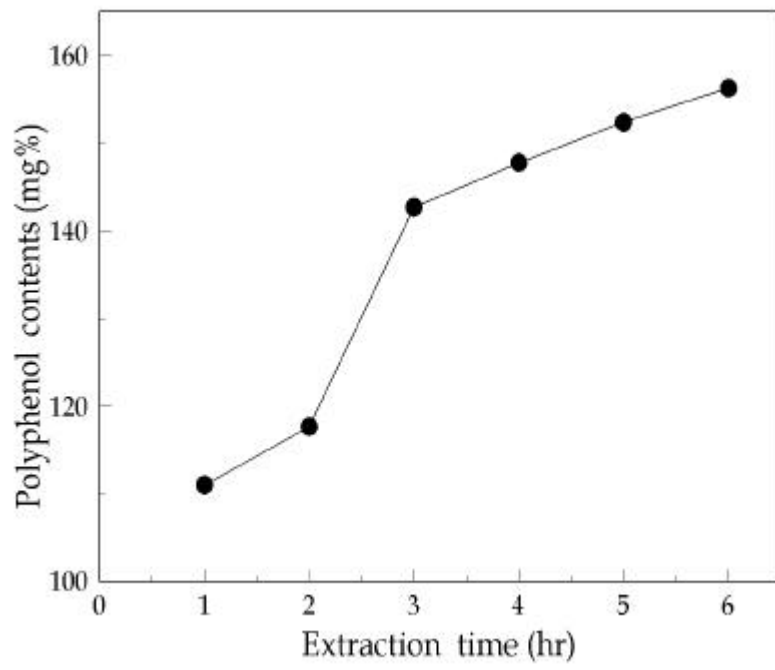
가 0.5 g/100 mL



9.



10.



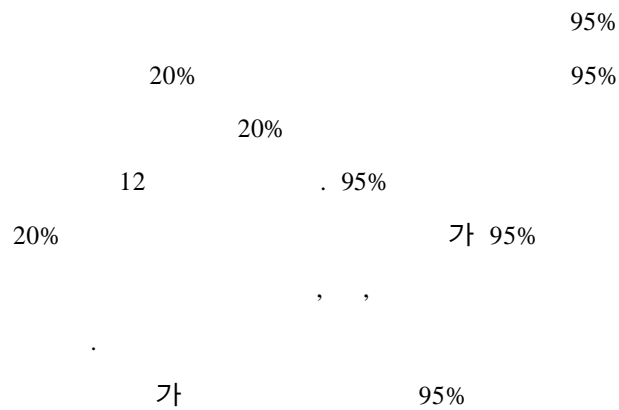
11.

11. 가

가				
0.1 g/100mL	2.0	1.2	1.5	1.8
0.5 g/100mL	1.4	1.2	1.4	1.4
1.0 g/100mL	2.8	1.2	3.0	3.0
1.5 g/100mL	3.0	1.4	3.6	3.2
2.0 g/100mL	3.8	1.6	3.2	3.6

a) values are mean : 1;very good, 2;good, 3;fair, 4;poor, 5;very poor

2)



12.

	3.7	1.8
	3.2	2.0
	3.5	2.2
	3.6	2.0

a) values are mean : 1;very good, 2;good, 3;fair, 4;poor, 5;very poor

3)

95%

13

15% v/v, 20% v/v 가

20%

13.

	15%	20%	25%	30%
	2.1	1.8	3.1	3.7
	2.2	2.0	2.3	2.9
	2.4	1.9	3.7	4.0
	2.4	2.0	3.5	3.9

a) values are mean : 1;very good, 2;good, 3;fair, 4;poor, 5;very poor



5.

1) 가  
가 가 가가  
가  
30 14 가  
가  
L, a, b 가  
가 가 가  
가 , 가  
polyphenol

2) 가  
가 가 가가  
가 30  
15 가  
가가 가  
L a  
가가 가  
가 0.01% 가 0.005% 가  
가 , - C,  
C

가 .

- C C .

3)

가

. , 16 5

가

, , disodium succinate, , ,

C, , ,

300

, dipeptide

150

가 . Disodium

succinate monosodium glutamate (MSG)

A가 가 C

가 MSG 가 E

14. 가

	3 day			135 day		
	Hunter value			Hunter value		
	L	a	b	L	a	b
가	87.19	- 6.41	23.67	84.39	- 3.73	23.52
0.5%	86.11	- 6.34	24.47	82.13	- 3.07	23.12
1.0%	86.60	- 6.51	24.02	82.04	- 3.49	24.88
1.5%	86.27	- .627	23.83	83.40	- 3.27	22.99
0.5%	86.17	- 6.25	24.38	78.72	- 2.16	27.54
1.0%	86.63	- 6.42	23.98	81.15	- 2.90	24.00
1.5%	86.50	- 6.34	23.94	82.95	- 3.44	22.21
0.5%	86.22	- 6.44	24.50	81.96	- 3.26	23.83
1.0%	86.94	- 6.31	23.47	82.95	- 3.52	22.84
1.5%	87.17	- 6.34	23.30	83.60	- 3.73	22.55
0.5%	86.63	- 6.18	24.57	81.81	- 3.06	24.59
1.0%	86.05	- 6.12	24.66	81.31	- 2.97	24.67
1.5%	86.36	- 6.08	24.15	82.74	- 3.41	23.14
0.5%	86.61	- 6.33	24.45	81.14	- 2.97	24.96
1.0%	86.06	- 6.22	24.36	81.97	- 3.22	24.03
1.5%	86.84	- 6.41	24.20	83.86	- 3.66	22.61
0.001%	86.53	- 6.44	24.35	81.72	- 3.32	24.69
0.005%	86.80	- 6.34	23.88	83.24	- 3.61	23.43
0.01%	86.80	- 6.54	24.44	84.49	- 3.91	21.96

15. 가

	3 day			140 day		
	Hunter value			Hunter value		
	L	a	b	L	a	b
가	86.55	- 6.21	24.42	84.57	- 3.77	23.96
0.005%	87.46	- 5.59	20.89	82.03	- 5.06	20.53
0.01%	86.99	- 5.51	21.02	84.31	- 5.26	18.31
0.005%	86.45	- 5.58	22.48	83.39	- 4.31	23.01
0.01%	85.95	- 5.45	22.16	82.41	- 4.08	23.65
0.005%	86.86	- 5.47	20.52	83.46	- 4.18	23.20
0.01%	87.24	- 5.49	20.27	83.09	- 4.16	24.90
0.005%	87.48	- 5.60	20.42	84.21	- 4.41	21.67
0.01%	87.32	- 5.55	20.48	85.78	- 4.85	22.22
- C 0.005%	87.34	- 5.60	21.01	85.23	- 6.23	20.26
0.01%	87.47	- 5.64	21.03	84.98	- 6.40	21.64
C 0.005%	87.25	- 5.64	21.30	85.89	- 5.69	19.06
0.01%	87.40	- 5.66	21.38	86.79	- 5.54	19.20

16.

---

A	stevioside	%
	vitamin C	%
	disodium succinate	%
	alanine	%
	oligosaccharide	%
		%
B	stevioside	%
	vitamin C	%
	disodium succinate	%
	alanine	%
	oligosaccharide	%
		%
C	aspartam	%
	vitamin C	%
	disodium succinate	%
	alanine	%
	oligosaccharide	%
		%
D	stevioside	%
	iso- vitamin C	%
	disodium succinate	%
	alanine	%
	oligosaccharide	%
	solbitol	%
		%
E	stevioside	%
	vitamin C	%
	MSG	%
	alanine	%
	oligosaccharide	%
		%

---

6.

가

가

tannase

, 가

가

tannase

가

1) 가

가

0.5%

가 10

0.45  $\mu$ m membrane filter

20%

,

17

100 400 mesh

. 100 400 mesh

12

가 PVPP

PVPP 가 가

HPLC 13 18 ,  
EGC, EC, EGCg, ECg 18  
ECg 가

, PVPP EGC, EC,  
EGCg, ECg 가 EC  
5.2 mg%, 5.5 mg%, 4.8  
mg% 가 12.0 mg%

PVPP  
EGCg 가 PVPP  
PVPP

PVPP 가 PVPP

17.

		A value ( 665 nm )	Hunter		
			L	a	b
Control		0.42	73.65	- 14.01	45.47
PVPP		0.40	73.86	- 14.11	45.26
Bentonite		0.40	72.17	- 13.38	44.44
Diatomaceous earth		0.41	72.73	- 13.65	44.83
Silica gel		0.41	72.65	- 13.56	44.73
Charcoal	4 8mesh	0.39	73.84	- 13.91	45.09
	100 400mesh	0.01	92.29	- 5.66	20.07

18.

Control		4.6	1.4	4.2	4.2
PVPP		2.4	1.2	2.4	2.4
Silica gel		2.8	1.2	3.0	3.0
Bentonite		3.0	1.4	3.6	3.2
Diatomaceous earth		3.8	1.6	3.2	3.6
Charcoal	4 8mesh	4.4	1.6	3.6	4.0
	100 400mesh	4.2	4.8	3.2	4.6

a) values are mean : 1;very good, 2;good, 3;fair, 4;poor, 5;very poor



19.

a)

		Contents of catechins (mg%)			
		EGCb)	ECc)	EGCgd)	ECge)
Control		18.3	12.0	43.1	3.4
PVPP		5.9	2.0	5.5	0.7
Silica gel		15.8	5.2	37.1	3.2
Bentonite		16.3	5.5	34.7	3.1
Diatomaceous earth		18.1	4.8	40.6	3.2
Charcoal	4 8mesh	17.4	11.9	41.8	3.2
	100 400mesh	18.0	12.1	39.0	3.4

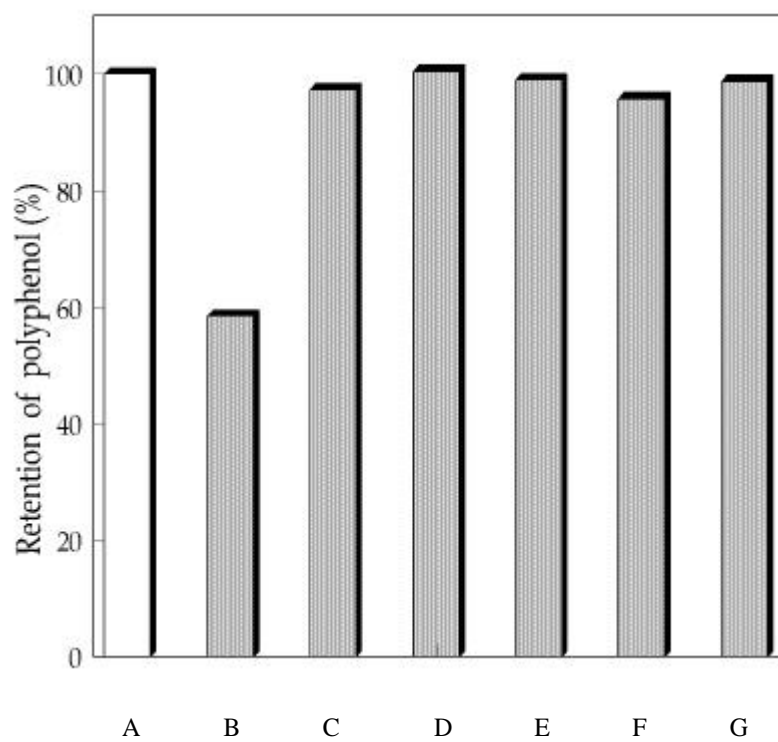
a) treated with 0.2 % filter aids in green tea liquor

b) EGC : epigallocatechin

c) EC : epicatechin

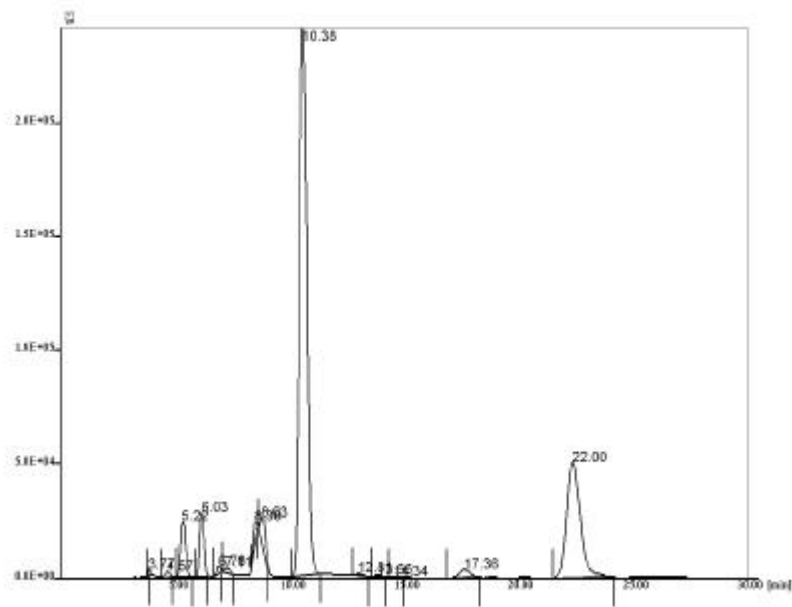
d) EGCg : epigallocatechin gallate

e) ECg : epicatechin gallate



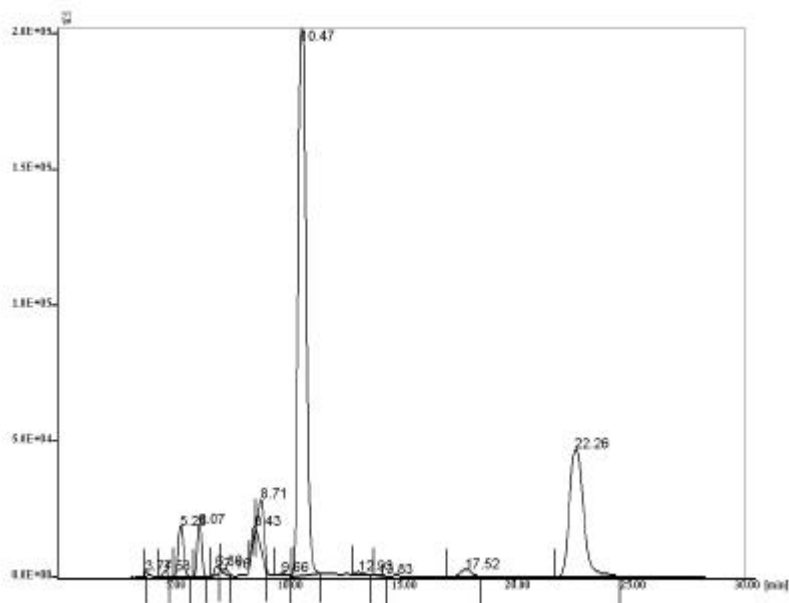
12.

A;Control, B;PVPP C;Diatomaceous earth D;Silica gel  
E;Bentonite, F;Charcoal(4-8mesh), G;Charcoal(100-400mesh)



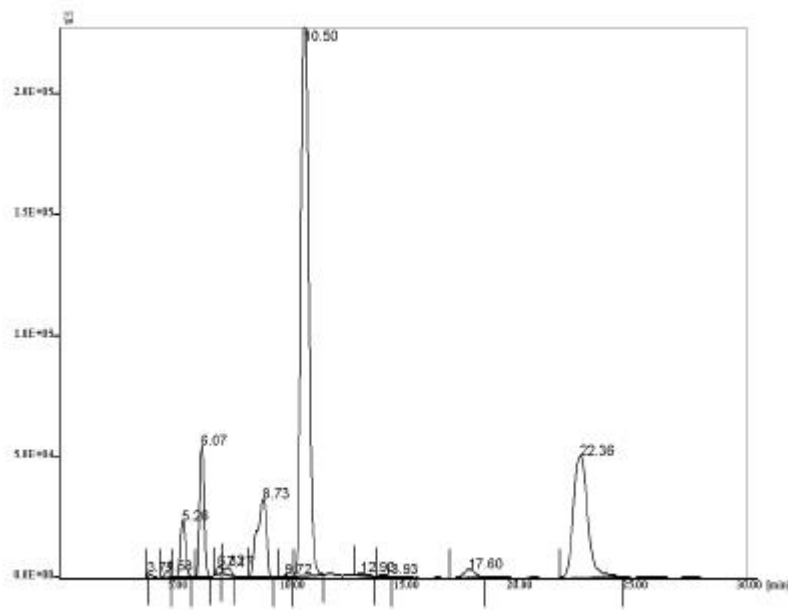
13.

HPLC chromatogram

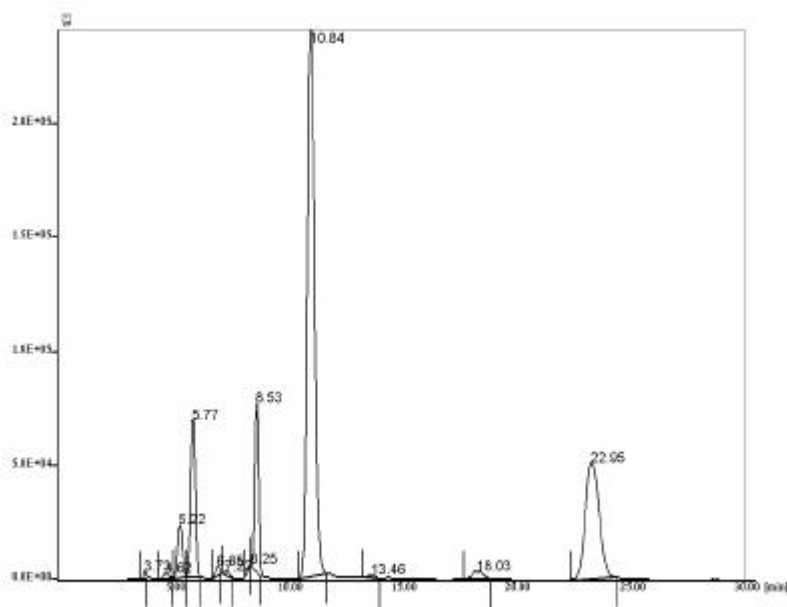


14.

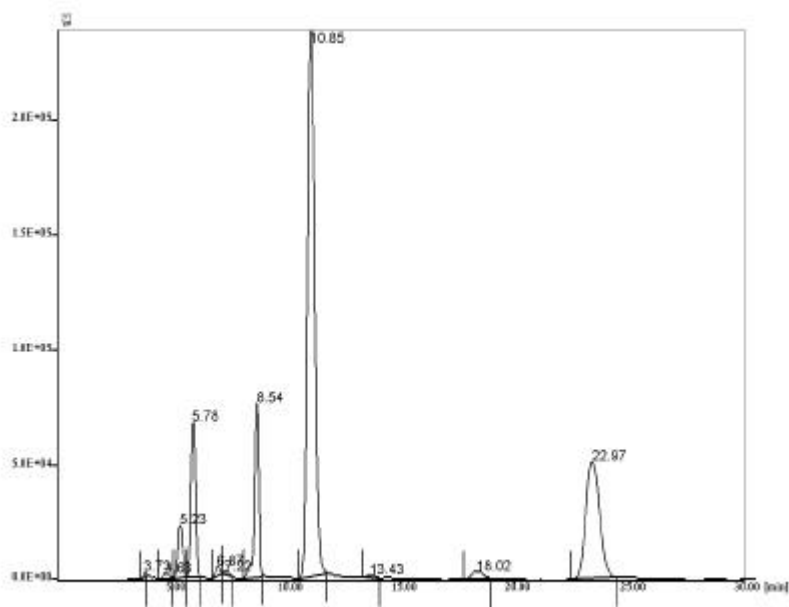
HPLC chromatogram



15. (100 400 mesh)  
HPLC chromatogram

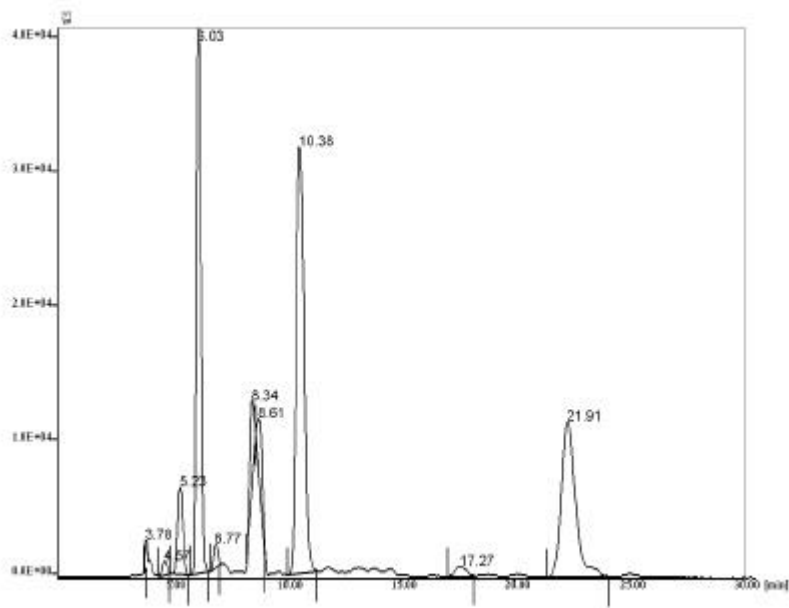


16. (4 8 mesh)  
HPLC chromatogram



17.

HPLC chromatogram



18. PVPP

HPLC chromatogram



2) PVPP 가

(1) PVPP 가

20 PVPP 가 665 nm ,  
가 665 nm

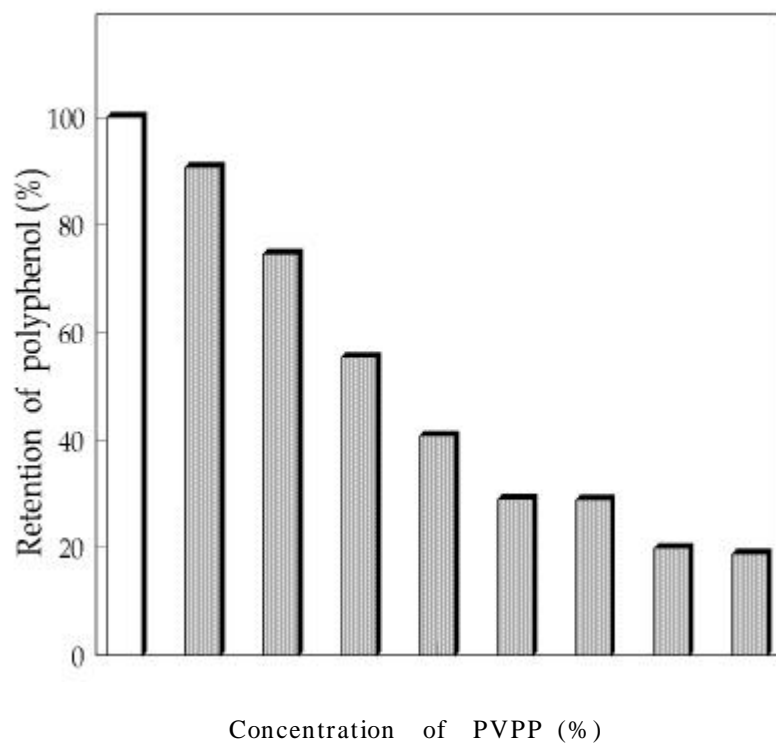
PVPP 가

. PVPP 가 가 PVPP 0.05%  
가 가 가 PVPP  
가 가 . 19 PVPP  
가 가 가

20. PVPP , Hunter L,a,b  
value

PVPP (%)	value (665 nm)	Hunter value			sensory evaluation <sup>a)</sup>
		L	a	b	
0	0.42	73.65	- 14.01	45.47	4.0
0.05	0.40	73.63	- 14.11	45.00	1.9
0.1	0.40	73.62	- 14.14	44.94	2.0
0.2	0.40	73.86	- 14.28	44.99	2.0
0.3	0.40	73.92	- 14.38	45.00	2.2
0.4	0.40	73.99	- 14.44	45.04	2.2
0.5	0.40	74.09	- 14.46	45.02	2.2
0.8	0.40	74.28	- 14.46	44.94	2.2
1.0	0.40	74.33	- 14.45	44.95	2.4

a) values are mean : 1;very good, 2;good, 3;fair, 4;poor, 5;very poor



19. PVPP

(2) PVPP

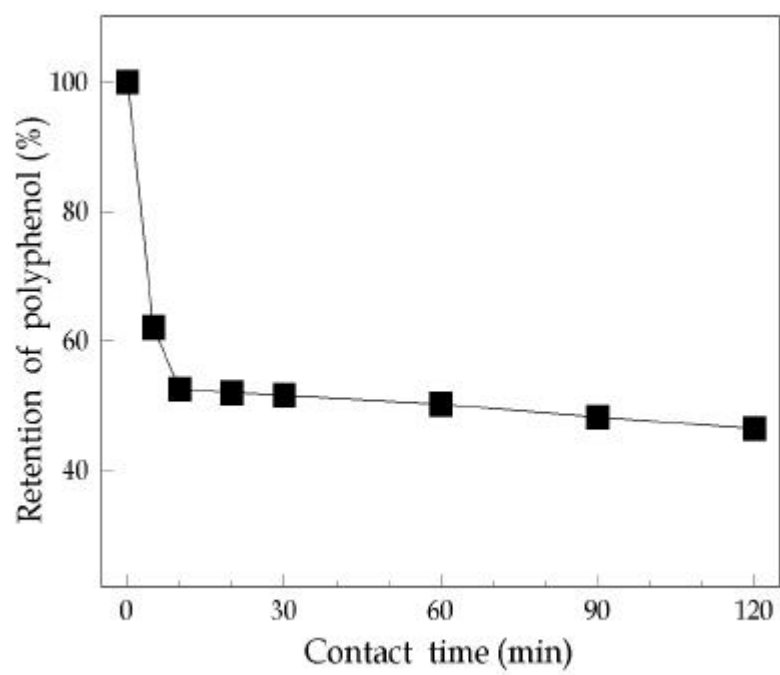
20 PVPP  
10 52.51%,  
120 46.41% 10

(3) PVPP

PVPP  
tannic acid 8 mg% 가  
PVPP 0.1% 가 5 min , 5 min 0.2 μm pore size  
filter pad  
21 가

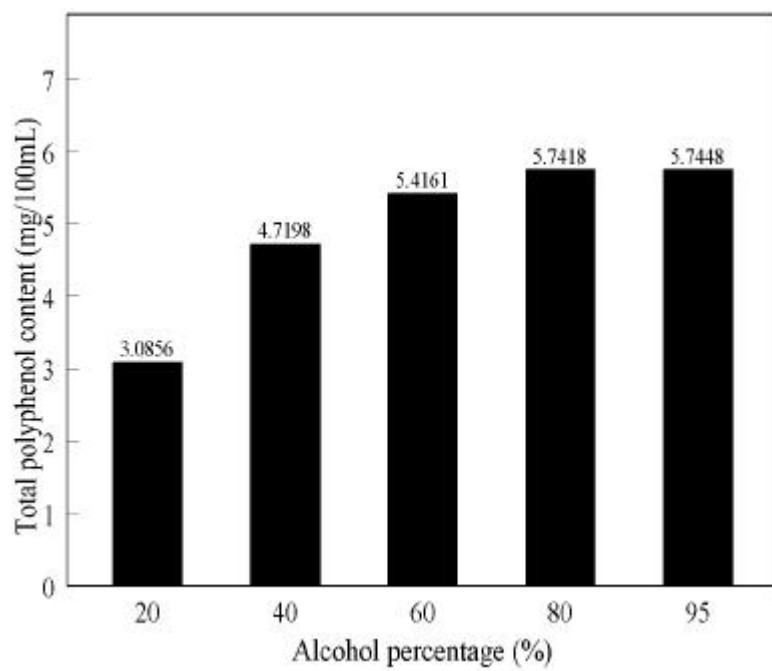
PVPP  
가 '14020'  
, 22 0 120  
, ,  
23 21 22 PVPP  
(c)가 PVPP

가 PVPP  
(a)

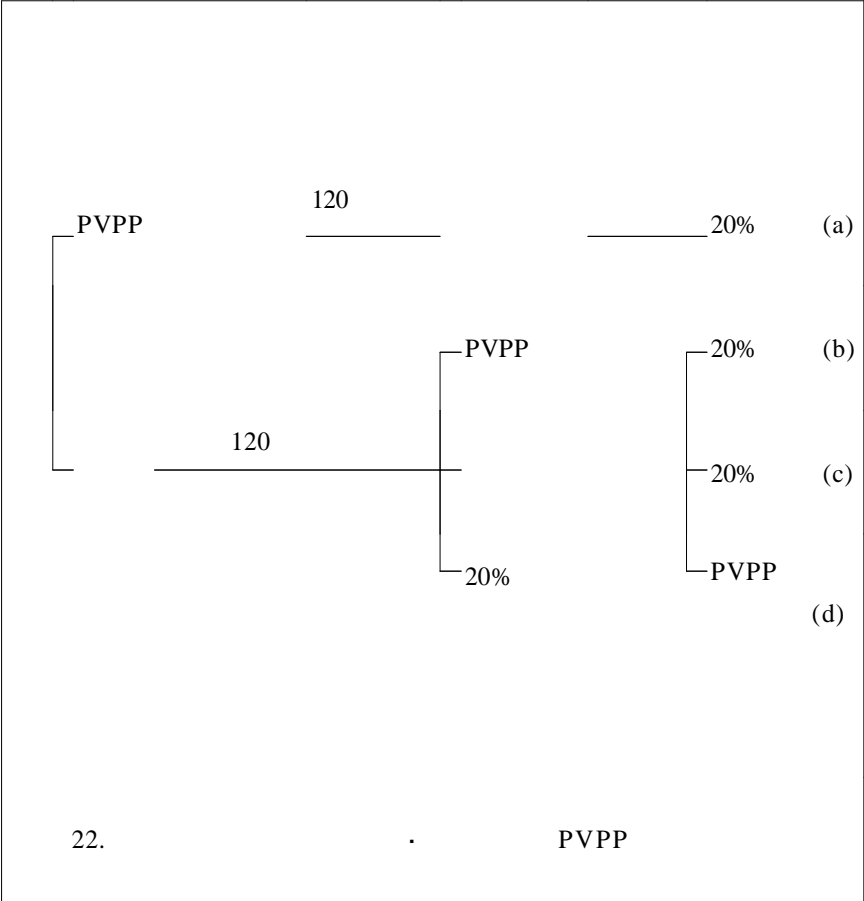


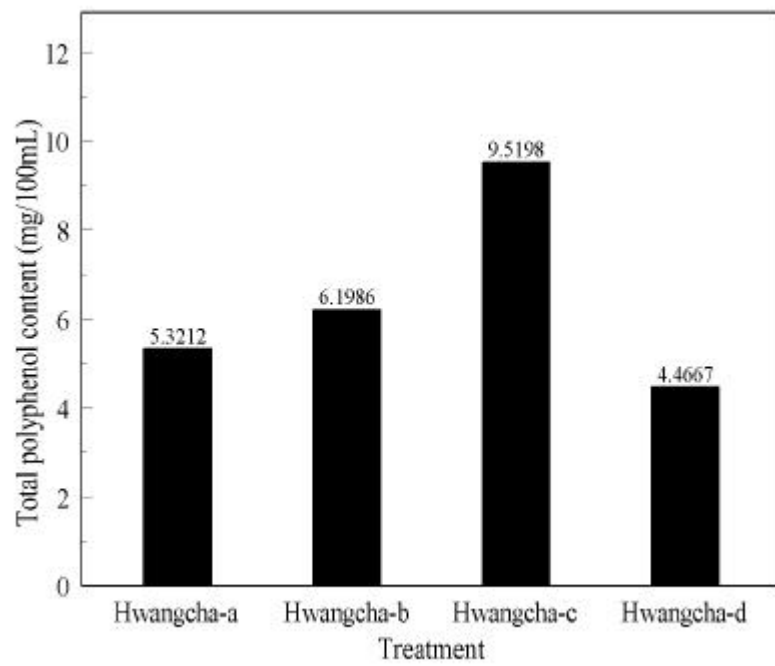
20.

0.2% PVPP



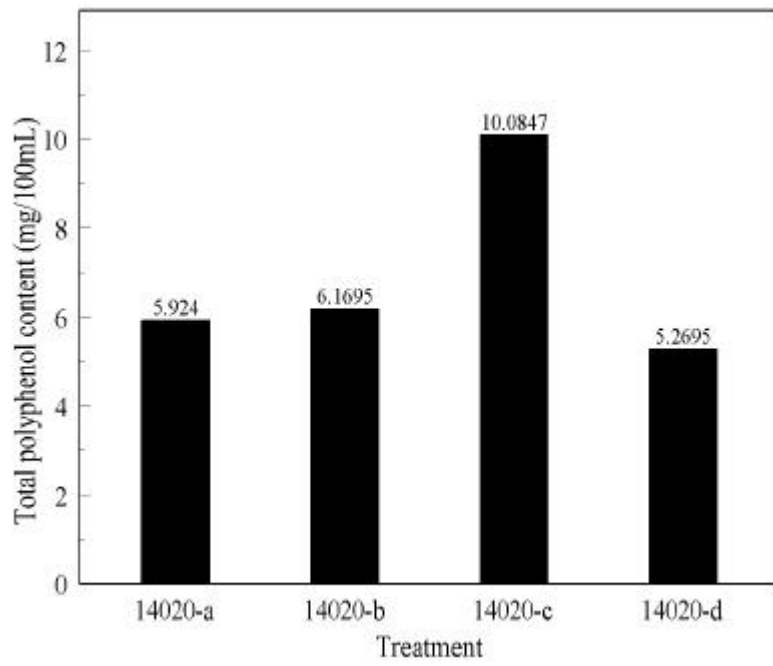
21. PVPP





23.

PVPP ( )



24.

PVPP (14020)



21.		PVPP	
	Turbidity(%)	Sensory evaluation1)	
14020- a	82.97	1.6	
14020- b	81.26	1.6	
14020- c	78.85	4.5	
14020- d	65.15	2.9	
Hw angcha- a	78.85	1.7	
Hw angcha- b	75.79	1.8	
Hw angcha- c	73.61	4.7	
Hw angcha- d	80.17	3.0	

1) values are mean : 1;very good, 2;good, 3;fair, 4;poor, 5;very poor

22.	PVPP			Hunter L, a, b
	Hunter			E
	L	a	b	
14020- a	85.95	- 7.18	26.37	30.73
14020- b	85.91	- 3.63	27.29	54.13
14020- c	85.06	- 8.16	28.51	33.19
14020- d	53.38	- 3.63	27.29	54.13
Hwangcha- a	84.97	- 9.00	30.25	34.95
Hwangcha- b	84.23	- 9.11	30.34	35.38
Hwangcha- c	83.35	- 9.51	31.98	37.28
Hwangcha- d	81.88	- 6.08	27.33	33.35

3)

가

(molecular weight cut- off) 30,000

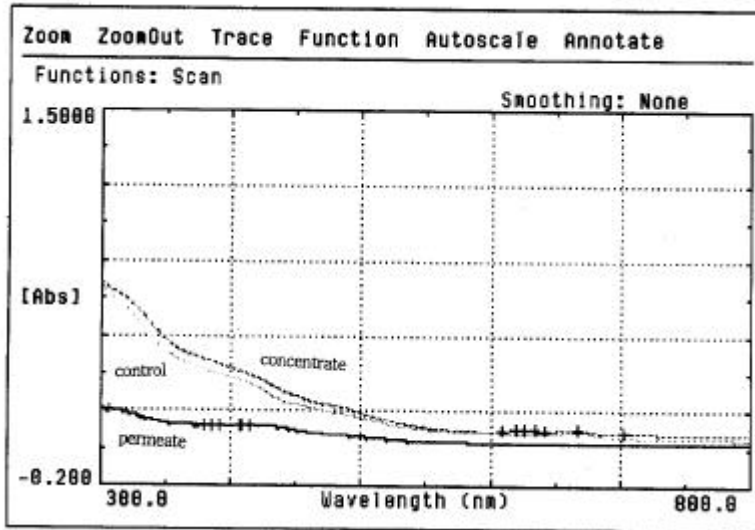
polyacrylonitrile

23

25

23.

	%T	Hunter		
		L	a	b
control	89.55	77.97	1.49	24.75
concentrate	56.95	42.92	6.77	24.81
permeate	99.41	86.62	- 0.18	24.73



25.

7.

95% PVPP 0 120  
 20% 16 100  
 mL sonicator 5 sealing 가  
 40 incubator ,  
 24  
 가  
 95% 20% 가

24.

Day	Turbidity	Hunter			Bacteria (CFU/mL)	Yeast and molds (CFU/mL)
		L	a	b		
0	96.29	92.21	-2.57	9.12	0	0
15	96.70	91.73	-2.23	8.78	0	0
50	96.70	90.93	-1.81	9.70	0	0
80	96.28	90.98	-1.78	9.26	0	0
100	96.41	90.54	-1.69	9.60	0	0
120	96.54	90.43	-1.68	9.60	0	0
180	97.97	88.60	-0.88	8.57	0	0

8.

1) 가

, , 95%

가 72

25

가

25. 가

	Hunter		
	L	a	b
Control	61.73	- 18.71	39.76
Apple juice	48.89	- 5.61	31.78
Apple flesh	62.66	1.26	40.31
Pear juice	60.23	- 12.30	38.55
Pear flesh	60.51	- 1.38	38.74

2)

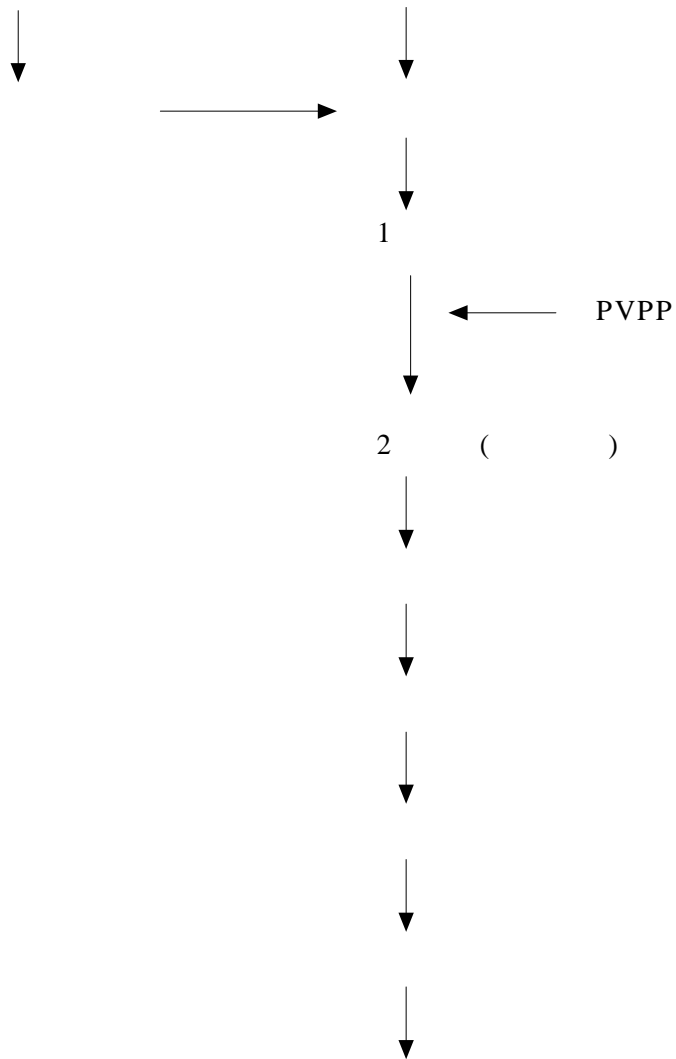


1) 가  
 가  
 가 ('99, 10)  
 가 ('99, 10)  
 가 offer ('99, 10)  
 , , , label  
 ( , )

27. 360 mL 가

가	(g)	가 ( )
		27.0
(95%)	95.8 mL	90.0
가		10.0
	1	60.0
		25.0
		212

2)





3)

28.

---

			가	( )
<b>A Extraction and Blending System</b>				
1.	Tank	5 set	1,200,000	6,000,000
2.	Filter	3 set	800,000	2,400,000
3.	Storage Tank	40 set	4,000,000	160,000,000
4.	Blending Tank	2 set	4,800,000	9,600,000
5.	Line Filter	1 set	1,000,000	1,000,000
<b>B</b>				
1.	Turn Table	1 set	4,000,000	4,000,000
2.	Rotaty Washer	1 set	45,000,000	45,000,000
3.	Filler and Sealer	1 set	95,000,000	95,000,000
4.	Cap Feeder	1 set	5,000,000	5,000,000
5.	Cap Sorter	1 set	6,000,000	6,000,000
6.	Glue Labeller	1 set	50,000,000	50,000,000
7.	Case Packer	1 set	48,000,000	48,000,000
8.		1 set	32,000,000	32,000,000
9.	Control System	1 set	26,000,000	26,000,000
10.	Ink Jet Printer	1 set	14,000,000	14,000,000
			<b>336,800,000</b>	

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26.

1. . . . . (1996)
2. Nakabayashi, T., Ina, K. and Sakata, K. Chemistry and function of green, black and oolong tea. Kogagu press, Japan (1994)
3. Lee, M.G., Lee, S.W., Kim, S.S., Lee, S.H. and Oh, S.L. Changes in testing constituents (tannin, free sugar, total nitrogen) of green tea by leaching condition. Korean J. Dietary Culture. 4: 411-416 (1984)
4. Park, J.H. Studies on chemical composition in Korean native tea plants. Ph.D. Thesis, Chonnam National Univ., Korea (1997)
5. Suematsu, S., Hisanobu, Y., Saigo, H., Matsuda, R., and Hara, K. Effects of pH on stability of constituents in canned tea drinks. J. Japanese Soc. Food Sci. Tech. 39: 178-182 (1992)
6. Komatsu, Y., Suematsu, S., Hisanobu, Y., Saigo, H., Matsuda, R. and Hara, K. Effects of pH and temperature on reaction kinetics of catechins in green tea infusion. Biosci. Biotech. Biochem. 57: 907-910 (1993)
7. Ikegaya, K., Takayanagi, H. and Anan, T. Quantitative analysis of tea constituents. Bull. Natl. Res. Tea. 71: 43-74 (1990)
8. Matsuzaki, T.L. and Hara, Y. Antioxidative activity of the leaf catechins. J. Agric. Chem. Soc. Japan. 59: 129-134 (1985)
9. Francis, F.J. Colorimetry of liquids. Food Technol. 26: 39-48 (1972)

10. Yoshioka, H., Sugiura, K., Kawahara, R., Hujita, T., Makino, M., Kamiya, M. and Tsuyumu, S. Formation of radicals and chemiluminescence during the autoxidation of the catechins. *Agric. Biol. Chem.* 55: 2717-2723 (1991)
11. Suematsu, S., Hisanobu, Y., Saigo, H., Matsuda, R., and Komatsu, Y. A new extraction procedure for determination of caffeine and catechins in green tea. *J. Japanese Soc. Food Sci. Tech.* 42: 419-424 (1995)
12. Nakagawa, M. Correlation of the constituents with the organoleptic evaluation of green tea liquors. *J. Japanese Soc. Food Sci. Tech.* 16: 12-18 (1974)
13. Masuda, S. and Nakagawa, M. General chemical and physical analysis on various kind of green tea. *Tea Res. J.* 46: 73 (1978)
14. Nakagawa, M., Buruya, G.M. Differences in amino acid, tannin, total nitrogen contents in leaves of cultivated species of green tea. *Bull. Japan Tea-Technician's association.* 48: 84-95 (1975)
15. Komatsu, Y., Hisanobu, Y., Suematsu, S. and Saigo, H. The method of making canned tea drinks. Japanese Patent 5-168407 (1993)
16. Terada, S., Maeda, Y., Masui, T., Suzuki, Y. and Ina, K. Composition of caffeine and catechin components in infusion of various tea(green, oolong and black tea) and tea drinks. *J. Japanese Soc. Food Sci. Tech.* 34: 20-27 (1987)
17. Iwaasa, S. Studies on bio-synthesis of catechin in tea. *Bull. Natl. Res. Tea.* 13: 101-135 (1977)

18. Yamamoto, T., Juneja, L.R., Chu, D.C. and Kim, M. Chemistry and applications of green tea. CRC Press, Boca Raton, New York, USA (1997)
19. . (1980)
20. . (1995)