

636.085
L 2937

GOVP 12007942

최종연구보고서

**가축의 환경친화형 사료개발과 무공해
축산물 생산을 위한 천연 항생물질의 개발**

**Development of Environmentally Sound
Livestock Feeds and Natural Antibiotics to
Produce Pollutants-free Animal Products**

주관연구기관 : 강원대학교
위탁연구기관 : (주)천보 실업

농 립 부

“가

”

.

1999 . 10 . 31 .

:

:

:

:

: ()

:

.

가

.

flavonoid propolis

가

steroid

.

.

가

cross - contamination 가

가

가

가

.

가

가

가

·
가

가

가

가

가

가

sarsaponin

가

,
(probiotics)

가

가

sarsaponin

· sarsaponin 가 ammonia
가

propolis

sarsaponin

·
sarsaponin

가

.

.

1. saponin
가 (:)

가. steroid ,
가

1) steroid

2) Yucca extract propolis 가가 E. coli

3) Propolis 가

.

1)

2) steroids 가

3) VOC(Volatile organic compounds)

. Sarsaponin

2. propolis 가 (:)

가. Propolis 가

. Propolis

. Propolis 가가

1) Propolis 가가

2) Propolis 가가

3) Propolis 가가

. 가가

. propolis 가가 propolis

. 가가

3. propolis 가 ()
:

가. Propolis 가

1) Propolis

2) propolis 가 *in vitro*

. Propolis 가

1) Propolis *in vivo* 가

2) Propolis가 T- Interleukin-
in vitro

3) Propolis 가 ICR mice Sarcoma 180

4) Propolis가

5) *in vitro*

propolis

. HPLC propolis

. Propolis 가

1) Propolis

2) Propolis

. Propolis

1) propolis 가

2) TTC(Triphenyl tetrazolium chloride) reduction test
propolis 가

4. 가 (

: ())

가.

가

1)

2)

가

.

1)

2)

3)

.

가

가

1)

가

가

2)

.

1.

가. saponin
가

1) steroid
가

가 가 가
가 3가 . 1
가

Yucca (YS), (PGR),
(CLPW), 가 (EEGB) (EEWB)
saponin .
saponin 가 *Streptococcus faecium*
cernelle 68(SF68) *Saccharomyces cerevisiae*(Yeast)
가 .

2 가

propolis yucca extract
SF68 E. coli

가 가 5가 3 propolis
(Lactose, Casein, Na-casein,
Red dog Cookie byproduct)
가 .

PGR 30mg 가

CLPW 가 0.1mM

YS 가

. EEGB WEGB 12mg
50% . PGR, CLPW EEGB
WEGB YS

saponin 가 SF68
 Yeast YS 1500ppm 가
 CLPW 500ppm Yeast
 PGL EEGB Yeast WEGB 100ppm
 가 0.5% SF68 Yeast
 가
 steroid 가 Yeast SF68
 YS 가 PGR 500ppm 가
 CLPW EEGB WEGB 100ppm 가 15, 33, 24
 가 SF68
 SF68 propolis 100ppm 가
 E coli 500ppm propolis 가
 가 Yucca extract 가 SF68 E coli
 가 propolis cookie byproduct 가
 70 가 90
 Na-casein propolis 가
 saponin
 가 yucca
 가 가 가 steroid saponin
 propolis
 SF68 steroid
 propolis 가

가 .

2)

steroid

H2O 50% ethanol

odor intensity ,

GC GC Mass

VOC

(PGR)

(GB)

saponin

PGR

GB 1%

가

1 , 3 , 7 , 14

가 .

가

odor

intensity가 ,

가

odor

intensity가 가

odor intensity

14

가 .

odor intensity

가

50%

ethanol

PGR 1%

GB 1%

7 , 14

odor intensity VOC

GC Mass

VOC dimethyl sulfide, methyl

chloride, 2-methyl pentane, 3-methyl pentane, dimethyl disulfide,

2-methyl hexane, heptane, toluene, acetic acid, ethyl benzene,

p-xylene, styrene, o-xylene

VOC acetic acid, ethyl benzene, p-xylene, styrene, o-xylene
dimethyl sulfide, dimethyl disulfide

가 PGR 1%, GB 1% 가

acetic acid,

ethyl benzene, p-xylene, styrene, o-xylene

dimethyl sulfide, dimethyl disulfide

steroid saponin PGR GB down stream
가 가 가 .

3) Sarsaponin

+ propolis 가

가 가 .

propolis 가 NH3 2

가 4

가

NH3 0.7% 1.4% 가

가 50 80%

H2S 0.7% 1.4% 가

+ propolis 가

가 가

propolis가 가

가 가 1.93 1.98

가

propolis

가

2) Propolis

propolis 가가

propolis 100, 200, 300, 400ppm

500ppm

가 6

가

1937g

propolis 가 가

1,986 2,041g

(p<.05) 가

300ppm 가 D

가

4,127g

propolis 가 가 4,154 4,183g

2.13

propolis

가 2.11 2.04

(p<.05)

, D

가

propolis

가 가

3) Propolis

가가

Propolis

propolis

300ppm

. , , pH
가 .
가
18. 25kg propolis
가 18. 39 18. 36kg
. , ,
가 .
가 .

4) 가가

가 propolis 가

가
가

가

가 가

5) propolis 가가

propolis

Propolis 가

Propolis 가

Propolis

60

, 가

Propolis

가

가

propolis

가

가

가

Pro-600

pro-300

Pro-300 600 가

Pro-600

Pro-300 600

Pro-300 600 가

Propolis

propolis가

6) 가가
 deodorizer 가 ,
 deodorizer 가
 deodorizer 가
 ,
 , , ,
 가
 가 0.7% 가 DO-50
 , , 가
 deodorizer 가 가

propolis 가
 1) Propolis 가
 97% ethanol 60%
 30g 14g propolis 46.6%
 50% 50%
 ethanol 60% .

Staphylococcus uberis, Staphylococcus aureus, Klebsiella pneumoniae, Proteus vulgaris, Staphylococcus agalactiae, E. coli

2) Propolis 가
 propolis ,

T-cell 가 IL-receptor
 propolis
 propolis 50ppm 10.25% 2
 IL-receptor
 ICR mice Sarcoma 180 propolis
 가

MCF7 0.1µg/ml 10%
 가
 propolis 가
 propolis neutrophil

3) HPLC propolis
 Antiviral activity antibacterial activity caffeic
 acid p-coumaric acid *Staphylococcus aureus*
 cinnamic acid propolis
 anti-viral gastric ulcer flavone
 glycoside rutin HPLC
 propolis 가
 propolis caffeic acid,
 p-coumaric acid, cinnamic acid rutin 가

propolis
 가

4) Propolis 가

E coli propolis 2 300
mg *E coli* 가

가

propolis

. *E coli*

propolis

300mg

6

가

24

propolis

flavonoids

가

propolis

가

가

.

5) Propolis

TTC-reduction test

Staphylococcus

aureus

propolis

500ppm

Staphylococcus agalactiae

Staphylococcus aureus

propolis가

50,000ppm

5,000ppm

500ppm

가 5,000ppm

4mm

Staphylococcus aureus

가

propolis가

Gram (+)

Staphylococcus uberis, *Staphylococcus aureus*, *Klebsiella pneumoniae*,

Proteus vulgaris, *Staphylococcus agalactiae*, *E coli*

가 HACCP

IMF
가

2002 HACCP 2001
HACCP
가

HACCP

가 2002

가

2.

가. flavonoids propolis

1)

가

2)

brand propolis
가 . - 97

(“ 가 가 (: 97-56291)”

3) 가

4) 가

5) propolis

6) Propolis , , , 가 가

1) saponin

가 deodorizer(Deodo)[®] flavonoid propolis 가 (complex feed supplement : Deodo-pro[®]) NH3 가

2) VOC(Volatile organic compounds) 가

3)

S U M M A R Y

In recent, the odorous substance like NH₃ and H₂S from manure play a critical role in livestock industry. And antibiotics residues in carcass and dairy product by drug abuse show a serious public health. Therefore, the purpose of these studies is to develop a dual purpose environmentally sound feed additives from the domestic natural substances which has both anti-microbial activity and odor control capability to produce pollutants-free animal products.

Part 1. Development of feed supplement from domestic saponin containing substances and natural anti-microbial substance

1. Development of natural bio-active substance to improve productivity and to control manure odor in animal production system

To develop a dual purpose feed additive that has both anti-microbial activity and odor control capability from the domestic natural substances, a series of basic nutraceutical evaluation studies have been executed.

In trial 1, to scrutinize potential odor control substance, five plant steroids extracted from *Yucca shidigera*(YS), *Platycodon grandiflorum*s root(PGR), *Condonopsis laceolata*'s processing waste(CLPW), ethanol-extracted Ginseng byproduct(EEGB) and water-extracted Ginseng byproduct(WEGB) were employed to be evaluated ammonia binding capacity. Two probiotic microorganisms, *Saccharomyces*

Cerevisiae(Yeast) and *Streptococcus faecium cernelle* 68(SF68) were then employed to test whether there is either synergistic or antagonistic action with the extracted plant steroids. In trial 2, propolis, a natural anti-microbial substance was evaluated to replace conventional antibiotics whether there is an interaction with probiotic SF68 and there is inhibition for *E. coli*. To develop multipurpose additive, the microbial growth assay was performed simultaneously with the yucca extract. In trial 3, melting point and morphological change of propolis after heat treatment were measured after adding one of the five carriers, casein, lactose, Na-casein, red dog and cookie byproduct to the propolis.

The results are summarized as follows,

Plant steroids extracted from PGR, CLPW, EEGB and WEGB were superior to the commercial YS in terms of ammonia binding capacity. Over 30mg of PGR had the highest ammonia binding capacity under higher ammonia dose. Over 12mg of EEGB and WEGB can bind over 50% of ammonia from any level of ammonia dose.

There was no influence of extracted plant steroids on the growth of SF68. 1500ppm of YS decreased the growth of Yeast. The growth of yeast was stimulated over control with 500ppm of CLPW. Although 100ppm of WEGB can stimulate the growth of Yeast, the yeast growth pattern was not different from those shown under 500ppm to 2000ppm of WEGB. PGL and EEGB had no effect on the growth of Yeast.

When the various concentrations of ammonia water were added in the broth of SF68 and Yeast, the growth of both probiotic microorganisms was retarded once the ammonia level exceeds 0.5%.

With more than 0.5% ammonia water in the broth, the extracted

plant steroids can not compensate the growth of the already inhibited Yeast. 1000ppm of PGR stimulates the growth of SF68, while the growth of SF68 was not affected by YS addition. All concentrations of CLPW, EEGB and WEGB can stimulate the growth of SF68 respectively after 15hrs, 24hrs and 24hrs.

Propolis has reduced the viability of SF68 although the viability of E. coli was reduced only by adding over 500ppm of propolis. There was no inhibition effect by yucca extract on the growth both SF68 and E. coli.

Melting initiation temperature of propolis incorporated with the cookie byproduct meal was lower than propolis incorporated with the other carrier but higher than that of pure propolis. Morphological change of carrier-incorporated propolis observed after heat treatment revealed the most efficient adsorption of propolis to the Na-casein.

In conclusion, the tested plant steroids were superior to YS in terms of ammonia binding which represents potential odor control capability. The microbial growth promoting or inhibiting effect of the plant saponins was subjective to the type of both saponins and microbes.

Since the propolis inhibits the growth of probiotic microbe SF68, it should be very cautious to combined add both propolis and probiotics simultaneously to the feed.

2. Identification of odorous substances from swine manure and down stream odor control using plants steroids

To examine the major odorous substance from swine manure and to evaluate plant steroids as an odor depressing agent, both olfactory

and GC/ GC Mass analysis have been executed.

The sample of swine manure have been fractioned with H₂O and 50% ethanol to categorize the possible odorous substances into either soluble or insoluble fraction. Swine manures mixed with either 1 % *Platycodon grandiflorum's* root(PGR) or 1% *Ginseng* byproduct(GB) and none of those have been stored 1 day, 3 days, 7 days, and 14 days at room temperature.

To scrutinize major odor generating substances from volatile organic compounds(VOCs) both GC and GC Mass analysis using specific absorbent tube, and olfactory odor intensity evaluation by specific sniffing detector have been executed. To develop the possible down stream odor depressing agent both PGR and GB were evaluated by the above odor analysis procedure.

The results can be summarized as follows :

After centrifugation of swine manure, the supernatant fraction exert stronger odor than the precipitates fraction. With increasing dilution of swine manures into water olfactory odor intensity was also increased, which indicate that the odor generating substances are generally water soluble and at least water mixible.

Odor intensity was relatively strong in soluble fraction of swine manure after 3 days at field temperature. However, the intensity was extremely strong in the insoluble fraction compare to the soluble after 14 days of storage.

Between soluble and insoluble fraction by 50% ethanol, olfactory odor intensity was relatively strong in ethanol insoluble fraction of swine manure.

In swine manure mixed with either 1% PGR or 1% GB, both olfactory

odor intensity and VOCs were decreased as storage time goes by 7 days to 14 days.

Major odorous substances of VOCs in swine manure analyzed by GC Mass were dimethyl sulfide, methyl chloride, 2-methyl pentane, 3-methyl pentane, dimethyl sulfide, 2-methyl hexane, heptane, toluene, acetic acid, ethyl benzene, p-xylene, styrene and o-xylene.

Among the odorous VOCs, acetic acid, ethyl benzene, p-xylene, styrene and o-xylene from hydrocarbon compounds and dimethyl sulfide and dimethyl disulfide from sulfide compounds were decreased with either 1% PGR and 1% GB addition after 7 to 14 days of storage at field temperature.

In conclusion, major odorous substances of swine manure considered to be ethyl benzene, p-xylene, styrene and o-xylene in hydro carbon compounds and dimethyl sulfide and dimethyl disulfide in sulfide compounds. Both PGR and GB were effective to control odor of swine manure at down stream process.

3. Market-ready production technology of feed additives imparting odor controlling and natural antibiotic compounds

To be ready for commercial production of feed additives to reduce fecal odor emission, two type of feed supplements were developed throughout this project.

The first market ready product is deodorizer(Deodo)[®] which are composed of several natural polymers including plant saponin. The second product is a dual purpose mixture(Deodo-pro)[®] combined both deodorizing substances and natural anti-microbial agent, propolis.

Both products were developed as a ready-to-add type of powder that

are well mixible with traditional feed ingredients and formula mash diet. The products are also heat stable and easy to apply for pellet processing of the feeds.

Dietary addition of both products were effective ($P < 0.05$) to reduce down to 50% of odor emission represented by fecal NH_3 and H_2S . As far as odor reduction is concerned, the product, Deodo[®] was more effective than the combined product of deodorizer and propolis (Deodo-pro)[®].

Part 2. Animal bio-evaluation study for natural antibiotics, propolis and odor controlling compounds

1. Effect of feeding propolis extracts and *Yucca Schidi gera* on body performance of broiler chicks

This study was carried out to determine the effect of feeding ethanol extract propolis (EEP) and yucca extracts (YE) supplementation on body weight gain, feed intake, feed efficiency and carcass composition. A total 500 broiler chicks were assigned to 10 treatments. Each treatment had 10 chicks with 5 replications. The supplementation levels of EEP and YE used in this experimental were 40, 80, 120 and 160ppm for EEP and 150 and 300ppm for YE respectively. The results obtained summarized as follows: The body weight gain (BWG) of chicks fed different levels of EEP and YE were similar that of control. Also, Feed intake in each treatment was similar over entire period. Feed efficiency of chick fed EEP and YE were slightly improved compare with control, but not significantly

different among treatments. There was no significantly different in carcass composition among the treatments.

2. Effect of feeding of various level of propolis on body performance of broiler chicks

This study was carried out to determine the effect of feeding ethanol extract propolis (EEP) supplementation on body weight gain, feed intake, feed efficiency. A total 280 broiler chicks were assigned to 7 treatments. Each treatment had 10 chicks with 4 replications. The supplementation levels of EEP used in this experimental were 100, 200, 300, 400 and 500ppm. The results obtained summarized as follows: The body weight gain (BWG) of chicks fed different levels of EEP significantly increased ($p < .01$) compare with that of control. Especially the treatment with 300ppm of EEP produced higher BWG among other treatments. Feed intake was not significantly different among treatments. Feed efficiency (FE) of chick fed EEP was significantly ($p < .05$) improved compare with control. and the highest in the treatment with 300ppm of EEP. Therefore, the most efficient result were found in the chicks of EEP treated 300ppm for BWG and FC.

3. Effect of feeding propolis extracts and Yucca Schidigera on rumen characteristics, milk production and growth performance of cattle

This study was carried out to evaluate the effect of feeding Yucca shidigera extract (YSE) and ethanol extract propolis (EEP) on rumen characteristic, lactation performance of Holstein cows and

growing performance of Korean native bulls. The supplementation levels of YSE and EEP used in this experimental were 300ppm respectively. The results obtained summarized as follows: Concentration of total amino acid (μM), ammonia, rumen microbial synthesis were not significantly difference among treatments. In cows trial, daily intake of dry matter was not significantly different among treatment. Milk production showed slightly increased for YSE and EEP than for control. But the significant difference among each treatment were not observed. In Korean bull trial, daily body gain and feed intake were not significantly different among treatments. Also, the carcass composition of bulls were not different among the treatments.

4. Effect of supplementing complex additive for both odor control and antibiotic action on the growing performance of pigs

Biological efficacy of feed supplements(deodo-pro) which have both deodorizing capability and natural antimicrobial activity was evaluated using growing swine with duplicated doses of the supplement to the diet.

Addition of complex feed supplement which have both deodorizer and natural anti-microbial propolis did not improve growth performance of finishing pigs although there was a numerical advantage due to the high level addition of the supplement. However, protein digestibility of growing pigs diet was improved($P<0.05$) by the addition of complex supplement. Both DM and energy digestions were also numerically improved by the addition.

Amino acid digestibility were also improved($P<0.05$) by the

addition of the complex supplement to the growing pig diet.

5. Effect of dietary propolis addition on the growing performance of pig and propolis impartation to the pork

Supplementation of propolis(pro) into swine diet as a natural anti-microbial agent and therefore possible growth promotor was evaluated. Two levels(300 and 600ppm) of propolis were added to swine finisher diet then both growth performance, nutrient digestibility and biological deposition of function substances into the meat were analyzed.

There was no marked effect of dietary propolis medication in growth rate, feed conversion and feed consumption of pigs. However, the nutrients digestions were improved($P < 0.05$) by both low and high level of propolis addition to the diet. Digestions of several essential and non-essential amino acids were also improved($P < 0.05$) by the propolis medication.

There was no biological transformation and bio-accumulation of propolis into the meat due to the feeding of the substance.

6. Dietary supplementation of odor controlling substances and their effect on the growing performance of the pig

Supplementation of deodorizing substances(Deodo) into the swine diet was evaluated with swine fed two levels of mixed deodorize. Low level addition of deodorizer has numerically improved growth rate and feed conversion rate although higher level addition negate the advantage.

However the high level addition of the deodorizer certainly

improved($P < 0.05$) dietary DM protein and GE digestions from no added control and low level added diet.

Amino acid digestions were also improved($P < 0.05$) by the high level addition of deodorizer to the diet.

Part 3. Isolation of propolis and their application for the disease control

1. Development of optimal isolation and evaluation of anti-bacterial effect for propolis

Propolis is a specialized, natural resinous, waxy material with which honey bees seal their colonies from the outside world. Besides its resinous character, propolis has a second property which is important in the beehive: it prevents the growth of bacteria.

Propolis has proved to be a highly efficient first-line protection against a wide spectrum of bacterial infections

First, in order to optimize the method to determine the optimal isolation method of crude propolis, we used different concentrations of ethanol. The optimal concentration of ethanol to separate a best quantity of propolis was 60%.

Second we determined the bactericidal or bacteriostatic effect of the natural propolis introduced in the growth media as well as the bactericidal and bacteriostatic effect of ethanol extracted propolis. The results of this study proved that all propolis has pronounced antibacterial effect.

2. Immunological effects of propolis

The aims of this study are to know the immunological and tumoricidal activity of propolis.

1) Effect of propolis on the *in vivo* immune function of mice

In order to determine the influence of propolis on immune function, various concentrations of propolis were administered to mice for 10 weeks. And the function of various immune cells are analyzed by monoclonal antibodies. The propolis administered group showed an enhanced immune function to compared with untreated control group.

Especially, the expression of interleukin- receptor in propolis treated group was activated in compare with control group.

These results suggested that long term administration of propolis modulate the immune function *in vivo*.

T-cell antigen receptor(TCR) is a critical role to defense the body against tumor. The expression of TCR also increased in the propolis administered group. This result indicated that propolis may use a bactericidal as well as a immune enhancing purpose for the livestock.

2) Effect of propolis on the Interleukin- receptor expression of human T-cell *in vitro*

To analyse the effect of propolis on the human T cell *in vitro*, we isolated T cells from peripheral blood and these were incubated with 50ppm propolis for 16hrs. After that T-cell were labelled with anti-human CD25-FITC(IL-2 receptor) for analysis by flowcytometry.

The expression of IL- receptor in the propolis treated T-cell was increased with 2 fold in compare with control group.

These data indicated also that propolis may use as feed additive not only for a bactericidal but a immune enhancing purpose to the livestock.

3) Effect of propolis on the Sarcoma 180 bearing mice

In order to investigate the curative activity of per oral administration of propolis on tumor, ICR mice was subcutaneously implanted Sarcoma 180.

In the 300mg/kg propolis administered group on day 28th, development of implanted tumors was strongly inhibited by 83.5%.

The regression rate of tumor was dose dependent.

The data suggested propolis shows antitumor effect.

4) Effect of propolis on the established tumor cell lines

The antitumor activity of propolis examined against five kinds of established tumor cell lines. For the test we prepared human gastric carcinoma (KATO III), human lung carcinoma (A549), human breast adenocarcinoma (MCF-7), mouse lymphoma (Sarcoma 180), and mouse lymphoma (Yac-1). Propolis was highly effective at all treated doses (0.1µg/Me, 1µg/Me, 5µg/Me and 25µg/Me).

This result suggest propolis may have tumoricidal activity.

5) Effect of propolis on human neutrophil function *in vitro*

In order to determine the effect of propolis on the phagocytic activity of human neutrophil, we prepared human neutrophil and cocultured with propolis (10ppm, 50ppm and 100ppm) *in vitro*. And an phagocytic activity was measured by luminometer. After 10hrs

incubation neutrophil treated with 10ppm propolis showed moderately enhanced phagocytic activity(36%). And the enhanced phagocytic activity is time dependent.

3. Identification of pharmacologic components of propolis by using HPLC

This study was carried out to investigate of major pharmacologic constituents of propolis by HPLC.

The HPLC analysis revealed the occurrence of at least fifteen flavonoids. Four of major constituents of propolis, caffeic acid (3,4-dihydroxycinnamic acid), p-coumaric acid(4-hydroxycinnamic acid), cinnamic acid (3-phenyl-2-propenoic acid) and rutin(flavone glycoside) were identified by HPLC.

The major constituents of propolis differed from country to country. It has been suggested that the different geographical origin influenced to the efficacy and the constituents of propolis.

4. Prophylactic and therapeutic effects of propolis on neonatal diarrhea in calves and piglets

To determine the action of ethanol extract of propolis(EEP) as feed additives on the protection and the treatment of *E. coli* caused neonatal diarrhea in calves and piglets.

To know whether EEP can be substituted for commercial antibiotics on therapy of *E. coli* caused neonatal diarrhea in calves and piglets.

Group A calves and piglets which were fed EEP prior to *E. coli* administration showed no attack of diarrhea. And in group B calves and piglets were fed EEP for 3-5 days after the artificial induction

of diarrhea by *E. coli* administration, EEP showed an excellent curative effect. Group C calves and piglets as a positive control which were treated by enrofloxacin showed also a curative effect.

In the result of group B and C, EEP an equivalent effect to enrofloxacin for the treatment of neonatal diarrhea in calves and piglets.

Therefore we could notice that EEP has not only prophylactic but therapeutic effects on *E. coli* induced neonatal diarrhea in calves.

These result showed that EEP without any antibiotics residues can be substituted for commercial antibiotics on neonatal diarrhea in calves and piglets.

5. Effect of propolis on mastitis and TTC-reduction test

Mastitis remains the most costly disease in the dairy industry. Antibiotic residues following therapy during lactation remain a serious concern for a safe milk and meat supply. Therefore, in this study we determined whether the ethanol extract of propolis(EEP) could substituted for commercial antibiotics on mastitis therapy. For the test we used various strains of bacteria such as *Staphylococcus aureus*, *Staphylococcus epidermidis*, *Streptococcus agalactiae*, *Staphylococcus dysgalactiae*, *Staphylococcus uberis* and *Escherichia coli*.

Propolis showed anti-bacterial effects against all tested mastitis caused bacterial strains. In addition, propolis was sensitive to *E. coli* which was resistant to broad spectrum antibiotics like ampicillin,

A unique advantage of propolis is that it enhances the effectiveness

of antibiotics such as penicillin and streptomycin. The combination of propolis with commercial antibiotics can reduce drug dosages, minimize drug side effects, and decrease chances of drug resistance. These results concluded that propolis could substituted for commercial antibiotics. In order to know whether propolis is detected by TTC-reduction test, various concentrations of propolis, 1 100 ppm were reacted with TTC. In this test TTC-reduction in all concentrations of propolis showed negative reaction. The results indicated that propolis without any antibiotics residues may safely substituted for commercial antibiotics on mastitis therapy.

C O N T E N T S

Development of Environmentally Sound Livestock Feeds and Natural Antibiotics to Produce Pollutants-free Animal Products

Chapter . Introduction	51
1.1. Purpose and scope of study	51
1.2. Object and content of research and application of research results	54
1.3. Expected effects of research results	59
Chapter . Development of feed supplement from domestic saponin containing substances and natural anti-microbial substance	61
2.1. Development of natural bio-active substance to improve productivity and to control manure odor in animal production system	61
2.1.1. Introduction	61
2.1.2. Materials and methods	63
2.1.2.1. Ammonia gas absorption capacity and interaction with feed microbials of the several phytosteroids	63
2.1.2.2. Individual and compounding addition of yucca extract and/or propolis to the diet and its effect on growth of feed microbes and E. coli.	67
2.1.2.3. Evaluation of several bio-carriers for binding and emulsification capacity with propolis	68
2.1.3. Results and discussion	69

2.1.3.1. Ammonia gas absorption capacity and interaction with feed microbials of the several phytosteroids -----	69
2.1.3.2. Individual and compounding addition of yucca extract and/or propolis to the diet and its effect on growth of feed microbes and E.coli.-----	72
2.1.3.3. Evaluation of several bio-carriers for binding and emulsification capacity with propolis -----	73
2.1.4. Conclusions -----	93
2.1.5. References -----	95
2.2. Identification of odorous substances from swine manure and down stream odor control using plants steroids -----	102
2.2.1. Introduction -----	102
2.2.2. Materials and methods -----	104
2.2.3. Results and discussion -----	110
2.2.4. Conclusions -----	144
2.2.5. References -----	146
2.3. Market-ready production technology of feed additives imparting odor controlling and natural antibiotic compounds -----	154
2.3.1. Introduction -----	154
2.3.2. Materials and methods -----	155
2.3.3. Results and discussion -----	159
2.3.4. Conclusions -----	165
2.3.5. References -----	166

Chapter . Animal bioevaluation study for natural antibiotics,
propolis and odor controlling compounds ----- 168

3.1. Effect of feeding propolis extracts and *Yucca schidigera* on

body performance of broiler chicks -----	168
3. 1. 1. Introduction -----	168
3. 1. 2. Materials and methods -----	171
3. 1. 3. Results and discussion -----	176
3. 1. 4. Conclusions -----	180
3. 1. 5. References -----	181
3. 2. Effect of feeding of various level of propolis on body performance of broiler chicks -----	186
3. 2. 1. Introduction -----	186
3. 2. 2. Materials and methods -----	188
3. 2. 3. Results and discussion -----	190
3. 2. 4. Conclusions -----	194
3. 2. 5. References -----	195
3. 3. Effect of feeding propolis extracts and Yucca schidigera on rumen characteristic, milk production and growth performance of cattle -----	198
3. 3. 1. Introduction -----	198
3. 3. 2. Materials and methods -----	199
3. 3. 2. 1. Effect of both propolis and euca extract addition on ruminal function -----	199
3. 3. 2. 2. Effect of dietary propolis and euca extract addition on milk production of dairy cow -----	201
3. 3. 2. 3. Effect of dietary propolis and euca extract addition on growing performance of beef cattle -----	203
3. 3. 3. Results and discussion -----	205
3. 3. 3. 1. Effect of both propolis and euca extract addition on ruminal function -----	205

3.3.3.2. Effect of dietary propolis and eucca extract addition on milk production of dairy cow -----	209
3.3.3.3. Effect of dietary propolis and eucca extract addition on growing performance of beef cattle -----	211
3.3.4. Conclusions -----	215
3.3.5. References -----	216
3.4. Effect of supplementing complex additive for both odor control and antibiotic action on the growing performance of pigs -	219
3.4.1. Introduction -----	219
3.4.2. Materials and methods -----	220
3.4.3. Results -----	223
3.4.4. Conclusions -----	226
3.4.5. References -----	227
3.5 Effect of dietary propolis addition on the growing performance of pig and propolis impartation to the pork -----	229
3.5.1. Introduction -----	229
3.5.2. Materials and methods -----	230
3.5.3. Results -----	235
3.5.4. Conclusions -----	245
3.5.5. References -----	246
3.6 Dietary supplementation of odor controlling substances and their effect on the growing performance of the pig -----	248
3.6.1. Introduction -----	248
3.6.2. Materials and methods -----	249
3.6.3. Results -----	252
3.6.4. Conclusions -----	256
3.6.5. References -----	257

Chapter 4. Isolation of propolis and their application for the disease control	259
4.1. Introduction	259
4.2. Development of optimal isolation and evaluation of anti-bacterial effect for propolis	260
4.2.1. Introduction	260
4.2.2. Optimization of isolation for propolis	261
4.2.2.1. Materials and methods	261
4.2.2.2. Results and discussion	261
4.2.3. Anti-bacterial effect of propolis on main pathogenic microorganisms	263
4.2.3.1. Materials and methods	263
4.2.3.2. Results and discussion	264
4.3. Immunological effects of propolis	270
4.3.1. Effect of propolis on the in vivo immune function of mice	270
4.3.1.1. Materials and methods	270
4.3.1.2. Results and discussion	272
4.3.2. Effect of propolis on the Interleukin- receptor expression of human T-cell in vitro	276
4.3.2.1. Materials and methods	276
4.3.2.2. Results and discussion	276
4.3.3. Effect of propolis on the Sarcoma 180 bearing mice ----	281
4.3.3.1. Materials and methods	281
4.3.3.2. Results and discussion	282
4.3.4. Effect of propolis on the different tumor cell lines --	284
4.3.4.1. Materials and methods	284

4.3.4.2. Results and discussion	286
4.3.5. Effect of propolis on the human neutrophil function in vitro	288
4.3.5.1. Materials and methods	288
4.3.5.2. Results and discussion	289
4.4. Identification of pharmacologic components of propolis by using HPLC	291
4.4.1. Materials and methods	291
4.4.2. Results and discussion	292
4.5. Prophylactic and therapeutic effects of propolis on neonatal diarrhea in calves and piglets	302
4.5.1. Prophylactic and therapeutic effects of propolis on neonatal diarrhea in calves	302
4.5.1.1. Materials and methods	302
4.5.1.2. Results and discussion	304
4.5.2. Prophylactic and therapeutic effects of propolis on neonatal diarrhea in piglets	307
4.5.2.1. Materials and methods	307
4.5.2.2. Results and discussion	309
4.6. Effect of propolis on mastitis and TTC-reduction test ----	311
4.6.1. Effect of propolis on mastitis control	311
4.6.1.1. Materials and methods	311
4.6.1.2. Results and discussion	312
4.6.2. Reaction of propolis on TTC-reduction test	315
4.6.2.1. Materials and methods	315
4.6.2.2. Results and discussion	316
4.7. References	317

Chapter . Economic evaluation and marketing strategy of natural antibiotics and odor controlling compounds -----	326
5.1. Analysis of marketing situation for environmentally sound livestock product in the future -----	326
5.1.1. Analysis of the present market condition for antibiotics -----	326
5.1.2. Analysis of the present market condition for environmentally sound livestock product -----	335
5.2. Preparing of resources and economic analysis for propolis and odor controlling compounds -----	340
5.2.1. Preparing of resources and economic analysis for propolis -----	340
5.2.2. Preparing of resources and economic analysis for odor controlling compounds -----	356
5.2.3. Establishment of marketing strategy for propolis and odor controlling compounds -----	361
5.3. Preparing of resources and economic analysis for propolis and odor controlling compounds -----	371
5.3.1. Introduction -----	371
5.3.2. Economic analysis after feeding of dual purpose mixture (Deodo-pro) [®] -----	372
5.3.3. Marketing strategy of market ready product -----	375
5.3.4. Conclusion -----	386
5.3.5. References -----	387

1	51
1	51
1.	51
가.	51
2	54
1.	54
3	59
1.	59
2.	•	59
3. 가	60
2	saponin	
가	61
1	steroid ,	
가.	61
1.	61
2.	63
가.	steroid	
	63
.	Yucca extract propolis 가가 E. coli	
	67
.	Propolis 가	
	68
3.	69
가.	steroid	

	69
. Yucca extract propolis	가가 E. coli	
	72
. Propolis	가	73
4.	93
5.	95
2		
	102
1.	102
2.	104
가.	104
.	105
. Saponin	106
.	(Olfactory analysis)	106
.	VOC(Volatile organic compounds)	107
. GC Mass	109
3.	110
가.	(Olfactory analysis)	110
.	steroids 가	115
. VOC(Volatile organic compounds)	125
4.	144
5.	146
3 Sarsaponin	154
1.	154
2.	155
3.	159
4.	165

. Propolis	가가	.. 209
. Propolis	가가	
	211
4.	215
5.	216
4	가가	
	219
1.	219
2.	220
3.	223
4.	226
5.	227
5	propolis 가가	
	propolis	229
1.	229
2.	230
3.	235
4.	245
5.	246
6	가가	
	248
1.	248
2.	249
3.	252
4.	256
5.	257

4	propolis	가	259
.....			
1			259
2	Propolis	가	260
1.			260
2.	Propolis		261
가.			261
.			261
3.	propolis	가	in vitro
			263
가.			263
.			264
3	Propolis	가	270
1.	Propolis	in vivo	가
			270
가.			270
.			272
2.	Propolis가	T-	Interleukin-
	in vitro		
			276
가.			276
.			276
3.	Propolis	가 ICR mice	Sarcoma 180
			281
가.			281
.			282
4.	Propolis가		284
가.			284
.			286

5. in vitro		
propolis	288
가.	288
.	289
4 HPLC propolis	291
1.	291
2.	292
5 Propolis 가	302
1. Propolis	302
가.	302
.	304
2. Propolis		307
가.	307
.	309
6 Propolis	311
1. Propolis	가	311
가.	311
.	312
2. TTC(Triphenyl tetrazolium chloride) reduction test	propolis	
가	315
가.	315
.	316
7	317
5	가	
.....		326
1		

	가	326
1.	326
가.	가	326
1)	326
2)	327
.	가	278
1)	328
2)	329
.	Propolis	331
1)	331
2)	331
.	Sarsaponin	334
1)	334
2)	334
2.	가	335
가.	335
.	337
2		340
1.	340
가.	340
.	341
.	343
.	가	347
.	가	349
.	351
2.	356
가.	356

.	357
.	359
3.	361
가.	361
.	364
1)	364
2) Propolis	368
3 가 가	371
1.	371
2. 가 가	372
3.	375
가.	375
.	377
.	379
. HACCP	383
.	385
4.	386
5.	387

1

1

1.

가.

가

가

가

.

가

가

.

가

.

가

가

가

.

가

가

.

가

.

sarsaponin

가

, 가
(probiotics) 가

sarsaponin

sarsaponin 가 ammonia
가

propolis

sarsaponin

sarsaponin
가

1)

가)

steroid

가 가 .

)

)

가 가

)

가

, , ,

2)

가) 275

)

)

가

2)

가)

)

)

가) 1997 7 1

flavonoids

propolis

HACCP

가

)

brand

brand

propolis
가

propolis가
가

” propolis
brand
가 가

“propolis

3)

가

가

,

,

ammonia

가

가

가

가

가

2

1.

가.

flavonoid propolis

가

sarsaponin

steroid

.

1)

50%

2)

가

flavonoid

3)

cross-contamination 가

4)

5)

.

2.

<p>1 (1996)</p>	<p>Sarsaponin , 가</p> <p>Propolis 가</p> <p>propolis 가</p>	<p>o Sarsaponin o 가</p> <p>o Sarsaponin 가</p> <p>o Propolis o propolis 가 in vitro</p> <p>o Propolis in vivo 가</p> <p>o 가 가</p> <p>o Propolis 가 가 가</p> <p>o 가</p> <p>o o Sarsaponin propolis 가</p> <p>o</p>

2 (1997)	<p style="text-align: center;">sarsaponin</p> <p>Propolis 가</p> <p>가 sarsaponin propolis 가</p> <p>가</p> <p style="text-align: center;">()</p>	<p>o</p> <p>o 가</p> <p>o Sarsaponin 가</p> <p>o ()</p> <p>o in vitro propolis</p> <p>o propolis 가</p> <p>o Propolis ()</p> <p>o 가</p> <p>o 가</p> <p>o</p> <p>o 가</p>

3 (1998)	Sarsaponin propolis 가	o 가 o 가 o propolis o propolis 가 o TTC propolis 가 o 가 o 가 o 가 o () o 가 가 가 o 가 가

3. 가

가

,

.

,

가가

.

,

가 가

,

가

,

,

가 가

.

2

saponin

가

1

steroid

가

1.

가

가 가

(antibiotics)

가

(probiotics)가

가

가

propolis

가

(Aga et al, 1994; Grange et

al., 1990; Krol et al., 1993; Park et al., 1998; Takai si - ki kuni and Schilcher, 1994).

가 가

가
 .
 , 가 , 가
 가 . 가
 가
 가
 . 가
 가 가 (Cole,
 1993). 가
 . 가
 가 가

Yucca shidigera

steroid sarsaponin (Johnston et al., 1981;
 Wu et al, 1994). Sarsaponin saponin
 saponin , , , ,
 가 (Cheeke, 1971; Milgate
 and Roberts, 1995). Yucca saponin
 가 가
 . yucca saponin 가가
 saponin 가
 .
 yucca saponin ammonia
 (Headon et al., 1991; Ma et al., 1993)
 urease (Hussain et al., 1996; Yeo and
 Kim 1997; Preston et al., 1987) . yucca extract

phyto saponin 가 (Cole and Tuck, 1995) 가 .

가 .
ammonia .
가

가
가
가
가
)

sarsaponin
saponin .)

yucca extract
propolis 가
가

가가 가 .)
propolis 가
가 .

2.

가. steroid

1) Saponin

steroid (*Yucca shi digera*; YS),

(*Platycodon grandiflorum* s root; PGR), (*Condonopsis
laceolata*'s processing waste; CLPW)

가 (ethanol-extracted ginseng byproduct; EEGB)
(water-extracted ginseng byproduct; WEGB)

Fig. 2-1 .

2) 가
steroid 가
Wallace et al. (1994) NH₄Cl 0.1mM 0.2mM 0.3mM 0.4mM
1ml 가
steroid 가 37 1
0.4ml phenol-hypochlorite-nitroprusside method
(Weatherburn, 1967) Spectrophotometer (UVIKON 942, Italy)
650nm

3) steroid 가
Streptococcus faecium cernelle 68(SF68) *Saccharomyces
cerevisiae*(Yeast) .

4) steroid
0.22μm membrane filter
가 .
가
0.5% 가 steroid

가 . SF68
Lactobacillus MRS broth(DIFCO Co.) Yeast Tryptic
 soy broth(DIFCO Co.) .

5)

SF68 *Lactobacillus* MRS broth Bacto-agar 1.5%
 가 1 *Lactobacillus* MRS broth
 20ml 35 ± 1 24 Yeast
 Potato dextrose agar(DIFCO Co.) 1
 Tryptic soy broth(DIFCO Co.) 30 24 100rpm

6)

SF68 Yeast 0.1ml 가 200ml 500ml
 SF68 35 ± 1 Yeast 30 ± 1

7)

SF68 Yeast Spectrophotometer(UVIKON 942, Italy)
 520nm 660nm (Optical
 density)

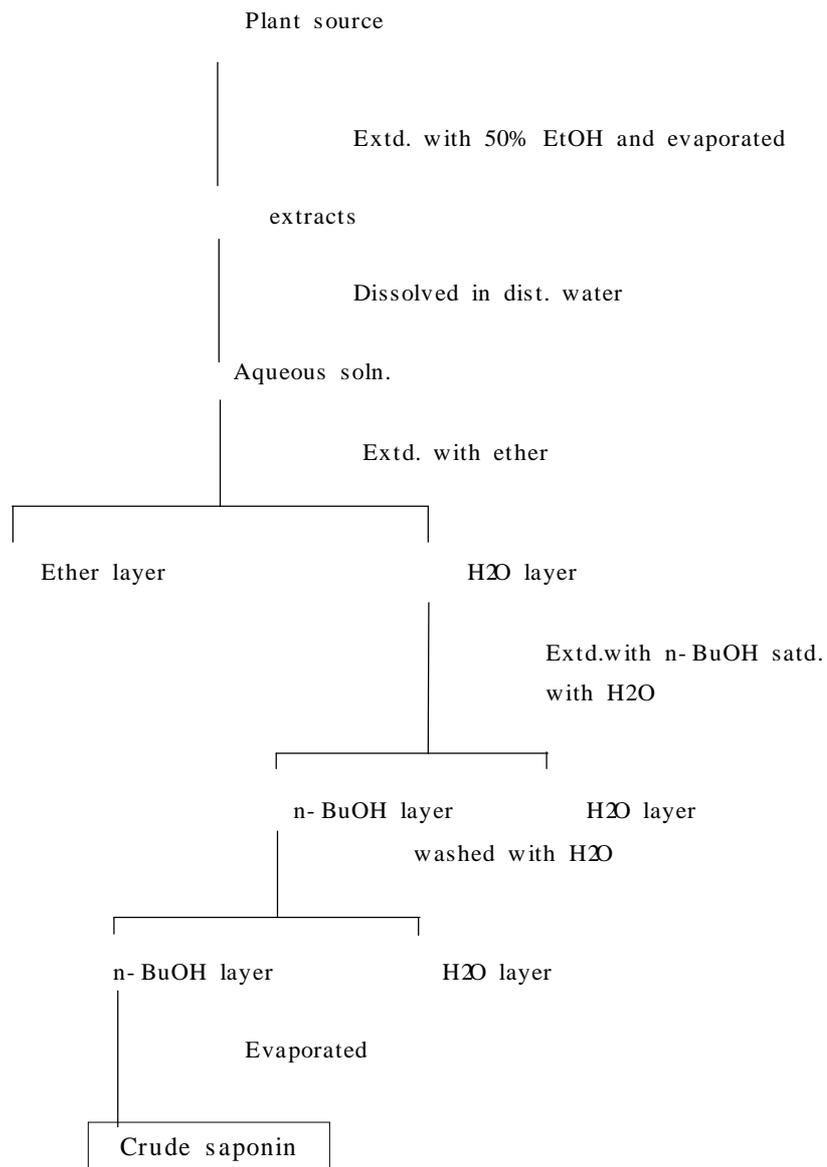


Fig. 2-1. Extraction procedure for the saponin fraction of several plant sources

. Yucca extract propolis 가가 E. coli

1) Propolis

propolis propolis
 (97%) 300ml 30g 2
 magnetic stirrer Whatman filter No. 4
 evaporator ethanol
 propolis

2)

1 SF68 E. coli

3)

가
 Propolis ethanol extract DMSO(Dimethyl sulfoxide)
 SF68
 1 Nutrient broth(DIFCO
 Co.) E. coli EC broth(DIFCO Co.)
 1 2
 10ml propolis yucca extract 0.22μm
 membrane filter 가
 가 35±1 , 37±1 24

4)

Propolis yucca extract 가 24

serial dilution 10 SF68

E. coli Brain heart infusion agar MacConkey agar

24

colony colony 30 300 colony

가 petri dish .

. Propolis 가

1)

Propolis 2 Red dog

cookie byproduct

casein, lactose Na-casein .

2)

Propolis 1:3(W/W) AOAC(1990)

oil

capillary tube 가 water bath 1 가

propolis가 .

3) propolis

mixture 60 2 propolis가

.

3.

가. steroid

saponin

Fig. 2-2 2-6

. YS 가

(Fig. 2-2) PGR 30mg

(Fig. 2-3). CLPW 0.1mM

가

YS

(Fig. 2-4). EEGB(Fig. 2-5)

WEGB(Fig. 2-6)

12mg

가

50%

saponin 가

CLPW

PGR, EEGB

WEGB

saponin

42.1mg/g

22.9mg/g

13.1mg/g

(1987)

saponin

steroid

saponin

가

가 yucca

saponin

saponin

Wallace et al. (1994)가

28.4mg

yucca saponin 가

saponin

saponin

가

. YS 가 SF68
 Yeast 1500ppm 가 SF68
 (Fig. 2-7). PGR SF68 Yeast
 (Fig. 2-8) CLPW WEGB 500ppm 가
 Yeast
 (Fig. 2-9 2-11). EEGB 가 saponin 가
 SF68 Yeast
 100ppm .
 YS Yeast
 saponin 가 Yeast
 가 YS
 가 . Nonaka(1986)
Trichoderma viride
 saponin 가
 saponin
 Shimoyamada et al.(1996) *Asparagus officinalis* 가
 saponin 가 *Candida albicans*
 saponin fraction 가
 . (1975) *Saccharomyces*
cerevisiae
 saponin Yeast 가
 .
 (1978) saponin
 (1980) saponin

saponin
가 (YS), (PGR), (CLPW) 가
(WEGB EEGB) , Price et al.(1987)
saponin

saponin
Yeast SF68 가

가

SF68 Yeast
가 . Fig.
2-12 가 0.5% SF68 Yeast
가 0.5% 가
1 saponin 가 SF68 Yeast

Yeast saponin 가
SF68 YS 가

가
(Fig. 2-13). PGR
100ppm 500ppm 가
(Fig. 2-14) CLPW 가 15
가 (Fig.
2-15). EEGB 33 (Fig. 2-16) WEGB 24
(Fig. 2-17)

saponin 가
가

Yeast 가 ,
 Nonaka(1986)가 alfalfa saponin *Trichoderma viride*
 , Shimoyamada et al.(1996) asparagus saponin *Candida*
albicans saponin Yeast
 가 .

. Yucca extract propolis 가가 E.
 coli

propolis 가 yucca extract
 가가 E. coli Fig.
 2-18 2-19 . Fig. 2-18 SF68

propolis 100ppm 가 가
 propolis yucca extract 가
 . Fig. 2-19 E. coli

propolis 250ppm
 500ppm 가 E. coli 90%
 propolis E. coli

가 . Mirzoeva et al.(1997)
Bacillus subtilis, *Escherichia coli* *R. sphaeroides*
 propolis propolis가
 가 , 가
 . , SF68
 E. coli

propolis 가
 . Aga et al. (1993) propolis 가 3
 가

propolis

propolis 가

가

Propolis 가 yucca extract 가 가

yucca extract 가 500ppm 1,000ppm 가

E. coli

SF68 yucca extract 가

100ppm 1,000ppm 가 가

propolis

가 가 propolis

가

가 yucca extract

Propolis 가

Propolis 가

가

Table 2-1.

propolis 49 54 60 가

propolis가

가

propolis가

cookie

byproduct 가

capillary tube

propolis가 cookie byproduct

capillary tube

가

propolis가

(90)

propolis

1:3

6

0 2

cookie byproduct mixture

propolis가

propolis

Na-casein

가

가

Lactose casein

(Table 2-2).

propolis

cookie byproduct

Na-casein propolis

가

Na-casein

가

propolis

가

Table 2-1. Capillary melting temperature() of carrier-based propolis

Propolis+carriers	Melting initiation temp. ()
Propolis	49 ± 2
Propolis + Lactose	over 90
Propolis + Casein	over 90
Propolis + Na-casein	over 90
Propolis + Red dog	over 90
Propolis + Cookie byproduct	70 ± 2

Table 2-2. Morphological scores of propolis-incorporated carriers after heat treatment

Carriers	Binding capability ^a
Lactose	***
Casein	***
Na-casein	*****
Red dog	**
Cookie byproduct	*

^a Increased number of asterisks represents the superiority of binding capability of carriers

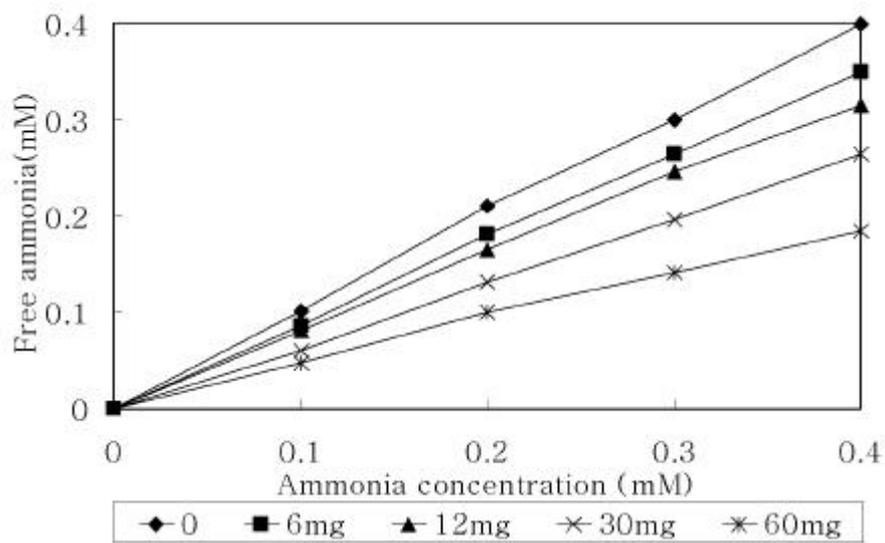


Fig. 2-2. Ammonia binding capacity of saponin fraction of YS

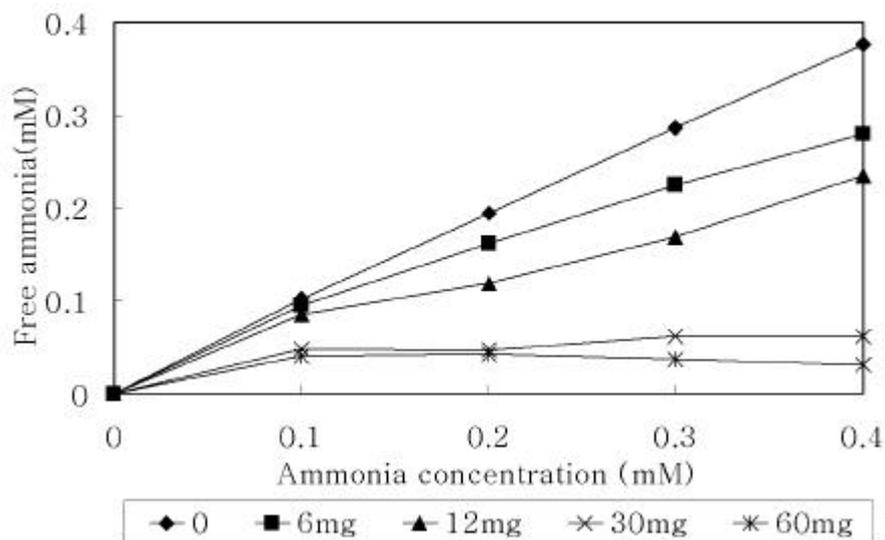


Fig. 2-3. Ammonia binding capacity of saponin fraction of PGR

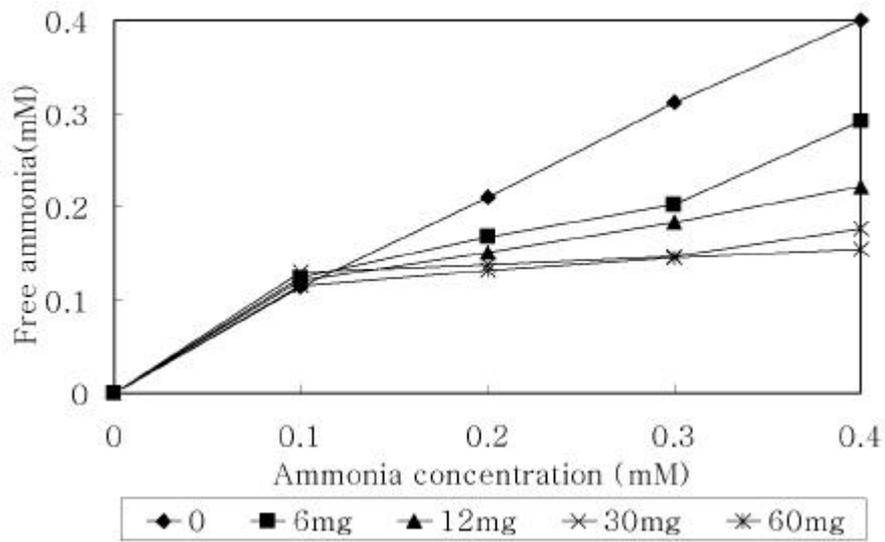


Fig. 2-4. Ammonia binding capacity of saponin fraction of CLPW

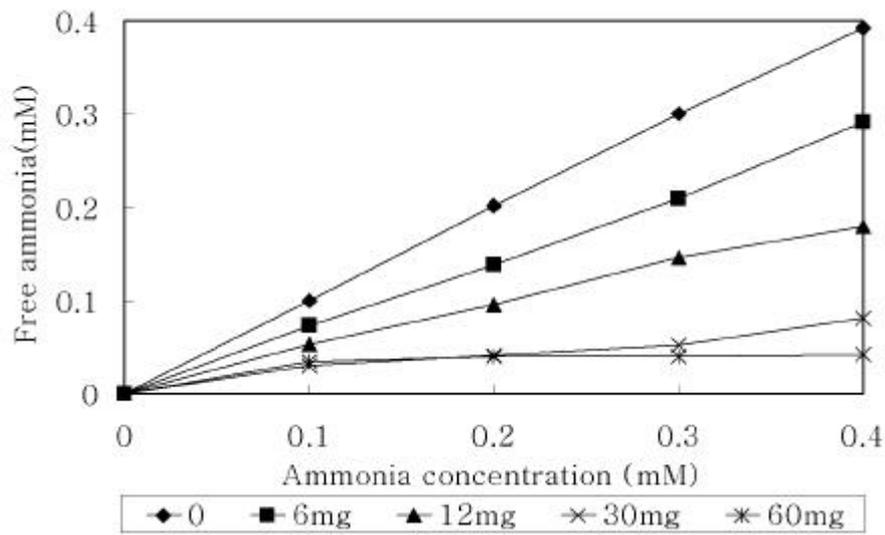


Fig. 2-5. Ammonia binding capacity of saponin fraction of FFCB

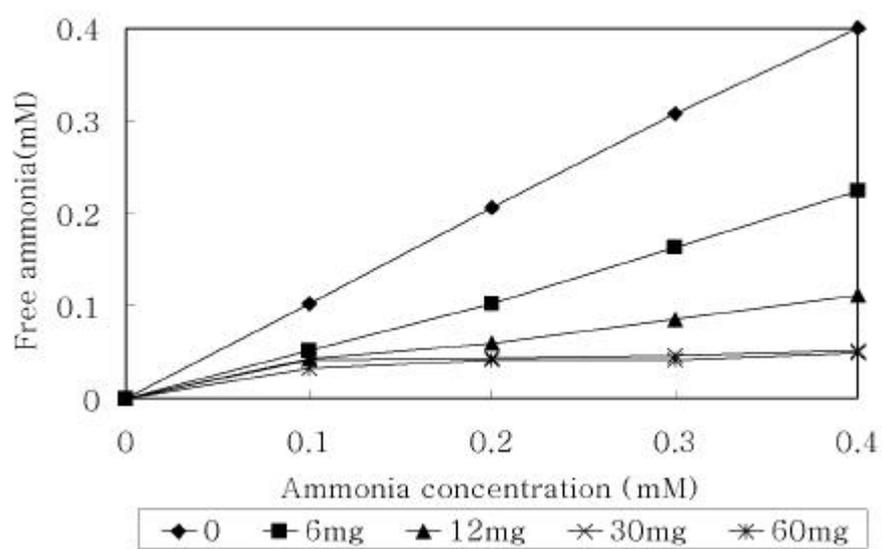


Fig. 2-6. Ammonia binding capacity of saponin fraction of VECB

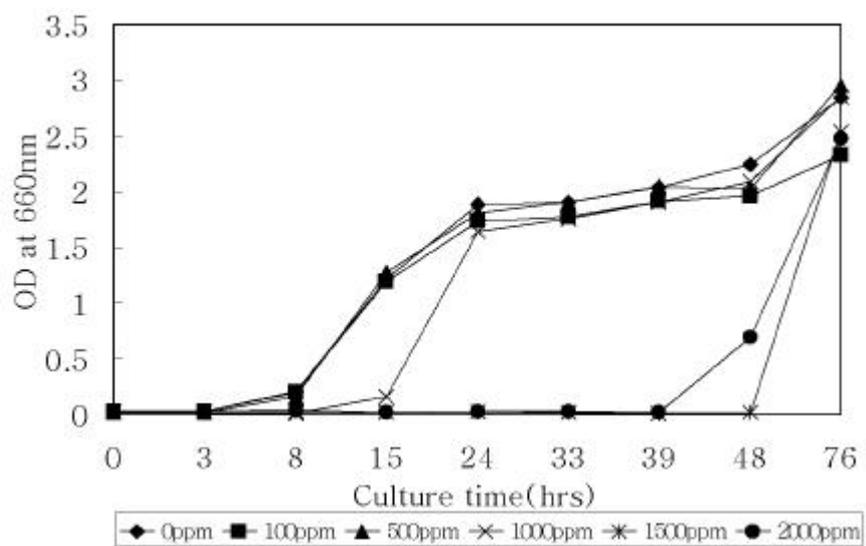
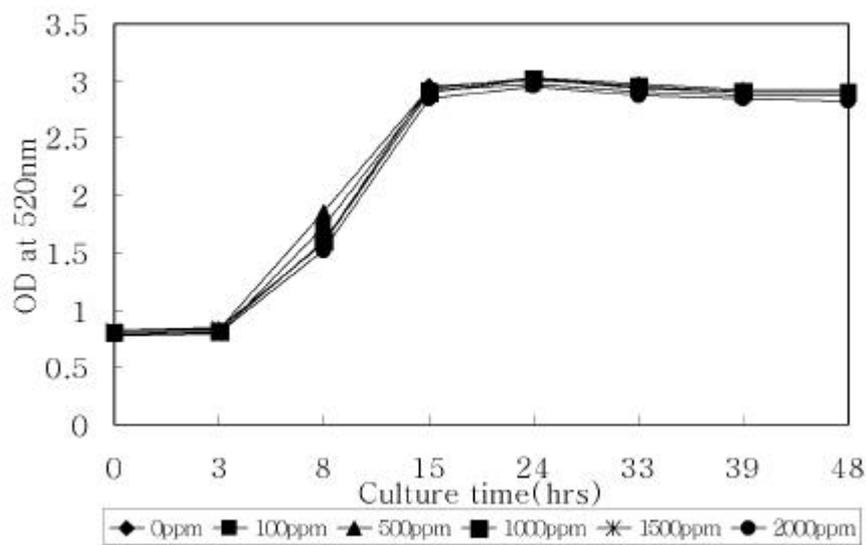


Fig. 2-7. Effect of saponin fraction of YS on the growth of SF68 (above) and Yeast (below)

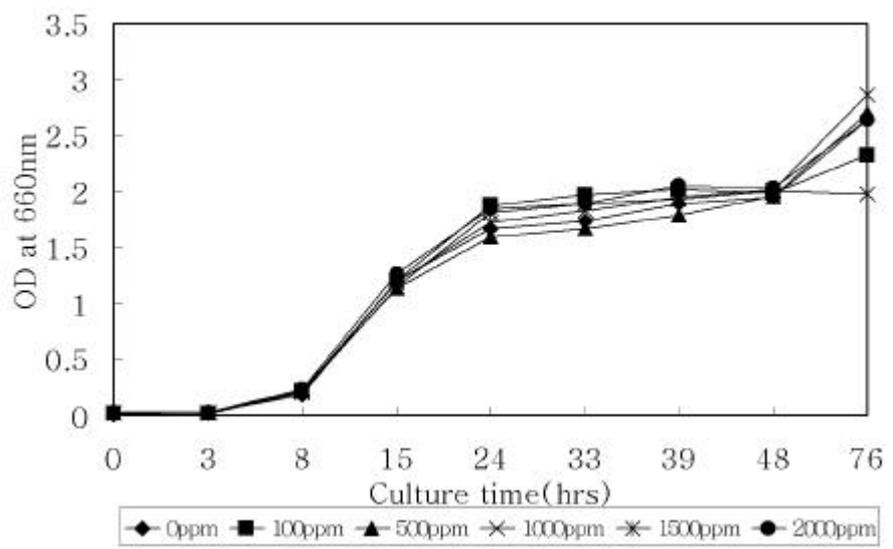
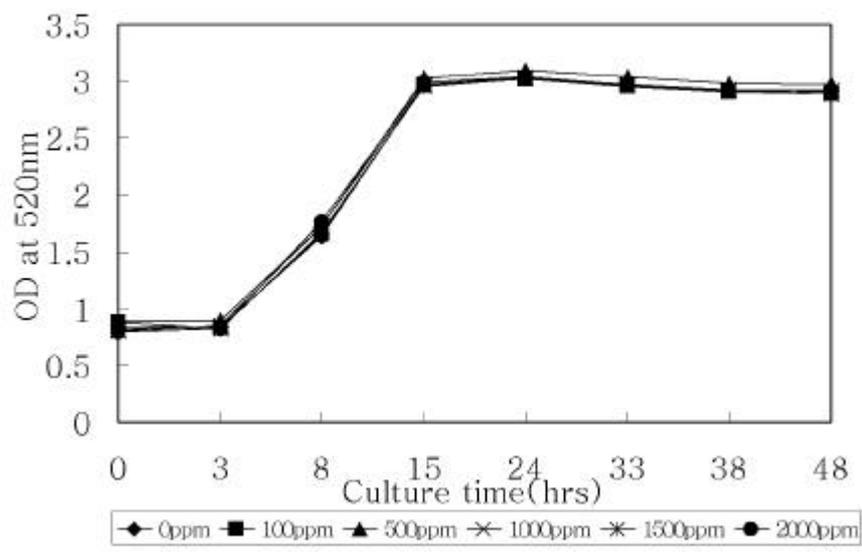


Fig. 2-8. Effect of saponin fraction of PGR on the growth of SF68 (above) and Yeast(below)

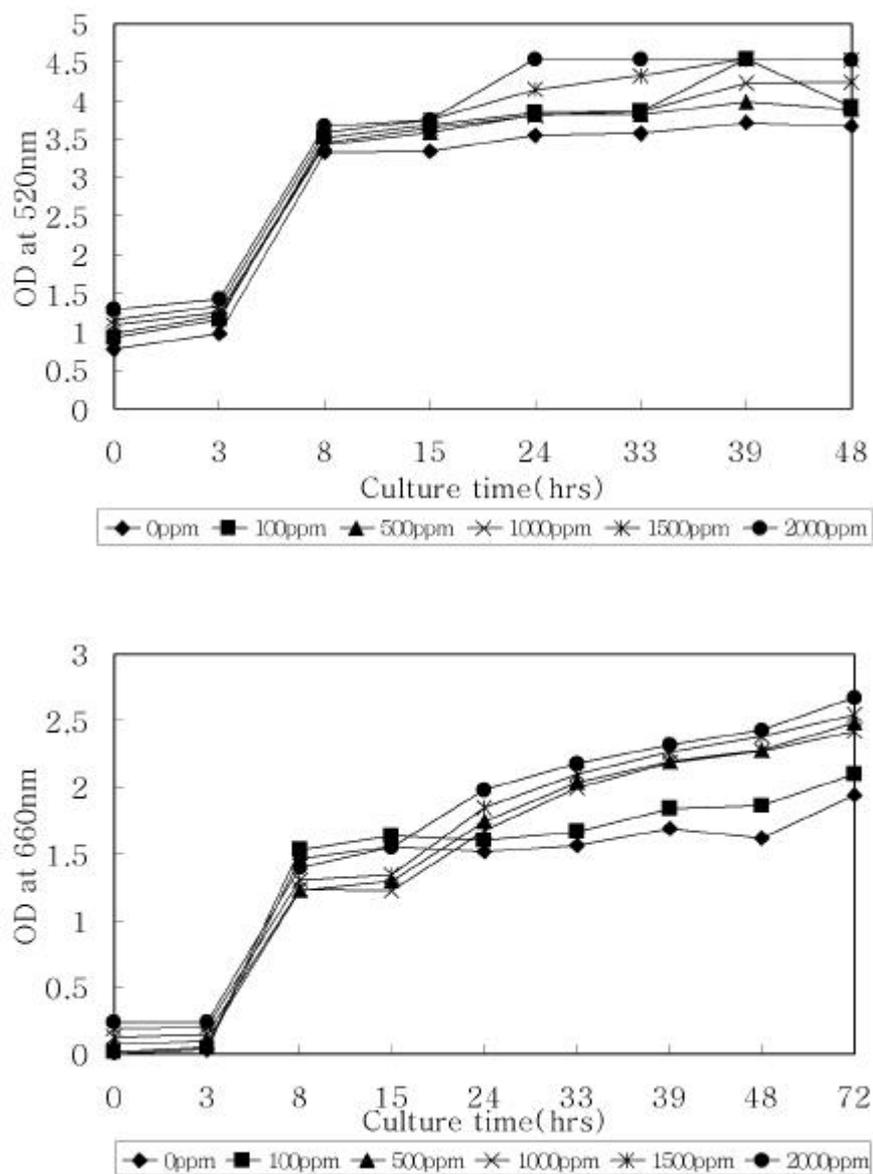


Fig. 2-9. Effect of saponin fraction of CLPW on the growth of SF68 (above) and Yeast(below)

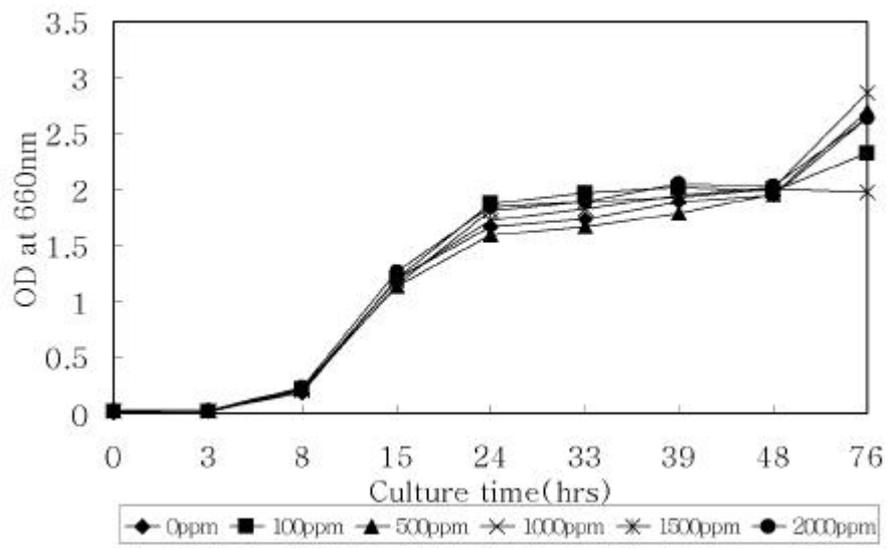
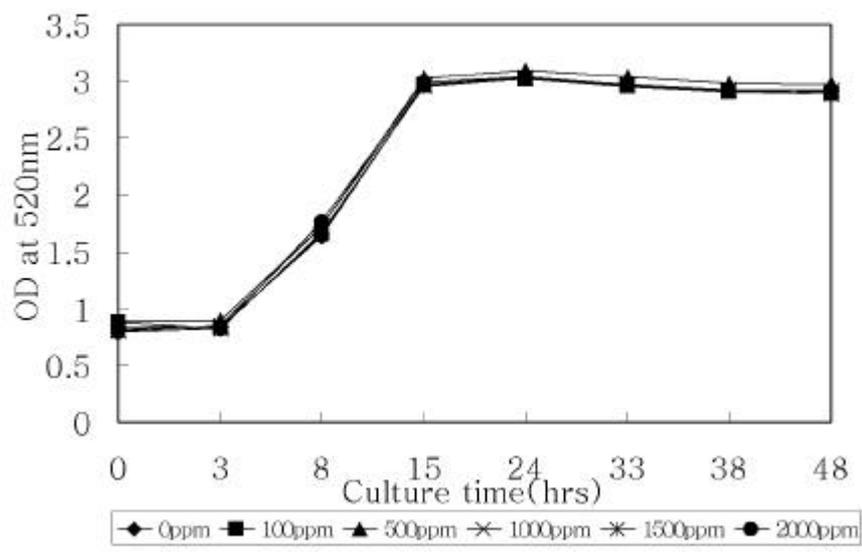


Fig. 2-10. Effect of saponin fraction of EEGB on the growth of SF68 (above) and Yeast(below)

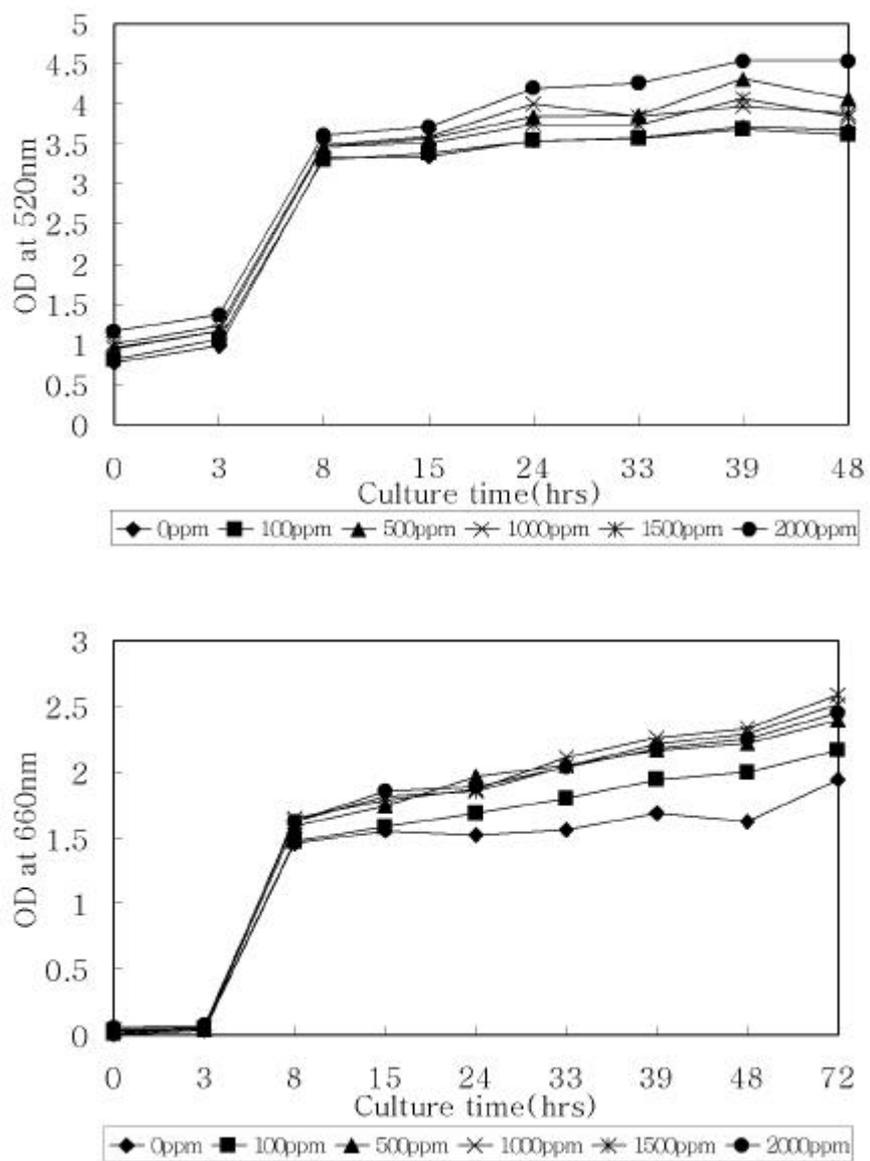


Fig. 2-11. Effect of saponin fraction of WEGB on the growth of SF68 (above) and Yeast(below)

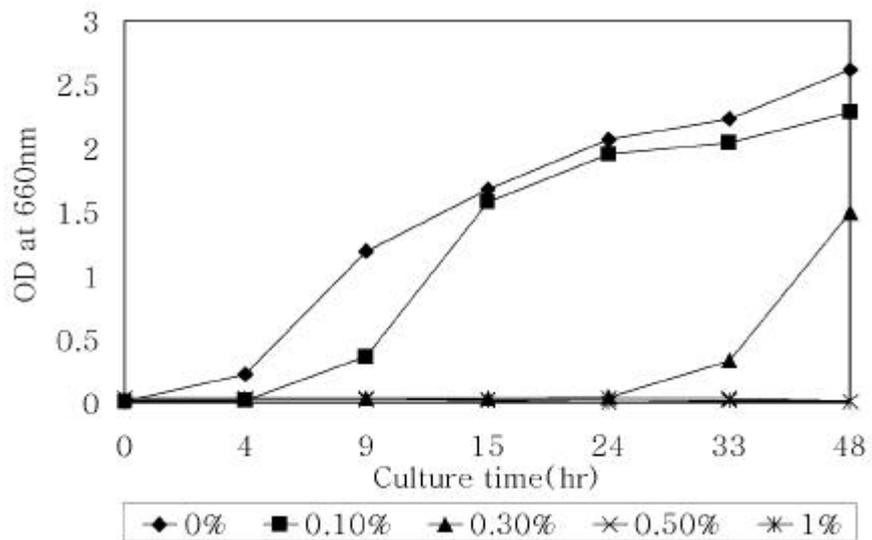
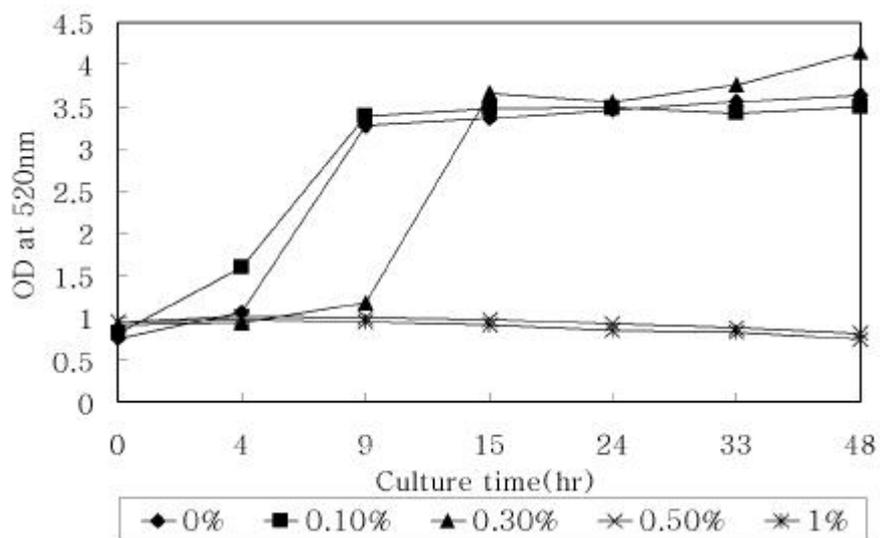


Fig. 2-12. Growth pattern of SF68(above) and Yeast(below) cultured in media with the various ammonia concentrations

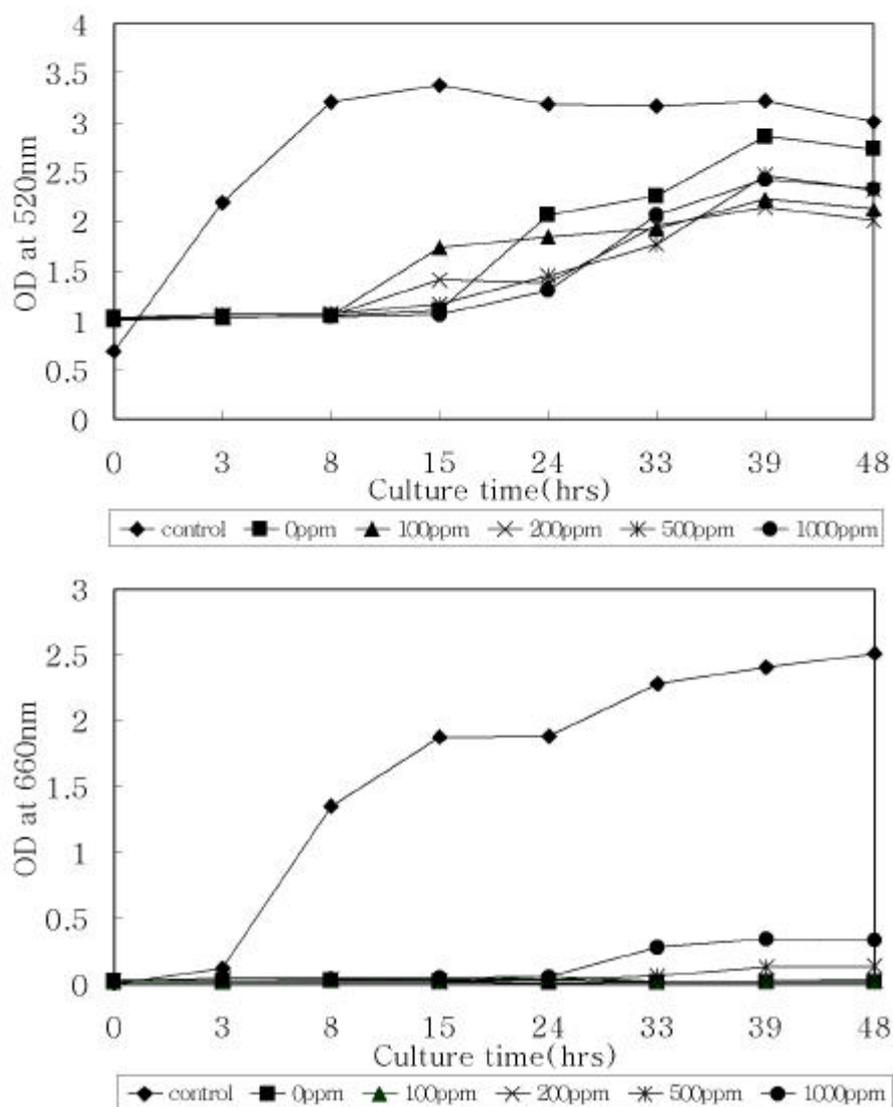


Fig. 2-13. Effect of saponin fraction of YS on the growth of SF68 (above) and Yeast (below) in media with ammonia water (0.5% V/V)

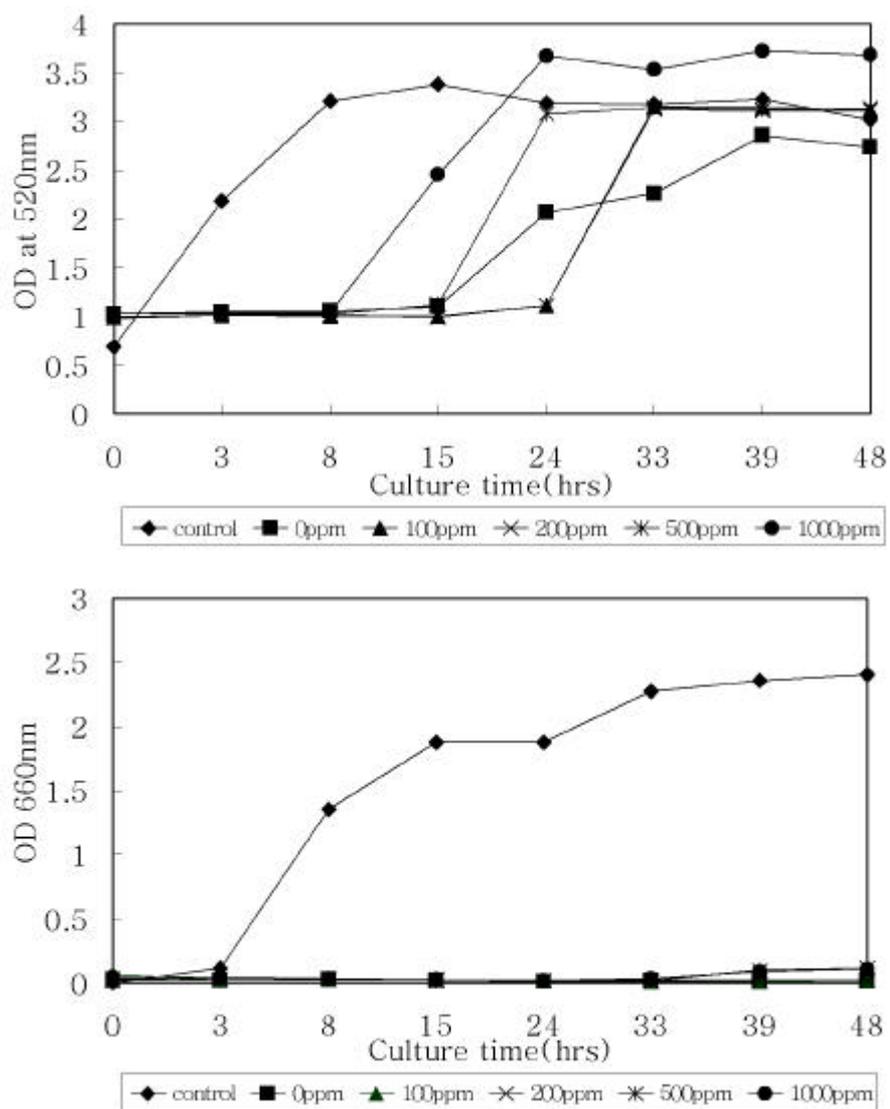


Fig. 2-14. Effect of saponin fraction of PCR on the growth of SF68 (above) and Yeast (below) in media with ammonia water (0.5% V/V)

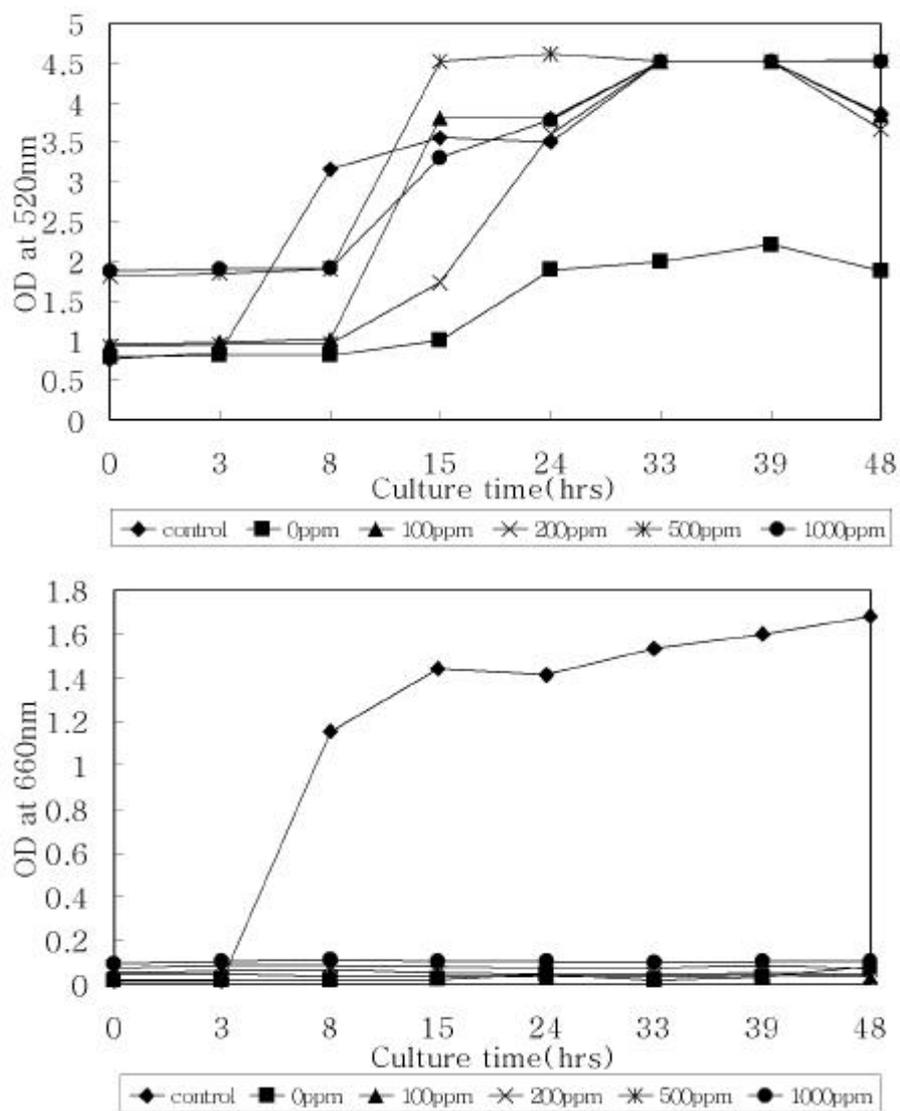


Fig. 2-15. Effect of saponin fraction of CLPW on the growth of SF68 (above) and Yeast (below) in media with ammonia water (0.5% V/V)

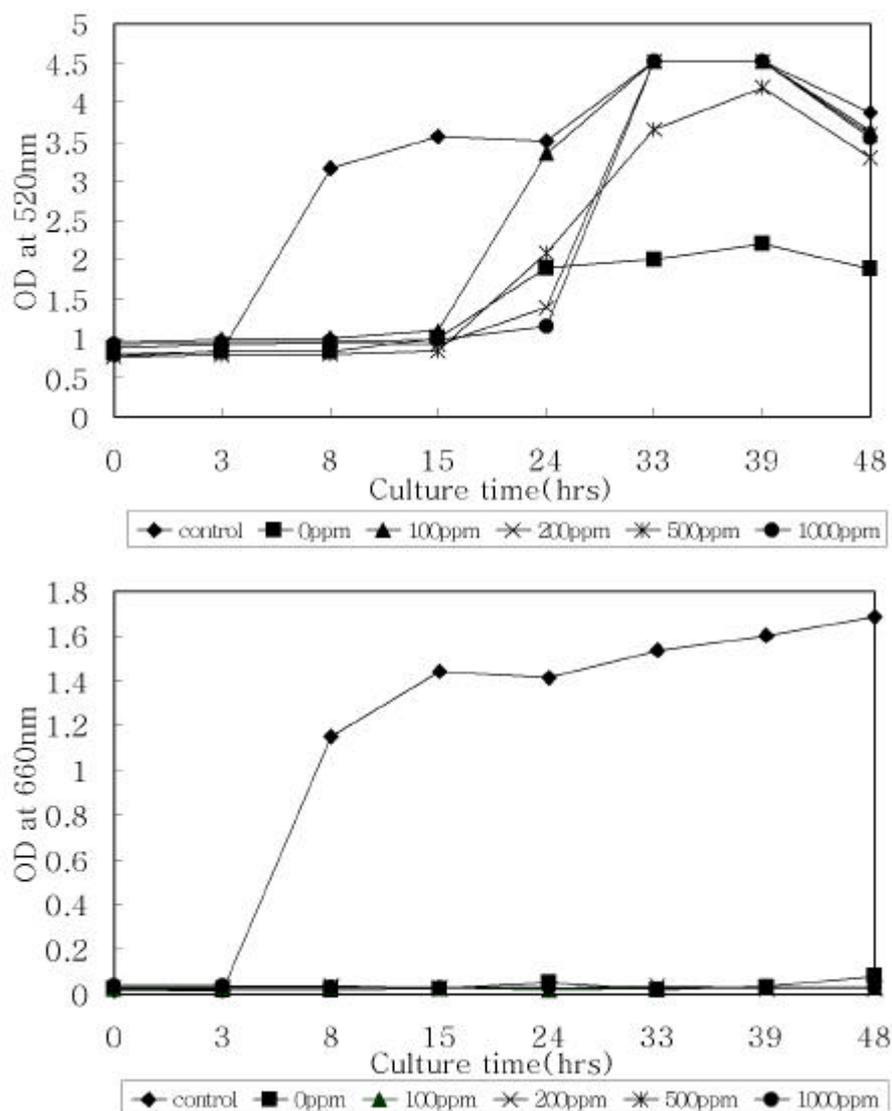


Fig. 2-16. Effect of saponin fraction of EEGB on the growth of SF68 (above) and Yeast (below) in media with ammonia water (0.5% V/V)

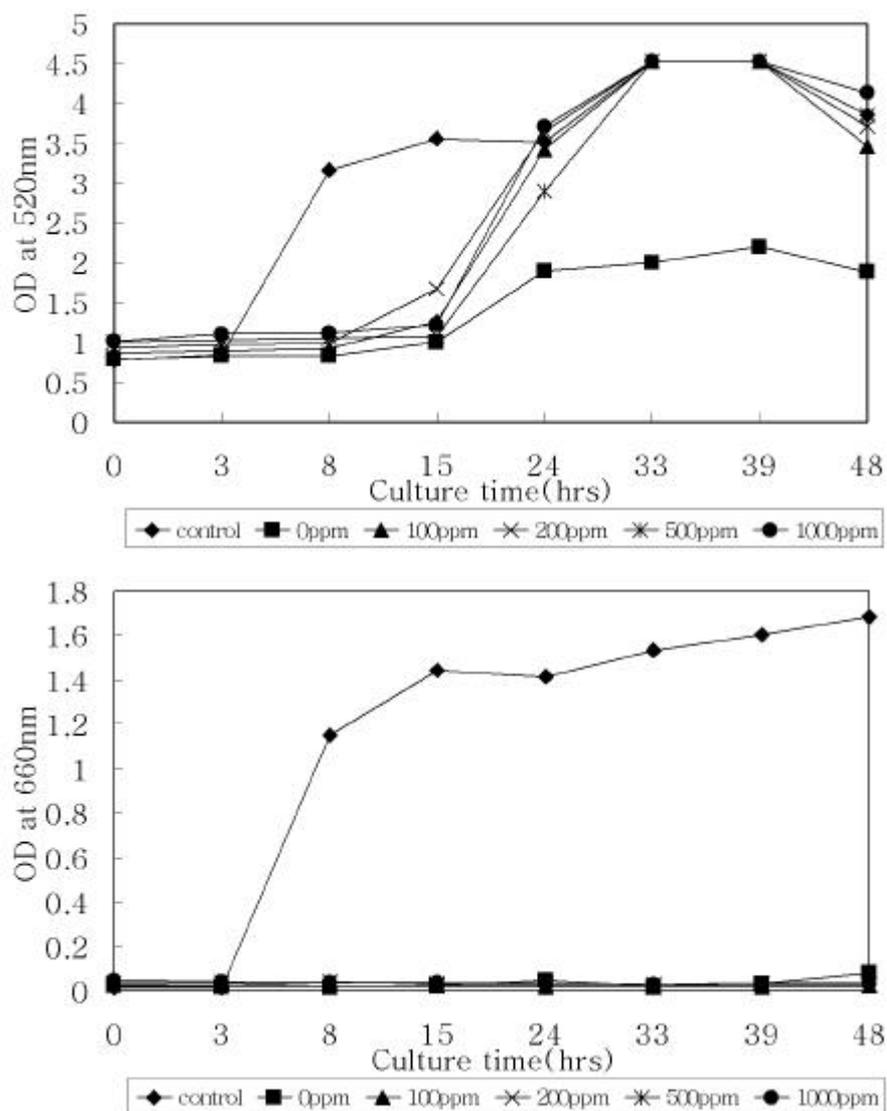


Fig. 2-17. Effect of saponin fraction of WEGB on the growth of SF68 (above) and Yeast (below) in media with ammonia water (0.5% V/V)

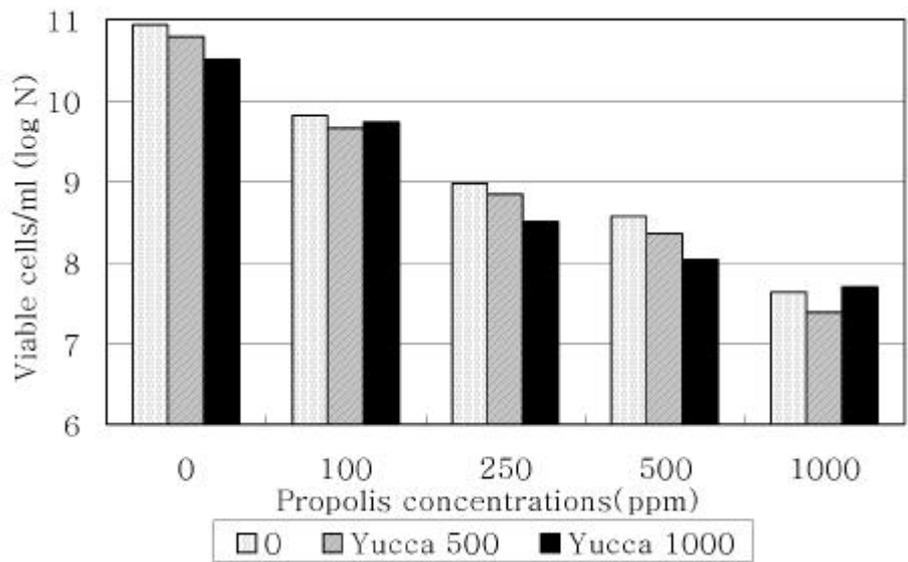


Fig. 2-18. Effect of both/either propolis and/or yucca extract on the viability of SF68

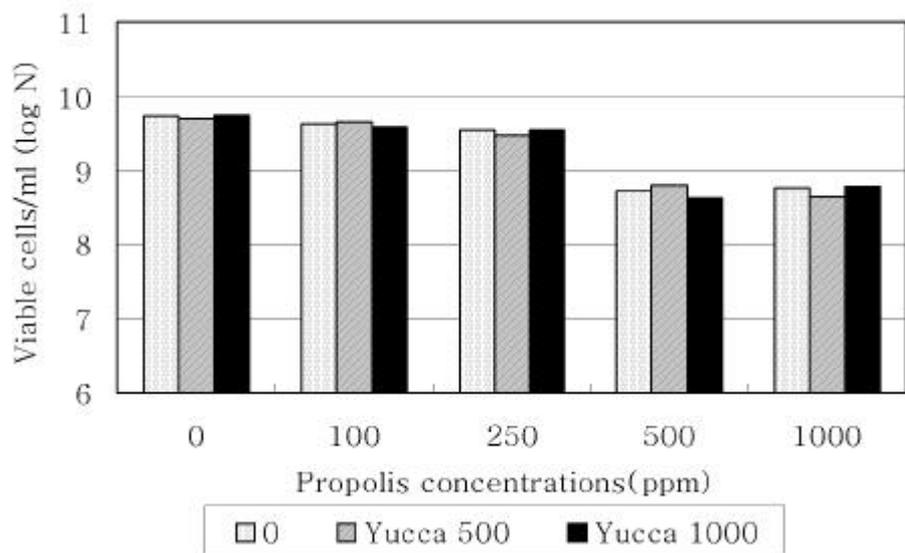


Fig. 2-19. Effect of both/either propolis and/or yucca extract on the viability of E. coli

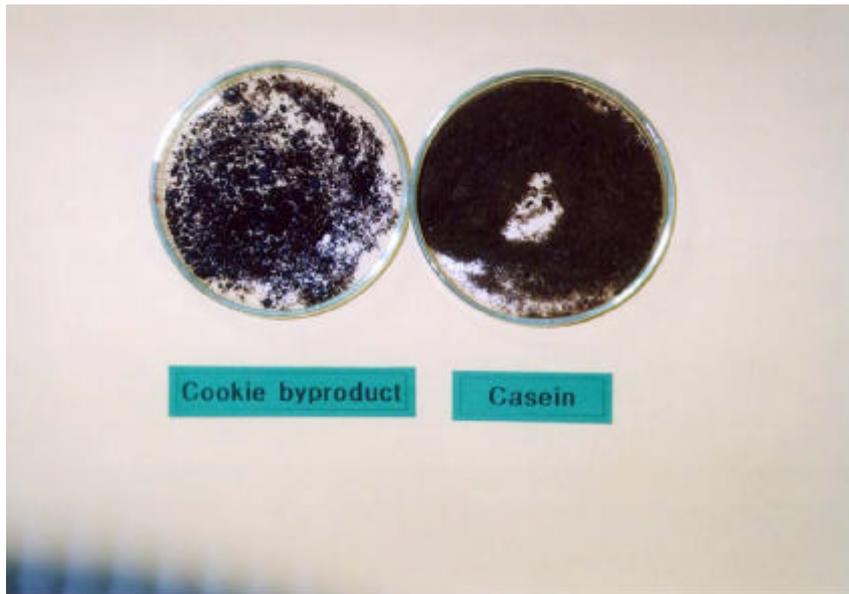


Fig. 2-20. Photograph of morphology of casein compared with cookie byproduct after heat treatment



Fig. 2-21. Photograph of morphology of lactose compared with cookie byproduct after heat treatment

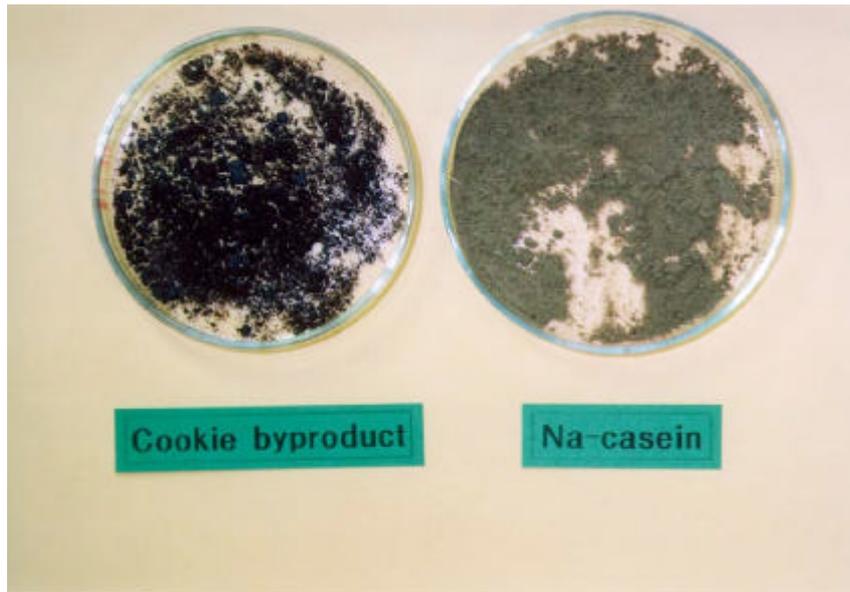


Fig. 2-22. Photograph of morphology of Na-casein compared with cookie byproduct after heat treatment

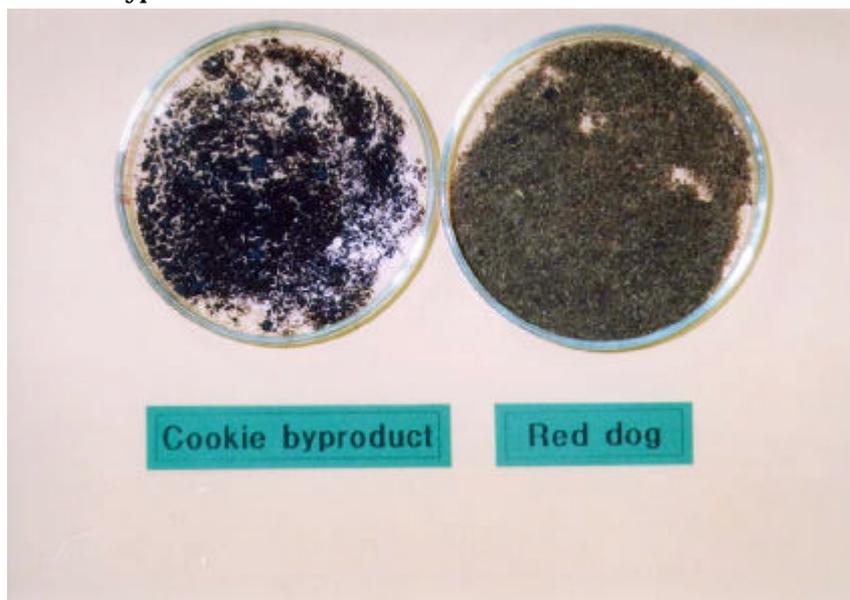


Fig. 2-23. Photograph of morphology of red dog compared with cookie byproduct after heat treatment

4.

가 가
가 3가 . 1
가
Yucca (YS), (PGR),
(CLPW), 가 (EEGB) (EEWB)
saponin .
saponin 가 *Streptococcus faecium*
cernelle 68(SF68) *Saccharomyces cerevisiae*(Yeast)
가 .

2 가
propolis yucca extract
SF68 E. coli
가 가 3 propolis
Red dog Cookie byproduct) (Lactose, Casein, Na-casein,
가 .

1. PGR 30ng 가
CLPW 가 0.1nM
YS 가
. EEGB WEGB 12ng
50% . PGR, CLPW, EEGB
WEGB YS

2. saponin 가 SF68
 Yeast YS 1500ppm 가
 CLPW 500ppm Yeast
 PGL EEGB Yeast . WEGB 100ppm
3. 가 0.5% SF68 Yeast
4. 가
 steroid 가 Yeast SF68
 YS 가 PGR 500ppm 가
 . CLPW, EEGB WEGB 100ppm 가 15, 33, 24
 가 SF68
5. SF68 propolis 100ppm 가
 E. coli 500ppm propolis 가
 가 . Yucca extract 가 SF68 E. coli
6. 가 propolis cookie byproduct
 가 70 가 90 .
 Na-casein propolis 가
 .
 saponin
 가 yucca

가 가 가 . steroid saponin
propolis
SF68 steroid
propolis 가
가 .

5.

AOAC. 1990. Official Methods of Analysis (15th Ed.). Association of Official Analytical Chemists, Washington, D. C.

Aga, H., Shibuya, T., Sugimoto, T., Kurimoto, M. and Nakajima, S. 1994. Isolation and identification of antimicrobial compounds in brazilian propolis. Biosci. Biotech. Biochem. 58 : 945-946

Brunfitt, W., Hamilton-niller, J.M. and Franklin, I. 1990. Antibiotic activity of natural products : 1. Propolis. Microbios. 62 : 19-22

Cheeke, P.R. 1971. Nutritional and physiological implications of saponins : a review. Can. J. Anim. Sci. 51 : 621-632

Cole, D.J.A. 1993. Controlling the impact of nitrogen waste products on animal health, performance and the environment. In : Biotechnology in the Feed Industry (T.P. Lyons, ed), Alltech Technical Publications. 9 : 293-305

Cole, D. J. A., Tuck, K. 1995. Using yucca to improve pig performance

while reducing ammonia. In : *Biotechnology in the Feed Industry* (T.P.Lyons, eds.). Alltech Technical Publications. 11 : 421-426

Ellenberger, M. A., Rumpler, W. V., Johnson, D. E. and Goodall, S. R. 1985. Evaluation of the extent of ruminant urease inhibition by sarsaponin and sarsaponin fractions. *J. Anim. Sci.* 61(suppl. 1) : 491

Ender, K., Kuhn, G., Nunberg, K. and Jacques, K. A. 1996. Effect of *Yucca schidigera* extract on performance, carcass evaluation and skatole content of fattening boars. *J. Anim. Sci.* 74(suppl. 1) :

Ferwick, D. E. and Oakenfull, D. 1983. Saponin content of food plants and some prepared foods. *J. Sci. Food Agric.* 34 : 186-191

Grange, J. M. and Davey, R. W. 1990. Antibacterial properties of propolis (bee glue). *J. R. Soc. Med.* 83 : 159-160

Headon, D. R., Buggle, K., Nelson, A. and Killeen, G. 1991. Glycofractions of the *Yucca schidigera* extract. In : *Biotechnology in the Feed Industry* (T. P. Lyons, ed), Alltech Technical Publications. 7 : 95-108

Hussain, I., Isnail, A. M. and Cheeke, P. R. 1996. Effects of feeding *Yucca schidigera* extract in diets varying in crude protein and urea contents on growth performance and cecum and blood urea and ammonia concentrations of rabbits. *Anim. Feed Sci. Tech.* 62 : 121-129

Johnston, N. L., Quarles, C. L., Fagerberg, D. J. and Caveny, D. D. 1981.

Evaluation Yucca saponin on broiler performance and ammonia suppression. Poul. Sci. 60 : 2289-2292

Johnston, N. L., Quarles, C. L. and Fagerberg, D. J. 1982. Broiler performance with DSS40 yucca saponin in combination nonensin. Poul. Sci. 61 : 1052-1054

Joo, C. N., Cho, Y. D. and Kwon, H. Y. 1978. Biochemical studies on ginseng saponins() : The effect of ginseng saponin on bacterial growth. Kor. J. Bichen. 11 : 113-125

Killeen, G. K., Buggle, K. A., Hynes, M. J., Walsh, G. A., Power, R. F. and Headon, D. R. 1994. Influence of *Yucca schottigera* preparations on the activity of urease from *Bacillus pasteurii*. J. Sci. Food Agric. 65 : 433-440

Krol, W., Scheller, S., Shani, J., Pietsz, G. and Czuba, Z. 1993. Synergistic effect of ethanolic extract of propolis and antibiotics on the growth of *Staphylococcus aureus*. Drug Res. 43 : 607-609

Ma, M. D., Vung, L. C., Huang, Y. T. and Fu, C. M. 1993. Effect of adding deodorizers to diets on the performance of pigs and the deodorization of pig wastes . Comparison between sarsaponin and zeolite powder. Chin. J. Anin. Sci. 22 : 229-235

Metzner, J., Bekeneier, H., Paintz, M. and Schneidewind, E. 1979. On the antimicrobial activity of propolis and propolis constituents.

Pharmazie. 34 : 97-102

Milgate, J. and Roberts, D. C. K. 1995. The nutritional and biological significance of saponins. Nutri. Res. 15 : 1223-1249

Mirzoeva, O. K., Grishanin, R. N. and Calder, P. C. 1997. Antimicrobial action of propolis and some of its components : the effects on growth, membrane potential and motility of bacteria. Microbiol. Res. 152 : 239-246

Nevel, C. J. V. and Denehy, D. I. 1990. Effect of antibiotics, a deaminase inhibitor and sarsaponin on nitrogen metabolism of rumen contents in vitro. Anim. Feed Sci. Tech. 31 : 323-348

Noraka, M. 1986. Variable sensitivity of *Trichoderma viride* to *Mecicago sativa* saponins. Phytochem. 25 : 73-75

Park, Y. K., Koc, M. H., Abreu, J. A., Ikegaki, M., Cury, J. A. and Rosalen, P. I. 1998. Antimicrobial activity of propolis on oral microorganisms. Curr. Microbiol. 36 : 24-28

Pedersen, M. W., Zimmer, D. E., McAllister, D. R. and Anderson, J. O. 1967. Comparative studies of saponin of several alfalfa varieties using chemical and biochemical assays. Crop Sci. 7 : 349-352

Pepeljnjak, S., Jalsenjak, I. and Maysinger, D. 1982. Growth inhibition of bacillus subtilis and composition of various propolis extracts.

Pharmazie. 37 : 864-865

Preston, R. I. Bartle, S. J. , May, T. and Goodall, S. R. 1987. Influence of sarsaponin on growth, feed and nitrogen utilization in growing male rats fed diets with added urea or protein. J. Anim. Sci. 65 : 481-487

Price, K. R. , Johnson, I. T. and Fenwick, G. R. 1987. The chemistry and biological significance of saponins in foods and feedingstuffs. Crit. Rev. Food Sci. Nutr. 26 : 27-135

Price, K. R. , Curl, C. L. and Fenwick, G. R. 1986. The saponin content and saponin composition of the seed of 13 varieties of legume. J. Sci. Food Agric. 37 : 1185-1191

Shinoyanada, N. , Suzuki, M. and Maruyama, N. 1996. An antifungal saponin from white asparagus (*Asparagus officinalis* L) bottoms. J. Sci. Food Agric. 72 : 430-434

Sucharita, S. , Harinder, P. S. , Makkar and Klaus, B. 1998. Alfalfa saponin and their implications in animal nutrition. J. Agric. Food Chem. 46 : 131-140

Takasi-kukuni, N. B. and Shilcher, H. 1994. Electron microscopic and microcalorimetric investigations of the possible mechanism of the antibacterial action of a defined propolis provenance. Planta Med. 60 : 222-227

Wallace, R. J., Arthaud, L. and Newbold, C. J. 1994. Influence of *Yucca schidigera* extract on ruminal ammonia concentrations and ruminal microorganisms. *Appl. Environ. Microbiol.* 60 : 1762-1767

Weatherburn, M. V. 1967. Phenol-hypochlorite reaction for determination of ammonia. *Anal. Chem.* 39 : 971-974

Wu, Z., Sadik, M., Sleiman, F. T., Sinas, J. M., Pessaraki, M. and Huber, J. T. 1994. Influence of yucca extract on ruminal metabolism in cows. *J Anim. Sci.* 72 : 1038-1042

Yeo, J. M. and Kin, K. I. 1997. Effect of feeding diets containing an antibiotic, a probiotic, or yucca extract on growth and intestinal urease activity in broiler chicks. *Poult. Sci.* 76 : 381-385

. 1995. , . 가 .
22 : 105-115

. 1980.
(1). . 4 : 121-132

. 1992.

. 1979. extracts가
. 3 : 113-126

. 1987. , .

, . 1979. saponin
. 3 : 144-155

. 1975. 가 .
. 1 : 49

1.

가 가 .
가

,
, 가 .
가 가 가 가 .
가 가

. 가
(, 1996)
,

.
, 가 가
. 20 가

가 .
가
(odor threshold)

level) (Dravenieks, 1972).

가 .

of factory analysis system

.

GC GC Mass .

가 가

up stream

down stream .

,

yucca extract

가 steroids (Panax

ginseng) (*Platycodeon grandiflorum*)

가 down stream 가 .

가가

가 .

2.

가.

가

GC Mass

가

H₂O

50% EtOH ,

steroid saponin

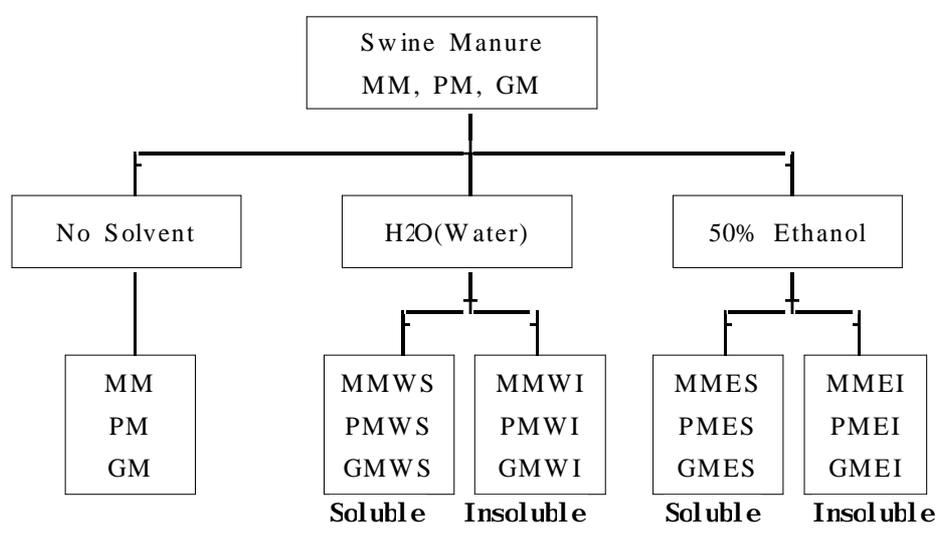
(*Platyodon grandiflorum's* root) (*Farax ginseng*
byproduct) 1% 가 down stream
가

Table 2-3. Experimental design

Down stream Odor control additives	Manure fractions			
	H ₂ O		50% EtOH	
	soluble	insoluble	soluble	insoluble
None PGR 1% GB 1%	duration days in room temp. 1, 3, 7, 14 Odor analyses : olfactory analysis GC, GC Mass analysis			

(GB) 1% 가 (PGR) 1%,
 1, 3, 7, 14 H₂O 50% ethanol
 72 μ m sieve
 MM, 가 PM, 가
 GM, H₂O -VS, -VI,
 50% EtOH -ES, -EI

Fig. 2-24 .



Legends :
 M: manure P: Platycodon grandiflorum G: Ginseng S: Soluble
 I: Insoluble W: Water E: 50% ethanol

Fig. 2-24. Separation diagram of swine manure

. Saponin

saponin fig. 2-24
 1kg 50% EtOH
 extracts ether, n-butanol
 n-butanol crude saponin (, 1992).
 Fig. 2-24 saponin HPLC
 (Waters 486) , column μ boundpak C18 3.9 x 150mm
 . saponin 30% nthanol 100% MeOH
 , column 30 flow rate 1.0ml/min .

(Olfactory analysis)

(soluble) (insoluble) H₂O 50% ethanol
 (olfactory analysis) . 가
 가
 Fig. 2-25 Table
 2-4 .
 sample tube
 , 20 20
 .

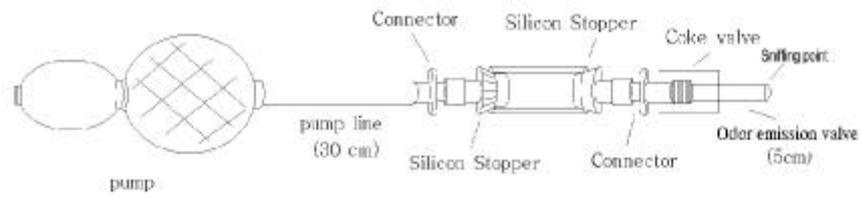


Fig. 2-25. Simplified diagram of olfactory detector

Table 2-4. Physical specification of olfactory detector

Parts	Specification
Air tube volume(ml)	470
Sample holder	
diameter(mm)	6
length(cm)	5
Sniffing point	
diameter(mm)	4
Air flow rate(ml/min) at sniffing	300 200

VOC(Volatile organic compounds)

VOC

. VOC

fig. 2-26 100nm, 4.7mm, 6.45mm

0.5g Tenax-GR(60/80mesh) frit

. Column stainless steel capillary

column(UA5, Frontier Lab 30m x 0.25mm, 0.3µm)

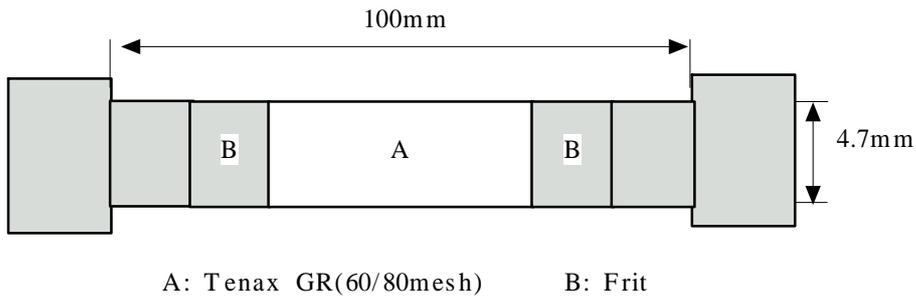
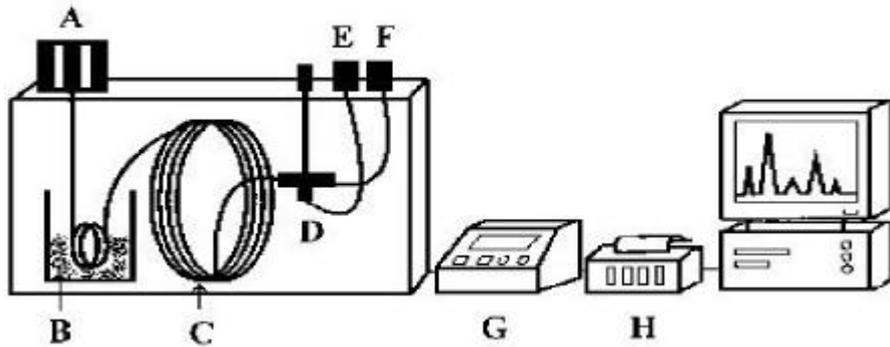


Fig. 2-26. Schematic diagram of 100mm adsorption tube

VOC fig. 2-27
 200 가 UA5
 column -196 8
 outlet splitter 가 1:3 dual
 detector system flame ionization detector(FID)
 , flame photometric detector(FPD)



A: Thermal desorption B: Liquid N2 C: Stainless column D: Outlet splitter
 E: FPD F: FID G: Electronic pressure control H: Integrator

Fig. 2-27. Schematic diagram of on-column cryofocusing gas chromatography system
 . GC Mass

UA-1HT(Frontier Lab.) fig. 2-27 200 가
 0.5 mm 가 20 cm
 on-column
 3cm 가
 20 ml/min , 5 가
 1.5 Ml/min fused silica
 HP-5 (0.32 mm i.d., 30m long, 0.25 μm film thickness)
 EI mode(70 eV) GC Mass(JMS-AM 150, Japan)
 GC Mass table 2-5

Table 2-5. Operating conditions of the on-column cryofocusing GC Mass

Purge & cryofocusing

Purge 200 , (He, 20 Ml/min, 5 min)
 Coolant Liquid- N2 (- 196)
 Concentration loop UA- 1HT (0.5 mm i.d., 20 cm long)

GC_Mass

Gas Chromatography HP- 5890 series
 Column HP- 5(0.32 mm i.d., 30 m long, 0.25 μm film thickness)
 Carrier gas He, 1.5 Ml/min
 Temperature 30 (5min)-3 /min-60 (5min)-5 /min-200 (10min)
 Mass spectrometer Quadrupole(JMS- AM 150, JEOL)
 Ionization EI(70 eV)
 Scan m/z 35 to m/z 400, 400 ms/scan

3.

가. (Olfactory analysis)

20 panel

Table 2-6

Table 2-6. Odor intensity of both water soluble and insoluble fraction of swine manure

Dilution w/H ₂ O	H ₂ O solubility	Odor intensity*				Intensity index**	
		(1)	(2)	(3)	(4)	class	composite
1 : 1	Soluble	2	10	5		54	65
	InSoluble		1	2		11	
1 : 10	Soluble		3	11	6	83	83
	InSoluble					-	
1 : 30	Soluble	2	8	7	2	66	70
	InSoluble			1		4	
1 : 50	Soluble	3	9	5	1	58	64
	InSoluble		2			6	
1 : 100	Soluble	7	6	2		25	33
	InSoluble	2	3			8	

* : Very faintly offensive odor; : Faintly offensive odor; : Definitely offensive odor; : Strongly offensive odor; : Very strongly offensive odor

**Intensity index : No. of detection multiplied by scores in parenthesis × (1), (2), (3), (4), (5)

water

soluble portion .

water soluble portion × 10 , × 30 , × 50 가

100 가 .

water soluble portion

가 가 가 .

100 가

가 .

가 10

10

1 10

.

3 , 7 14 5 10

H₂O

table 2-24 2-25 .

Table 2-7. Odor intensity of supernatant portion of swine manure

separated by centrifuge(27,000g, 4)

Storage	Dilution	Odor class	Odor intensity*					Intensity index**	
								class	composite
3 days	1 : 5	Soury	2	8				18	
		Fishy			1			3	40
		Beany		8	1			19	
	1 : 10	Soury							
		Fishy			10			30	61
		Beany			9	1		31	
7 days	1 : 5	Soury			5			15	
		Fishy			6	2		26	63
		Beany		1	4	2		22	
	1 : 10	Soury			1	8		35	
		Fishy		2	2	7		38	73
		Beany							
14 days	1 : 5	Soury	2	1	3			19	
		Fishy			5	4		31	67
		Beany			3	2		17	
	1 : 10	Soury		1	1	4	1	26	
		Fishy			2	7	2	44	76
		Beany			2			6	

* : Very faintly offensive odor; : Faintly offensive odor; : Definitely offensive odor; : Strongly offensive odor; : Very strongly offensive odor
 **Intensity index : No. of detection multiplied by scores in parenthesis × (1), (2), (3), (4), (5)

Table 2-8. Odor intensity of precipitate portion of swine manure separated by centrifuge(27,000g, 4)

Storage	Dilution	Odor class	Odor intensity*			Intensity index**	
						class	composite
3 days	1 : 5	Soury				-	
		Fishy	2	5	1	15	35
		Beany	4	8		20	
	1 : 10	Soury			1	3	
		Fishy		6	2	18	45
		Beany	2	5	4	24	
7 days	1 : 5	Soury		3	1	13	
		Fishy		4	5	32	67
		Beany		2	4	22	
	1 : 10	Soury				-	
		Fishy		4	3	21	54
		Beany		5	5	33	
14 days	1 : 5	Soury		1	1	9	
		Fishy		5	2	23	69
		Beany		7	4	37	
	1 : 10	Soury		3	1	13	
		Fishy		6	3	30	66
		Beany		5	2	23	

* : Very faintly offensive odor; : Faintly offensive odor; : Definitely offensive odor; : Strongly offensive odor; : Very strongly offensive odor
 **Intensity index : No. of detection multiplied by scores in parenthesis × (1), (2), (3), (4), (5)

Table 2-7 2-8
가

가
Williams(1984)가 10 20 10,000g
VFA(C2-C6)가 , Spoelstra(1977)가 GC indole
penol

Yasuhara Fuwa(1977) butyric,
isovaleric, benzoic, penylacetic acid p-cresol
butyric, isovaleric penylacetic acid
, Bell (1970) VFA

table 2-24

1:5 1:10

가 가

가

가

()

1:10

1:5

가

(table 2-25)

가 가

Spoelstra(1980) Hammond et al.(1989)

(soury odor)가 가 (fishy odor)가 가

. Spoelstra(1980)

steroids 가

down stream

(PGR)

(GB)

가

가 . ,

PGR 1%, GB 1% 가 1 , 3 , 7 , 14

H₂O 50% ethanol

Tables 2-9

2-12 .

가

steroids

가

가

가

. PGR GB saponin

28. 4ng/g 19. 2ng/g . (1987)

saponin 22. 9ng/g , 42. 1ng/g ,

saponin .

Table 2-9. Odor intensity of swine manure stored for 1 day

Partition	pH	Odor intensity *					Intensity index**
MM	8.53	3	7	6	2	2	53
MMWS	7.06	3	5	6	4	2	57
MMWI	7.43	2	2	11	3	2	61
MMES	7.78	2	3	6	7	2	64
MMEI	7.14		2	7	9	2	71
PM	7.88	8	8	2	2		38
PMWS	6.68	7	9	2	2		39
PMWI	6.43	3	6	9	2		50
PMES	7.65	2	4	2	7	5	69
PMEI	7.38		1	2	13	4	80
GM	8.56	6	9	3	2		41
GMWS	7.00	3	6	7	4		52
GMWI	6.71		9	5	4	2	59
GMES	7.93		3	8	6	3	69
GMEI	7.32		3	4	8	5	75

* : Very faintly offensive odor; : Faintly offensive odor; : Definitely offensive odor;

: Strongly offensive odor; : Very strongly offensive odor

**Intensity index : No. of detection multiplied by scores in parenthesis × (1), (2), (3), (4), (5)

Table 2-10. Odor intensity of swine manure stored for 3 days

Partition	pH	Odor intensity *					Intensity index**
MM	7.73		3	5	12		69
MMWS	7.69		2	3	11	4	77
MMWI	7.63		6	9	5		59
MMES	8.21		11	5	2	2	55
MMEI	7.36			4	6	10	86
PM	6.29		5	3	8	4	71
PMWS	7.71	3	10	5	2		46
PMWI	7.54	6	8	4	2		42
PMES	6.95	4	3	6	7		56
PMEI	6.66		3	5	9	3	72
GM	7.31		2	5	6	7	78
GMWS	7.18	1	4	8	4	3	64
GMWI	6.95	5	8	3	4		46
GMES	7.24	5	9	4	2		43
GMEI	6.71		3	7	4	6	73

* : Very faintly offensive odor; : Faintly offensive odor; : Definitely offensive odor; : Strongly offensive odor; : Very strongly offensive odor

**Intensity index : No. of detection multiplied by scores in parenthesis × (1), (2), (3), (4), (5)

Table 2-11. Odor intensity of swine manure stored for 7 days

Partition	pH	Odor intensity *					Intensity index**
MM	7.77		6	5	9		63
MMWS	7.65	8	8	4			72
MMWI	8.01		10	3	7		57
MMES	8.12	2	5	4	4	5	65
MMEI	7.56		2		4	14	90
PM	6.52	5	7	6	2		45
PMWS	7.85	4	8	7	1		45
PMWI	7.78	3	4	7	4	2	58
PMES	6.72	4	4	4	6	2	58
PMEI	6.55			3	4	13	90
GM	6.34	2	5	6	7		58
GMWS	7.56	4	10	5	1		43
GMWI	8.48	6	6	8			42
GMES	8.57		5	11	4		59
GMEI	8.59		2	8	4	6	74

* : Very faintly offensive odor; : Faintly offensive odor; : Definitely offensive odor; : Strongly offensive odor; : Very strongly offensive odor

**Intensity index : No. of detection multiplied by scores in parenthesis × (1), (2), (3), (4), (5)

Table 2-12. Odor intensity of swine manure stored for 14 days

Partition	pH	Odor intensity *					Intensity index**
		(1)	(2)	(3)	(4)	(5)	
MM	8.10	2	3	8	7		75
MMWS	8.07		7	11	1	1	56
MMWI	8.25		7	6	5	2	62
MMES	7.92		6	5	6	3	66
MMEI	8.20		2	5	10	3	74
PM	8.27	6	8	6			40
PMWS	6.53		7	6	3	4	64
PMWI	6.42		2	6	10	2	72
PMES	8.43		5	4	6	5	71
PMEI	8.63			5	12	3	78
GM	8.33	5	10	3	2		42
GMWS	7.81		8	9	3		55
GMWI	8.18	2	5	8	5		56
GMES	6.67				5	15	95
GMEI	6.50				4	16	96

* : Very faintly offensive odor; : Faintly offensive odor; : Definitely offensive odor; : Strongly offensive odor; : Very strongly offensive odor

**Intensity index : No. of detection multiplied by scores in parenthesis × (1), (2), (3), (4), (5)

pH가 가 pH 7
 pH가 8 PM 14 GM
 50% ethanol pH가
 14 PMI pH Fig. 2-48
 sulfide disulfide
 pH 가
 sulfonic VOC 가 pH가

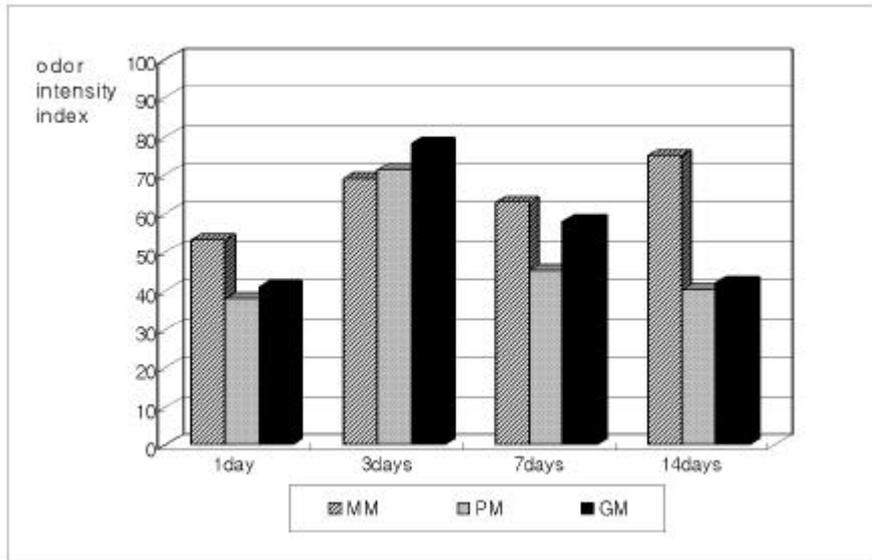


Fig. 2-28. Odor intensity of swine manure during field storage

Fig. 2-29

3

가

Fig. 2-30

가

(PGR) 1%

(GB) 1%

가

7

Fig. 2-28

가 (M)

가

Fig. 2-29 2-30

PGR GB 가

가 7

가

3

가

가

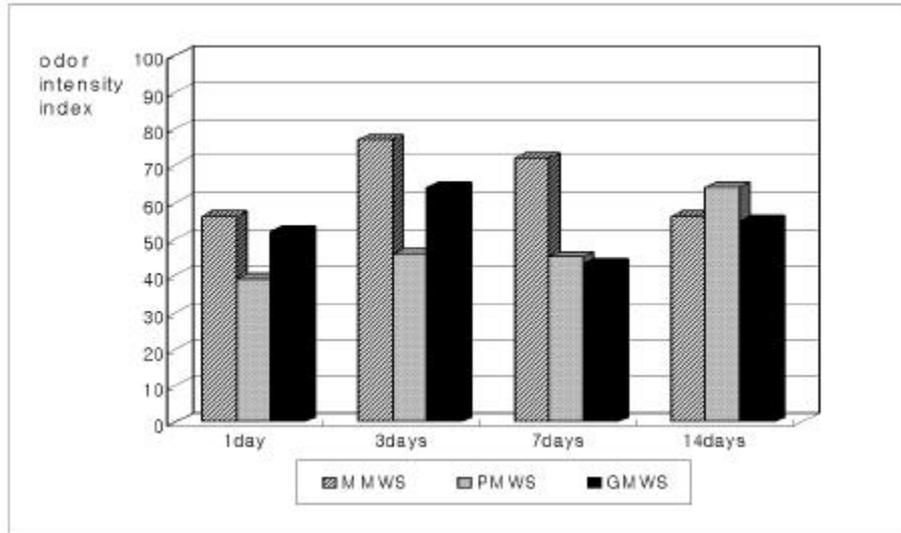


Fig. 2-29. Odor intensity of water soluble fraction of swine manure during field storage

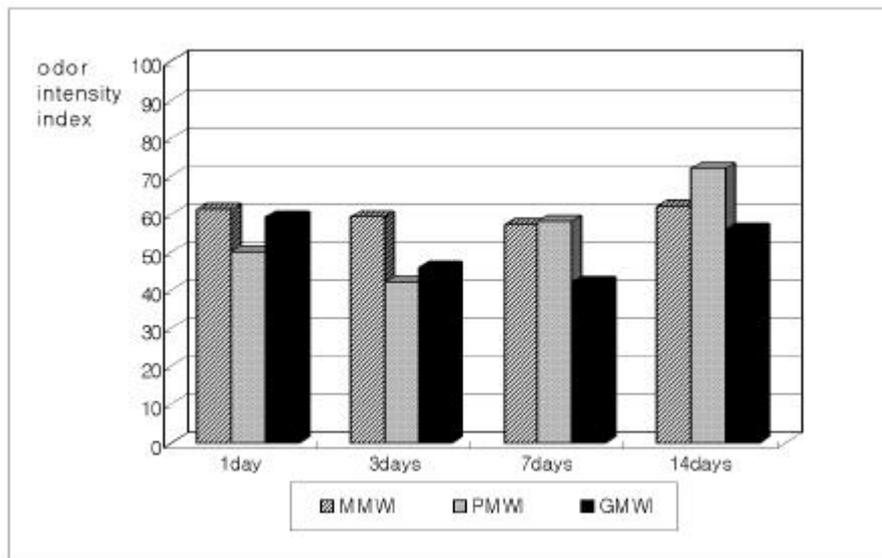


Fig. 2-30. Odor intensity of water insoluble fraction of swine manure during field storage

50% ethanol

Fig. 2-31 2-32 . Fig. 2-31

ethanol 가 .
가

alcohol 가

alcohol

Ethanol

가 (Fig. 2-32).

alcohol 가

가 가

PGR GB 가가 ethanol

ES EI

가 . ethanol 14

PGR GB 가 가

. GB 가 14 가 가 ethanol
가

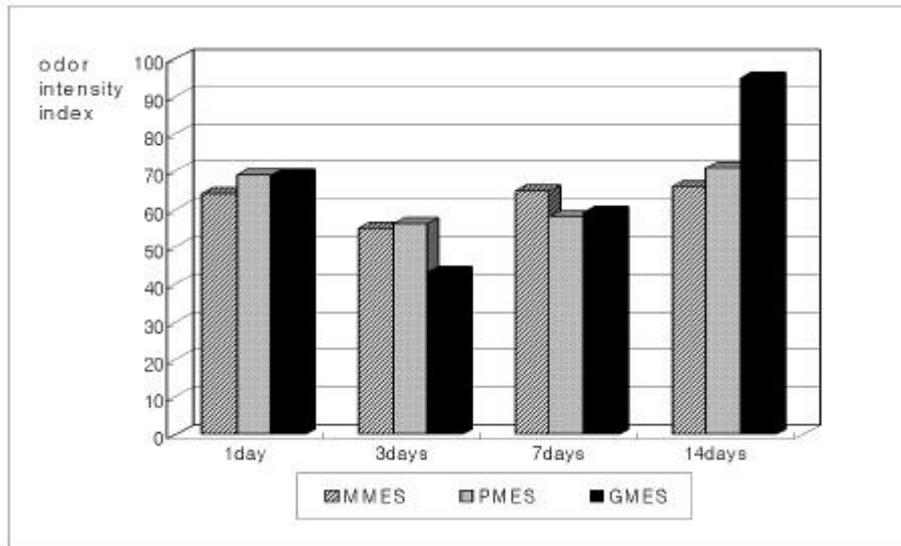


Fig. 2-31. Odor intensity of 50% ethanol soluble fraction of swine manure during field storage

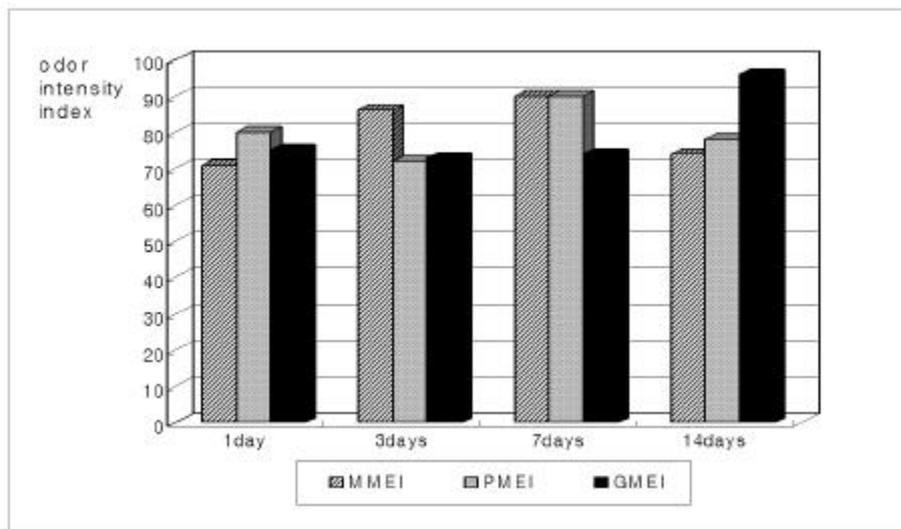


Fig. 2-32. Odor intensity of 50% ethanol insoluble fraction of swine manure during field storage

. VOC(Volatile organic compounds)

(olfactory analysis)

GC GC
Mass (Volatile organic compounds, VOC) VOC
profile .

(PGR) 1% (GB) 1% 가

1 , 3 , 7 , 14

VOC profile Figs. 2-33 48 .

Figs. 2-33 2-40 flame ionization detector(FID)

VOC , Figs. 2-41 2-48 curve

profile flame photometric detector(FPD) VOC

S- .

Figs. curve FID FPD GC

GC Mass curve profile

Figs. 2-33 2-48 VOC profile 가

FID FPD dimethyl sulfide,

dimethyl disulfide, methyl chloride .

olfactory analysis 가

FID ,

acetic acid, ethyl benzene, p-xylene, styrene

o-xylene 가

가 (1996) .

O'Neill Phillips(1992) Bell(1970)

disulfide FID 가
 가 . VOC
 가 PGR GB
 가 dimethyl sulfide dimethyl disulfide profile
 PGR GB 가 가

GC Mass VOC dimethyl sulfide,
 methyl chloride, 2-methylpentane, 3-methylpentane, dimethyl disulfide,
 2-methyl hexane, heptane, toluene, acetic acid, ethyl benzene, p-xylene,
 styrene, o-xylene .

GC Mass Hammond
 et al. (1989) GC GC Mass 가

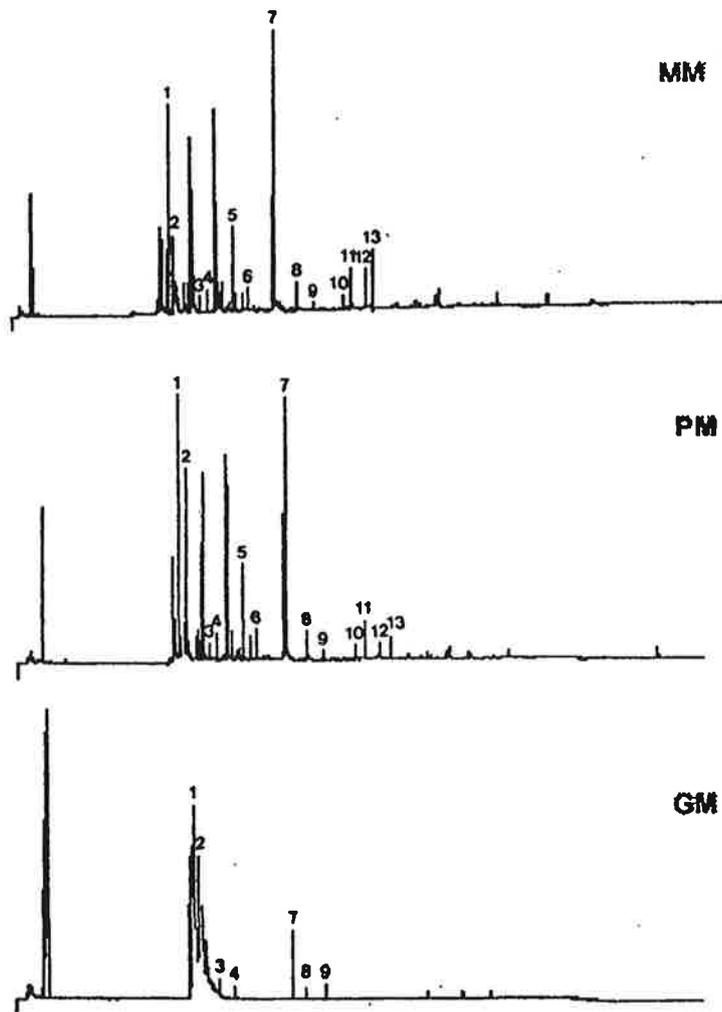
dimethyl sulfide, dimethyl disulfide ,
 methyl pentane, ethyl acetate, p-cresol, indole

. Hobbs et al. (1995) GC Mass dimethyl sulfide
 dimethyl disulfide, dimethyl trisulfide

methyl sulfide

methyl sulfide

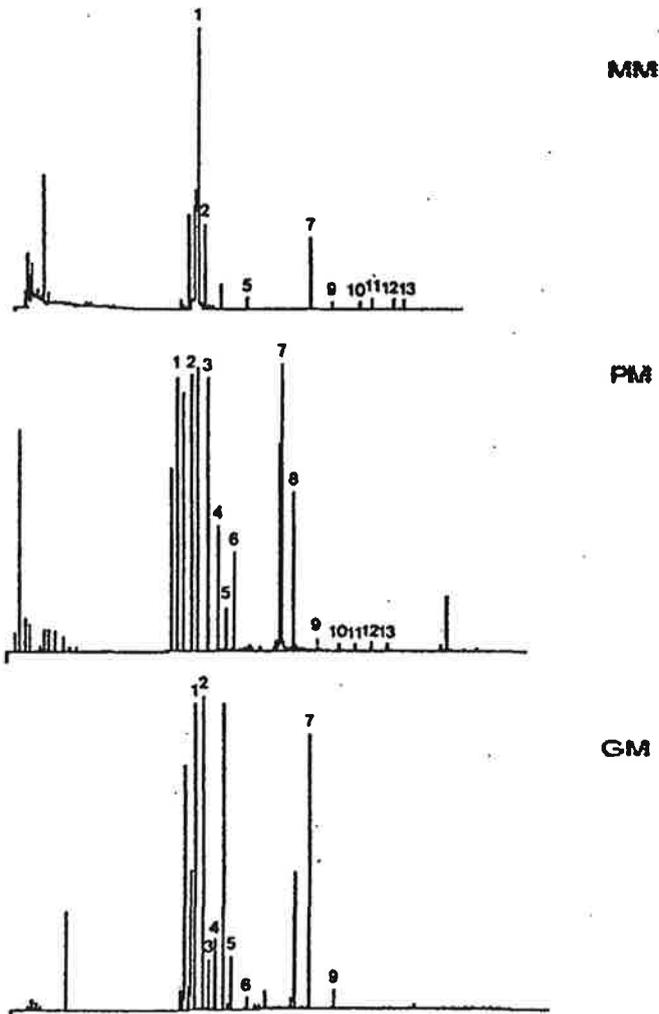
가



Legends :

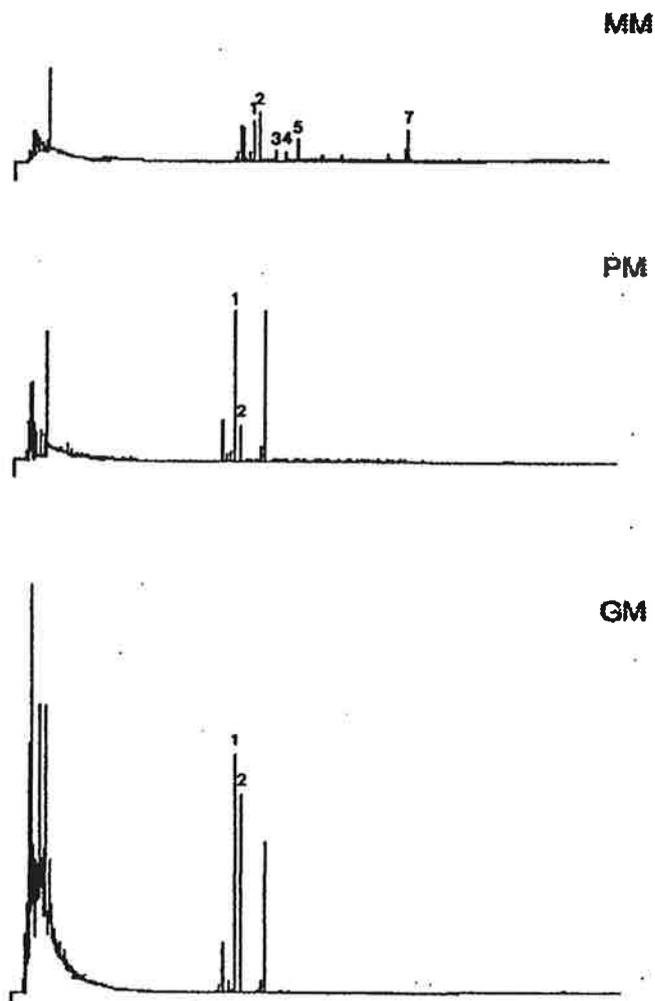
1: Dimethyl sulfide 2: Methyl chloride 3: 2-Methylpentane 4: 3-Methylpentane
 5: 2-Methyl hexane 6: Heptane 7: Dimethyl disulfide 8: Toluene 9: Acetic acid
 10: Ethyl benzene 11: p-Xylene 12: Styrene 13: o-Xylene

Fig. 2-33. Hydrocarbon VOC profile of swine manure stored 1 day



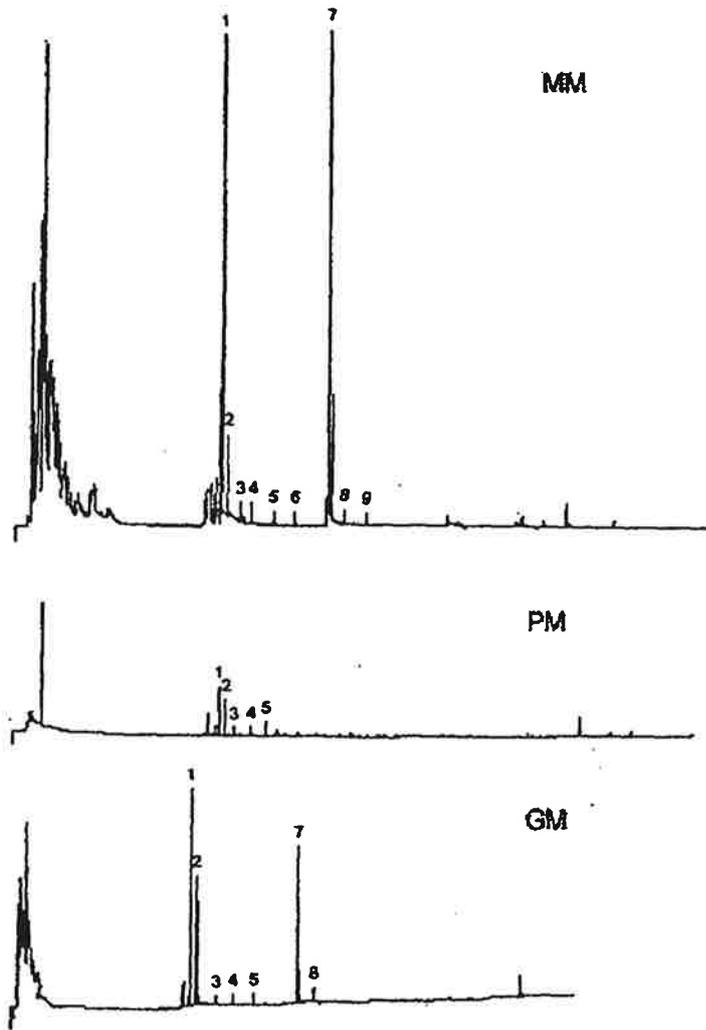
Legends :	
1:	Dimethyl sulfide
2:	Methyl chloride
3:	2-Methylpentane
4:	3-Methylpentane
5:	2-Methyl hexane
6:	Heptane
7:	Dimethyl disulfide
8:	Toluene
9:	Acetic acid
10:	Ethyl benzene
11:	p-Xylene
12:	Styrene
13:	o-Xylene

Fig. 2-34. Hydrocarbonic VOC profile of swine manure stored 3 days



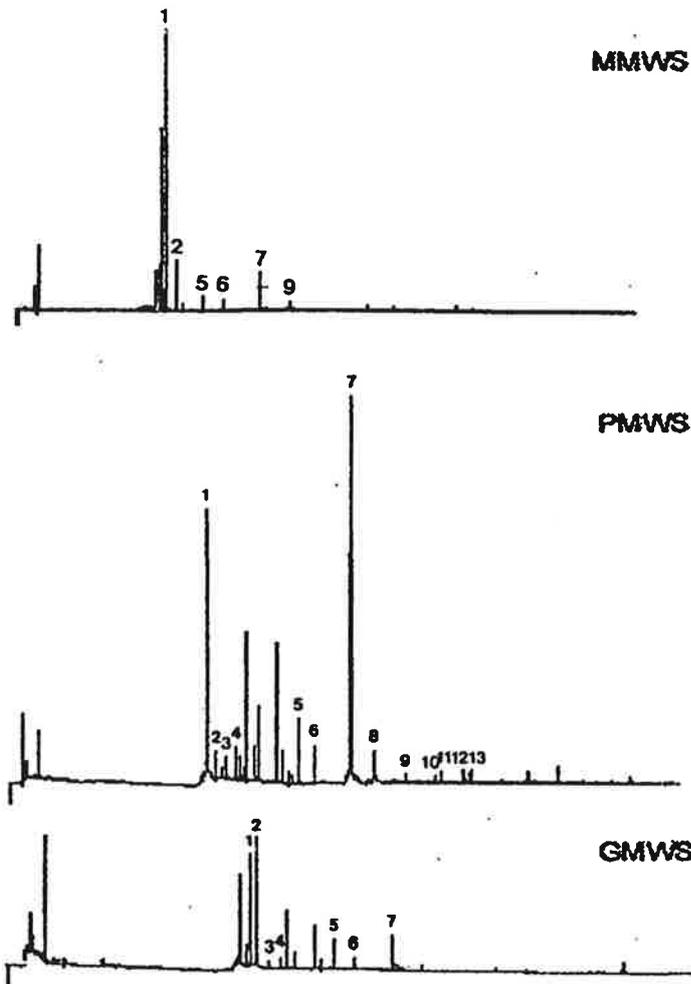
Legends :
 1: Dimethyl sulfide 2: Methyl chloride 3: 2-Methylpentane 4: 3-Methylpentane
 5: 2-Methyl hexane 7: Dimethyl disulfide

Fig. 2-35. Hydrocarbonic VOC profile of swine manure stored 7 days



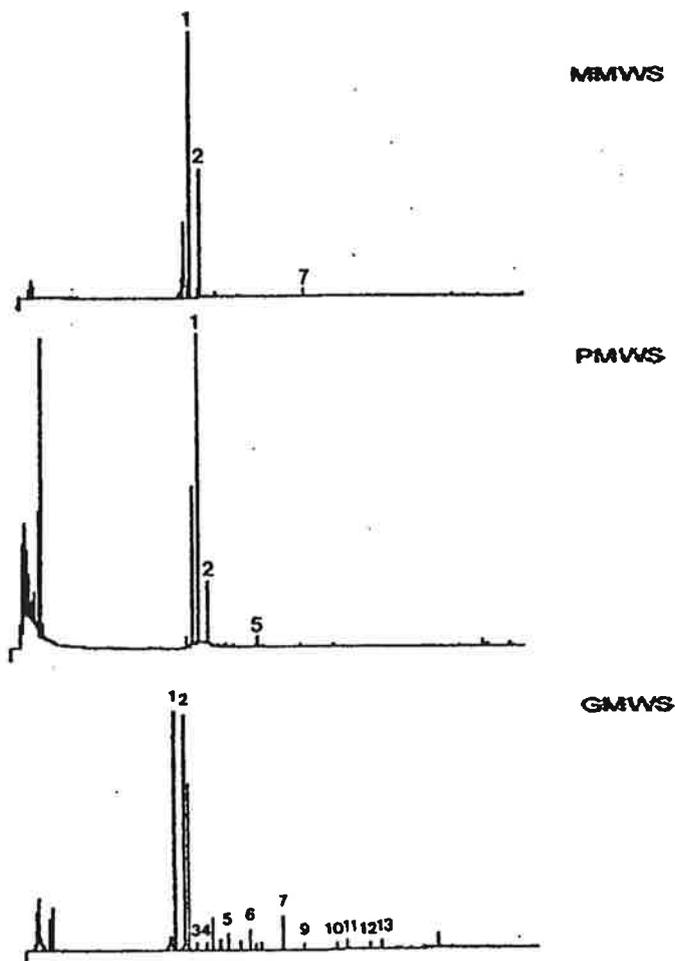
Legends :
 1: Dimethyl sulfide 2: Methyl chloride 3: 2-Methylpentane 4: 3-Methylpentane
 5: 2-Methyl hexane 6: Heptane 7: Dimethyl disulfide 8: Toluene 9: Acetic acid

Fig. 2-36. Hydrocarbonic VOC profile of swine manure stored 14 days



Legends :			
1:	Dimethyl sulfide	2:	Methyl chloride
3:	2-Methylpentane	4:	3-Methylpentane
5:	2-Methyl hexane	6:	Heptane
7:	Dimethyl disulfide	8:	Toluene
9:	Acetic acid		
10:	Ethyl benzene	11:	p-Xylene
12:	Styrene	13:	o-Xylene

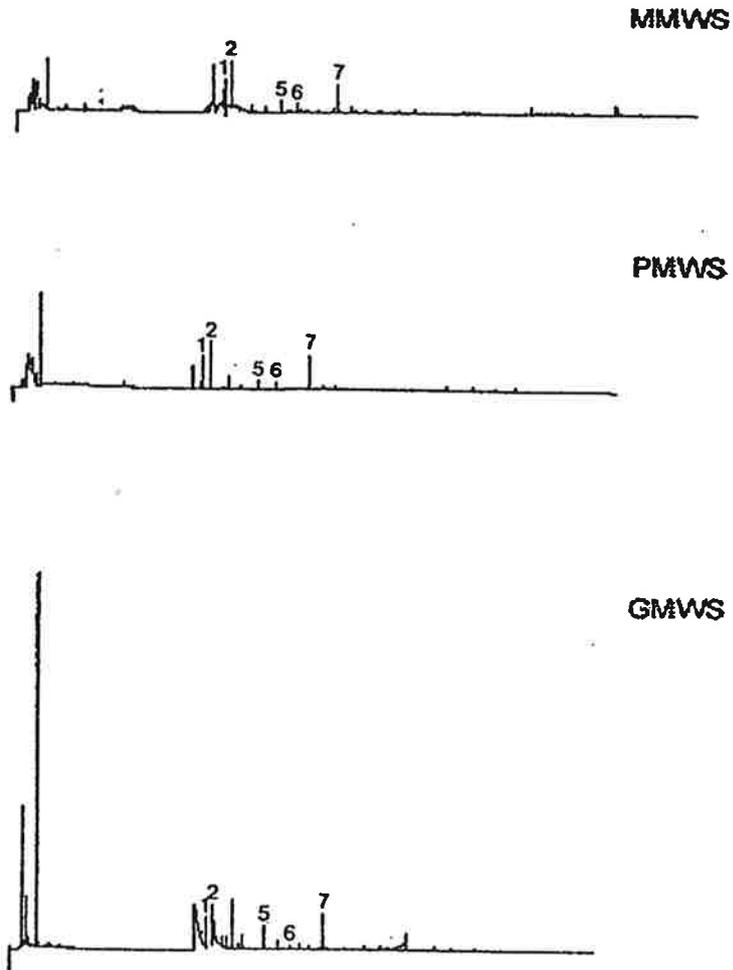
Fig. 2-37. Hydrocarbon VOC profile of water soluble manures stored 1 day



Legends :

1: Dimethyl sulfide 2: Methyl chloride 3: 2-Methylpentane 4: 3-Methylpentane
 5: 2-Methyl hexane 6: Heptane 7: Dimethyl disulfide 9: Acetic acid
 10: Ethyl benzene 11: p-Xylene 12: Styrene 13: o-Xylene

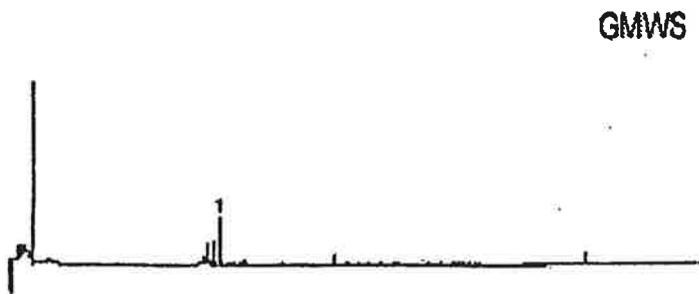
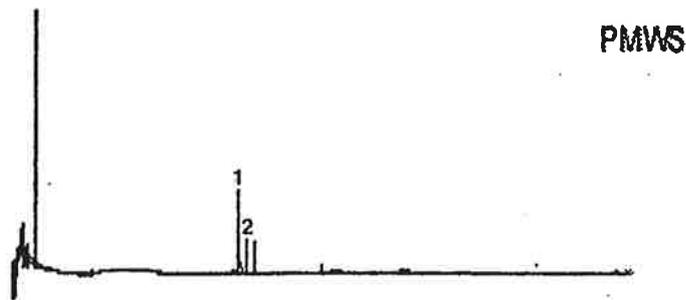
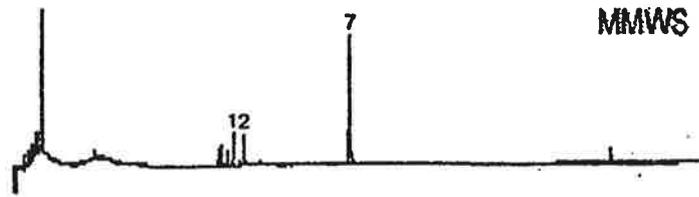
Fig. 2-38. Hydrocarbonic VOC profile of water soluble manures stored 3 days



Legends :

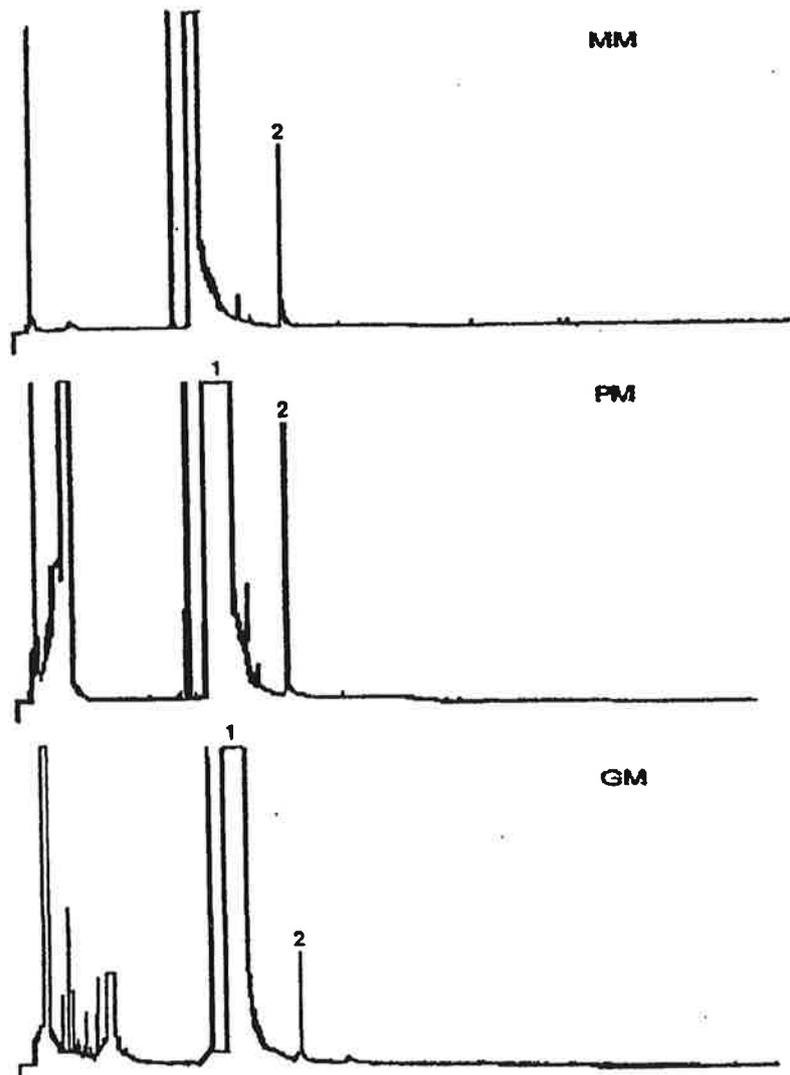
- 1: Dimethyl sulfide 2: Methyl chloride 5: 2-Methyl hexane 6: Heptane;
 7: Dimethyl disulfide

Fig. 2-39. Hydrocarbonic VOC profile of water soluble manures stored 7 days



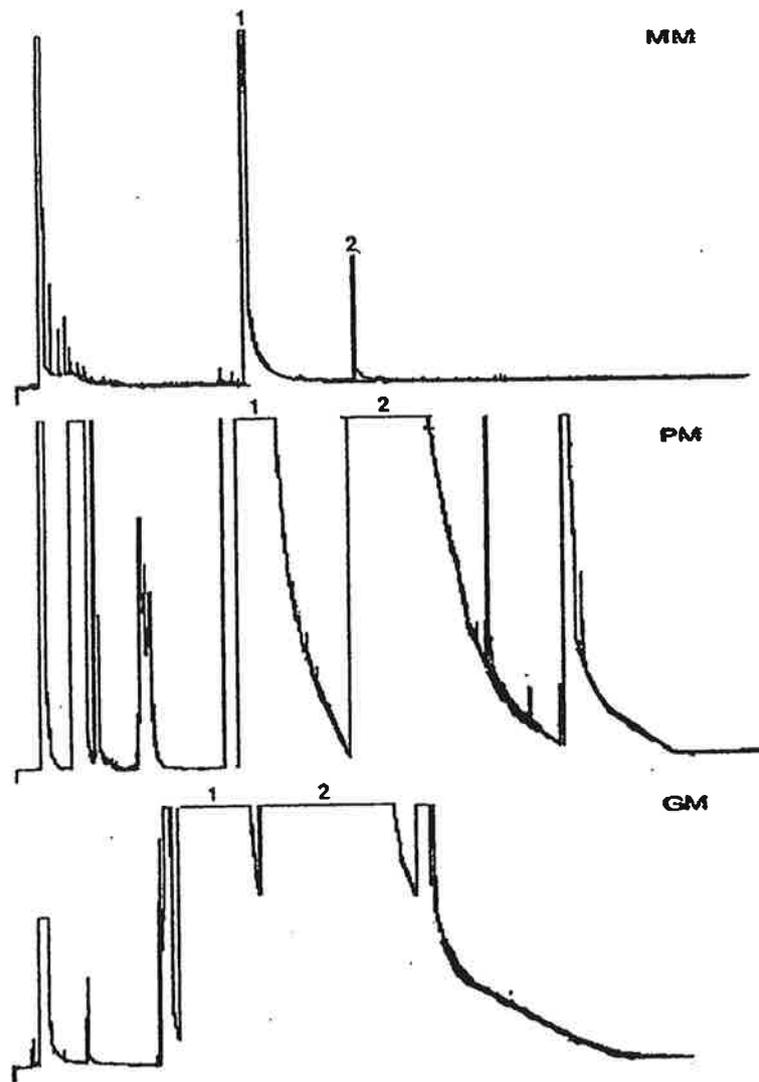
Legends :
 1: Dimethyl sulfide 2: Methyl chloride 7: Dimethyl disulfide

Fig. 2-40. Hydrocarbonic VOC profile of water soluble manures stored 14 days



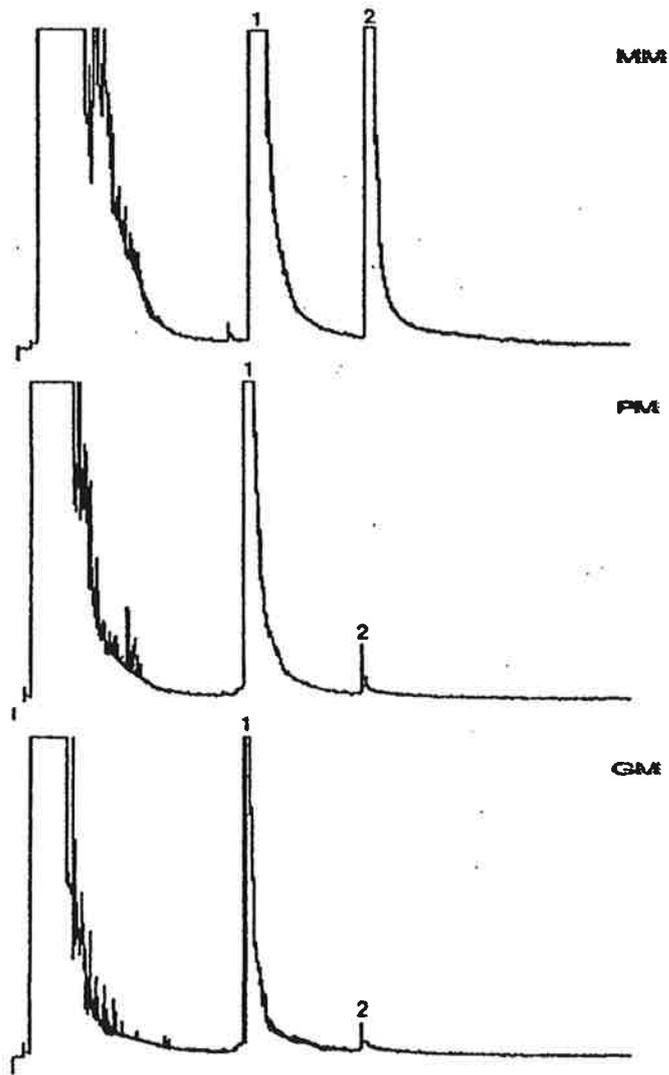
Legends:
 1: Dimethyl sulfide 2: Dimethyl disulfide

Fig. 2-41. Sulfonic VOC profile of swine manures stored 1 day



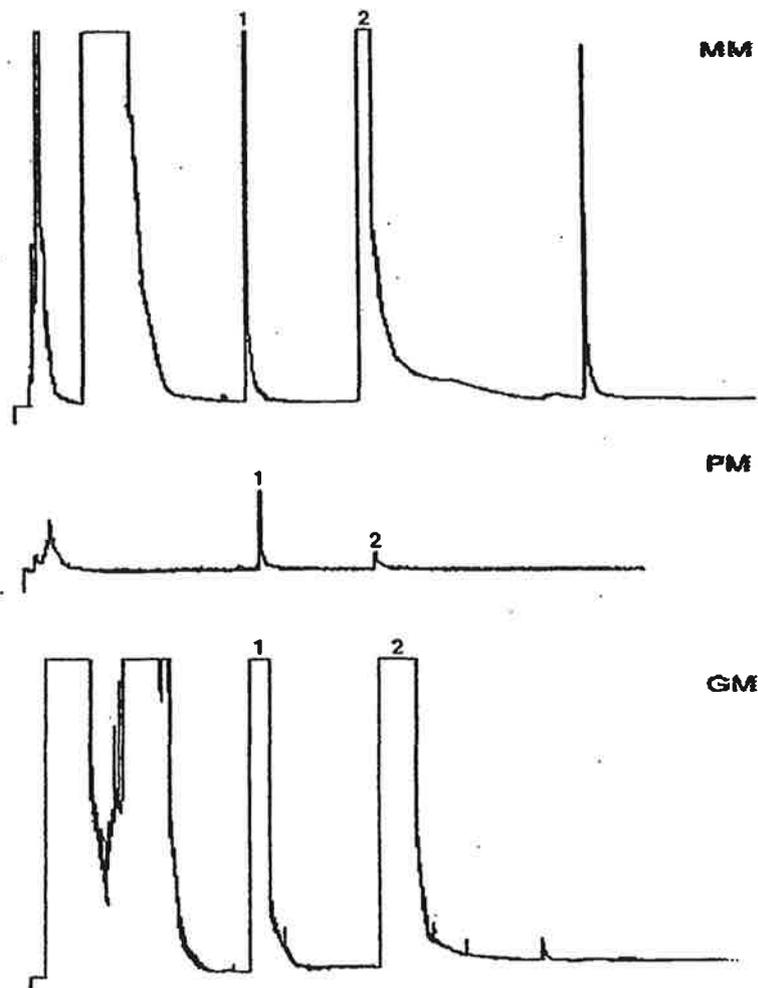
Legends:
 1: Dimethyl sulfide 2: Dimethyl disulfide

Fig. 2-42. Sulfonic VOC profile of swine manures stored 3 days



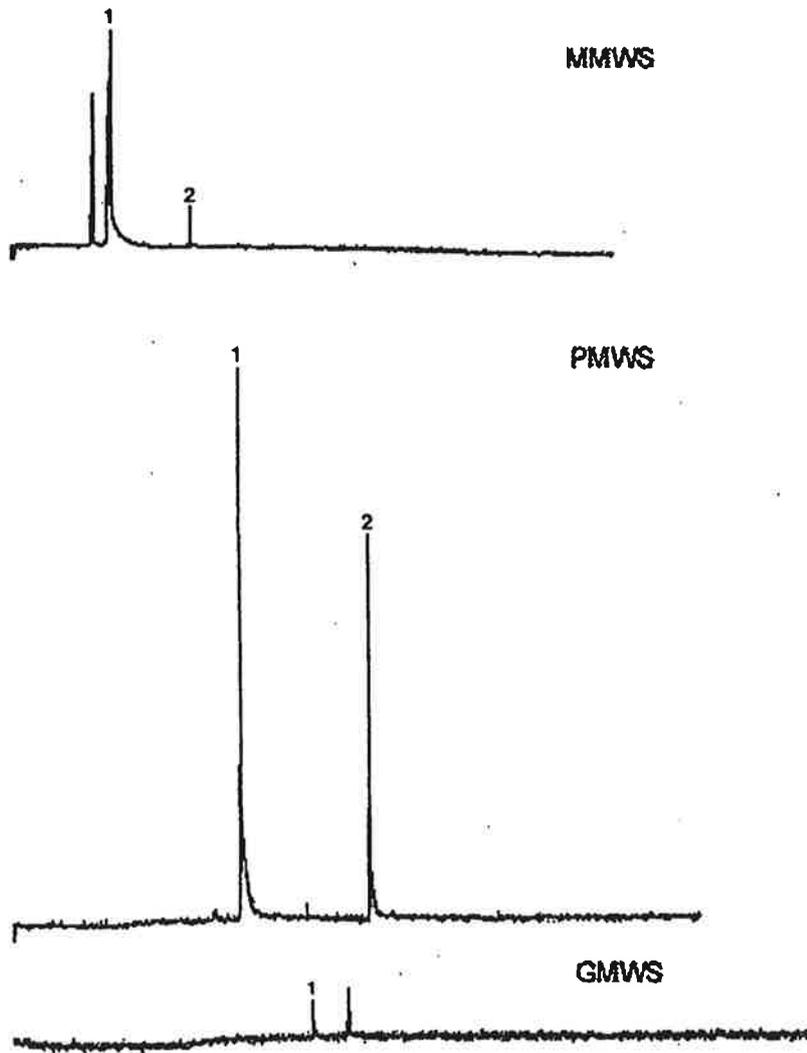
Legends:
 1: Dimethyl sulfide 2: Dimethyl disulfide

Fig. 2-43. Sulfonic VOC profile of swine manures stored 7 days



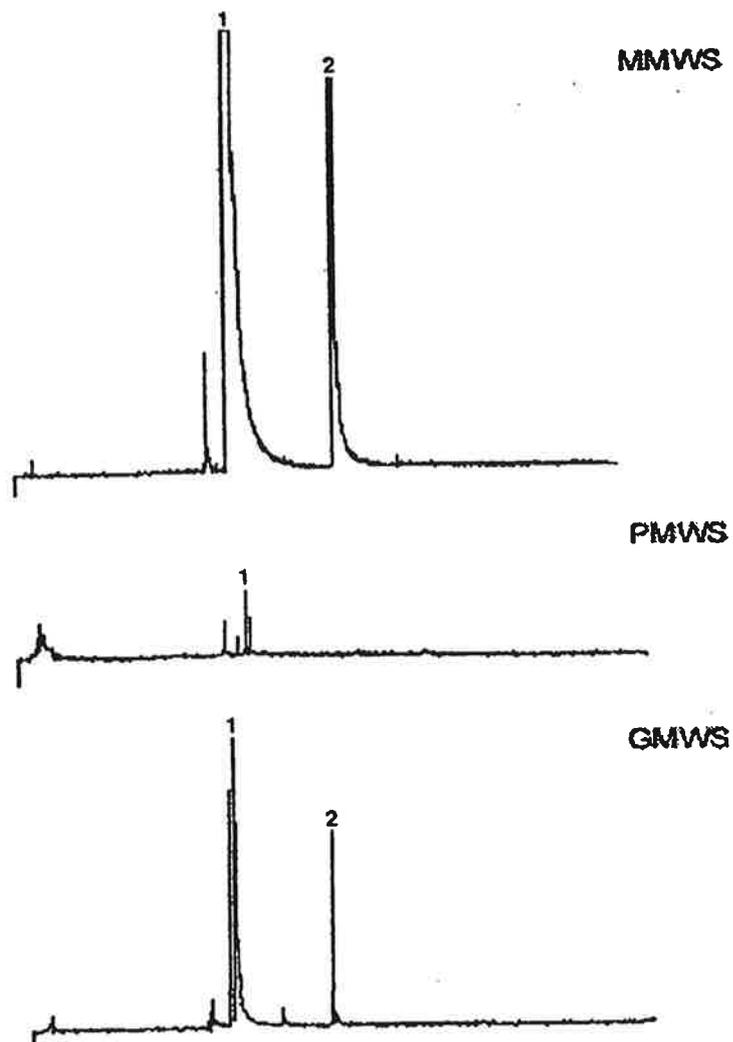
Legends:
 1: Dimethyl sulfide 2: Dimethyl disulfide

Fig. 2-44. Sulfonic VOC profile of swine manures stored 14 days



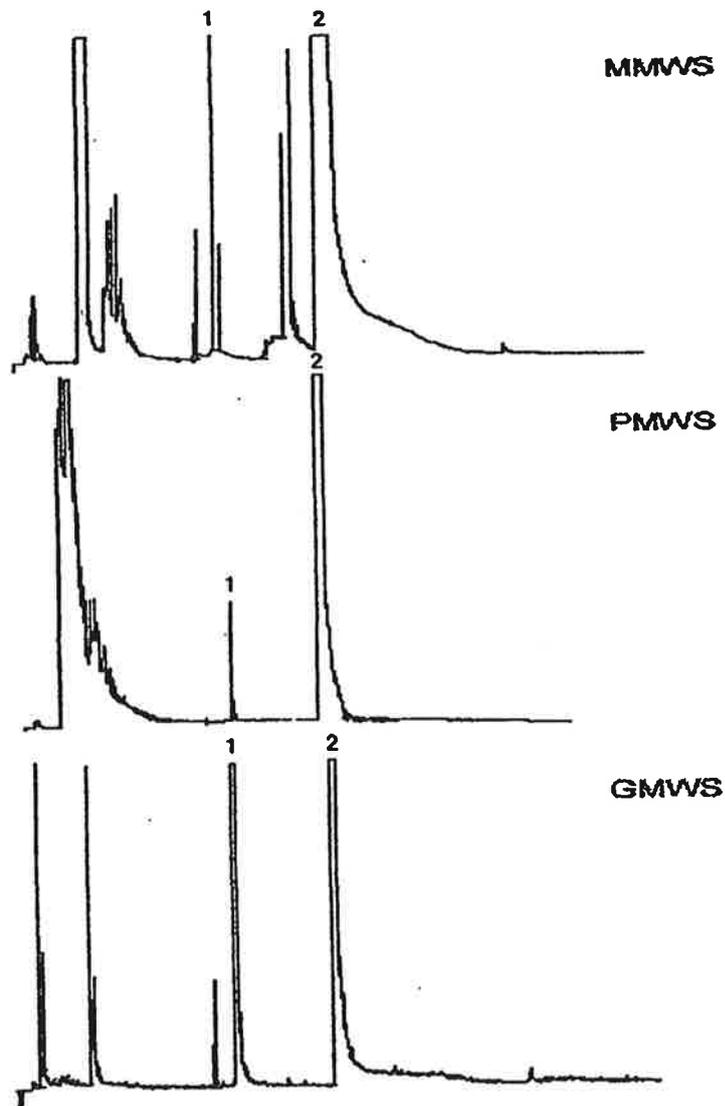
Legends:
 1: Dimethyl sulfide 2: Dimethyl disulfide

Fig. 2-45. Sulfonic VOC profile of water soluble manures stored 1 day



Legends:
 1: Dimethyl sulfide 2: Dimethyl disulfide

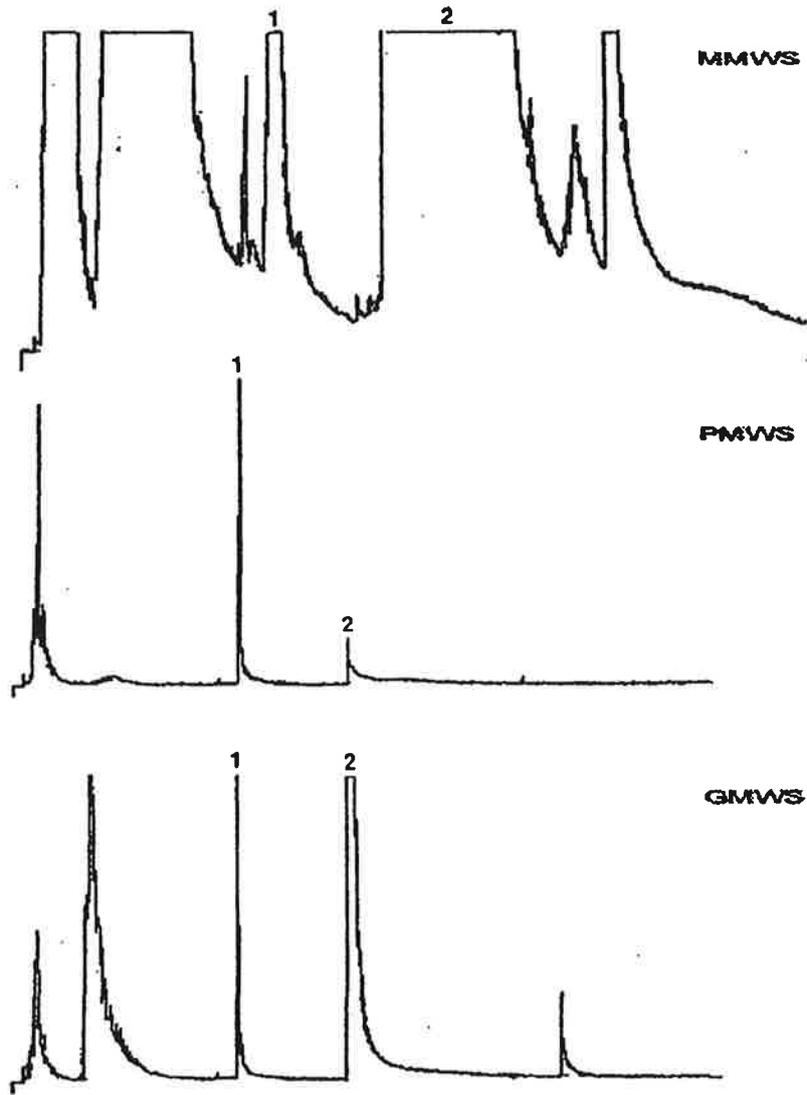
Fig 2-46. Sulfonic VOC profile of water soluble manures stored 3 days



Legends:

1: Dimethyl sulfide 2: Dimethyl disulfide

Fig. 2-47. Sulfonic VOC profile of swine manures stored 7 days



Legends:
 1: Dimethyl sulfide 2: Dimethyl disulfide

Fig. 2-48. Sulfonic VOC profile of swine manures stored 14 days

4. 요 약

돈분뇨에서의 악취를 일으키는 원인물질을 규명하고 도라지와 인삼부산물에서 추출한 식물성 steroid의 악취저감효과를 알아보려고 본 실험을 수행하였다.

돈분뇨에서의 악취를 일으키는 물질을 규명하고자 H₂O와 50% ethanol로 분획하여 수용성과 불용성으로 돈분을 분리하여 각 처리별 돈분뇨에서의 후각인지 측정을 통한 odor intensity를 분석하였고, 별도 제작한 흡착관을 이용하여 GC와 GC Mass에서 악취유발 VOC 물질을 탐색하고자 하였다. 또한 도라지(PGR)와 인삼부산물박(GB)의 saponin 함량을 분석하고 PGR와 GB 1%를 각각 돈분에 첨가하여 밀봉한후 1일, 3일, 7일, 14일 동안 실온에 방치한후 악취감소 정도를 상기 방법으로 평가하였다.

1. 돈분뇨를 원심분리하였을 때 상층액의 악취강도가 하층액보다 odor intensity가 강하게 나타났으며, 희석배율이 일정수준까지 증가할 때 odor intensity가 증가하는 경향을 보여 주요 악취성분이 수용성임을 나타내었다.

2. 후각분석에 의한 돈분 분획의 odor intensity는 방치초기에는 수용성 분획내에서 상대적으로 강하였으며 14일 경과후에는 비수용성 분획에서도 그 강도가 높아졌다.

3. 돈분의 에탄올 분획에서의 odor intensity는 비용해성 분획에서의 강도가 용해성 분획에 비하여 상대적으로 강한 것으로 나타나 악취성분이 50% ethanol에 의하여 일부 파괴됨을 보여주었다.

4. PGR 1% 또는 GB 1%를 처리한 분뇨의 경우 7일, 14일이 경과되면서 각 odor intensity와 VOC 함량이 감소되는 경향을 나타내었다.

5. GC Mass VOC dimethyl sulfide,
 methyl chloride, 2-methyl pentane, 3-methyl pentane, dimethyl
 disulfide, 2-methyl hexane, heptane, toluene, acetic acid, ethyl
 benzene, p-xylene, styrene, o-xylene .

6. VOC acetic acid, ethyl benzene, p-xylene, styrene,
 o-xylene dimethyl sulfide, dimethyl
 disulfide 가 PGR 1%, GB 1%
 가 .

acetic acid,
 ethyl benzene, p-xylene, styrene, o-xylene
 dimethyl sulfide, dimethyl disulfide .

steroid saponin PGR GB down stream
 가 가 가 .

5.

Anon, M., Dobeic, M., Misselbrook, T. H., Pain, B. F., Phillips, V. R. and Sneath, R. W. 1995. A farm scale study on the use of De-Odorase[®] for reducing odour and ammonia emissions from intensive fattening piggeries. *Biores. Tech.* 51 : 163-169

Austin, R. L., Steven, J. M. and William, J. T. 1988. Rapid direct extraction derivatization method for the determination of acylglycerol lipids in selected sample matrices. *J. Food Sci.* 53(3) : 940-946

Banwart, W. L. and Brenner, J. M. 1975. Formation of volatile sulfur compounds by microbial decomposition of sulfur containing amino acids in soils. *Soil. Biol. Biochem.* 7 : 359-364

Barth, C. L., Hill, D. T, Polkowski, L. B. 1974. Correlating odor intensity index and odorous components in stored dairy manure. *Trans. Am. Soc. Agric. Engrs.* 17 : 742-747

Bell, R. G. 1970. Fatty acid content as a measure of the odour potential of stored liquid poultry manure. *Poul. Sci.* 49 : 1126-1129

Bethea, R. M. and Narayan, R. S. 1972. Identification of beef cattle feedlot odors. *Trans. Am. Soc. Agric. Engrs.* 15 : 1135-1137

Burnett, W. E. 1969. Air pollution from animal wastes. Determination of malodors by gas chromatographic and organoleptic techniques. Environ. Sci. Tech. 3 : 744-750

Day, D. L., Hansen, E. L. and Anderson, S. 1965. Gases and odors in confinement swine buildings. Trans. Am. Soc. Agric. Engrs. 8 : 118-121

Dravnieks, A. 1972. Odor measurement. Environ. Letters, 3 : 81-100

Elliott, L. F. and Travis, T. A. 1973. Detection of carbonyl sulfide and other gases emanating from beef cattle manure. Soil Sci. Soc. Am. Proc. 37 : 700-702.

Hammond, E. G., Heppner, C. and Smith, R. J. 1989. Odors of swine waste lagoons. Agric. Ecosystems and Environment, 25 : 103-110

Hammond, E. G. and Smith, R. J. 1981. Survey of some molecularly dispersed odorous constituents in swine-house air. Iowa state Jour. Res. 55(4) : 393-399

Hammond, E. G., Junk, G. A., Kuczala, P and Kozel, J. 1974. Constituents of swine house odors. In : *Proc. Int. Symp. Livestock Environment*. 364-372

Hartung, J. and Hilliger, H. G. 1980. Odour characterisation in animal houses by gas chromatographic analysis on the basis of low

temperature sorption. *In : Effluents from livestock* , Appl. Scien. Pub. London. 561-578

Hartung, L. D. , Hammond, E. G. and Miner, J. R. 1971. Identification of carbonyl compounds in a swine building atmosphere. *In : Livestock waste management and Pollution Abatement. Proc. Int. Symp. Livestock Wastes*. An. Soc. Agric. Engrs. , St. Joseph, Michigan. 105-106

Headon, D. R. and Walsh, G. 1993. *Yucca schottigera* extracts and ammonia control. *In : Livestock Environment* , ed. E. Collins and C. Egan. An. Soc. Agric. Engng. St Joseph, Michigan, USA. 686-693

Hilliger, H. G. and Hartung, J. 1978, Geruchsbewertung im stall durch verknuffung von sensorischen und analytischen methoden. *Organische verunreinigungen in der umwelt*. 475-482

Hobbs, P. J. , Misselbrook, T. H. and Pain, B. F. 1997. Characterisation of odorous compounds and enissions from slurries produced from weaner pigs fed dry feed and liquid diets. *J. Scien. Food Agric.* 73 : 437-445.

Hobbs, P. J. and Pain, B. F. 1996. Reduction of odorous compounds in fresh pig slurry by dietary control of crude protein. *J. Scien. Food Agric.* 71 : 508-514

Hobbs, P. J. , Misselbrook, T. H. and Pain, B. F. 1995. Assessment of odours from livestock wastes by a photoionization detector, an electronic

nose, olfactometry and gas chromatography-mass spectrometry. J. Agric. Engng Res. 60 : 137-144

Janowski, T., Gawlik, J. and Zinnal, S. 1975. Gas chromatographische untersuchungen der stallluft. VDI-Berichte. 226 : 123-126

Kazutaka, K., Takahi, O., Mitihiro, Y., Akane, K. Takako, N. Sigenori, M. and Tonoko, K. 1996. Emissions of malodorous compounds and greenhouse gases from composting swine feces. Biores, Technol. 56 : 265-271

Kenne, P. A., Jongbloed, A. W., Ellaert, B. M. and Krolkraener, F. 1993. The use of a *Yucca shidaigera* extract as 'urease inhibitor' in pig slurry. In: *Nitrogen Flow in Pig Production and Environmental consequences*, ed. M. V. A. Verstegen et al., Pudoc Scientific Publis. Wageningen, The Netherlands. 330-335.

Klarenbeek, J. V., Jongebreuer, A. A, Beumer, S. C. C. 1982. Odour emission in pig fattening sheds. IMAG Report No. 48, Wageningen

Kowalewsky, H. H., Scheu, R. and Vetter, H. 1980. Measurement of odour emissions and immissions. In : *Effluents from livestock*. Applied Science Publishers, London. pp 609-626.

Liao, P. H., Chen, A. and Lo, K. V. 1995. Renoval of nitrogen from swine manure wastewater by ammonia stripping. Bioresource Technology 54 : 17-20.

Macrae, R., Robinson, R. K. and Sadler, M. J. 1993. Encyclopedia of food

science, food technology and nutrition Vol. 6. Academic Press. San Diego. CA

Mackie, R. I. 1994. Microbial production of odor components. In *Proceedings of International Round table on Swine Odor Control*, 18-19. Ames, IA : Iowa state University.

Merck. 1996. Merck Index 18th ed. Merck & Co., Inc. N. J. USA.

Merkel, J. A., Hazen, T. E and Miner, J. R. 1969. Identification of gases in a swine confinement building atmospheres. *Trans. Am. Soc. Agric. Engrs*, 12 : 310-316.

Miner, J. R. and Hazen, T. E. 1969. Ammonia and amines. Components of swine-building odor. *Trans. Am. Soc. Agric. Engrs.*, 12 : 772-774.

Miner, J. R., Kelly, M. D. and Anderson, A. W. 1975. Identification and measurement of volatile compounds within a swine building and measurement of ammonia evolution rates from manure-covered surfaces. *In : Managing Livestock Wastes. Proc. 3rd Int. Symp. Livestock Wastes.* Am. Soc. Agric. Engrs., St. Joseph, Michigan, 351-353.

Moncrieff, R. W. 1967. The chemical senses. Leonard Hill, London. pp. 142

Mosier, A. R., Andre, C. E. and Viets, F. G. 1973. Identification of aliphatic amines volatilized from cattle feedyard. *Environm. Sci.*

Techn., 7 : 642-644

O'Neill, D.H. and Phillips, V.R. 1992. A review of the control of odour nuisance from livestock buildings: Part 3, properties of the odorous substances which have been identified in livestock wastes on in the air around them. Journal of Agricultural Engineering Research 53 : 23-50

Schaefer, J. 1977. Sampling, characterization and analysis of malodours. Agriculture and Environment, 3 : 121-127

Schaefer, J., Benelans, J.M.H. and Ten Over de Brauw, M.C. 1974. Onderzoek naar de voor de stank van varkensnesterijen verantwoordelijke componenten. Landbouwk. Tijdschr., 86 : 228-232

Schreier, P. 1975. Gas chromatographische massenspectrometrische Untersuchungen von Geruchstoffen aus der Tierhaltung. VDI-Berichte, 226 : 127-130

Soon-Il, Y. and Yoshiyuki, O. 1997. Some physiological properties of microorganisms capable of deodorizing farm animal feces. Biores. Technol. 21-26

Spoelstra, S.F. 1980. Origin of objectionable odorous components in piggery wastes and the possibility of applying indicator components

for studying odour development. *Agric. and Envir.* 5(3) : 241-260

Spelstra, S.F. 1977. Simple phenols and indoles in anaerobically stored piggery wasters. *J. Sci. of Food and Agric.* 28 : 415-423

Williams, A.G. 1984. Indicators of piggery slurry odour offensiveness. *Agric. Wastes* 10 : 15-36

Williams, A.G. and Evans, M.S. 1981. Storage of piggery slurry. *Agricultural Wastes*, 3(4) : 311-322

White, R.K., Taiganides, E.P. and Cole, C.D. 1971. Chromatographic identification of malodors from dairy animal waste. *In : Livestock Wastes Management and Pollution Abatement. Proc. Int. Symp. Livestock Wastes.* An. Soc. Agric. Engrs. St. Joseph, Michigan, pp 110-113

Wu, Z., Sadik, M., Sleinan, F.T., Sinas, J.M., Pessaraki, M. and Huber, J.T. 1994. Influence of yucca extract on ruminal metabolism in cows. *J. Anim. Sci.* 72 : 1038-1042

Yasuhara, A. and Fuwa, K. 1977. Odor and volatile compounds in liquid swine manure. Carboxylic acids and phenols. *Bull. Chem. Soc. Japan*, 50(3) : 731-733

Zhu, J., Bundy, D.S., Li, X. and Rashid, N.R. 1997. A procedure and its application in evaluating pit additives for odor control. *Cana.*

Agri. Eng. (3) 39 : 207-214

. 1996,

. 1992.

. 1987. ,

, , , , . 1996. ,

가가 , . 38(1)

: 52-58

3 Sarsaponin

1.

가 가 ,
가
가 가 . 가
가 , , +
가 3 .
가 가 , ,
 , . 가
가 , ,
(uniform) (nash) .
가
(carrier)가 . 가
 .
stear , ,
가 .
가
가
 . ,

(coating), , ,

가

가

가

가

가

가

가

가

가

가

가 가

가

.

,

가

가

가

가

.

NH₃,

H₂S

VOC(Volatile organic compounds)

가 .

가

(deodorizer : deodo)

가 (complex feed supplement : deodo-pro)

가

가 .

2.

가.

(1)

propolis

가

1999 7 5 9 2

60 2 4 (I*Y*D) control,
 low complex high complex

(2) 가

1999 10 15 11 13 30
 2 4 ,
 (D*H*L*Y) control, D0-50 D0-100

(1) 가 NH3 VOC
 NH3 H2S

(2) NH3 H2S
 가 Konyo kitagawa precision gas
 detectors , 250M \emptyset 50g
 50M \emptyset 25 1 3
 NH3 H2S

(3) VOC(Volatile organic compounds)

VOC

. VOC

fig. 2-49 100nm, 4.7mm, 6.45mm
 0.5g Tenax-GR(60/80mesh) frit

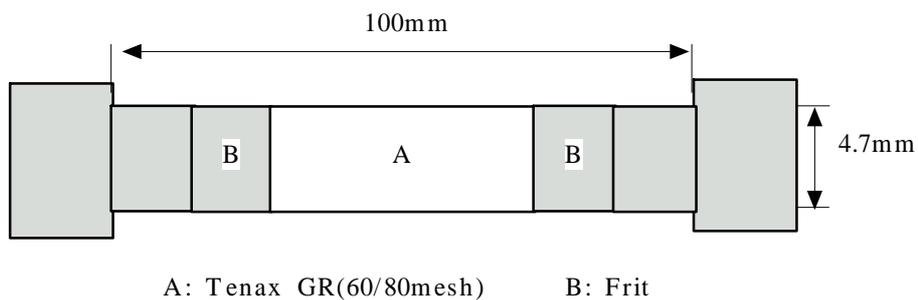
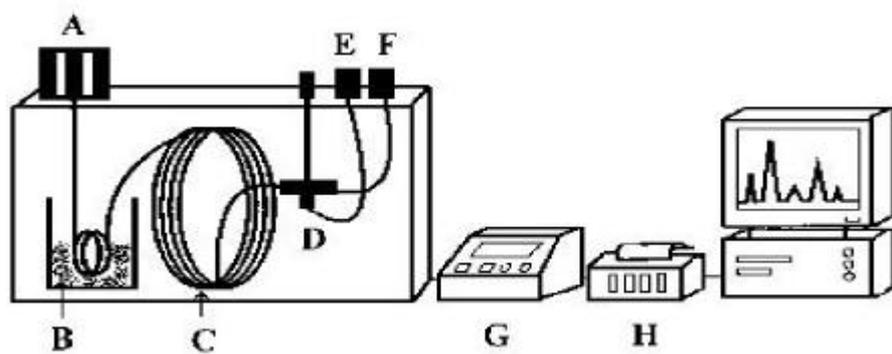


Fig. 2-49. Schematic diagram of 100mm adsorption tube



A: Thermal desorption B: Liquid N₂ C: Stainless column D: Outlet splitter E: FPD F: FID G: Electronic pressure control H: Integrator

Fig. 2-50. Schematic diagram of on-column cryofocusing gas chromatography system

Column stainless steel capillary
 column(UA5, Frontier Lab 30m x 0.25mm, 0.3μm)
 VOC fig. 2-50
 200 가 UA5
 column -196 8
 outlet splitter 가 1:3 dual
 detector system flame ionization detector(FID)
 , flame photometric detector(FPD)

(4)

SAS (1989)
 Duncan
 (Snedecor Cochran, 1980)

가 가
 가 가 ,
 가 , 가 가

(1) (particle size)

Ro-Tap testing sieve shaker 가
 shaking time 5 sieve
 U.S. standard testing sieve NO. 14, 45, 80, 100, 140, 170

(2) Caking action

Propolis caking 가
 가 cake 가 cake
 24 cake

(3) (mixability)
 가 chloride ion test(Quantab)
 가 CV 10

3.

	Saponin control	propolis control	saponin 가	NH ₃ saponin (P<0.05)
1	NH ₃	saponin propolis	가	2
NH ₃	가	3	가	가
4	2	NH ₃	가	가
3	NH ₃	,	가	가
sample	가		가	, NH ₃

Table 2-13. Effect of complexed mixture addition on ammonia gas emission from feces (unit : ppm)

Sampling week	Duration	Control	Low - complex	High- complex
2nd	1day	0.95 ± 0.19b	1.43 ± 0.38a	1.12 ± 0.25ab
	3days	1.16 ± 0.40	1.62 ± 0.48	1.06 ± 0.36
4nd	1day	0.53 ± 0.46	0.48 ± 0.50	0.43 ± 0.46
	3days	0.50 ± 0.35	0.18 ± 0.17	0.20 ± 0.14

* Value with different superscript differ(P<0.05)

Table 2-14. VOC profiles of swine manure stored 1day

VOCs	Elution area(mv.sec)	
	Low complex	High complex
Dimethyl sulfide	792.2	681.5
Methyl chloride	832.7	-
2- Methyl pentane	390.4	11.7
2- Methyl hexane	958.9	568.2
Dimethyl disulfide	462.3	363.3
Acetic acid	91.7	-
p- Xylene	65.3	-

Table 2-15. VOC profiles of swine manure stored 3days

VOCs	Elution area(mv.sec)	
	Low complex	High complex
Dimethyl sulfide	771.7	574.3
Methyl chloride	667.6	162.2
2- Methyl pentane	357.8	29.9
2- Methyl hexane	1082.4	557.3
Dimethyl disulfide	534.4	1132.6
Acetic acid	82.6	80.0
p- Xylene	63.8	41.9

Flame ionization detector(FID)

VOC flame photometric detector(FPD)

VOC S-

. VOC 가

dimethyl

sulfide, dimethyl disulfide, methyl chloride

가

low-complex high-complex 가

가

. low-complex 가

high-complex 가

VOC 가

high-complex deodorase 가

NH₃

Table 2-16

Table 2-16. Emissive NH₃ Content in the fecal sample (unit : ppm)

propolis and sole complex

	Feed mixture	Complex + pro	Complex
Dgw	839	227	82
Sgw	2.16	2.56	2.05
SA	73.0	312.5	714.8
P/G	18785	3518294	13699433

Dgw : Particle size(μm)

Sgw : Standard Dev

SA : Surface area(cm^2/gram)

P/G : Particles/gram

table 2-18

	839 μm	83.93%가	350 μm	gram
	73 cm^2 ,	gram	18785	.
propolis			227 μm	350 μm
		gram	312.5 cm^2 ,	88 μm
	3518294	.		82 μm
				sieve가
	88 μm	73.13%		82 μm
		.	gram	714.8 cm^2
gram	13699433		.	

table 2-19

propolis		silica	pro:silica가	4:1
(carrier)			가	
propolis		silica		
	zeolite	propolis	pro:zeolite가	2:1

1:1

Table 2-19. Caking index of propolis complex

Item	Zeolite	Silica			
Pro : Add					
1 : 1	A	A	B	B	A
2 : 1	A	A	B	B	B
3 : 1	C	A	C	C	C
4 : 1	C	A	C	C	C

A : No caking

B : Moderate caking

C : Extreme caking

table 2-20

CV 10%

Table 2-20. Mixing efficiency

	Low - complex	High - complex	DO- 50	DO- 100
CV(%)	7.13 ± 0.04	6.95 ± 0.02	6.88 ± 0.05	6.78 ± 0.08

4.

가. +propolis 가

가 가 .

. propolis 가 NH₃ 2
가 4
가

. NH₃ 0.7% 1.4% 가
가 50 80%

. H₂S 0.7% 1.4% 가

. +propolis 가
가 가
propolis가 가

. +propolis VOC VOC
가 Low High-complex

. 839μm, propolis

가 227 μ m 82 μ m ,
CV 10 .

. Cake 가 silica가 가
가 zeolite 가 cake
가 2:1 .

5.

Crober, D. 1991. Effect of De-odorase on ammonial levels in a deep pit cage layer house. P. 367-368. *In* T. P. Lyons(ed.), *Biotechnology in the feed industry*. Alltech, Inc., Nicholasville, Ky.

Giesy, R. G., B. Harris, Jr., J. G. Giesy and H. H. Van Horn, Jr. 1992. Effectiveness of De-odorase in reducing ammonia levels in daily barns during summer months, P. 16-18. *In* T. P. Lyons(ed.), *Biotechnology in the feed industry*. Alltech, Inc., Nicholasville, Ky.

Headon, D. R., K. Buggle., A. Nelson and G. Killeen. 1991. Glycofractions of the yucca plant and their role in ammonia control, P. 95-108. *In* T. P. Lyons(ed.), *Biotechnology in the feed industry*. Alltech, Inc., Nicholasville, Ky.

Jeon, B. S., J. H. Kwag., Y. H. Yoo., J. O. Cha and H. S. Park. 1996. Effect of feeding Enzymes, Probiotics or Yucca Powder on pig

Growth and Odor-Generating Substances in Feces. K. J. Anim. Sci.
38(1): 52-58.

Standefer, S. 1992. Biofiltration a new technology to minimize VOC
and odor emissions. PPC Biofilter. Long view. TX.

Tanaka, H., Kuroda, K. and Yonaga, M. 1992. Biological removal of VFA
from animal waste. J. Anim. Sci. Technol, (Jpn). 63(1): 54

3

propolis

가

1 Propolis

가

1.

가 ,
가
가
가 가

(Smith, 1962; Smith, 1975)

가 가

가 (Alder
Danassa, 1980; Barrow, 1992).

가

가

가

saponin

urease-activity

(Preston ,

1987) 가

가

(Goodall

Matsushine, 1978). Russell (1984)

, Johnston (1981)

Rowland Flyer(1976)

가 가 , Cronwell (1985)

.

가

가

, 가

가

propolis

.

propolis

가

. propolis

5km

가

(Meyer, 1956;

Cattorini, 1963), propolis

.

propolis

가

.

가

가

.

propolis

가 40 50%,

20 49%,

10%,

10%,

5%

(Ghisalberti, 1979, Konig Dustnan,

1989), propolis

,

,

,

,

,

,

,

.

propolis

propolis

(Harborne, 1982; Swain, 1977).

300

propolis

(Ghisalberti, 1979)

Propolis

()

가

(Meyer,

1956).

Propolis가

Karinovad(1961)

propolis

Lavie (1960),

Kivalkina(1969), Bolshakova(1964), Chernyak(1973)

Ikeno (1994), Matsuno(1992), Metzner (1979)

Takino Mochida(1982)

가

(Ghisalbert,

1979; Lindenfelser, 1979; Lavie, 1978).

propolis

Havsteen(1983),

Donadien(1987), Scheller (1988), Krol (1990)

propolis

propolis

propolis

(Grochowski , 1985),

(Christova, 1985)

, 가 (Scheller , 1988; Matsuno, 1992)

가 (Takahana, , 1984: Erben-Buss , 1987) 가
가 .

propolis 가 ,
propolis 가 가 가

, propolis 가
가

propolis 가 가

가
가 .

propolis 가 propolis

2.

가.

1997 3 15 4 26 6

, 5

10 5 31 3

500 , .

10 5 50
 10 500
 19.5%, 3,150 kcal/kg
 Table 3-1 가 가
 Table 3-2 Propolis

100% ethanol 48
 Whattnan filter paper rotary evaporator ethanol
 propolis
 30% ,
 가

1997 4 26 6 , 1997 3 15
 1997 4 26 6 , 3

1)

1

가

2)

1

3)

가

4) Abdominal fat pad

4

Abdominal fat pad

A. O. A. C(1995)

one way-ANOVA(analysis of variance)
(Steel Torrie, 1980)

Table 3-1. Feed formula of experimental diets

Ingredients	%
Corn	60.4
Wheat bran	0.1
Wheat flour	4.83
soybean meal	21.85
Rapeseed meal	3.80
Corn gluten meal	4.40
Yellow grease	0.55
Limestone	0.90
Tricalcium phosphate	1.95
Salt	0.11
HCl- lysin	0.17
DL- mehtionine	0.23
Vit.- Min. mixture	0.71
Total	100.0
Chemical composition	
Moisture, %	12.7
Crude protein,%	19.45
M e t a b o l i z a b l e	3134
Energy,kcal/kg	
Ether extracts, %	3.10
Crude fiber, %	3.50
Crude ash, %	5.60
Calcium, %	1.10
Phosphorus,%	0.70
Arginine, %	1.17
Lysin, %	1.04
Methionine, %	0.36

Table 3-2. Experimental design

Unit: PPM

Experimental diets	Treatments									
	A	B	C	D	E	F	G	H	I	J
Propolis		40	80	120	160					
Yucca extracts						150	300			
Antibiotics								5		
Arsonic acid									50	
Biominerals										50

3.

가.

propolis 가

(0 3), (4 6) (0 6)

Table 3-3, 3-4 3-5 .

Table 3-3 3 A

가 가 가 ,

propolis 160ppm 가 G 가 .

701 741g .

1,200g 1237g , 1.74

가 가 가 2 5% .

Table 3-4 1,448 1,473g

, propolis 120 160ppm 가 가

가 가 가 3,163g

2.16

가 2.16 2.17, propolis 가 2.11 2.07 4%

6 Table 3-5 2,170g

가 가 2,182g propolis

40ppm 80ppm 2,182g, 120ppm

2,199g 160ppm 2,192g .

2,176-2,175g

4,381g 가 가 가 4,239 4,295g
 . propolis 가 , 가
 . 2.03 가 가
 가 1.98 1.93 2 5% , propolis
 160ppm 가 G 가
 .
 가 가 가
 , (Rowland Plyer,
 1976) (Hays, 1978; Hawbaker , 1960; Powley , 1981)
 가
 . propolis 가
 가 . propolis
 .
 가 . propolis
 .

Table 3-3. Effect of feeding ethanol extract propolis, yucca extracts

and antibiotics on body performance of broiler chicken by starting periods(0 3weeks)

Treatments	Body gain(g)	Feed intake(g)	Feed efficiency
A	701 ± 25	1218 ± 29	1.74 ± 0.03
B	712 ± 12	1222 ± 13	1.72 ± 0.02
C	729 ± 16	1237 ± 19	1.70 ± 0.02
D	708 ± 19	1201 ± 26	1.70 ± 0.02
E	716 ± 14	1223 ± 7	1.71 ± 0.03
F	717 ± 7	1201 ± 17	1.68 ± 0.03
G	741 ± 13	1231 ± 19	1.66 ± 0.02
H	714 ± 15	1208 ± 20	1.69 ± 0.03
I	712 ± 13	1216 ± 6	1.71 ± 0.04
J	712 ± 14	1202 ± 29	1.69 ± 0.01

* There were no significant($p < 0.05$) difference among all treatments.

* Mean ± SE.

Table 3-4. Effect of feeding ethanol extract propolis, yucca extracts and antibiotics on body performance of broiler chicken by finishing periods(4 6weeks)

Treatments	Body gain(g)	Feed intake(g)	Feed efficiency
A	1461 ± 13	3163 ± 41	2.16 ± 0.02
B	1466 ± 17	3027 ± 56	2.06 ± 0.02
C	1458 ± 32	3021 ± 53	2.07 ± 0.05
D	1448 ± 11	3057 ± 49	2.11 ± 0.04
E	1463 ± 61	3072 ± 49	2.11 ± 0.06
F	1468 ± 19	3047 ± 31	2.08 ± 0.05
G	1473 ± 26	3051 ± 48	2.07 ± 0.03
H	1456 ± 24	3031 ± 32	2.08 ± 0.02
I	1465 ± 27	3037 ± 36	2.07 ± 0.04
J	1463 ± 28	3064 ± 31	2.10 ± 0.04

* There were no significant($p < 0.05$) difference among all treatments.

* Mean ± SE.

Table 3-5. Effect of feeding ethanol extract propolis, yucca extracts

and antibiotics on body performance of broiler chicken.

Treatments	Body gain(g)	Feed intake(g)	Feed efficiency
A	2170 ± 26	4381 ± 59	2.03 ± 0.01
B	2182 ± 27	4249 ± 66	1.95 ± 0.01
C	2172 ± 44	4258 ± 67	1.95 ± 0.04
D	2167 ± 23	4258 ± 71	1.97 ± 0.03
E	2182 ± 73	4295 ± 56	1.98 ± 0.04
F	2199 ± 24	4248 ± 44	1.95 ± 0.04
G	2192 ± 35	4282 ± 63	1.93 ± 0.01
H	2174 ± 34	4239 ± 45	1.95 ± 0.02
I	2176 ± 36	4253 ± 30	1.95 ± 0.03
J	2175 ± 29	4266 ± 36	1.96 ± 0.03

* There were no significant($p < .05$) difference among all treatments.

* Mean ± SE.

propolis 가

propolis

가 가

가 가

가 , propolis 가 , ,

가 .

Propolis

가

, , ,

Table 6 .

72.7 73.3%, 20.6 21.1%, 4.

4 4.9%

Table 3-6. Effect of feeding propolis, yucca extracts and antibiotics

on carcass composition of broiler

Treatment	Carcass composition, %			
	Abdominal fat	Moisture	Protein	Fat
	pad (% of body wt.)			
A	2.12 ±0.11	73.7 ±0.23	20.6 ±0.14	4.6 ±0.17
B	2.39 ±0.14	73.4 ±0.29	20.6 ±0.16	4.8 ±0.19
C	2.16 ±0.14	72.9 ±0.38	21.0 ±0.10	4.9 ±0.12
D	2.47 ±0.16	72.8 ±0.18	21.0 ±0.17	4.7 ±0.17
E	2.36 ±0.11	72.7 ±0.30	20.7 ±0.18	4.8 ±0.15
F	2.32 ±0.15	73.0 ±0.17	20.9 ±0.11	4.4 ±0.17
G	2.22 ±0.11	72.9 ±0.29	21.0 ±0.12	4.9 ±0.11
H	2.47 ±0.15	73.2 ±0.30	21.0 ±0.13	4.3 ±0.12
I	2.32 ±0.15	72.9 ±0.39	21.1 ±0.16	4.6 ±0.17
J	2.47 ±0.14	73.6 ±0.31	20.7 ±0.18	4.6 ±0.15

* There were no significant(p<.05) difference among all treatments.

* Mean ± SE.

2.12 2.47%

가

4.

propolis

가가

propolis 40, 80,

120ppm

160ppm

150

300ppm

가

6

가

2, 170g 가 가 가 2, 174 2, 199g
, 4, 381g
가 가 가 4, 239 4, 295g
. propolis 가
. 2. 03 가
가 가 1. 93 1. 98 .
가 .
propolis 가 ,
.

5.

1. Alder, H. E. and Damassa, A. J. 1980. Effect of ingested Lactobacilli on Salmonella infantis and Escherichia coli and on intestinal flora, pasted vents and chick growth. Avian Disease 24:868.
2. A. O. A. C. 1990. Official methods of analysis(15th ed.). Association of Official Analytical Chemists. Washington. D. C.
3. Barrow, P. A. 1992. Probiotics for chickens. In R. Fuller(Ed.). Probiotics: The Scientific basis. Chapman & Hall, London.
4. Bolshakova, V. F., I. V. Vinogradova, 1964. Experience in the use of propolis salicylate ointment. Nauch. Zap. Gor 'kovskogo Nauchno-issled. Inst. Derm. Vener. 24:148.
5. Cattorini, P. E. 1963. Le API e glib uonini [Bees and Men]. Fitoterapia 34: 85.
6. Chernyak, N. F. 1973. On synergistic effect of propolis and some

anti-bacterial drugs. *Antibiotiki* 18:259.

7. Christova, V. M. 1985. A Propolis-enriched was therapy against inflammatory diseases of the articular-muscular system. The XXXth Internation Apicultural Congress. pp. 429-431. Apinondia.
8. Cronwell, G .L., T .S. Stahly and J. J. Nonogue. 1985. Efficacy of sarsaponin for weanling and growing-finishing swine housed at two animal densities. *J. Anim.* 61(suppl.1.):111.
9. Donadieu, Y. 1987. Propolis in natural therapeutics. *Honey bee Science* 82:67.
10. Erben-Russ, M., W. Bors and M. Saran. 1987. Reaction of linoleic and peroxy radicals with phenolic antioxidants pulse radiolysis study. *International Journal of Radiation Biology.* 52:393.
11. Ghisalberti, E.L. 1979. Propolis: A review. *Bee World* 60:59.
12. Goodall, S. R. and J. K. Natsushina. 1978. Sarsaponin in beef cattle rations. Pages 9-10 in *Beef nutrition research. Gen. Ser.* 979. Colorado State Univ. Exp. Sta.
13. Grochowsk, J., M. Bilinska, and D. Stankiewicz, (Poland). 1985. The therapeutic effect of 3% propolis treatment on mice of DBA/2H breed suffering from skin burns subsequently infected with *Pseudomonas aeruginosa*. The XXXth Internation Apicultural Congress. pp. 439-440. Apinondia.
14. Harborne, J. B. 1982. *Introduction to Ecological Biochemistry.* Academic Press. London.
15. Havsteen, B. 1983. Flavonoids, a class of natural products of high harm acological potency. *Biochenical Pharnacology.* 32: 1141.
16. Ikeno, K., T. Ikeno and T. Miyazawa. 1994. Effects of propolis

on dental caries in rats. Honeybee Science 15: 1.

17. Johnston, N. L., C. L. Quarles, D.J. Fagerberg, and D.D. Caveny. 1981. Evaluation of yucca saponin on broiler performance and ammonia suppression. Poultry Sci., 60: 2289.
18. Karinova, Z.K.H. 1961. About the medicinal qualities of propolis. Pchelovodstvo. 38;32. In Russian.
19. Kivalkina, V.P. 1969. Effect of propolis on immunological reactivity. 22 Int. Beekeep. Congr. Summ.: 136 In Russian
20. Krol, W., Z. Szuba, S. Scheller, J. Gabrys, S. Grabiec and J. Shani. 1990. Anti-oxident property of ethanolic extract of propolis (EEP) as evaluated by inhibiting the chemiluminescence oxidation of luminol. Vol. 21. No4. pp. 593-597.
21. Lavie, P. 1960. Les substances antibactériennes dans la colonie d'abeilles (*Apis mellifica* L.) Anmls. Abeille 3: 103.
22. Lavie, P. 1978. The antibiotic from propolis, Propolis, pp. 41-48. Apinondia Publishing House.
23. Mayer, W. 1956. "Propolis bees" and their activities. Bee World 37: 25.
24. Matsuno, T. 1992. Isolation and character isolation of the tumoricidal substances from Brazilian propolis. Honey bee Science 13: 49.
25. Netzner, J., H. Bekeneter, M. Paintz and E. Schneidewind. 1979. Pharmazie 34: 97.
26. Preston, R. L., S. J. Bartle, T. May and S. R. Goodall. 1987. Influence of sarsaponin on growth, feed and nitrogen utilization in growing male rats fed diets with added urea or protein. J. Anim. Sci. 65: 481.

27. Rowland, I., O. Jr., J. E. Plyler, and J. W. Bradley. 1976. Yucca schidigera extract effect on egg production and house ammonia levels. Poultry Sci., 55:2086.
28. Russel, J. B. 1984. The effect of sarsaponin on ammonia levels in rodent cages. District & Convention Amer. Assoc. Lab. Anim. Sci. (abstr.).
29. Scheller, S., G. Gazda, J. Gabrys, J. Szunias, L. Eckert and J. Shani, 1988. The ability of EEP to accumulate plaque formation in immunized mouse spleen cells. Pharmacological Research Communication. 20: 323.
30. Smith, H. W., 1962. The effects of the use of antibiotics on the emergence of antibiotic resistant disease producing organism in animals. Antibiotics in Agriculture. Proceedings of the University of Nottingham. Ninth Easter School in Agriculture Science. Butterworth. London. p 374.
31. Smith, H. W., 1975. Persistence of tetracycline resistance in pig E. coli Nature : 258 : 628.
32. Steel, R. G. D. and J. H. Torrie. 1980. Principles and procedures of statistics. A Biometrical Approach. McGraw-Hill, New York. pp. 223.
33. Swain, T. 1977. Secondary compounds as protective agents. Annual review of plant physiology. 28:479.
34. Takahana, U., R. J. Youngnan, and E. F. Elstner. 1984. Transformation of quercetin by singlet oxygen generated by photosensitized reaction. Photobiocchemistry and Photobiophysics. 7:175.
35. Takino, Y. and S. Mochida. 1982. Propolis, its chemical cons-

tituents and biological activities. Honeybee Science 3: 145.

2 Propolis

1.

가 ,
 , 가
 . 가
 가 가
(Smith, 1962; Smith, 1975)
propolis
 .
Propolis
 가 , propolis
 가
(Meyer, 1956; Cattorini, 1963),
propolis .
propolis
 가 .
가 가 . Propolis
가 40 50%, 20 49%, 10%, 10%,
5% (Ghisalberti, 1979, Konig Dustnan, 1989),
propolis , , , ,
 , , , .
propolis

propolis

(Harborne, 1982; Swain, 1977).

Propolis가

Karinovad(1961)

propolis

Lavie (1960),

Ki valkina(1969), Bolshakova(1964), Chernyak(1973)

Lindenfelser,

(1967)

Ikeno

(1994), Matsuno(1992),

Metzner (1979)

Takino

Nochi da(1982)

가

(Ghisalbert, 1979; . Lavie,

1978).

propolis

Havsteen(1983),

Donadien(1987), Scheller

(1988), Krol

(1990)

propolis

propolis

propolis

(Grochowski , 1985),

(Christova, 1985)

가 (Scheller , 1988, Matsuno, 1992)

가 (Takahana, , 1984:

Erben-Buss , 1987)

가

가

propolis

가

propolis

가

가

가

propolis

가가 가

100, 200, 300, 400ppm

500ppm

가

2.

가.

1997 9 27 11 8 6

97 9 25 280

7 4 28

10 280

20%,

3, 050kcal

, propolis

propolis

100, 200, 300, 400ppm

500ppm

가

가

1

10ppm

가

Table 3-7

1)

가 1 ,

2)

1 ,

3)

가

one way-ANOVA(analysis of variance)
(Steel Torrie, 1980).

Table 3-7. Feed formula of experimental diets

Ingredients	Control	A	B	C	D	E	F
Corn	59.7	59.7	59.7	59.7	59.7	59.7	59.7
Wheat bran	3.0	3.0	2.99	2.98	2.97	2.96	2.95
Soy bean meal	24.8	24.8	24.8	24.8	24.8	24.8	24.8
Rape seed meal	2.3	2.3	2.3	2.3	2.3	2.3	2.3
Fish meal	4.6	4.6	4.6	4.6	4.6	4.6	4.6
Yellow grease	2.55	2.55	2.55	2.55	2.55	2.55	2.55
Limestone	0.60	0.60	0.60	0.60	0.60	0.60	0.60
TCP(%)	1.50	1.50	1.50	1.50	1.50	1.50	1.50
Salt	0.10	0.10	0.10	0.10	0.10	0.10	0.10
Vit.- Min. mix.	0.45	0.45	0.45	0.45	0.45	0.45	0.45
HC - Lysin, 98%	0.06	0.06	0.06	0.06	0.06	0.06	0.06
D - Meth.50%	0.34	0.34	0.34	0.34	0.34	0.34	0.34
Antibiotics,ppm		10					
Propolis,ppm			100	200	300	400	500
Total	100.0	100.0	100.0	100.0	100.0	100.0	100.0
C h e m i c a l composition							
Crude protein, %	20.3	20.3	20.3	20.2	20.1	20.1	20.3
ME, kcal/kg	3055	3055	3055	3050	3048	3045	3055
Calcium, %	1.0	1.0	1.0	1.0	1.0	1.0	1.0
Phosphorus, %	0.7	0.7	0.7	0.7	0.7	0.7	0.7

3.

가.

propolis

1

,

(0 3),

(4 6)

(0 6)

Table

3-8, 3-9 3-10 .

Table 3-8 3

681g

propolis 가 가 681-713g

가

propolis 300ppm 가 D

가 . 1,186g

가 가 1,172-1,207g 가

. 1.74 propolis 가 가

, 300ppm 가 D 가 1.66 가

Table 3-9

1,257 1,330g

1257g

propolis 300ppm

가

D, E, Frn

가

가 A 1,265g

propolis 가 가

2.34

2.32-2.24

1-5%

propolis 300ppm

가

D, E F 가

6

Table 3-10

1,937g

가 가 1,967g

, propolis

100ppm 가 1,986g, 500ppm 가 F 1,993g, 200ppm 가

C 1999g, 400ppm 가 E 2021g, 300ppm 가 D

2041g 가 (p<.05). propolis

300ppm 가 가 300ppm

300ppm

Table 3-8. Effect of Propolis extracts on growth performance of

broiler chicks by starting period(0-3weeks)

Treatments	Body gain (g)		Feed intake (g)		Feed / Gain	
Control	681	± 14	1186	± 10	1.74	± 0.02
A	702	± 10	1178	± 10	1.68	± 0.01
B	688	± 5	1172	± 11	1.70	± 0.03
C	707	± 14	1207	± 12	1.71	± 0.02
D	713	± 8	1185	± 23	1.66	± 0.01
E	691	± 10	1198	± 23	1.73	± 0.01
F	681	± 7	1178	± 17	1.73	± 0.01

Means in the same column with different superscripts differ (p<.05). Mean ± SE

Table 3-9. Effect of Propolis extracts on growth performance of broiler chicks by finishing period(4-6weeks)

Treatments	Body gain (g)		Feed intake (g)		Feed / Gain	
Control	1257c	± 7	2941	± 10	2.34a	± 0.01
A	1265tc	± 26	2941	± 36	2.33a	± 0.02
B	1298atc	± 14	3009	± 5	2.32a	± 0.03
C	1292atc	± 6	2975	± 21	2.30ab	± 0.01
D	1328a	± 20	2969	± 10	2.24b	± 0.03
E	1330a	± 8	2983	± 12	2.24b	± 0.02
F	1312ab	± 4	3005	± 10	2.29ab	± 0.01

Means in the same column with different superscripts differ (p<.05). Mean ± SE

Table 3-10. Effect of Propolis extracts on growth performance of

broiler chicks

Treatments	Body gain (g)		Feed intake (g)		Feed / Gain	
Control	1937d	± 20	4127	± 20	2.13a	± 0.01
A	1967cd	± 16	4119	± 26	2.09ab	± 0.01
B	1986bc	± 9	4181	± 6	2.11a	± 0.01
C	1999abc	± 8	4182	± 10	2.09ab	± 0.01
D	2041a	± 12	4154	± 13	2.04c	± 0.02
E	2021ab	± 18	4181	± 11	2.07bc	± 0.01
F	1993abc	± 11	4183	± 8	2.10ab	± 0.01

Means in the same column with different superscripts differ (p < .05). Mean ± SE

propolis 300ppm 가 가 A
 3.7% . 4127g
 propolis 가 가 4154-4183g propolis
 가 .
 2.13 가 가 2.09 , propolis 가
 B 2.11 D 2.04 propolis 가
 (p < .05) . 300ppm 가 D
 2.04 가 .
 propolis 가 300ppm
 가 가
 . 500pp 가
 . 가
 .
 propolis (Karinovad,
 1961) (Natsuno, 1992; Taki no Mochi da, 1982)

가 .

300ppm

propolis ,

가 .

4.

propolis 가가

propolis 100, 200, 300, 400ppm

500ppm 가 6

가 ,

1937g propolis 가 가 1,986

2,041g (p<.05) 가 ,

300ppm 가 D 가 .

4,127g propolis 가 가 4,154 4,183g

. 2.13

propolis 가 2.11 2.04 (p<.05)

, D 가 .

propolis

가 가 .

5.

1. Bolshakova, V. F., I. V. Vinogradova, 1964. Experience in the use of propolis salicylate ointment. Nauch. Zap. Gor 'kovskogo Nauchno-issled. Inst. Derm. Vener. 24:148.
2. Cattorini, P. E. 1963. Le API e glib uonini [Bees and Men]. Fitoterapia 34: 85.
3. Chernyak, N. F. 1973. On synergistic effect of propolis and some anti-bacterial drugs. Antibiotiki 18:259.
4. Christova, V. M. 1985. A Propolis-enriched was therapy against inflammatory diseases of the articular-muscular system. The XXXth Internation Apicultural Congress. pp. 429-431. Apinondia.
5. Donadieu, Y. 1987. Propolis in natural therapeutics. Honey bee Science 82:67.
6. Erben-Russ, M., W. Bors and M. Saran. 1987. Reaction of linoleic and peroxy radicals with phenolic antioxidants pulse radiolysis study. International Journal of Radiation Biology. 52:393.
7. Ghisalberti, E.L. 1979. Propolis: A review. Bee World 60:59.
8. Grochowsk, J., M. Bilinska, and D. Stankiewicz, (Poland). 1985. The therapeutic effect of 3% propolis treatment on mice of DBA/2H breed suffering from skin burns subsequently infected with *Pseudomonas aeruginosa*. The XXXth Internation Apicultural Congress. pp. 439-440. Apinondia.
9. Harborne, J. B. 1982. Introduction to Ecological Biochemistry. Academic Press. London.
10. Havsteen, B. 1983. Flavonoids, a class of natural products of high harm acological potency. Biochemical Pharnacology. 32: 1141.

11. Ikeno, K., T. Ikeno and T. Miyazawa. 1994. Effects of propolis on dental caries in rats. *Honeybee Science* 15: 1.
12. Karinova, Z. K. H. 1961. About the medicinal qualities of propolis. *Pchelovodstvo*. 38; 32. In Russian.
13. Kivalkina, V. P. 1969. Effect of propolis on immunological reactivity. *22 Int. Beekeep. Congr. Summ.*: 136 In Russian
14. Krol, W., Z. Szuba, S. Scheller, J. Gabrys, S. Grabiec and J. Shani. 1990. Anti-oxident property of ethanolic extract of propolis (EEP) as evaluated by inhibiting the chemiluminescence oxidation of luminol. Vol. 21. No4. pp. 593-597. 20.
15. Konig, B and J. H. Dustman. 1989. *Anim Res. and Develop* 29: 21.
16. Lavie, P. 1978. The antibiotic from propolis, PROPOLIS, pp. 41-48. Apinondia Publishing House.
17. Lindenfelser, L. A. 1967. Antimicrobial activity of propolis. *American Bee Journal*. 107: 90.
18. Mayer, W. 1956. "Propolis bees" and their activities. *Bee World* 37: 25.
19. Matsuno, T. 1992. Isolation and character isolation of the tumoricidal substances from Brazilian propolis. *Honey bee Science* 13; 49.
20. Metzner, J., H. Bekeneter, M. Paintz and E. Schneidewind. 1979. *Pharnazie* 34: 97.
21. Scheller, S., G. Gazda, J. Gabrys, J. Szumias, L. Eckert and J. Shani, 1988. The ability of EEP to accumulate plaque formation in immunized mouse spleen cells. *Pharmacological Research Communication*. 20: 323.
22. Smith, H. W., 1962. The effects of the use of antibiotics on the

emergence of antibiotic resistant disease producing organism in animals. Antibiotics in Agriculture. Proceedings of the University of Nottingham. Ninth Easter School in Agriculture Science. Butterworth. London. p 374.

23. Smith, H.W., 1975. Persistence of tetracycline resistance in pig *E. coli* Nature : 258 : 628.
24. Steel, R. G. D. and J. H. Torrie. 1980. Principles and procedures of statistics. A Biometrical Approach. McGraw-Hill, New York. pp. 223.
25. Swain, T. 1977. Secondary compounds as protective agents. Annual review of plant physiology. 28:479.
26. Takahana, U., R. J. Youngnan, and E. F. Elstner. 1984. Transformation of quercetin by singlet oxygen generated by photosensitized reaction. Photobiocchemistry and Photobiophysics. 7:175.
27. Takino, Y. and S. Mochida. 1982. Propolis, its chemical constituents and biological activities. Honeybee Science 3: 145.

3 Propolis

가가

1.

Propolis

. 가
.
, 가 (Harvstten, 1983;
Donadien, 1987; Lavis, 1960; Ikeno , 1994; Matsuno, 1992;
Ghisalbet, 1979; Krol, 1990; Kivalkina, 1969; Taki no Mochida,
1982). propolis ,
가 (Margo Carvallo,
1994, Ivanovska , 1995). Propolis
가 150
가

Propolis

가 .
가 .
가 propolis 가
.
가 가
가

가 propolis propolis

.

2.

가. Propolis 가가

(1)

600 kg 가 3 propolis

가 300 ppm .

11.5%, TDN 71.5%, 0.8% 0.5%

2%()

30:70 .

(2)

3

1 2

2kg

8kg , .

(3)

(가) pH

가 0, 1, 2, 3, 4,

5, 6, 7 8 4 가
 20-30nl pH/Ion neter)
 pH .
 ()
 15nl 20nl 15000g, 4
 15 2nl
 (total amino
 acid: TAA) auto analyzer(Lachat instrument co.,)
 phenol reagent, hypochlorite
 reagent, buffer nitroprusside reagent quikchem
 method(Lachat instrument) hot bath
 60 , 630nm ,
 ninhydrin , hydrazin sultate hot
 bath 135 , cooling bath 10 570nm

()
 0, 2, 4, 6, 8
 500nl 4 가
 50% 50: 1 가 -20
 (crude protein) ,
 100nl

. Propolis

가가

(1)

600kg 9 3 x 3
 Latin square design 3 3
 2 6 . propolis
 300ppm 가 17.5%
 TDN 71.5% Table
 3-11 .

(2)

. 1 2
 , 40% ,
 . 3
 , .
 .

(3)

2 1
 , 41ng potassium dichronate
 . Kjeldahl
 , infrared milk analyzer
 .

Table 3-11. Formulae of experimental diets

Ingredient	Control	Treatment	
		Yucca 1)	Propolis 2)
Corn	29.60	29.60	29.60
Wheat	5.00	5.00	5.00
Wheat bran	11.80	11.77	11.77
Rice bran	5.00	5.00	5.00
Gluten feed	6.00	6.00	6.00
Corn germ meal	8.00	8.00	8.00
Corn gluten meal	0.00	0.00	0.00
Soybean hull	4.20	4.20	4.20
Soybean meal	11.00	11.00	11.00
Rapeseed oil meal	7.00	7.00	7.00
Cottonseed oil meal	4.00	4.00	4.00
Yellow grease	1.80	1.80	1.80
Molasses	3.00	3.00	3.00
Limestone	1.60	1.60	1.60
Calcium- phosphate	0.80	0.80	0.80
Salt	0.50	0.50	0.50
NaHCO ₃ - MgO mixture	0.44	0.44	0.44
Vit.- Min.mixture	0.25	0.25	0.25
Yucca shidigera extracts	0.00	0.03	0.00
Propolis	0.00	0.00	0.03
Total	100.0	100.0	100.0
Chemical composition(%)			
Crude protein	17.58	17.58	17.58
TDN ³⁾	71.56	71.56	71.56
Calcium	1.02	1.02	1.02
Phosphorus	0.77	0.77	0.77

1) Yucca = yucca shidigera extracts 300 ppm

2) Propolis = propolis 300 ppm.

3) TDN = total digestible nutrients

Table 3-12. Feed formula of experimental diets

Ingredient	Control	Treatment	
		Yucca1)	Propolis2)
Corn	31.00	31.00	31.00
Milo	11.00	11.00	11.00
Wheat	7.00	7.00	7.00
Wheat bran	21.00	21.00	21.00
Rice bran	5.17	5.17	5.17
Gluten feed	4.90	4.90	4.90
Tapioka	15.00	15.00	15.00
Yellow grease	0.40	0.40	0.40
Limestone	1.95	1.95	1.95
Calcium- phosphate	0.25	0.25	0.25
Salt	0.50	0.50	0.50
Urea	0.80	0.80	0.80
NaHCO ₃ - MgO mixture	0.40	0.40	0.40
Vit.- Min.mixture	1.00	0 1.00	0 1.00
Yucca shidigera extracts	0.00	0.00	0.00
Propolis	0.00	0.00	0.00
T total	100.0	100.0	100.0
Chemical composition(%)			
Crude protein	11.70	11.70	11.70
TDN ³⁾	70.48	70.48	70.48
Calcium	0.84	0.84	0.84
Phosphorus	0.54	0.54	0.54

1) Yucca = yucca shidigera extracts 300ppm

2) Propolis = propolis 300ppm

3) TDN = total digestible nutrients

() 가

12

가

(, 1993) 가 .

Table 3-13. Total amino acid concentration in the rumen fluid after feeding (unit: ng/100nl)

Time(hr)	Control	Treatments	
		A 1)	B 2)
0	11.7 ± 1.40	10.8 ± 0.50	10.4 ± 1.25
1	38.1 ± 5.98	35.6 ± 7.07	29.2 ± 1.08
2	26.6 ± 5.49	25.7 ± 5.85	21.7 ± 0.71
3	18.6 ± 1.57	17.7 ± 2.36	12.1 ± 1.15
4	11.1 ± 1.80	9.2 ± 0.48	7.2 ± 0.49
5	7.7 ± 0.66	7.4 ± 1.16	5.6 ± 2.48
6	6.0 ± 1.55	5.8 ± 0.09	6.2 ± 1.22
7	6.4 ± 1.57	5.9 ± 0.03	5.4 ± 0.09
8	7.8 ± 1.06	5.3 ± 0.59	5.3 ± 1.15
Average	14.9 ± 2.12	13.9 ± 1.96	11.7 ± 0.96

1) A = yucca shidigera extracts 300 ppm

2) B = propolis 300 ppm

There were no significantly(p<.05) difference among all treatments

Mean ± SE

, 13.9ng/100nl , A 가 13.4ng/100nl, B 가
10.4ng/100nl propolis 가

8 1.7 - 3.2 ng/100nl

pH (Table 3-15) 6.92, A 6.71, B
6.93 propolis 가 pH

Table 3-16

2

51.5

51.0

A

53.4, B

Table 3-14. Ammonium concentration in the rumen fluid after feeding

Time(hr)	control	Treatments	
		A 1)	B 2)
0	4.9 ± 0.20	4.9 ± 0.17	6.2 ± 0.74
1	13.9 ± 0.56	13.4 ± 1.66	10.4 ± 0.16
2	12.3 ± 1.15	13.9 ± 1.75	10.4 ± 0.17
3	9.5 ± 0.08	9.54 ± 1.29	8.5 ± 0.07
4	4.8 ± 1.12	4.5 ± 0.64	5.6 ± 0.71
5	2.8 ± 1.67	1.7 ± 0.57	2.7 ± 0.14
6	1.8 ± 0.25	1.0 ± 0.06	4.6 ± 0.51
7	2.0 ± 0.21	0.5 ± 0.09	3.4 ± 0.16
8	2.9 ± 0.50	1.7 ± 0.02	3.2 ± 0.63
Average	6.1 ± 0.64	5.7 ± 0.70	6.1 ± 0.36

1) A = yucca shidigera extracts 300 ppm

2) B = propolis 300 ppm

Mean ± SE. There were no significantly(p<.05) difference among all treatments

4, 6 8

propolis 가 pH,

가

가

Table 3-15. Change of rumen pH after feeding

Time(hr)	control	Treatments	
		A1)	B2)
0	7.00 ± 0.05	6.75 ± 0.42	6.98 ± 0.25
1	6.87 ± 0.05	6.83 ± 0.33	7.06 ± 0.09
2	6.95 ± 0.02	6.69 ± 0.27	7.05 ± 0.08
3	6.87 ± 0.06	6.61 ± 0.21	6.82 ± 0.25
4	6.90 ± 0.01	6.70 ± 0.22	6.84 ± 0.11
5	6.83 ± 0.01	6.68 ± 0.26	6.92 ± 0.15
6	6.89 ± 0.01	6.66 ± 0.30	7.05 ± 0.17
7	6.95 ± 0.05	6.77 ± 0.26	6.85 ± 0.22
8	7.05 ± 0.06	6.74 ± 0.26	6.77 ± 0.09
Average	6.92 ± 0.03	6.71 ± 0.28	6.93 ± 0.19

1) A = yucca shidigera extracts 300 ppm

2) B = propolis 300 ppm

Mean ± SE.

There were no significantly ($p < .05$) difference among all treatments

Table 3-16. Rumens microbial protein concentration after feeding

Time(hr)	control	Treatments	
		A1)	B2)
0	16.7 ± 1.20	15.8 ± 0.97	16.4 ± 1.48
2	51.0 ± 2.81	53.4 ± 4.21	51.5 ± 2.89
4	49.7 ± 3.81	50.2 ± 2.97	52.2 ± 2.99
6	34.0 ± 1.80	34.1 ± 3.19	37.9 ± 1.37
8	32.7 ± 3.52	33.4 ± 3.79	33.6 ± 3.35
Average	36.8 ± 2.63	37.4 ± 3.03	38.3 ± 2.42

1) A = yucca shidigera extracts 300 ppm

2) B = propolis 300 ppm

Mean ± SE. There were no significantly(p<.05) difference among all treatments

. Propolis 가가

Propolis

Table 3-17 3-18 . 1

15.43kg, YSE 가 15.48kg propolis 15.36kg

가

7.18-7.9kg

8.18-8.19kg

18.25kg

YSE propolis 가 18.39 18.36kg

가

3.42 3.45% , 3.41 3.42%

1. 18 YSE 1. 19 propolis 가 1. 20
 . 1. 51x10
 4 1. 52x104 /nl , 11. 6x04 11. 91x104
 . 1
 가 ,
 . 가

Table 3-17. Effect of feeding yucca shidigera extract(YSE) and propolis on dry matter intake of Holstein cows

Item	Treatments		
	Control	YSE	Propolis
	Mean SEM1)	Mean SEM	Mean SEM
Feed Intake, kg/cow/day	7.25 ± 0.26	7.29 ± 0.30	7.18 ± 0.32
Concentrate			
Silage	8.18 ± 0.38	8.19 ± 0.42	8.18 ± 0.46
Total DMI2)	15.43 ± 0.64	15.48 ± 0.72	15.36 ± 0.74

1) SEM = standard error of mean

2) DM = dry matter intake

There were no significantly(p<.05) difference among all treatments

Table 3-19. Effect of feeding yucca shidigera extract(YSE) and propolis extract on body performance of Hanwoo during finishing period

Item	Treatments					
	Control		YSE		Propolis	
	Mean	SE ¹⁾	Mean	SE	Mean	SE
Feed intake(kg)						
Concentrate	9.51	± 0.12	9.52	± 0.13	9.66	± 0.18
Rice straw	1.50	± 0.01	1.47	± 0.01	1.51	± 0.02
TDMI ²⁾	11.01	± 0.13	10.99	± 0.13	11.16	± 0.20
ADG ³⁾ (kg)	1.05	± 0.05	1.04	± 0.05	1.07	± 0.06
DMI per gain ⁴⁾ (kg/kg)	10.52	± 0.39	10.68	± 0.43	10.49	± 0.39

1) SEM = standard error of mean 2) TDMI = total dry matter intake

3) ADG = average daily gain 4) DMI = dry matter intake per gain

There were no significantly(p<.05) difference among all treatments

10.99kg propolis 11.165kg
 10.52 10.68 propolis
 10.49 가
 propolis 가

Table 3-20 3
 347 367kg (BFI)
 8.8 9.5mm 가 propolis 가 가
 (REA)

79cm² propolis 가 77.3 가 75.6cm²
가 가 가 .

68.0 68.7 cm .

, 3.25 가 4.0
propolis 가 4.38
가 .

4.25 4.50 , 2.50 2.75
가

Table 3-21 .

68% (1995) 21

71% .

三橋 (1987)

23

(1995)

. 20.1%, 20.2 20.5% 가

, 7.5% 8.2% , 1.3 1.45%

가 . (1995)

21

가 (

Anderson , 1984)

가 , 가

4

가 가

Table 3-20. Effect of feeding yucca shidigera extract(YSE) and propolis extract on carcass traits during finishing period in Korean native bulls.

Item	Treatments		
	Control	YSE	Propolis
	Mean SE1)	Mean SE	Mean SE
Slaughter weight, kg	575 ± 19	585 ± 9	615 ± 19
Yield traits			
Carcass, weight, kg	347 ± 12	349 ± 6	367 ± 10
BFT2), mm	8.8 ± 0.6	8.8 ± 1.1	9.5 ± 0.06
REA3), Cm2	79.0 ± 2.3	75.6 ± 2.4	77.3 ± 4.6
MPI4), score	68.7 ± 0.2	68.7 ± 0.5	68.0 ± 0.9
Quality traits			
MS5), score	3.25 ± 0.48	4.00 ± 0.41	4.38 ± 0.99
MC6), score	4.25 ± 0.25	4.50 ± 0.50	4.25 ± 0.25
FC7), score	2.50 ± 0.29	2.75 ± 0.25	2.75 ± 0.25

1) SE = standard error

2) BFT= back fat thickness(mm)

3) REA= rib eye area (cm)

4) MPI= neat production index

5) MS = marbling score

6) MC = neat color 7) FC = fat color

Table 3-21. Effect of feeding yucca shidigera extract(YSE) and propolis extract on chemical composition of strip loin.

Item	Treatments		
	Control	YSE	Propolis
			%
Moisture	68.25	67.88	67.60
Crude protein	20.45	20.48	20.10
Ether extract	8.91	9.39	9.66
Crude ash	1.42	1.39	1.44

4.

Propolis

300ppm

propolis

pH

18. 25kg

가

propolis

가

18. 39

18. 36kg

가

5.

1. Bolshakova, V. F., I. V. Vinogradova, 1964. Experience in the use of propolis salicylate ointment. Nauch. Zap. Gor 'kovskogo Nauchno-issled. Inst. Derm. Vener. 24:148.
2. Cattorini, P. E. 1963. Le API e glib uonini [Bees and Men]. Fitoterapia 34: 85.
3. Chernyak, N. F. 1973. On synergistic effect of propolis and some anti-bacterial drugs. Antibiotiki 18:259.
4. Christova, V. M. 1985. A Propolis-enriched was therapy against inflammatory diseases of the articular-muscular system. The XXXth Internation Apicultural Congress. pp. 429-431. Apinondia.
5. Donadiou, Y. 1987. Propolis in natural therapeutics. Honey bee Science 82:67.
6. Erben-Russ, M., W. Bors and M. Saran. 1987. Reaction of linoleic and peroxy radicals with phenolic antioxidants pulse radiolysis study. International Journal of Radiation Biology. 52:393.
7. Ghisalberti, E.L. 1979. Propolis: A review. Bee World 60:59.
8. Grochowski, J., M. Bilinska, and D. Stankiewicz, (Poland). 1985. The therapeutic effect of 3% propolis treatment on mice of DBA/2H breed suffering from skin burns subsequently infected with Pseudomonas aeruginosa. The XXXth Internation Apicultural Congress. pp. 439-440. Apinondia.
9. Harborne, J. B. 1982. Introduction to Ecological Biochemistry. Academic Press. London.
10. Havsteen, B. 1983. Flavonoids, a class of natural products of high harm acological potency. Biochemical Pharnacology. 32:

1141.

11. Ikeno, K., T. Ikeno and T. Miyazawa. 1994. Effects of propolis on dental caries in rats. *Honeybee Science* 15: 1.
12. Karinova, Z. K. H. 1961. About the medicinal; qualities of propolis. *Pchelovodstvo*. 38;32. In Russian.
13. Kivalkina, V.P. 1969. Effect of propolis on immunological reactivity. 22 *Int. Beekeep. Congr. Summ.*: 136 In Russian
14. Krol, W., Z. Szuba, S. Scheller, J. Gabrys, S. Grabiec and J. Shani. 1990. Anti-oxident property of ethanolic extract of propolis (EEP) as evaluated by inhibiting the chemiluminescence oxidation of luminol. *Vol. 21. No4.* pp. 593-597. 20.
15. Konig, B and J. H. Dustnan. 1989. *Anim Res. and Develop* 29:21.
16. Lavie, P. 1978. The antibiotic from propolis, PROPOLIS, pp. 41-48. Apinondia Publishing House.
17. Lindenfelser, L. A. 1967. Antimicrobial activity of propolis. *American Bee Journal*. 107:90.
18. Mayer, W. 1956. "Propolis bees" and their activities. *Bee World* 37:25.
19. Matsuno, T. 1992. Isolation and character isolation of the tumoricidal substances from Brazilian propolis. *Honey bee Science* 13;49.
20. Metzner, J., H. Bekeneter, N. Paintz and E. Schneidewind. 1979. *Pharmazie* 34:97.
21. Scheller, S., G. Gazda, J. Gabrys, J. Szumias, I. Eckert and J. Shani, 1988. The ability of EEP to accumulate plaque formation in immunized mouse spleen cells. *Pharmacological Research Communication*. 20:323.

22. Smith, H.W., 1962. The effects of the use of antibiotics on the emergence of antibiotic resistant disease producing organism in animals. *Antibiotics in Agriculture. Proceedings of the University of Nottingham. Ninth Easter School in Agriculture Science. Butterworth. London. p 374.*
23. Smith, H.W., 1975. Persistence of tetracycline resistance in pig *E. coli* *Nature* : 258 : 628.
24. Steel, R. G. D. and J. H. Torrie. 1980. *Principles and procedures of statistics. A Biometrical Approach. McGraw-Hill, New York. pp. 223.*
25. Swain, T. 1977. Secondary compounds as protective agents. *Annual review of plant physiology. 28:479.*
26. Takahana, U., R. J. Youngnan, and E. F. Elstner. 1984. Transformation of quercetin by singlet oxygen generated by photosensitized reaction. *Photobiochemistry and Photobiophysics. 7:175.*

4

가가

1.

2000 1
 BOD SS 350ppm 150ppn, 500ppm
 350ppm 가
 (Choi.,
 1999) 8. 15% 84. 8%

가 96%
 (Choi and Han. 1998). 가 가
 가 가
 가 가
 가

2.

가.

1999 7 5 9
 (L*Y*D)
 2 60
 50 60kg Control ,
 5 2 .

propolis 가 (complex supplement)
 가
 3 (control, ,
), 3 (L*Y*D)
 (7.3kg) (50kg) 9

가
 Cr2O3 0.25% 가 4 2
 1 2 60
 24 1

AOAC (1990) , bomb
 calorimeter (Parr Co., USA), spectrophotometer (Kontron 942)
 , 6 N HCl 24 105
 가 Phenyl isothiocyante(PITC)

Phenyl thiocarbarate (FTC) HPLC (Waters 486) UV
 (254nm) Cr2O3

=100- (100	Cr2O3 (%)	x	(%)
	Cr2O3 (%)		(%)

가
 가 propolis
 dry ice 4 : 1
 hammer-mill ,
 -10 가 가
 saponin
 가

Table 3-23

SAS program(1985) ANOVA

Table 3-22. Addition rate of complex supplement to the diet

	Control	Low complex	High complex
Propolis(%)	0	0.06	0.12
Deodorizer mix(%)	0	0.075	0.15
Sub total(%)	0	0.135	0.27
Feed mixture(%)	100	99.865	99.73
Total(%)	100	100	100

Table 3-23. Formula & chemical composition of experimental basal diet

Item	%
Ingredients	
Corn, yellow	25.00
Wheat	33.06
Soybean meal(44%)	25.62
Wheat shorts	5.00
Animal fat	5.42
Molasses	3.50
Tricalcium phosphate	1.00
Limestone	0.40
L- Lysine- HCl	0.09
Salt	0.26
Vit and mineral mixa	0.10
Antibioticsb	0.05
Additives	0.50
Total	100.00
Chemical composition	
Crude protein	17.70
Crude fat	7.33
Crude fiber	2.76
Crude ash	4.40
Calcium	0.60
Total phosphous	0.55
Lysine	0.97
DE(kcal/kg)	3,500

a Supplied per kg diet 8,000 IU vitamin A, 1,500 IU vitamin D3, 40 IU vitamin E, 1.5ng vitamin K3, 1.0ng thiamine, 4.0ng riboflavin, 2ng vitamin B6, 20µg vitamin B12, 12ng pantothenic acid, 20ng niacin, 0.1ng biotin, 0.6ng folic acid; 60ng Fe, 15ng Cu, 25ng Mn, 0.2ng I, 0.25ng Se.

b Flavonycin 5g

procedure

5%

Duncan

3.

가.

Table 3-24

가

가 가

가

가 가 2.34,

가 가 2.26

0.13, 0.05kg

가

가 가 2.56

가

가

Table 3-24. Growth performance of finishing pigs fed diet supplemented with complexed medicated mixture

Item	Control	Low complex	High complex
Average daily gain(kg)	0.84 ± 0.06	0.83 ± 0.01	0.89 ± 0.07
Daily feed intake(kg)	2.21 ± 0.16	2.34 ± 0.01	2.26 ± 0.00
F/G	2.66 ± 0.4	2.82 ± 0.06	2.56 ± 0.19

가 가

Table 3-25 3-26

Table 3-25

가 가

(P<0.05). sarsaponin 가
가

Table 3-26

가 가 (P<0.05). 가
가 가 . ,
가 가 가 가 가
(P<0.05). 가
가 .

Table 3-25. Effect of feeding complex supplement on nutrient digestibility of starter and growing pigs diet.

	Control (No medication)	Complex supplement (mg/kg)		SE1)
		Low	High	
Starter				
Dry matter	76.44	74.83	75.12	1.91
Crude protein	70.68	70.61	72.25	1.20
Gross energy	76.11	75.38	76.61	1.87
Grower				
Dry matter	69.54	70.86	70.13	1.51
Crude protein	66.63b	69.65a	72.43a	2.94
Gross energy	67.09	68.82	68.27	1.72

1) Pooled standard error.

ab Values on the same line without a common superscript differ (P<0.05).

Table 3-26. Apparent fecal digestibility of amino acids as affected by feeding complex supplement in growing pigs.

Amino acid	Control (No medication)	Complex supplement		SE1)
		Low	High	
Indispensable, %				
ARG	83.93b	85.36ab	88.03a	1.93
HIS	75.73	78.92	84.80	4.48
ILE	80.66b	88.75a	88.68a	4.06
LEU	60.24b	81.47a	75.90a	9.64
LYS	80.79	80.07	80.65	1.81
MET	78.82	76.92	78.38	1.22
PHE	80.54b	88.14a	88.53a	3.95
THR	83.25	83.64	86.05	1.52
VAL	69.95b	78.20a	74.72ab	3.80
Sub- mean	77.10b	82.38a	82.86a	2.94
Dispensable, %				
ALA	59.36b	69.37a	72.28a	6.07
ASP	61.78b	77.70a	78.92a	8.36
GLU	42.39b	62.17a	63.14a	10.29
GLY	78.51	74.18	77.14	2.23
PRO	71.40	76.25	73.72	2.59
SER	77.67b	85.31a	85.75a	4.05
TYR	67.68	73.85	72.64	3.36
Sub- mean	65.54b	74.12a	74.80a	4.66
Total, %	71.32b	78.25a	78.83a	3.79

1) Pooled standard error.

ab Values on the same line without a common superscript differ significantly ($p < 0.05$).

4.

가 propolis 가

가 .

가.

가

가

.

.

가

.

.

,

, ,

가 가

, ,

.

5.

Choi, H. L and J. D. Han. 1998.

가

Brumm, M. C., R , E. R. Jr. and Mader, T. L. 1985. Effect of dietary sarsaponin and chlorotetracycline on pig performance. *J. Anim. Sci.* 61(Supple. 1) : 312

Brumm, M. C., Peo, E. R. Jr and Mader, T. L. 1985. Effect of dietary sarsaponin and chlorotetracycline on pig performance. *J. Anim. Sci.* 61(supple. 1) : 312

Cronwell, G. L., Stahly, T. S and Monegue, H. J. 1985. Efficacy of sarsaponin for weaning and growing-finishing swine housed at two animal densities. *J. Anim. Sci.* 61(suppl. 1) : 111

Presten, R. L., Bartles, S. J., May, J., Goodhall, S. R. 1987. Influence of sarsaponin on growth, feed and nitrogen utilization in growing male rats fed diets with added urea or protein. *J. Anim. Sci.* 65 : 481

Cronwell, G. L., Turner, I. W., Gates, R. S., Lindenann, M. D., Traylor, S. L., Dozier, W. A and Monegue, H. J. 1998. Manipulation of swine diets to reduce odors and harmful gaseous emissions from manure.

Macrae, R., Robinson, R. K and Sadler, M. J. *Encyclopaedia of food*

science food technology and nutrition. Vol.4. 2276-2278

Foster, J. R. 1983. Sarsaponin for growing-finishing swine alone and in combination with an antibiotic at different oig densities. J. Anin. Sci. 57(suppl.) : 245

Jacques, K. A. 1988. Yucca extract inproves air quality and pig performance. Pigs. 4(5):8

Na, M. D., Vung, L. C., Huang, Y. T and Fu, C. M. 1993. Effect of adding deodorizers to diets on the performance of pigs and the deccorization of pig wastes. 1. Preliminary observation of deodorizers in diets for pigs. Pig News and Information. 14(1) : 64

Tanaka, H., Kuroda, K. and Yonaga, M. 1992. Biological removal of VFA from aninal waste. J. Anin. Sci. Technol, (Jpn). 63(1):54

B. S. Jeon., J. H. Kwng., Y. H. Yoo., J. O. Cha and H. S. Park. 1996. Effects of feeding enzyne, probiotics or yucca powder on pig growth and odor-generating substances in feces. korean. J. Anin. Sci., 38(1)52-58

5

propolis 가가

propolis

1.

가

(issue)

가

가

가

가

가

가

가

(Council Directive 70/524/EEC, '70. 11.23)

(Council

Regulation EC 2821/98) 1998 12

가

가

가

propolis

propolis

propolis

-glucosidase flavonoid flavonoid
 aglycones 가 .

(Grange et al., 1990; Krol et al., 1993; Aga et al., 1994; Park et al., 1998). (樹脂) 50%, (蜜蠟) 30%, (油性) 10%, (花粉) 5% 5%

가 propolis (Park, 1998). propolis , , , 가 가 가 propolis 가 propolis propolis 가 가 .

2.

가.

		1999	7	5	9
2	60		50	60kg	
	(L*Y*D)	Control,	Propolis	300ppn	(pro-300)
	Propolis	600ppn	(pro-600)	3	5
2					

Propolis 가
 propolis 3 (Control,
 pro-300, pro-600), 3
 (I*Y*D) (7.1 kg) (50.2 kg)
 9

가
 Cr2O3 0.25% 가 4 2
 1 2 60
 24 1

AOAC (1990) , bomb
 calorimeter (Farr Co., USA), spectrophotometer (Kontron 942)
 , 6 N HCl 24 105
 가 Phenyl isothiocyanate(PITC)
 Phenyl thiocarbamate (PIC) HPLC (Waters 486) UV
 (254nm) _____ . Cr2O3

=100- (100	Cr2O3 (%)	x	(%)
	Cr2O3 (%)		(%)

conial tube 2Mℓ ethylacetate 5g
 400rpm 5 4.2Mℓ 6Mℓ 가
 hexane 1:1(v/v) 1.4Mℓ 가 chloroform
 0.7Mℓ 가
 HPLC Injection volume 20μℓ , μ
 Bondapak C18 ,10μm column (3.9×150mm) Kontron 422 HPLC
 pump, detector 430A, thermostat controller 402(50c)
 CH₃OH, CH₃COOH, H₂O 60:35:5
 chromatogram 272nm

propolis
 dry-ice 4 : 1 hammer-nill
 -10

Propolis Table 3-27 control
 propolis 가 , Pro-300,
 Pro-600 가
 가
 Table 3-28

Table 3-27. Propolis addition rate to the growing pig diet

(unit : %)

	Control	Pro- 300	Pro- 600
Propolis	0	0.06	0.12
Feed mixture	100	99.94	99.88
Total	100	100	100

Table 3-28. Formula and chemical composition of experimental basal diet

Ingredients	(%)
Corn, yellow	25.00
Wheat	33.06
Soybean meal(44%)	25.62
Wheat shorts	5.00
Animal fat	5.42
Molasses	3.50
Tricalcium phosphate	1.00
Limestone	0.40
L- Lysine- HCl	0.09
Salt	0.26
Vit and mineral mixa	0.10
Antibioticsb	0.05
Additives	0.50
Total	100.00
Chemical composition(%)	
Crude protein	17.70
Crude fat	7.33
Crude fiber	2.76
Crude ash	4.40
Calcium	0.60
Total phosphous	0.55
Lysine	0.97
DE(kcal/kg)	3,500

a Supplied per kg diet : vit-A 8,000 IU, vit-D3 1,500 IU, vit-E40 IU, vit-K3 1.5ng, thianine 1.0ng, riboflavin 4.0ng, vit-B6 2ng, vit-B12 20 μ g, pantothenic acid 12ng, niacin 20ng, biotin 0.1ng, folic acid 0.6ng; Fe 60ng, Cu 15ng, Mn 25ng, I 0.2ng, Se 0.25ng.

b Flavonycin 5g

* Calculated value

. SAS program(1985) ANOVA
 procedure , 5% Duncan
 (Snedecor Cochran, 1980) .

3.

가.

Table 3-29 Propolis 가 .
 propolis 가 ,
 propolis 600 가 .
 가 .
 propolis 300 600 가
 propolis 600 가 가 propolis 300 가 가

. propolis 600

가

Propolis 600 가

Table 3-29. Growth performance of finishing pigs fed diet supplemented with propolis

Item	Control	Propolis	
		300	600
Average daily gain(kg)	0.84 ± 0.06	0.85 ± 0.13	0.86 ± 0.13
Daily feed intake(kg)	2.21 ± 0.16	2.25 ± 0.00	2.07 ± 0.00
F/G	2.66 ± 0.4	2.68 ± 0.4	2.40 ± 0.29

Propolis 가 Table 3-30 Table 3-31

Table 3-30 ,

propolis 600 ng/kg 가

propolis 300 ng/kg (P<0.05)

가 ,

propolis 가 (P<0.05).

propolis 가 가 .

Table 3-31 ,

propolis 600ng/kg 가

(P<0.05) , propolis 300 600 ng/kg

(P<0.05) .

propolis 가 가

(P<0.05). 가 가 .

Table 3-30. Effect of feeding propolis on nutrient digestibility in starter and growing pigs

	Control	Propolis (mg/kg)		SE 1)
	(No sarsaponin)	300	600	
Starter				
Dry matter	75.77b	74.50b	77.49a	0.87
Crude protein	71.32	70.36	71.94	1.94
Gross energy	76.97b	77.03b	78.11a	0.97
Grower				
Dry matter	69.87b	74.86a	73.72a	3.02
Crude protein	67.44b	73.51a	73.72a	4.42
Gross energy	68.13b	73.65a	72.12a	2.87

1) Pooled standard error.

ab Values on the same line without a common superscript differ (P<0.05).

Table 3-31. Apparent fecal digestibility of amino acids as affected by feeding propolis in growing pigs.

Amino acid	Control	Propolis (mg/kg)		SE 1)
	(No Propolis)	300	600	
Indispensable, %				
ARG	83.93b	84.40b	87.10a	1.44
HIS	75.73c	83.43b	86.01a	4.67
ILE	80.66	86.59	86.70	4.18
LEU	60.24b	70.45ab	76.06a	7.35
LYS	80.79	80.61	81.15	3.48
MET	78.82	64.51	71.17	7.42
PHE	80.54	84.96	85.35	3.42
THR	83.25	79.56	80.40	5.68
VAL	69.95b	80.47a	81.48a	5.74
Sub- mean	77.10	79.44	81.71	2.62
Dispensable, %				
ALA	59.36b	69.63a	72.24a	6.07
ASP	61.78b	79.15a	80.26a	9.45
GLU	42.39b	61.55a	64.68a	10.78
GLY	78.51	78.19	78.42	1.15
PRO	71.40	81.42	83.19	5.87
SER	77.67	83.36	84.70	3.79
TYR	67.68	68.52	71.24	2.20
Sub- mean	65.54b	74.55a	76.39a	5.13
Total, %	71.32b	76.99a	79.05a	3.66

1) Pooled standard error.

ab Values on the same line without a common superscript differ significantly ($p < 0.05$).

caffeic acid (3,4-dihydroxycinnamic acid), p-coumaric acid (4-hydroxy-cinnamic acid), cinnamic acid(3-phenyl-2-propenoic acid) and rutin (flavone glycoside) 4가 HPLC

(FAO, 1996)

가

가

(Lavie, 1976).

160 가

(Green-way et al., 1990) ,

4가

, caffeic acid (3,4-dihydroxy cinnamic-acid)가

Caffeic acid antiviral activity(Konig and Dustmann, 1985), anti-bacterial activity gran-positive and gran-negative micro-organisms(Villanueva et al., 1970) , anti-inflammatory activity.(Bankova et al., 1983)

Table 3-32. Fig. 1, 2

가

caffeic acid (3,4-dihydroxycinnamic acid)가

0.25ppm, 0.19ppm , p-coumaric acid (4-hydroxycinnamic acid), cinnamic acid(3-phenyl-2-propenoic acid) rutin (flavone glycoside)

Fig. 3.

caffeic acid (3,4-dihydroxycinnamic acid), p-coumaric acid(4-hydroxy-cinnamic acid), cinnamic acid(3-phenyl-2-propenoic acid) rutin(flavone glycoside) , peak가 . 가

peak

Fig. 4, 5

caffeic acid (3,4-dihydroxycinnamic acid), p-coumaric acid (4-hydroxy-cinnamic acid), cinnamic acid(3-phenyl-2-propenoic acid) rutin (flavone glycoside) , peak 가 . 가

peak

Table 3-32. Composition of caffeic acid (3,4-dihydroxycinnamic acid) from chicken's chest and chicken's leg

	caffeic acid(3,4- dihydroxycinnamic acid)
chicken's chest	0.25ppm
chicken's leg	0.19ppm

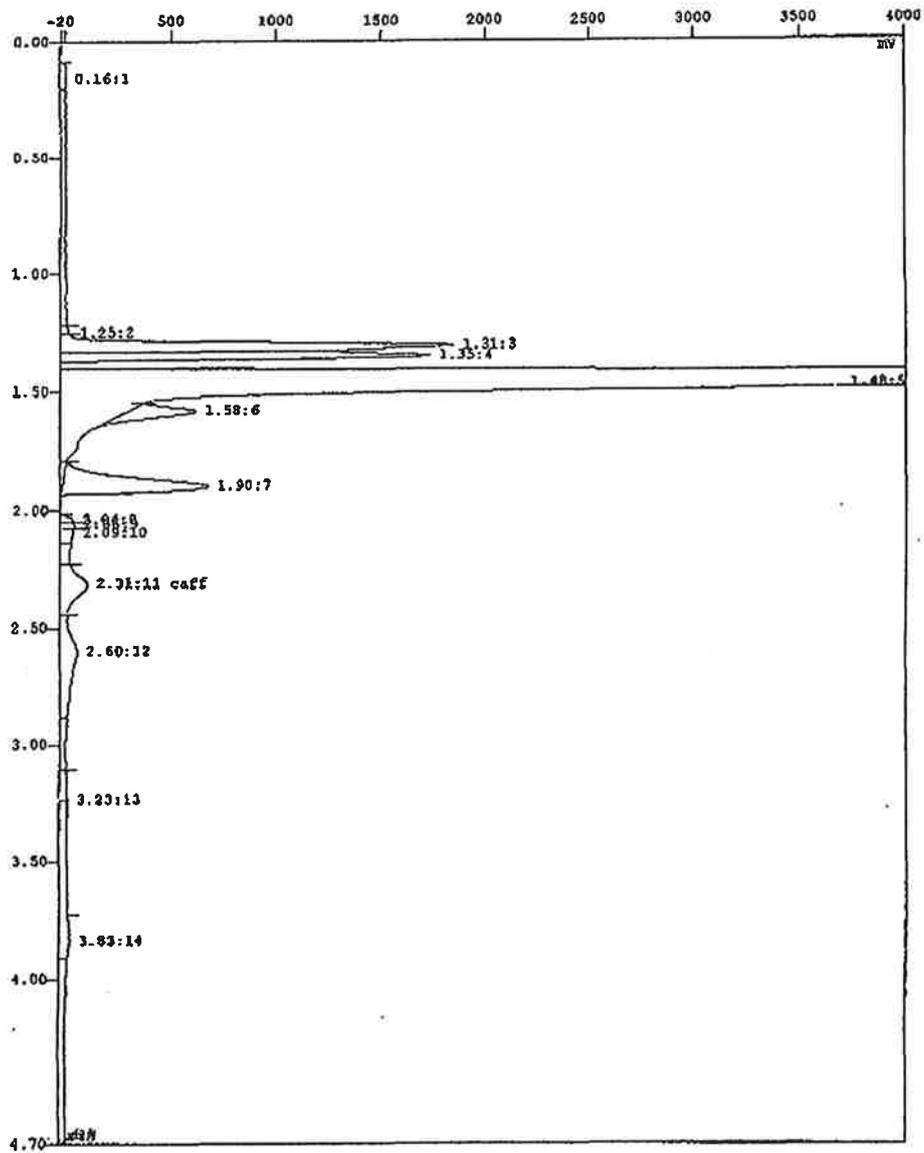


Fig 3-1. HPLC diagram for breast muscle of broiler fed propolis
 * 2.31 CAFF : caffeic acid (3,4-dihydroxycinnamic acid)

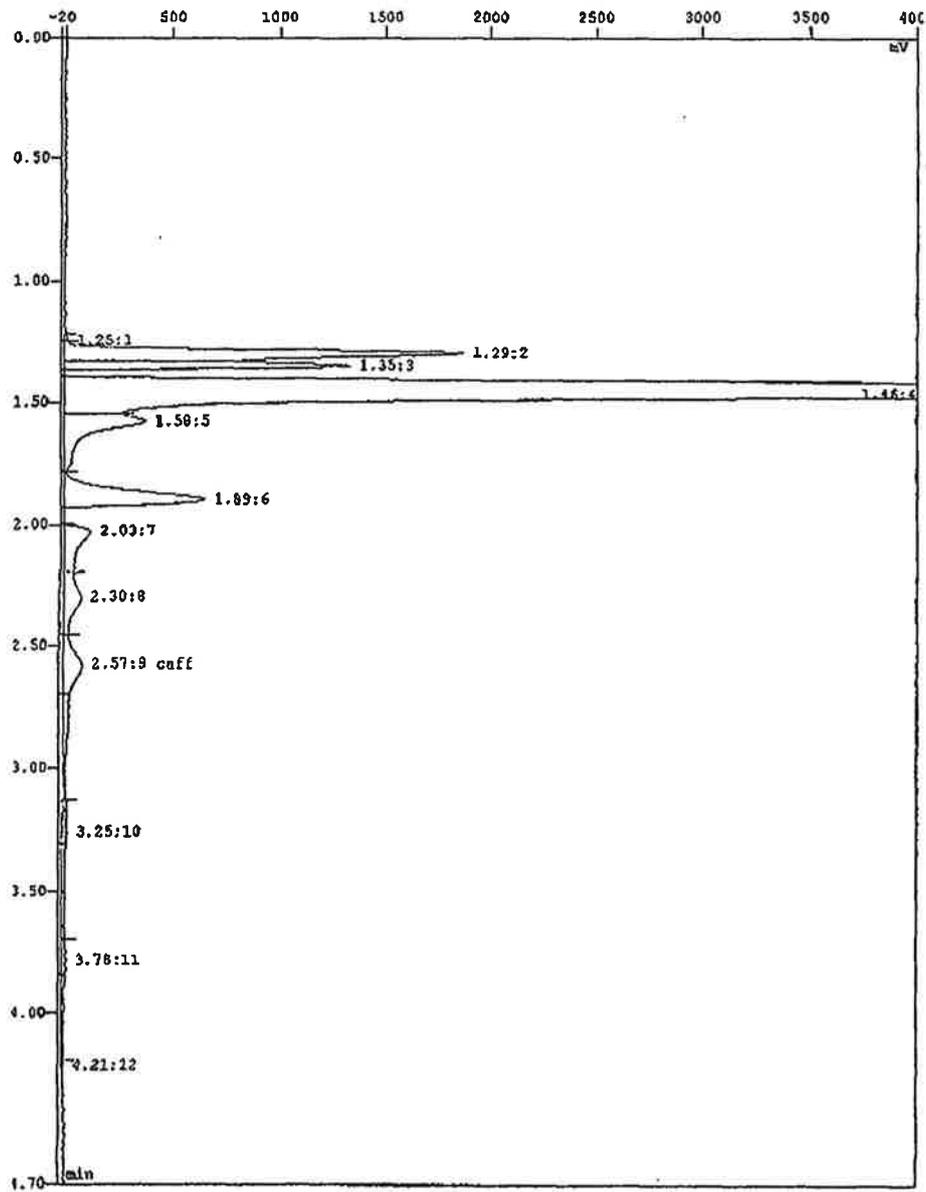


Fig 3-2. HPLC diagram for drumstick of broiler fed propolis
 * 2.31 CAFF : caffeic acid (3,4-dihydroxycinnamic acid)

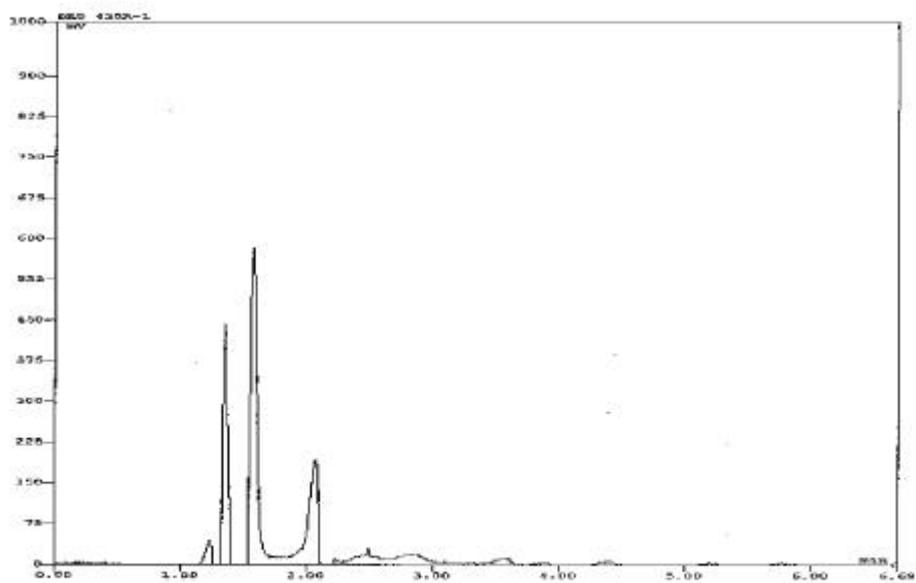


Fig 3-3. HPLC profiles for belly muscle of pigs fed propolis

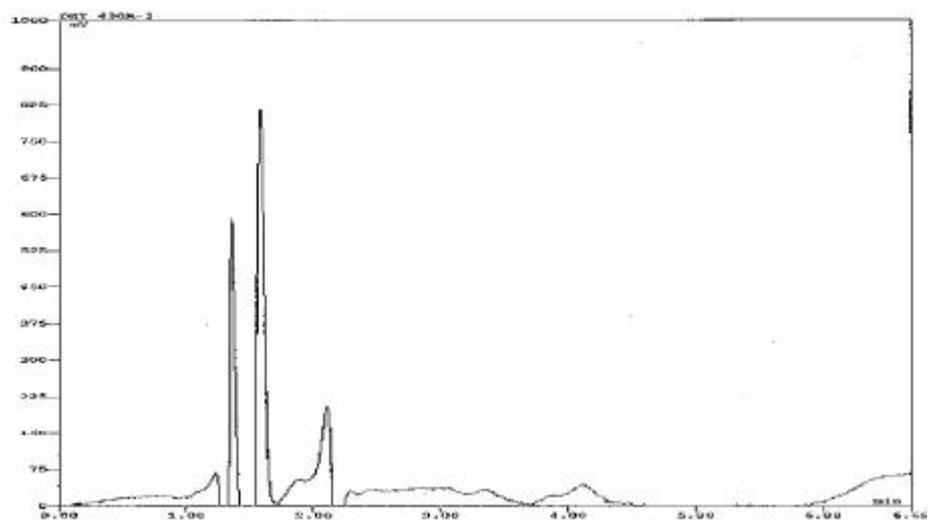


Fig 3-4. HPLC profile with small intestine of pigs fed propolis.

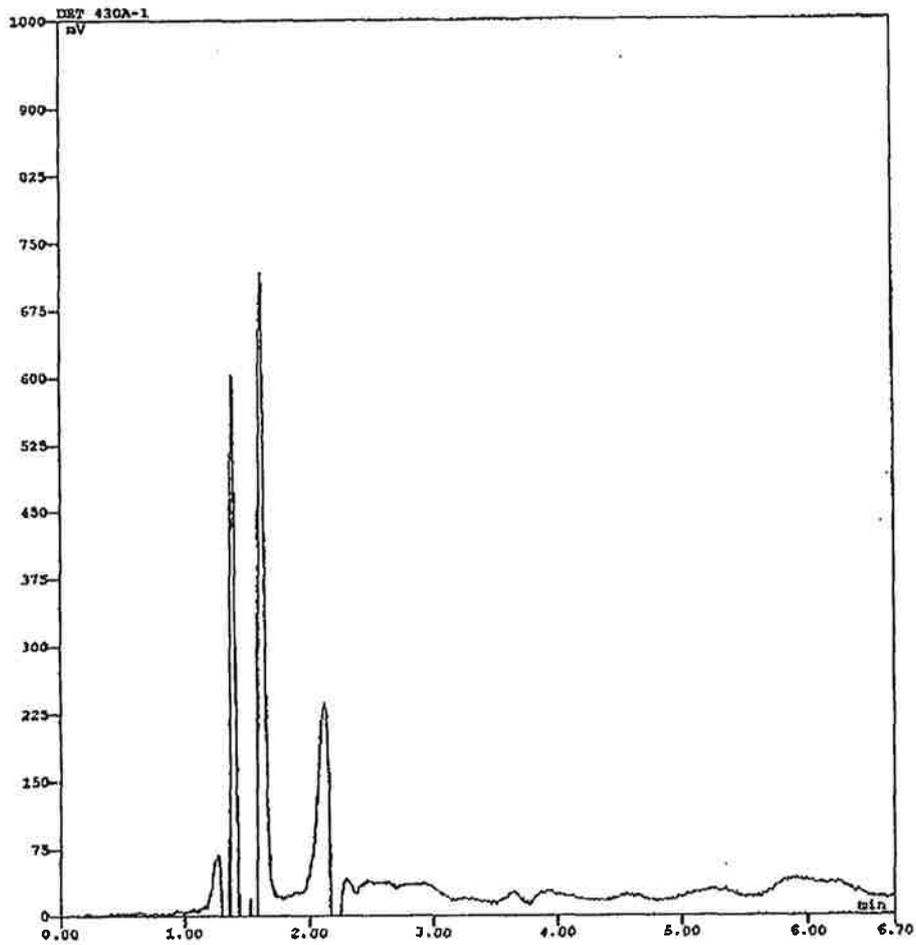


Fig 3-5. HPLC profile for liver of pigs fed propolis.

4.

Propolis 가

.

가. Propolis 가

Propolis

60

, 가

Propolis

가

가

propolis

가

가

가

Pro-600

pro-300

Pro-300

600

가

Pro-600

Pro-300

600

Pro-300

600

가

5.

Takino, Y. and S. Mochida. 1982. Propolis, its chemical constituents and biological activities. Honeybee Science 3: 145.

Lavie, P. 1978. The antibiotic from propolis, PROPOLIS, pp.41-48. Apinondia Publishing House.

Christova, V. M. 1985. A Propolis-enriched was therapy against inflammatory diseases of the articular-muscular system. The XXXth Internation Apicultural Congress. pp. 429-431. Apinondia.

Brum, M. C., Peo, E. R. Jr and Mader, T. L. 1985. Effect of dietary sarsaponin and chlorotetracycline on pig pergormance. J. Anin. Sci. 61(supple. 1) : 312

Cronwell, G. L., Stahly, T. S and Monegue, H. J. 1985. Efficacy of sarsaponin for weaning and growing-finishing swine housed at two animal densities. J. Anin. Sci. 61(suppl. 1) : 111

Presten, R. L., Bartles, S. J., May, J., Goodhall, S. R. 1987. Influence of sarsaponin on growth, feed and nitrogen utilization in growing male rats fed diets with added urea or

protein. *J. Anim. Sci.* 65 : 481

Cronwell, G. L., Tuner, I. W., Gates, R. S., Lindenann, M. D., Traylor, S. L., Dozier, W. A and Nonegue, H. J. 1998. Manipulation of swine diets to reduce odors and harmful gaseous emissions from manure.

Macrae, R., Robinson, R. K and Sadler, M. J. *Encyclopaedia of food science food technology and nutrition*. Vol.4. 2276-2278

Foster, J. R. 1983. Sarsaponin for growing-finishing swine alone and in combination with an antibiotic at different pig densities. *J. Anim. Sci.* 57(suppl.) : 245

Jacques, K. A. 1988. Yucca extract improves air quality and pig performance. *Pigs*. 4(5):8

Ma, M. D., Vung, L. C., Huang, Y. T and Fu, C. M. 1993. Effect of adding deodorizers to diets on the performance of pigs and the deodorization of pig wastes. 1. Preliminary observation of deodorizers in diets for pigs. *Pig News and Information*. 14(1) : 64

1.

가

가 가 ,

가 eucca-extract 가 (Cronwell et al. 1985). 가

eucca-extract 가 saponin (Deodorizer) 가 gas 가 .

2.

가. (Deodorizer :D0)

saponin

,

alginate

cheto-oligosaccharide(cos)

가

가 1999 10 15 11

13 30 (D*H*L*Y)

Control, D0-50, D0-100 3 . 36

(62.02 ± 7.16kg) 6

가

가

D0-50 D0-100

가

가

가 가

Table 3-33

Table 3-33. 가 (%)

Item \ Treatment	Control	Deodo- 50	Deodo- 100
Deodorizer	0	0.7	1.4
Feed mixture	100	99.3	98.6
Total	100	100	100

Table 3-34.

	(%)
	35.7
	14.2
	15.0
	12.0
	13.68
	3.0
	3.0
	0.54
	0.26
3	1.34
	0.28
	0.555
()	0.2
()	0.075
	0.07
C.T.C(16.7%)	0.06
	0.04
	100.0
	11.71
	16.67
	7.27
	4.10
	4.46
	0.65
	0.56

Table 3-34

가 100kg Multiple mixer Pellet
 bach 5% 가 .
 Pellet 가 5.7mm die feeder

1200rpm
 pelleting pellet 가
 3 cooling .
 pellet hardness durability table 3-35

Table 3-35. Hardness and durability of pellet diet

	Control	DO- 50	DO- 100
hardness(kg/cm ²)	1.84 ± 0.31	2.8 ± 0.21	3.85 ± 0.42
durability(%)	93.8 ± 0.1	93.15 ± 0.05	92.57 ± 0.03

pellet DO-100>DO-50>
 가 Zeolite illite 가 ,
 . DO-50
 DO-100 가 .

Cr₂O₃ 0.25% 가 4 2
 1 2 60
 24 1

AOAC (1990) , bomb
 calorimeter (Parr Co., USA), spectrophotometer (Kontron 942)
 , 6 N HCl 24 105

가 Phenyl isothiocyante (PITC)
 Phenyl thiocarbanate (PTC) HPLC (Waters 486) UV
 (254nm) Cr2O3

=100- (100	Cr2O3 (%)	x	(%)
	Cr2O3 (%)		(%)

SAS (1985)
 Duncan
 (Snedecor Cochran, 1980)

3.

가.

가 0.94 ± 0.02, DO-50
 1.01 ± 0.02 DO-100 0.98 ± 0.03 DO-50>DO-100>
 가 2.57 ± 0.06, DO-50 2.71 ± 0.14 DO-100 2.63 ±
 0.13 DO-50>DO-100>
 가 2.73, DO-50 2.62 ± 0.09 DO-100 2.68
 ± 0.06 DO-50>DO-1000>

Table 3-36. Growth performance of swine

Item \ Treatment	Control	DO- 50	DO- 100
ADG(kg)	0.94 ± 0.02	1.01 ± 0.02	0.98 ± 0.03
ADFI(kg)	2.57 ± 0.06	2.71 ± 0.14	2.63 ± 0.13
F/G	2.73 ± 0.00	2.62 ± 0.09	2.68 ± 0.06

Deodorizer 가 Table 3-37 3-38
 Table 3-37 ,
 D0-100 가 D0-50
 가 (P<0.05)
 D0-100
 가 , D0-50 가
 (P<0.05).
 Table 3-38 ,
 deodorizer 가
 (P<0.05).
 deodorizer 가
 D0-100 가
 (P<0.05).
 , , , deodorizer 가
 가 (P<0.05).
 D0-100 가 (P<0.05)

Table 3-37. Effect of feeding deodorizer on nutrient digestibility in starter and growing pigs

	Control (No sarsaponin)	DO- 50	DO- 100	SE1)
Starter				
Dry matter	75.12b	75.35b	76.37a	1.33
Crude protein	70.15	69.58	70.22	2.12
Gross energy	77.21	77.14	76.15	1.79
Grower				
Dry matter	68.75b	68.66b	74.77a	2.75
Crude protein	67.83b	65.25b	74.57a	3.54
Gross energy	67.54b	66.70b	72.88a	3.41

1) Pooled standard error.

abc Values on the same line without a common superscript differ (P<0.05).

Table 3-38. Apparent fecal digestibility of amino acids as affected by feeding deodorizer in growing pigs

Amino acid	Control (No sarsaponin)	DO- 50	DO- 100	SE1)
Indispensable, %				
ARG	83.93	82.60	82.68	0.85
HIS	75.73b	81.24a	82.62a	3.23
ILE	80.66a	69.77c	73.49b	4.81
LEU	60.24b	70.71a	73.70a	6.17
LYS	80.79	80.30	82.05	1.77
MET	78.82a	71.34b	77.39a	3.59
PHE	80.54a	75.60b	79.97a	2.48
THR	83.25a	73.04c	78.46b	4.49
VAL	69.95c	73.83b	77.00a	3.10
Sub- mean	77.10ab	75.38b	78.59a	1.49
Dispensable, %				
ALA	59.36c	64.99b	68.31a	3.96
ASP	61.78b	85.31a	87.90a	12.49
GLU	42.39c	50.04b	55.99a	5.99
GLY	78.51a	74.34b	77.22a	1.88
PRO	71.40b	82.99a	84.78a	6.34
SER	77.67b	79.92ab	82.48a	2.19
TYR	67.68	62.08	66.83	2.87
Sub- mean	65.54c	71.38b	74.79a	4.08
Total, %	71.32b	73.38b	76.69a	2.39

1) Pooled standard error.

ab Values on the same line without a common superscript differ significantly ($p < 0.05$).

4.

가. deodorizer 가 ,
deodorizer 가
($P < 0.05$).

. deodorizer 가
, , ,
.

. 가 가 0.7% 가 D0-50
, ,
가
deodorizer 가 가 .

5.

Brumm, M. C., Peo, E. R. Jr and Mader, T. L. 1985. Effect of dietary sarsaponin and chlorotetracycline on pig performance. *J. Anim. Sci.* 61(supple. 1) : 312

Cronwell, G. L., Stahly, T. S and Monegue, H. J. 1985. Efficacy of sarsaponin for weaning and growing-finishing swine housed at two animal densities. *J. Anim. Sci.* 61(suppl. 1) : 111

Presten, R. L., Bartles, S. J., May, J., Goodhall, S. R. 1987. Influence of sarsaponin on growth, feed and nitrogen utilization in growing male rats fed diets with added urea or protein. *J. Anim. Sci.* 65 : 481

Cronwell, G. L., Turner, I. W., Gates, R. S., Lindenann, M. D., Traylor, S. L., Dozier, W. A and Monegue, H. J. 1998. Manipulation of swine diets to reduce odors and harmful gaseous emissions from manure.

Macrae, R., Robinson, R. K and Sadler, M. J. *Encyclopaedia of food science food technology and nutrition.* Vol.4. 2276-2278

Foster, J. R. 1983. Sarsaponin for growing-finishing swine alone and in combination with an antibiotic at different oig densities. *J. Anim. Sci.* 57(suppl.) : 245

Jacques, K. A. 1988. Yucca extract improves air quality and pig performance. Pigs. 4(5):8

Ma, M. D., Vung, L. C., Huang, Y. T and Fu, C. M. 1993. Effect of adding deodorizers to diets on the performance of pigs and the deodorization of pig wastes. 1. Preliminary observation of deodorizers in diets for pigs. Pig News and Information. 14(1) : 64

Tanaka, H., Kuroda, K. and Yonaga, M. 1992. Biological removal of VFA from animal waste. J. Anim. Sci. Technol, (Jpn). 63(1):54

B. S. Jeon., J. H. Kwng., Y. H. Yoo., J. O. Cha and H. S. Park. 1996. Effects of feeding enzymes, probiotics or yucca powder on pig growth and odor-generating substances in feces. Korean. J. Anim. Sci., 38(1) 52-58

4

propolis

가

1

가

가

가

가

가

가

cross-contamination

1997 7 1

flavonoids

propolis

HACCP

가

brand

propolis

brand

propolis가

가

가

propolis

brand

가

가

.

2 Propolis

가

1.

가

가

가

97 7

가 **VIO**

가

.

-

flavonoids propolis - 가

(, 가 , ,)

crude propolis **ethanol**

가

propolis

가 가

2. Propolis

가.

propolis
(97%)
97%, 90%, 80%,
70%, 60%, 50%, 40%, 30%, 20% 10%
300ml 30g
2 magnet stirrer Whatnan filter No. 4
evaporator ethanol

1) Ethanol 가 propolis

Ethanol	propolis	Table 4-1
97%	ethanol	60%
30g	14g propolis	46.6%
50%	50%	
50% ethanol	6g propolis	20%
40% ethanol	4g propolis	13.3%
30% ethanol	10%	2g
6.6%	.	
ethanol	60%	.

Table 4-1. (D) propolis(30g) ethanol

(ethanol)	(%)	- (g)	- (%)
97		14	46.6
90		14	46.6
80		14	46.6
70		14	46.6
60		14	46.6
50		6	20.0
40		4	13.3
30		2	6.6
20		2	6.6
10		2	6.6

Table 4-2. 97% ethanol propolis

	(A)	(B)	(C)	(D)	가			()	()
(%)	59%	52	39	46.6	48	40	56	30	43

a) = %

2) 97% ethanol

propolis

Table 4-2

propolis

97% ethanol

propolis

A

59%

가

56%

propolis

C

39%

30%

3. propolis 가

in vitro

가.

1)

Table 4-3. Propolis

Strains	
1. <i>Staphylococcus aureus</i> - NCTC 5663 - hem	Brain-heart-infusion broth Brain-heart-infusion agar
2. <i>Staphylococcus epidermidis</i> - ATCC 12228	Brain-heart-infusion broth Brain-heart-infusion agar
3. <i>Staphylococcus agalactiae</i> - ATCC 13813	Brain-heart-infusion broth Brain-heart-infusion agar
4. <i>Streptococcus pyogenes</i> - ATCC 21059	Brain-heart-infusion broth Tryptic soy agar
5. <i>Staphylococcus uberis</i> - ATCC 27958	Brain-heart-infusion broth Tryptic soy agar
6. <i>Fructus vulgaris</i> -	Escherichia coli broth MacConkey agar
7. <i>Clostridium perfringens</i> - ATCC 36295-4	Cook-Neat broth Brain-heart-infusion agar
8. <i>Corynebacterium pyogenes</i> - IVR 2001	Brain-heart-infusion broth Brain-heart-infusion agar
9. <i>Mycobacterium pneumoniae</i>	Escherichia coli broth MacConkey agar
10. <i>Escherichia coli</i> - ATCC 10536	Escherichia coli broth MacConkey agar

Table 4-3

108/M0

DMSO(dinethyl sulfoxide)

propolis	6mm	paper disc 15 μ l
35 \pm 1		
positive control (disc potency : 10 μ g)	penicillin	ampicillin
norfloxacin (disc potency : 10 μ g)		quinolone
50,000ppm 10	5ppm	propolis

1) propolis

Fig. 4-1

	<i>Staphylococcus uberis</i>	50,000ppm
	가	
	penicillin	
ampicillin	propolis	norfloxacin
	ampicillin	
	가	
	propolis	disc
	propolis	
	가	flavonoids

Fig. 4-2

Staphylococcus aureus

50,000ppm
 50,000ppm 5,000ppm
 50,000ppm 5ppm

Fig. 4-3 *Mlebsiella pneumoniae* ampicillin

Fig. 4-1 propolis 50,000ppm

Fig. 4-4 *Ercteus vulgaris* propolis 50,000ppm ampicillin

Fig. 4-5 *Staphylocccccus agalactiae* *Staphylocccccus uberis* 3 propolis가 50,000ppm 5,000ppm propolis가 Gram (+)

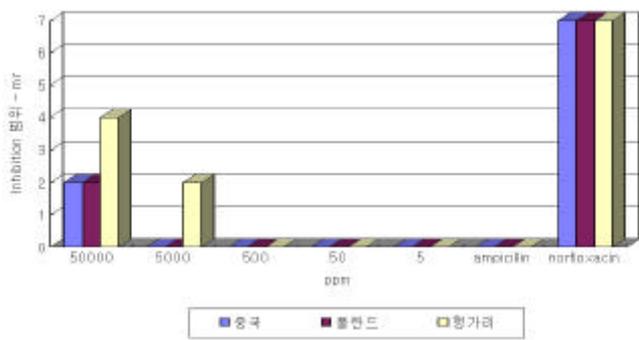


Fig. 4-1. *Staphylocccccus uberis*(10⁸/ml) propolis

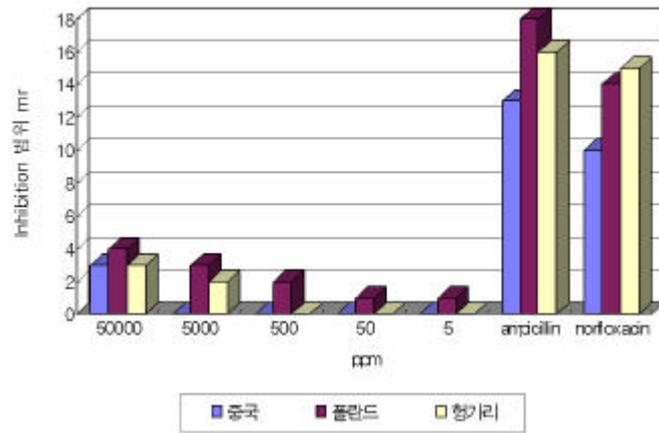


Fig. 4-2. *Staphylococcus aureus*(10⁸/nl) propolis

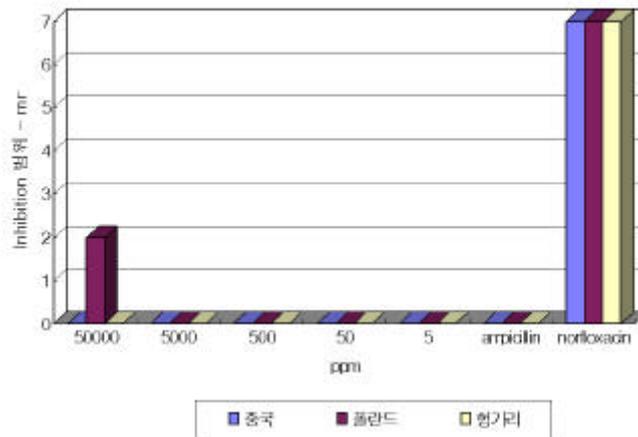


Fig. 4-3. *Klebsiella pneumoniae*(10⁸/nl) propolis

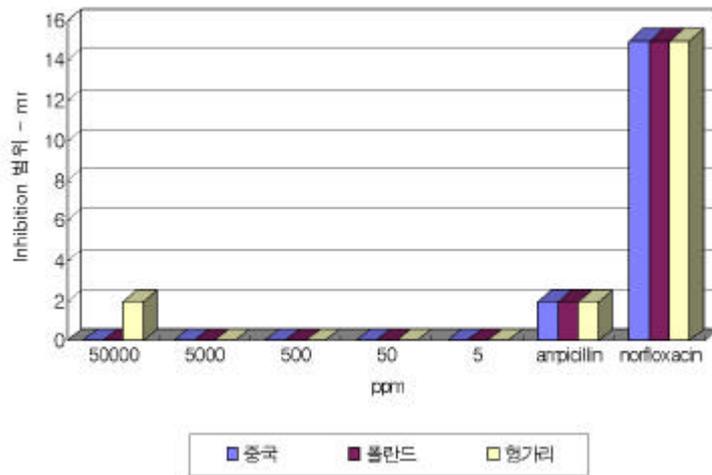


Fig. 4-4. *Proteus vulgaris*(10⁸/ml) propolis

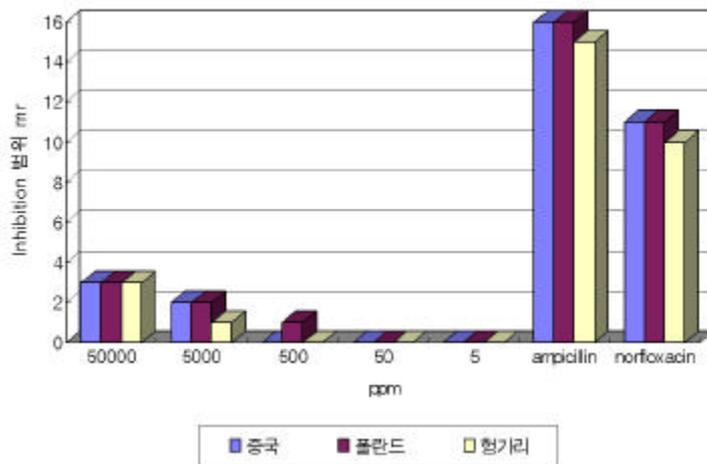


Fig. 4-5. *Staphylococcus agalactiae*(10⁸/ml) propolis

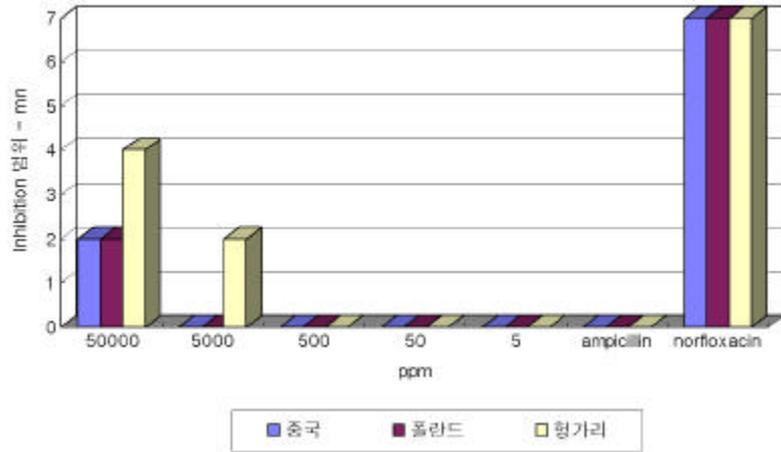


Fig. 4-6. *E. coli* (10⁸/nl) propolis

Fig. 4-6 Gram (-) *E. coli*
 ampicillin 3 propolis
 가 50,000ppm 4mm
 5,000ppm .

2) ethanol propolis

Propolis ethanol
 Table 4-4, 4-5 4-6
 80% ethanol propolis가 gran(+) 가
 . 80% ethanol
 propolis
 flavonoids 가 가 .

Table 4-4. ethanol propolis *Staphylococcus agalactiae*

Ethanol	97%	90%	80%	70%	60%	100%	propylen-glycol
inhibition- (mm)	2	2	4	2	1	N. D.	N. D.

N. D. = no detectable

Table 4-5. ethanol propolis *Staphylococcus aureus*

Ethanol	97%	90%	80%	70%	60%	100%	propylen-glycol
inhibition- (mm)	2	1.8	3	2	2	N. D.	1

N. D. = no detectable

Table 4-6. ethanol propolis *Streptococcus pyogenes*

Ethanol	97%	90%	80%	70%	60%	100%	propylen-glycol
inhibition- (mm)	1.5	2	3	1	1	N. D.	0.8

N. D. = no detectable

Table 4-7. ethanol propolis *Proteus vulgaris*

Ethanol	97%	90%	80%	70%	60%	100%	propylen-glycol
inhibition- (mm)	N. D.						

N. D. = no detectable

Table 4-8. ethanol propolis *Klebsiella pneumoniae*

Ethanol	97%	90%	80%	70%	60%	100%	propylen-glycol
inhibition- (mm)	N. D.						

N. D. = no detectable

Table 4-9. ethanol propolis *E. coli*

Ethanol	100%	90%	80%	70%	60%	100%	propylen -glycol
inhibition- (mm)	N. D.						

N. D. = no detectable

3 Propolis 가

propolis

Balb/C mice propolis 10 *in vivo*

T

cytokine interleukin-II *in*

vitro

1. Propolis *in vivo* 가

가.

1)

6 Balb/C mice male

2) : Table 4-10 .

3)

thymus, spleen, liver, blood, serum

4) Flowcytometry

Propolis가 mice T-

propolis

Table 4-10.

: MSK-97010 : 10

: Pronolis(가) : Per Oral

Vehicle : Tween 80 : Balb/c Mice. Male

: 97. 6. 21 - 97. 8. 10

[] < >

(ng/kg)	(/group)	(g)	(nl/kg)	(ng)	(nl) 5nl
300	20	20	10nl/kg	150ng + 300ul Tween80 + 4. 7nl PBS	
100	20	20	10nl/kg	75ng + 150ul Tween80 + 4. 7nl PBS	
50	20	20	10nl/kg	25ng + 100ul Tween80 + 4. 9nl PBS	
	15	20	10nl/kg	0ng + 100ul Tween80 + 4. 9nl PBS	
	15	20	10nl/kg	0ng + 300ul Tween80 + 4. 7nl PBS	
	15	20	10nl/kg	0ng	--> 5. 0nl PBS
R =			1 : 225ng	--> 15. 0nl	
			7 : 1. 575ng	--> 105. 0nl	
			2 : 3. 150ng	--> 210. 0nl	
			3 : 4. 725ng	--> 315. 0nl	
			4 : 6. 300ng	--> 420. 0nl	
			5 : 7. 875ng	--> 525. 0nl	
			10 : 15, 750ng	--> 1, 050. 0nl	

[] < >

(ng/kg)	(/group)	(g)	(nl/kg)	(ng)	(nl)
300	20	20	10nl/kg	30ng/nl	5nl
150	20	20	10nl/kg	15ng/nl	5nl
50	20	20	10nl/kg	5ng/nl	5nl
	15	20	10nl/kg	0ng/nl	5nl
	15	20	10nl/kg	0ng/nl	5nl
	20	20	10nl/kg	0ng/nl	5nl
Vehicle / day					
150ng + 300ul Tween80 + 4. 70nl PBS					
75ng + 150ul Tween80 + 4. 75nl PBS					
25ng + 100ul Tween80 + 4. 90nl PBS					
0ng + 100ul Tween80 + 4. 90nl PBS					
0ng + 300ul Tween80 + 4. 70nl PBS					
0ng + 0ul Tween80 + 5. 0nl PBS					

2 x 10⁶ anti-mouse CD3-FITC(pan T cell marker), anti-mouse CD-4-FITC(T-helper cell), anti-mouse CD-8 (T-suppressor cell) , anti-mouse CD-25-FITC(interleukin II receptor), anti-mouse TNF- α -FITC(TNF- α receptor), anti-mouse TCR-FITC (T-cell antigen receptor) Flowcytometry(Becton Dickinson)

1) Propolis 가 nice T-

가 T-

antigen

10 propolis Balb/C nice 50ng/kg, 150ng/kg 300ng/kg T-

Table 4-11 . propolis

, T-cell 가

CD-3(pan T-cell marker) 가

6.46% 150ng/kg propolis 10.78% 가

T-cell subset T-helper cell (CD-4)

25.06% 150ng/kg 40.88% 가

T-suppressor cell (CD-8)

18.18% 50ng/kg 가 150ng/kg

26.19% 가 .

T- 가 50ng/kg 150ng/kg

가

T-helper cell (CD-4) T-suppressor cell (CD-8) 2:1

300ng/kg

150ng/kg

가

T-

propolis 가 가

T- 가 T-helper cell

T-suppressor cell 가

2:1 1:1

T- 가 2가 marker

T-

propolis가 T- 가

propolis 10 가

2) Propolis가 cytokine receptor Interleukin-receptor tumor necrosis factor-receptor

가

cytokine interleukin

tumor necrosis factor 가

T- interleukin-receptor

macrophage tumor necrosis factor- ()

receptor propolis 가

Table 4-11. 10 propolis 가 Balb/C nice

marker	control	vehicle 1	vehicle 2	50mg/kg	150mg/kg	300mg/kg
CD- 3	6.46 ± 1.29	5.39 ± 1.47	4.92 ± 2.54	5.88 ± 4.71	10.78 ± 1.30	7.95 ± 1.20
CD- 4	25.06 ± 6.22	26.43 ± 4.83	23.55 ± 3.83	27.31 ± 4.12	40.88 ± 6.28	29.60 ± 4.61
CD- 8	18,18 ± 3,80	16.56 ± 2.99	17.23 ± 3.95	24.88 ± 3.12	26.19 ± 3.46	18.94 ± 2.73
IL- II receptor	6.80 ± 1.31	7.82 ± 1.35	6.44 ± 2.76	8.79 ± 2.65	11.86 ± 1.79	8.89 ± 1.53
TNF- receptor	9.09 ± 1.67	8.05 ± 2.87	9.54 ± 4.87	10.55 ± 2.57	12.14 ± 2.19	9.66 ± 1.79
TCR receptor	19.31 ± 2.81	14.27 ± 3.54	13.95 ± 5.98	17.32 ± 4.55	31.81 ± 4.91	18.94 ± 1.85

% 10, 000 flowcytonetry . n = 10 marker

Table 4-12. 10 propolis 가 Balb/C nice

marker	control	vehicle 1	vehicle 2	50mg/kg	150mg/kg	300mg/kg
CD- 3	2.11 ± 1.43	2.77 ± 1.71	2.09 ± 0.15	4.15 ± 1.74	3.98 ± 1.11	2.92 ± 1.43
CD- 4	73.04 ± 5.32	70.33 ± 3.91	63.85 ± 8.28	78.13 ± 5.14	82.25 ± 4.28	76.92 ± 2.61
CD- 8	78,14 ± 5,30	79.12 ± 5.29	77.82 ± 1.21	82.48 ± 5.32	85.15 ± 5.14	78.94 ± 5.17
IL- II receptor	1.39 ± 1.31	1.18 ± 1.93	1.44 ± 1.65	1.97 ± 1.60	2.19 ± 1.57	1.89 ± 1.35
TNF- receptor	2.39 ± 0.67	1.05 ± 1.71	1.54 ± 1.70	2.15 ± 0.57	3.20 ± 1.79	1.25 ± 0.79
TCR receptor	28.92 ± 4.23	25.42 ± 5.14	27.54 ± 3.01	33.98 ± 4.32	48.98 ± 4.11	28.14 ± 4.38

% 10, 000 flowcytonetry . n = 5 marker

Interleukin-

가 . II- receptor propolis

T- 6.8% receptor
150ng/kg 11.86% 2 가
II- receptor propolis *in vitro*
가
II- receptor T-
propolis

macrophage가 TNF- receptor
가

IL-II 10
nice propolis TNF-
receptor 9.09% 150ng/kg
2.14% 가
macrophage TNF- receptor

3) Propolis가 T-cell antigen receptor(TCR)

T-cell antigen receptor T- 가 virus

cytotoxicity

Table 4-11 4-12

propolis		가
19.31%	150ng/kg	31.81%
28.92%		150ng/kg
		48.98%

propolis

2. Propolis가 T- Interleukin-
in vitro

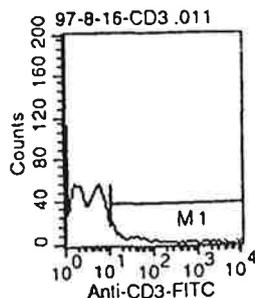
가.

Propolis가	T-	Interleukin-
Ficoll isopaque		2 x 10 ⁶
	PBS	propolis 50ppm
16	CO2 incubator	anti-human
CD25-FITC(IL-2 receptor)		Flowcytonetry (Becton Dickinson)

Propolis		anti-human CD25-FITC(IL-2
receptor)	Fig. 4-8	spontaneous IL- receptor
4.9%	propolis 50ppm	10.25%
2		
<i>in vitro</i>		propolis <i>in vivo</i>

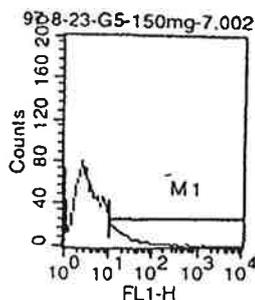
가 propolis가

가 가 가
가 .



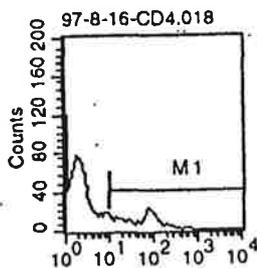
File: 97-8-16-CD3 .011 Total Events: 10000

Marker	% Total	Mean	SD	CV
All	100.00	11.17	192.56	1634.79
M1	6.96	113.07	694.27	605.15



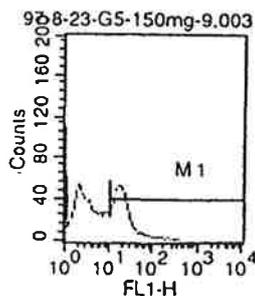
File: 97-8-23-G5-150mg-7.002 Total Events: 10000

Marker	% Total	Mean	SD	CV
All	100.00	6.92	20.38	294.41
M1	12.95	26.72	52.19	195.32



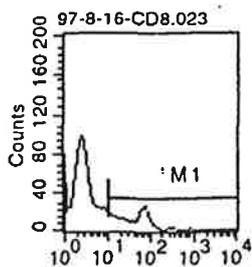
File: 97-8-16-CD4.018 Total Events: 10000

Marker	% Total	Mean	SD	CV
All	100.00	13.48	33.45	248.11
M1	19.18	60.02	56.07	93.41



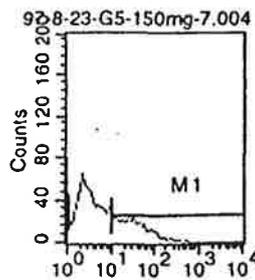
File: 97-8-23-G5-150mg-9.003 Total Events: 10000

Marker	% Total	Mean	SD	CV
All	100.00	12.73	23.00	180.65
M1	41.71	24.97	31.66	126.79



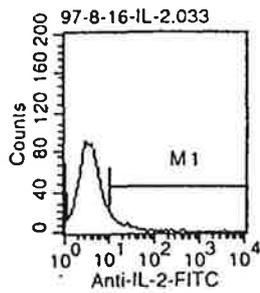
File: 97-8-16-CD8.023 Total Events: 10000

Marker	% Total	Mean	SD	CV
All	100.00	10.89	21.59	198.29
M1	18.41	44.79	33.26	74.25



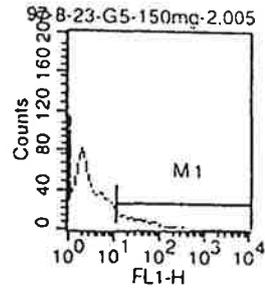
File: 97-8-23-G5-150mg-7.004 Total Events: 10000

Marker	% Total	Mean	SD	CV
All	100.00	14.40	26.83	186.35
M1	29.52	39.43	39.21	99.46



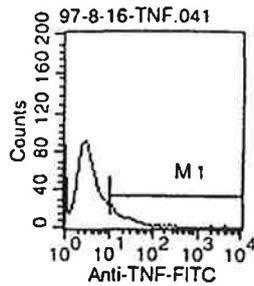
File: 97-8-16-IL-2.033 Total Events: 10000

Marker	% Total	Mean	SD	CV
All	100.00	5.04	29.90	593.40
M1	6.44	23.99	116.10	484.00



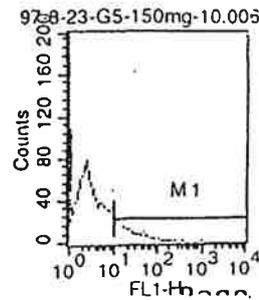
File: 97-8-23-G5-150mg-2.005 Total Events: 10000

Marker	% Total	Mean	SD	CV
All	100.00	6.45	14.39	222.99
M1	11.62	31.16	32.48	104.23



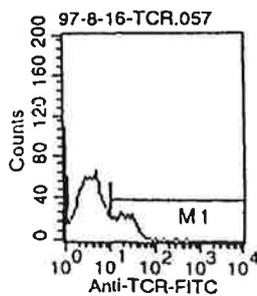
File: 97-8-16-TNF.041 Total Events: 10000

Marker	% Total	Mean	SD	CV
All	100.00	5.97	39.05	654.11
M1	9.87	27.20	122.19	449.18



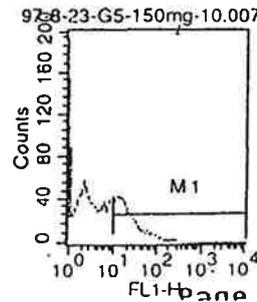
File: 97-8-23-G5-150mg-10.006 Total Events: 10000

Marker	% Total	Mean	SD	CV
All	100.00	7.86	90.91	1157.29
M1	12.31	39.00	256.98	658.88



File: 97-8-16-TCR.057 Total Events: 10000

Marker	% Total	Mean	SD	CV
All	100.00	8.01	11.10	138.50
M1	20.46	23.59	16.72	70.88

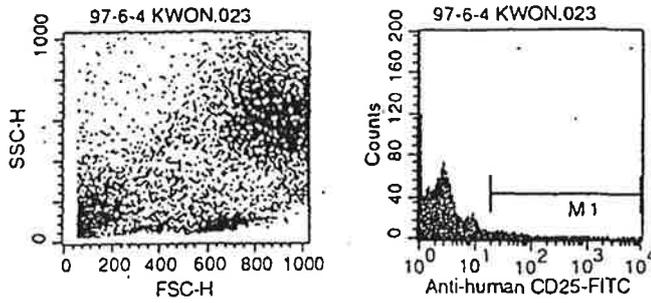


File: 97-8-23-G5-150mg-10.007 Total Events: 10000

Marker	% Total	Mean	SD	CV
All	100.00	11.19	17.81	159.14
M1	32.54	25.80	25.36	98.31

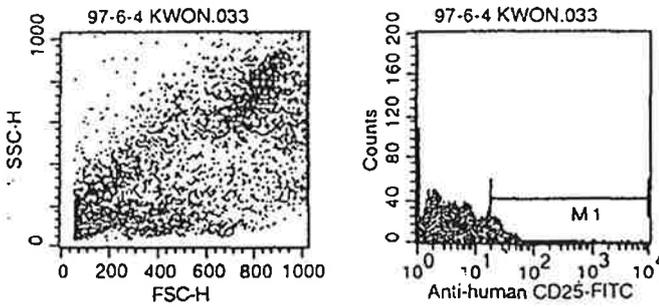
Fig.4-7. Representative histogram of Balb/C splenocytes specially reactive with different monoclonal antibody using flowcytometry for detection of the effect of propolis

1) Control



Marker	Left, Right	Events	% Gated	% Total	Mean	Geo Mean	CV	Median	Peak Ch
All	1, 9910	10000	100.00	100.00	15.66	2.82	1446.46	2.39	1
M1	20, 9910	490	4.90	4.90	258.38	69.12	384.48	48.26	24

2) Propolis 50ppm



Marker	Left, Right	Events	% Gated	% Total	Mean	Geo Mean	CV	Median	Peak Ch
All	1, 9910	10000	100.00	100.00	11.10	4.95	829.70	4.41	1
M1	20, 9910	1025	10.25	10.25	60.11	32.91	470.48	26.90	21

Fig.4-8. Representative histogram of Human T-lymphocytes specially reactive with CD-25(Interleukin-II receptor) monoclonal antibody using flowcytometry for detection of the effect of propolis

3. Propolis 가 ICR mice Sarcoma 180

propolis *in vivo* propolis가
가

가.

1)

ICR 5 × 10⁶ Sarcoma 180

, 1

가

cutting , PBS(phosphate buffered saline)

sarcoma 180

50nl cornical tube . PBS

1,000rpm 4 5 washing 3

. 5 ICR 5 × 10⁶ Sarcoma 180

2) Propolis 가

propolis

가

Sarcoma 180 7 , calliper

5mm가

propolis

, propolis 300mg/kg 600mg/kg

zonde

(1)

propolis sarcona 180 (2)

$$(1) = 4/3 \times a^2b/2 \quad (a: \quad , b: \quad)$$

$$(2) = \frac{C - T}{\text{control}} \quad T :$$

Table 4-13 4-14 propolis 가

Table 4-13. Propolis 가

days	8	16	22	29	36
	68.5	148.6	127.1	118.3	102.7
300mg/kg	69.6	87.8	62.9	57.5	63.9
600mg/kg	65.3	47.7	23.5	19.5	38.6

8 , 10

a) mm3

16 300mg/kg propolis

148.6mm3 87.8mm3

40.9% 600mg/kg

47.7mm3 67.9% 29

118.3mm3 300mg/kg 57.5

mm3 51.4% 600mg/kg propolis

19.5mm3 83.5%

가
 가 ICR mice xenograft
 propolis
 propolis
 Table 4-16 propolis

Table 4-14. Propolis가

days	8	16	22	29	36
	-	0%	0%	0%	0%
300mg/kg	-	40.9%	50.5	51.4	37.8
600mg/kg	-	67.9	81.5	83.5	62.4

a) $\frac{(C - T)}{\text{control}}$ T :

Table 4-15. Propolis

days	8	16	22	29	36	43
	37.2%	37.6	39.1	40.4	40.6	41.5
300mg/kg	37.2	38.6	38.4	39.7	40.4	40.8
600mg/kg	37.2	39.8	39.7	41.2	42.5	41.9

a) g

4. Propolis가

가.

1)

Propolis가 *in vitro*

human gastric carcinoma (KATO III), human lung carcinoma (A549), human breast adenocarcinoma (MCF-7), mouse lymphoma (Sarcoma 180), mouse lymphoma (Yac-1) Korea Cell Line Bank(KCLB)

Hep3B (Human hepatocellular carcinoma)

American Tissue Culture Collection(ATCC, HB8064)

Hep 3B() Sarcoma 180 RPMI-1640

10% Fetal bovine serum, 37 , 5% CO₂

Dulbecco's modified Eagle's medium(DMEM, 6.7g)

nutrient mixture F-12(5.3g) 1:1(V/V) mixture, gentamicin sulfate 40ng, sodium bicarbonate 2g, hepes buffer 2g 3

pH 7.0-7.2 20% FBS 37 (5% CO₂

incubator) 24 . Hep 3B() Sarcoma 180

RPMI-1640 10% Fetal bovine serum, 37 , 5%

CO₂

2) *in vitro* SRB(sulforhodanine B) method

가

SRB(sulforhodanine B)

96well plate

well 5 × 10⁴/ml cell line(MCF-7, A549, Hep3B)

100μl 가 37 (5% CO₂ incubator) 24

0.1µg/Me, 1µg/Me, 5µg/Me 25µg/Me A
propolis가 100µl 가 48 . 48
aspirator cooling TCA(trichloro acetic acid,
10%) 100µl 가 4 1 .
5 well 1% acetic acid SRB
(sulforhodanine B) 100µl 가 . 30
1% acetic acid 5 . 10nM Tris
buffer 150µl ELISA reader 540nm

3) MTT(3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromine)
method propolis cell viability

MTT assay cell colorimetric assay
cell dehydrogenase enzyme
MTT dark blue formazan .

Table 4-16.

Cell line	Target organ	cell
Sarcoma 180	mouse lymphoma	DMEM
YAC-1	T-lymphoma	RPMI-1640
MCF7		RPMI-1640
KATO		RPMI-1640
Hep 3B		DMEM
A549		RPMI-1640

96 well microtiter plate KATO III , Sarcoma 180, Yac-1 cell 10%
fetal bovine serum RPMI 1640 1 × 10⁵/ml

well 180 μ l 37 , 5% CO₂ 24 incubation 0.1 μ g/
 M ℓ , 1 μ g/M ℓ , 5 μ g/M ℓ 25 μ g/M ℓ A propolis
 가 volume 200 μ l가
 48 incubation MT (2mg/M ℓ) 50 μ l
 well 가 fornazan 4 incubation
 aspirator 150 μ l DMSO(dinethylsulfoxide) 가
 fornazan ELISA reader 540nm

4)
 propolis
 가
 가
 O. D. (100%) O. D.

Table 4-17 4-18 SRB method MT method
 5 \times 10⁵/well, 1 \times 10⁷/well 2 가

4-17 MCF7
 0.1 μ g/M ℓ 10%가
 Table 4-18 KATOIII 25 μ g/M ℓ
 propolis 1 \times 10⁷/well 49.4%, Hep3B
 25 μ g/M ℓ propolis 1 \times 10⁷/well 50.7%
 A549 25 μ g/M ℓ 1 \times 10⁷/well

44. 3%가

4-18

Sarcona 180 Yac-1 5µg/Ml 1 × 10⁵/well
 27.9% 29.8% 25µg/Ml

Table 4-17. propolis I

	Sarcona 180		Yac-1		MCF7	
	5 × 10 ⁵ /well	1 × 10 ⁷ /well	5 × 10 ⁵ /well	1 × 10 ⁷ /well	5 × 10 ⁵ /well	1 × 10 ⁷ /well
control	-	-	-	-	-	-
0.1µg/Ml	2.6	8.4	1.4	1.0	14.0	12.1
1µg/Ml	13.4	14.8	7.6	5.2	19.9	12.6
5µg/Ml	19.2	28.9	19.9	29.8	17.2	25.4
25µg/Ml	15	21.3	14.9	22.6	36.8	33.6

a) %, triplicate

Table 4-18. propolis II

	KATO		Hep 3B		A549	
	5 × 10 ⁵ /well	1 × 10 ⁷ /well	5 × 10 ⁵ /well	1 × 10 ⁷ /well	5 × 10 ⁵ /well	1 × 10 ⁷ /well
control	-	-	-	-	-	-
0.1µg/Ml	7.2	8.7	5.0	11.8	4.3	3.6
1µg/Ml	11.8	12.1	7.5	13.9	6.3	18.2
5µg/Ml	25.9	28.0	18.4	23.1	35.1	33.5
25µg/Ml	38.1	49.4	50.1	50.7	27.3	44.3

a) %, triplicate

5. *in vitro*

propolis

propolis가

polymorphonuclear granular neutrophil (PMN :)

가

propolis가

neutrophil

propolis

in vivo

가

가.

Double-density Percoll[®]

neutrophil chemiluminescence media phenol
red가 1% BSA가 가 RPMI-1640 1 × 10⁶/ml
propolis 10ppm, 50ppm, 100ppm 가
neutrophil 30 , 1.5 , 10 , 11.5 13.5
respiratory burst reactive
oxygen(RO) luminol amplifier 5
O₂⁻(superoxide anion) C3b opsonization
zynosan particle stimulator 40 luminometer(Berthold
LB-9505) chemiluminescence(CL)

Table 4-19 propolis

	30	1.5	50ppm	100ppm	propolis
10	CL	36.85%	가	CL	.
10ppm	propolis	가	CL	11.5	.
	30	1.5	가	46.51%	가
	10ppm	neutrophil		108.40%	가
	13.5			가	.

Table 4-19. Propolis가 neutrophil chemiluminescence

	Control(%)	100ppm(%)	50ppm(%)	10ppm(%)
0.5	100a)	102.56	100.43	103.19
1.5	100	100.96	101.31	107.19
10.0	100	101.39	102.14	136.85
11.5	100	104.36	135.40	146.51
13.5	100	118.24	156.82	208.40

a) = 4.044 × 109 cpm

50ppm	neutrophil	11.5	35.40%
가	CL , 13.5	56.82%	가 , 100ppm
	11.5	가	13.5
	18.24%	가	.
	propolis		

가

propolis

neutrophil

propolis

가

가

.

.

4 HPLC propolis

, ,
caffeic acid
(3,4-dihydroxycinnamic acid), p-coumaric acid(4-hydroxycinnamic
acid), cinnamic acid(3-phenyl-2-propenoic acid) rutin (flavone
glycoside) 4가 HPLC .

(FAO, 1996)

가

가 (Lavie, 1976)

160 가

(Greenway et al., 1990) ,

4가

가

1.

가. Propolis

Propolis ()

8 11

가

HPLC methanol (CH₃OH), acetic acid(CH₃COOH), water(H₂O)

ethanol (C₂H₅OH) J. T. Baker .

가 10g propolis 99% ethanol

72 evaporator(Hei dol ph
VV2000)

ethanol, water propolis volume 1ng/ul

, injection volume 5μl . μ Bondapak C18 ,
10μm column(3.9 × 150mm) Kontron 422 HPLC pump, detector
430A, thermostat controller 402 .

CH₃OH, CH₃COOH, H₂O 60 : 35 : 5 .

chromatogram 272nm .

2.

가.

Caffeic acid antiviral activity(Konig and Dustnann, 1985),
antibacterial activity Gram (+) Gram (-) (Villanueva
et al., 1970) , anti-inflammatory activity(Bankova et
al., 1983) .

Cinnamic acid

Staphylococcus aureus . p-coumaric acid
anti-microbial and anti-mycotic activities(Metzner et al., 1979)

. Rutin flavone glycoside anti-viral (Konig and
Dustnann, 1985) gastric ulcer (The hive and the honey
bee, 1992) .

Table 4-20 Fig. 4-9 propolis
caffeic acid (3,4-dihydroxycinnamic acid), p-coumaric

acid(4-hydroxycinnamic acid), cinnamic acid (3-phenyl-2-propenoic acid) rutin(flavone glycoside) 99% ethanol
 (Fig. 4-10) propolis caffeic acid p-coumaric acid
 2-3 cinnamic acid rutin .
 propolis propolis 가
 propolis

Table 4-20 ethanol propolis
 propolis caffeic acid (3,4-dihydroxycinnamic acid),
 p-coumaric acid (4-hydroxycinnamic acid), cinnamic acid
 (3-phenyl-2-propenoic acid) rutin(flavone glycoside)
 가

Table 4-20 Fig. 4-11 ethanol propolis
 cinnamic acid(3-phenyl-2-propenoic acid)
 3-5 가

Table 4-20 Fig. 4-12 ethanol propolis
 p-coumaric acid(4-hydroxycinnamic acid)
 ethanol propolis
 caffeic acid (3,4-dihydro-xycinnamic acid), cinnamic
 acid(3-phenyl-2-propenoic acid) rutin (flavone glycoside)
 Fig. 4-11

Table 4-20 ethanol propolis

caffeic acid (3,4-dihydroxycinnamic acid), p-coumaric acid(4-hydroxycinnamic acid), cinnamic acid(3-phenyl-2-propenoic acid) rutin(flavone glycoside)

propolis

Table 4-20. Fig. 4-13 ethanol propolis
rutin(flavone glycoside) 가

ethanol propolis 3가 , caffeic acid (3,4-dihydroxycinnamic acid), p-coumaric acid (4-hydroxycinnamic acid), cinnamic acid(3-phenyl-2-propenoic acid)

Table 4-20 ethanol propolis ,
caffeic acid (3,4-dihydroxy- cinnamic acid), p-coumaric acid (4-hydroxycinnamic acid), cinnamic acid(3-phenyl-2-propenoic acid), rutin(flavone glycoside)

ethanol propolis

가

가

Table 4-20 ethanol propolis rutin (flavone glycoside) caffeic acid(3,4-dihydroxy-cinnamic acid), p-coumaric acid (4-hydroxycinnamic acid), cinnamic acid (3-phenyl-2-propenoic acid)

Table 4-20 Fig.4-14 ethanol propolis
caffeic acid (3,4-dihydroxy-cinnamic acid), p-coumaric acid(4-hydroxycinnamic acid), cinnamic acid (3-phenyl-2-propenoic acid) rutin(flavone glycoside)

poplar

polyphenols

(Aga

et al., 1993)

propolis

(FAO,

1996) propolis 가

propolis 가 , propolis
가

Table 4-20. caffeic acid(3,4-dihydroxycinnamic acid), p-coumaric acid(4-hydroxycinnamic acid), cinnamic acid (3-phenyl-2-propenoic acid) rutin(flavone glycoside)

Country	caffeic acid ($\mu\text{g/g}$)	p-coumaric acid ($\mu\text{g/g}$)	cinnamic acid ($\mu\text{g/g}$)	rutin ($\mu\text{g/g}$)
China A(w)	331.59(33.82)	101.92(15.44)	18.35(8)	110.60(9)
China A(e)	102.02(15.12)	52.95(12.53)	45.23(24.78)	151.0(11.35)
China S(e)	33.25(12.88)	18.41(11.39)	14.47(20.72)	62.65(12.31)
Newzealand(e)	91.95(9.080)	50.12(7.75)	112.28(40.97)	103.11(5.16)
Poland(e)	46.68(6.46)	234.30(58.44)	17.92(7.28)	64.24(3.52)
Korea J(e)	37.69(15.38)	19.10(14.04)	26.24(31.43)	60.37(9.75)
Korea K(e)	86.91(9.35)	50.44(8.67)	44.75(17.80)	n.d
Spain(e)	22.14(9.29)	54.53(36.51)	10.69(16.58)	19.62(4.17)
Eastern-europe	88.66(16.79)	45.78(13.84)	37.84(26.48)	14.45(13.88)
Brasil(e)	30.07(13.56)	1.2(0.88)	5.80(9.68)	n.d

* (w) - water extracted propolis, (e) - ethanol extracted propolis.

* (w) - water extracted propolis, (e) - ethanol extracted propolis.

* China A - A level propolis,

China S - Sanghi propolis,

Korea J - Junnan,

Korea K - Kangwon.

* () - Rel. Ar %.

* N. D. - not detected.

* Linear regression : $y = a + bx$ ($a = -0.96065$, $R = 0.99801$)

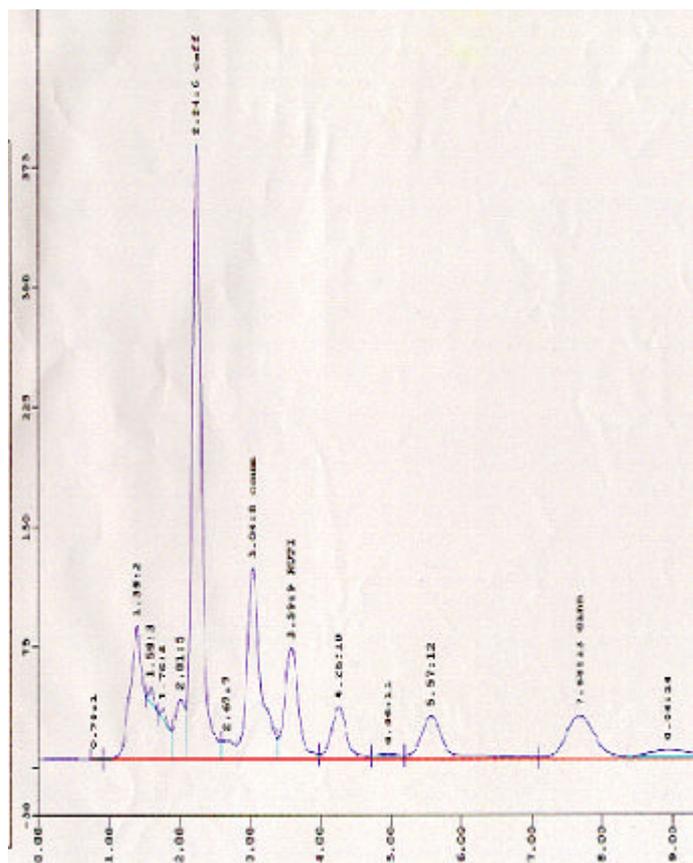


Fig. 4-9.

propolis

- 1) caffeic acid (3, 4-dihydroxycinnamic acid) - 2.24(s)
- 2) p-coumaric acid(4-hydroxycinnamic acid) - 3.04(s)
- 3) rutin(flavone glycoside) - 3.59(s)
- 4) cinnamic acid(3-phenyl-2-propenoic acid) - 7.68(s)

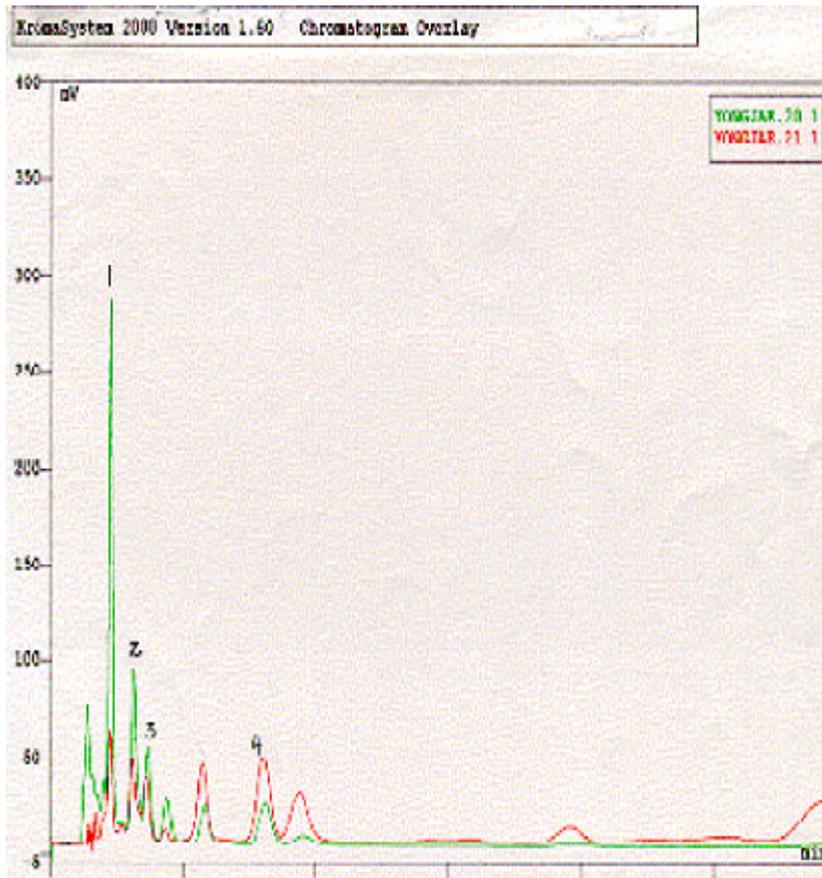


Fig. 4-10. ethanol propolis propolis

- * Green colour -
- * Yellow colour - ethanol propolis
- 1) 1-caffeic acid (3,4-dihydroxycinnamic acid) - 2.21(s)
- 2) 2-*n*-coumaric acid(4-hydroxycinnamic acid) - 3.04(s)
- 3) 3-rutin(flavone glycoside) - 3.59(s)
- 4) 4-cinnamic acid(3-phenyl-2-propenoic acid) - 7.68(s)

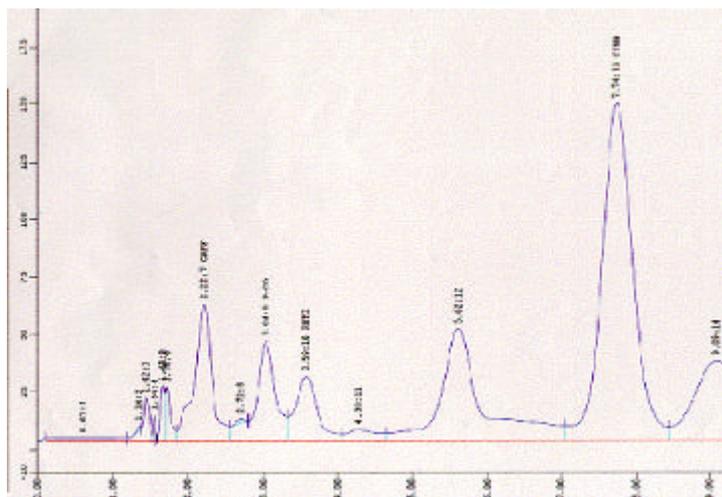


Fig. 4-11. ethanol propolis
 1) caffeic acid (3,4-dihydroxycinnamic acid) - 2.22(s)
 2) p-coumaric acid(4-hydroxycinnamic acid) - 3.04(s)
 3) rutin(flavone glycoside) - 3.59(s)
 4) cinnamic acid(3-phenyl-2-propenoic acid) - 7.74(s)

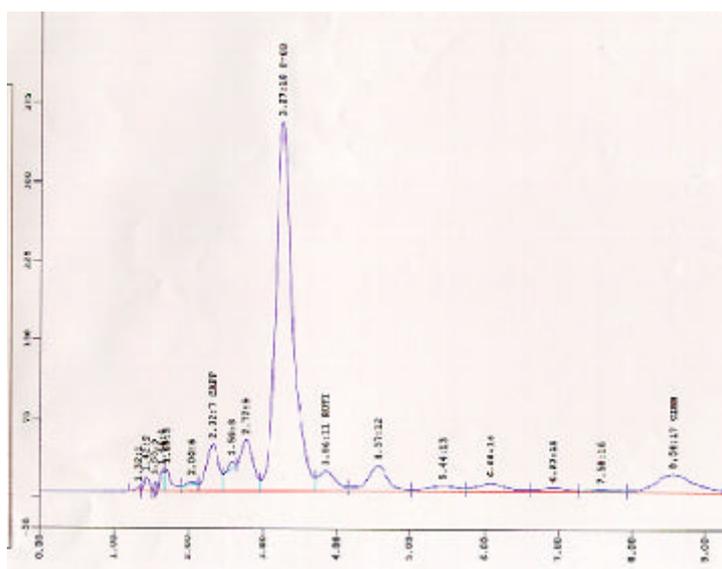


Fig. 4-12. ethanol propolis
 1) caffeic acid (3,4-dihydroxycinnamic acid) - 2.32(s)
 2) p-coumaric acid(4-hydroxycinnamic acid) - 3.27(s)
 3) rutin(flavone glycoside) - 3.86(s)
 4) cinnamic acid(3-phenyl-2-propenoic acid) - 8.54(s)

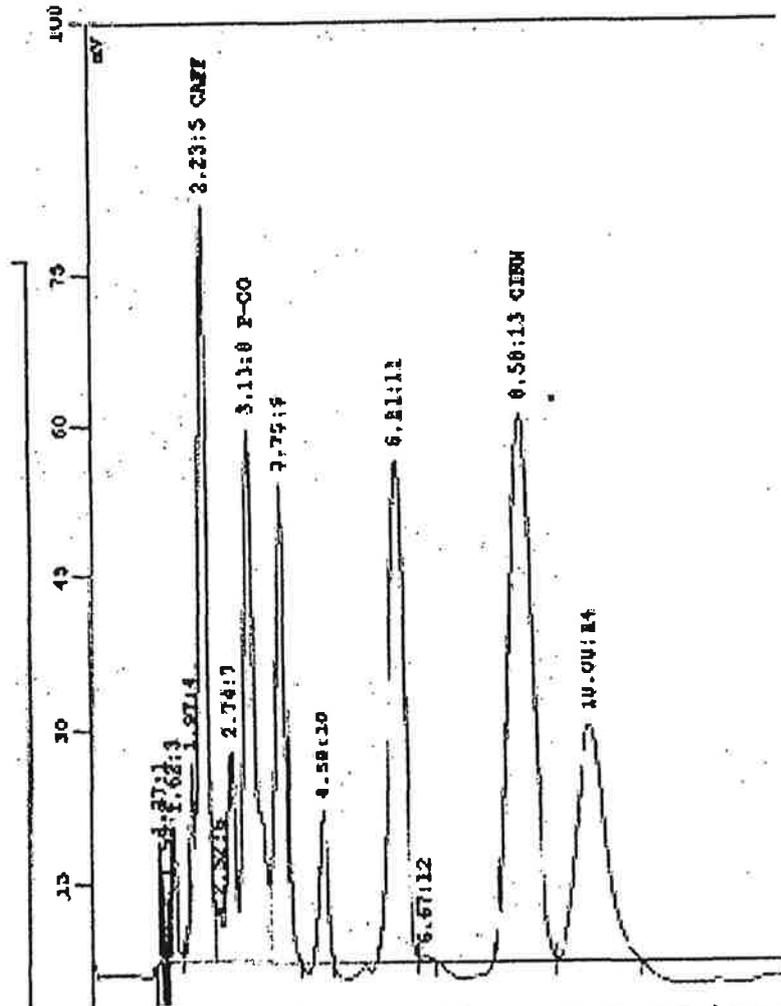


Fig. 4-13. 한국 강원산 ethanol 추출 propolis의 크로마토그램

- 1) caffeic acid (3,4-dihydroxycinnamic acid) - 2.23(s)
- 2) p-coumaric acid(4-hydroxycinnamic acid) - 3.13(s)
- 3) rutin(flavone glycoside) - no detectable
- 4) cinnamic acid(3-phenyl-2-propenoic acid) - 8.58(s)

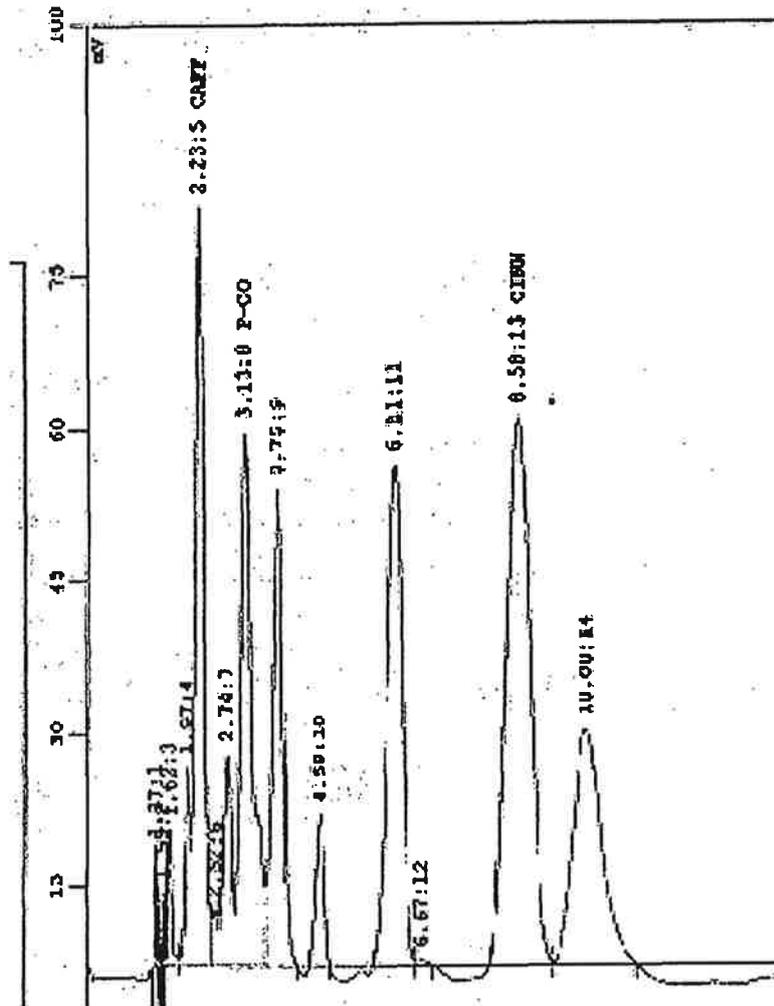


Fig. 4-14. 브라질산 ethanol 추출 propolis의 크로마토그램

- 1) caffeic acid (3,4-dihydroxycinnamic acid) - 2.08(s)
- 2) p-coumaric acid(4-hydroxycinnamic acid) - 2.93(s)
- 3) rutin(flavone glycoside) - no detectable
- 4) cinnamic acid(3-phenyl-2-propenoic acid) - 6.89(s)

5 Propolis 가

1. Propolis

in vitro propolis
propolis 가
가
propolis

가.

1)

1998 6 8 2
5 5
,
Propolis ,
propolis , propolis
, 5
1 3
propolis .

2)

10
1 2 .

3)

E. coli K88ac
가 tryptic soy broth 37
24 , Minca medium
37 24 . *E. coli* 1 ×
10⁹/Mℓ 1:1 1 1.5

4) propolis

가 propolis 300mg 10Mℓ ethyl alcohol
100Mℓ가 saline

5)

, , propolis
Culturette™ (Becton Dickinson Co.) *E.*
coli Petrifilm™ (3M Co.) colony count

6)

Baytril
2.5Mℓ/100kg
propolis 15g 1 2

1) *E. coli*

Table 4-21

		<i>E. coli</i>		가	가
		2		가	가
		propolis		가	가
		propolis		가	가
		<i>E. coli</i>		가	가
					1
					3

Table 4-21. *E. coli*

(6)	
	0/6
Propolis 2) + <i>E. coli</i> b)	0/6
<i>E. coli</i> + propolis ,	6/6
<i>E. coli</i> + propolis) d)	6/6
<i>E. coli</i> + e) e)	6/6

a) = 300mg/day

b) = 1, 500M ℓ (1x10⁹/M ℓ)

c) = 150mg

2

d) =1M ℓ /100kg/day

e) = 2. 5M ℓ /100kg

2) propolis

Table 4-21 propolis 2
 300mg *E. coli* 가
 가 propolis

Table 4-22 propolis *in vitro* propolis 가 Gram (-) Gram (+) *in vitro*
 1 가
 가
 enrofloxacin Baytril positive control
E. coli propolis 300mg
 6
 가 24

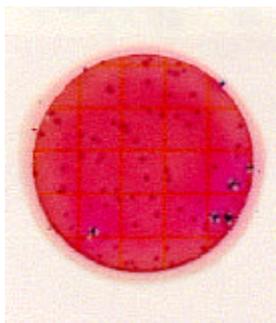
Culturette™(Becton Dickinson Co.)
E. coli Petrifilm™(3M Co.) colony count . Fig. 4-15
 5.0×10^4 CFU/g
 1.4×10^6 CFU/g 가
 propolis 2 2.4×10^5 CFU/g
E. coli propolis 150mg 1Mℓ/100kg
 6 propolis 6 *E. coli*
 propolis
 enrofloxacin 2.5Mℓ/100kg 6

Table 4-22. *E. coli*

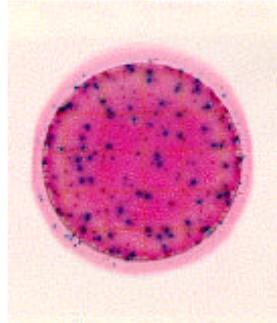
propolis

(6)	
	-
propolis a)(2) + <i>E. coli</i> b)	6/6
<i>E. coli</i> + propolis ,	6/6
<i>E. coli</i> + propolis c)	6/6
<i>E. coli</i> + e)	6/6

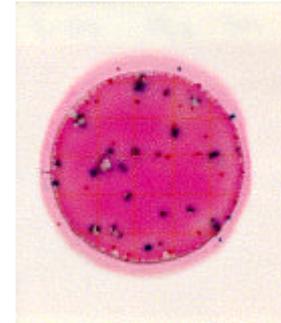
a) = 300mg/day h) = 1.500Ml(1x10⁹/Ml) c) = 150mg
 d) = 1Ml/100kg/day e) enrofloxacin(Baytril) 100mg/Ml = 2.5Ml/100kg



A)
(5.0 × 10⁸CFU/g)



B) *E. coli*
(1.4 × 10⁶CFU/g)



C) propolis
(2.4 × 10⁶CFU/g)

Fig. 4-15. Propolis

propolis

flavonoids

가 propolis 가

가 .

in vitro propolis

E. coli Gram (-)

가 *in vivo*

가 propolis

가 spectrum

.

2. Propolis

propolis 가

가

.

가.

1)

1999 6 11 70

.

, Propolis

, propolis ,

7 5 2

propolis .

2)

35

7

1 2

1

2

3)

E. coli K99

tryptic soy broth 37 24

가

Minca medium 37 48 .

E. coli K99 1 × 10⁹/Mℓ 1:1

100 200ml .

4) propolis

B propolis 150mg 10Mℓ ethanol

100Mℓ가 saline .

5)

, propolis

Culturette™(Becton Dickinson Co.)

E. coli

Petri film™(3M Co.)

colony count

6)

Baytril

2.5

Mℓ/100kg

propolis

15g 1 2

Table 4-23. *E. coli* K99

propolis

(7 2)		
	0/14	-
2 propolis a)	0/14	14/14
+ <i>E. coli</i> b)		
<i>E. coli</i> K99	14/14	14/14
+ propolis ,		
<i>E. coli</i> K99 +	14/14	14/14
propolis c) d)		
<i>E. coli</i> K99 +	14/14	14/14
e)		

a) = 150mg/day

b) = 100-200Mℓ (1x10⁵/Mℓ)

c) = 150mg , , 3

d) = 1Mℓ/100kg/day

e) = 2.5Mℓ/100kg/day

Table 4-23

E. coli

2

propolis

150mg

E. coli

가 가

propolis

propolis 가
in vitro propolis 가 Gram (-)
 Gram (+) *E. coli*
in vivo
 가
 enrofloxacin Baytril positive control
E. coli K99 가
 propolis 150mg , 3
 6
 가 24
 48 -72
 , propolis
 Culturette™(Becton Dickinson Co.) *E.*
coli Petrifilm™(3M Co.) colony count
 6.0 × 10⁴CFU/g
E. coli 1.8 × 10⁶CFU/g *E. coli* 가
 propolis propolis
 2 1.0 × 10⁵CFU/g
 propolis
 flavonoids
 가 propolis 가
 가

6 Propolis

1. Propolis

가

propolis

propolis

가

in vitro

가.

1)

Table 4-24

가

10⁸/M^l

DMSO(dimethyl sulfoxide)

propolis

6mm

paper disc 15 μ l

35 \pm 1

positive control

penicillin

ampicillin(disc potency : 10 μ g)

quinolone

norfloxacin(disc potency : 10 μ g)

propolis

50,000ppm 10

5ppm

Table 4-25. Propolis *Staphylococcus uberis*

	Propolis					ampicillin	norfloxacin
	50,000ppm	5,000ppm	500ppm	50ppm	5ppm	(10 μ g)	(10 μ g)
inhibition (mm)	4	1.8	1	1	N.D.	N.D.	6

N. D. = not detectable

Staphylococcus

aureus

propolis 500ppm

Table 4-26. Propolis *Staphylococcus aureus*

	Propolis					ampicillin	norfloxacin
	50,000ppm	5,000ppm	500ppm	50ppm	5ppm	(10 μ g)	(10 μ g)
inhibition (mm)	4	2.8	2	1	N.D.	15	13

N. D. = not detectable

Table 4-26 *Staphylococcus agalactiae* *Staphylococcus*

aureus propolis가 50,000ppm, 5,000ppm 500ppm
 가 5,000ppm 4mm *Staphylococcus*
aureus 가
 propolis가 Gram (+)

Table 4-27. Propolis *Staphylococcus agalactiae*

	Propolis					ampicillin	norfloxacin
	50,000ppm	5,000ppm	500ppm	50ppm	5ppm	(10 μ g)	(10 μ g)
inhibition (mm)	4	4	2	1	N.D.	13	9

N.D. = not detectable

Table 4-27 Gram(-) *E. coli*
 ampicillin propolis

propolis가 propolis

Table 4-28. Propolis *E. coli*

	Propolis					ampicillin	norfloxacin
	50,000ppm	5,000ppm	500ppm	50ppm	5ppm	(10 μ g)	(10 μ g)
inhibition (mm)	4	3	1	N.D.	N.D.	N.D.	7

N.D. = not detectable

2. TTC(Triphenyl tetrazolium chloride) reduction test
propolis 가

가.

1)

가) : *Staphylococcus thermophilus*

) 10%

) (10% 12

10% 1:1 .)

) , Incubator, , pipette etc.

2) Propolis 가 test

Zero control TMP control tube skim milk(10%)

8nl propolis가 가 skim

milk 8nl . zero control 1nl , TMP control

가 1nl TMP . 82±2

2 30 가 37

1nl 가 ,

Incubator(37) 2 300μl TIC 가

incubator(37) 60 . TMP control zero control

TMP control ,

TMP control

Table 4-29 propolis TTC-reduction test
 1ppm 100ppm propolis
 가 propolis

Table 4-29. Propolis *Staphylococcus thermophilus* TTC

Propolis		
1ppm	(-)	
5ppm	(-)	
10ppm	(-)	
50ppm	(-)	
100ppm	(-)	

- 1) Vollenweber.E., Hausen. B.M., Greenaway. W. : Phenolic constituents and sensitizing properties of propolis, poplar balsam and balsam of peru. *Bull. Groupe. Polyphenols*. 15, 112-120 (1990)
- 2) Ayala. F., Lenbo.G., Nappa.P. and Balato.N. : Contact dermatitis from propolis. *Contact dermatitis* 12, 181-182 (1985)
- 3) Bankoba. V., Christov.R., Kujungiev.A. : Chemical Composition and Antibacterial Activity of Brazilian Propolis, *Z. Naturforsch.* 50c, 167-172 (1995)
- 4) Greenaway, W., May, J., Scaysbrook. T. and Whatley, F. R. : Identification by gas-chromatography mass spectrometry of 150 compounds in propolis. *Z. Naturforsch.* 111-121. (1990)
- 5) Gabrys.J., Konecki. J., Krol. W., Scheller.S. : Free amino acids in bee hive product as identified and quantified by gas-chromatography. *Phaarm. Res. Commun.* 18, 513-518(1986)
- 6) Bankova.V.S. : A study of flavonoids of propolis. *J. Nat. Products.* 4, 471-475 (1982)
- 7) Scheller, S., Gazda, G., Krol, W., Czuba, Z., and Shani, J. The ability of ethanolic extract of propolis to protect mice against gamma irradiation. *Zeitschrift fur Naturforschung* 1049-1052

- 8) Serkedjieva.J., Manolova.N. : Anti-influenza virus effect of propolis constituents and their analogues. *J.Nat. Products* 55, 294-297 (1992)
- 9) Anoros.M., Sinoes.C.M.O.: Synergistic effect of flavonoids and flavonols against herpes simplex virus type I in cell culture. *J. Nat. Products* 55, 732-740 (1992)
- 10) Grange.J.M., Davey.R.V. : Antibacterial properties of propolis(bee glue). *J. Royal Soc. Med.* 83, 159-160 (1990)
- 11) Scheller.S., Szaflarski.J., Tustanowski.J., Nolewajka.E. and Stojko.A. : Biological properties and clinical application of propolis I. Some physicochemical properties of propolis I. Some physicochemical properties of propolis. *Arzneim. Forsch/Drug Res.* 27, 889-890.
- 12) Scheller.S., Krol.W., Swiacik.J., Owczarek.S., Gabrys.J. : Antitumoral property of ethanol extract of propolis in mice-bearing ehrlich carcinoma as compared to bleomycin. *Z.Naturforsch* 44, 1063-1065 (1989)
- 13) Grunberger.J., Banerjee.R., Eisinger.K., Oltz. E.M. : Preferential cytotoxicity on tumor cells by caffeic acid phenethyl ester isolated from propolis. *Experientia* 44, 230-232 (1988)
- 14) Krol.W., Czuba.Z., Scheller.S., Gabrys. J., Grabi ec.S. and Shani.

J. : Anti-oxidant properties of ethanol extract of propolis as evaluated. *Biochemistry international*. 21, 4, 593-597 (1990)

15) Sud'ina. G. F., Mirzoeva. O. K., Pushkareva. M. A., Korshunova. G. A., Sunbatyan. N. V. and Varfolomeev. S. D.: Caffeic acid phenethyl ester as a lipoxigenase inhibitor with antioxidant properties. *FEES* 329, 1, 2, 21-24 (1993)

16) Krol. W., Scheller. S., Czuba. Z., Matsuno. T., Zydowicz. G., Shani. J., Nos. M. : Inhibition of neutrophils' s chemiluminescence by ethanol extract of propolis and its phenolic components. *Journal of Ethnopharmacology* 55, 19-25 (1996)

17) Mirella Nardini, Fausta Natella, Vincenzo Gentili, Maurizio Di Felice, and Cristina Scaccini : Effect of caffeic acid dietary supplementation on the antioxidant defense system in rat. *Archives of biochemistry and biophysics* Vol. 342, No. 1, 157-160, (1997)

18) Scheller. S., Wilczok. T., Inielski. S., Krol. W., Gabrys. J. and Shani. J. : Free radical scavenging by ethanol extract of propolis. *Int. J. MALAI. EICL*. 57, 3 461-465 (1990)

19) Misic. V., Ondrias. K., Gergel. D., Bullova. D. : Lipid peroxidation of lecithin liposomes depressed by some constituents of propolis. *Fitoterapia LX2* 215-220 (1991)

20) Krol. W., Czuba. Z., Scheller. S., Gabrys. J., Grabi ec. S. and Shani.

J. : Antioxidant properties of ethanol extract of propolis as evaluated. *Biochemistry international*. 21, 4, 593-597 (1990)

21) Volpert.R. and Elstner.E.F. : Biochemical activities of propolis extracts. I. Standardization and antioxidative properties of ethanolic and aqueous derivatives. 2. *Naturforsch*. 48c 851-857 (1993)

22) Tracey KJ, Beutler B, Lowry SF, Merryweather J, Wolpe S, Milsark IW, Hariri RJ, Fahey TJ, Zentella A, Albert JD, et al. Shock and tissue injury induced by recombinant human cachetin. *Science* 234: 470-474.

23) Calandra, T., Baugartner, J.D., Grau, G.E. *et al.* Prognostic values of tumor necrosis factor/cachetin, interleukin-1, interferon- γ , and interferon- β in the serum of patients with septic shock. Swiss-Dutch J5 immunoglobulin study group. *J. Infect. Dis.* 161, 982-987(1990)

24) Van der Poll, T. and Lowry, S.F. Tumor necrosis factor in sepsis: mediator of multiful organ failure or essential part of host defense? *Shock* 3, 1-12(1995)

25) Auphan, N., Di Donato, J.A., Rosette, C. *et al.* Immunosuppression by glucocorticoids : inhibition of NF- κ B activity through induction of I κ B synthesis. *Science* 270, 286-290(1995)

26) Biswas, D.K., Ahlers, C.M., Dezube, B.J. *et al.* Pentoxifylline

and other protein kinase C inhibitors down-regulate HIV-1 NF- κ B induced gene expression. *Mol. Med.* 1, 31-43.

27) Young, P., McDonnell, P., Laydon, J. *et al.* A novel MAP kinase regulates the production of IL-1 and TNF in LPS activated human monocytes. *Cytokine* 6, 564

28) Bianchi, M., Bloom, O., Raabe, T. *et al.* Suppression of proinflammatory cytokines in monocytes by a tetravalent guanylylhydrazone. *J. Exp. Med* 183, 927-936

29) Tappel, A.T., Lipid peroxidation damage to cell components. *Fed. Proc.*, 32(8), 1870-1874, 1973.

30) Misra, H.P. and Fridovich, I., Univalent reduction of oxygen by reduced flavins and quinones, *J. Biol. Chem.*, 247, 188, 1972.

31) Merrill, A. H., Lambeth, J. D., Edmondson, D.E., and McCormic, D. B., Formation and mode of action of flavoproteins, *Annu. Rev. Nutr.*, 1, 281, 1981.

32) Haber, F. and Weiss, J., The catalytic decomposition of hydrogen peroxide by iron salts, *Proc. R. Soc. London Ser. A*, 147, 332, 1934.

33) Bilinski, T., Krawiec, Z., Licznanski, A., and Litwinska, J., Is hydroxyl radical generated by the Fenton reaction *in vivo*, *Biochem. Biophys. Res. Commun.*, 130, 533, 1985.

- 34) Kanofsky, J. R., Singlet oxygen production by lactoperoxidase. Evidence from 1270nm chemiluminescence, *J. Biol. Chem.*, 258, 5991, 1983.
- 35) Khan, A. U., Gebauer, P., and Hager, L. P., Chloroperoxidase generation of singlet molecular oxygen observed directly by spectroscopy in the 1.0 to 1.6 μ m region, *Proc. Natl. Acad. Sci. U.S.A.*, 80., 5195, 1983.
- 36) Khan, A. U., Enzyme system generation of singlet molecular oxygen observed directly by 1.0-1.8 μ m luminescence spectroscopy, *J. Am. Chem. Soc.*, 105, 7195, 1983.
- 37) Vidigal Martinelli, C., Zimmer, K., Kachar, B., Duran, N., and Cilento, G., Emission from singlet oxygen during the peroxidase-catalyzed oxidation of malonaldehyde, *FEBS Lett.*, 108, 266, 1979.
- 38) Cadenas, E., Sies, H., Nastainczyk, W., and Ullrich, V., Singlet oxygen formation detected by low-level chemiluminescence during enzymatic reduction of prostaglandin G₂ TO H₂, *Hoppe-Seyler's Z. Physiol. Chem.*, 364, 519, 1983.
- 39) Rucker, R. R. and Mueller, G. C., Effect of oxygen tension of HeLa cell growth, *Cancer Res.*, 20, 944, 1960.

- 40) Drew, R. M., Painter, R. B., and Feinendegen, L. E., Oxygen inhibition of nucleic acid synthesis in HeLa S3 cells, *Exp. Cell Res.*, 36, 297, 1964.
- 41) Clément, A., Hubscher, U., and Junod, A. F., Effects of hyperoxia on DNA synthesis in cultured porcine aortic endothelial cells, *J. Appl. Physiol.*, 59, 1110, 1985.
- 42) Junod, A. F., Effects of oxygen intermediates on cellular functions, *An. Rev. Respir. Dis.* 135, S32, 1987.
- 43) Anes, B.N. and Saul, R.L., Oxidative DNA damage as related to cancer and aging, in Genetic Toxicology of Environmental Chemicals, Part A, Alan R. Liss, New York, 11., 1986.
- 44) Vuillaume, M., Reduced oxygen species, mutation, induction and cancer initiation, *Mutat. Res.*, 186, 43, 1987.
- 45) Sies, H., Ed., Oxidative Stress, Academic Press, London, 1985.
- 46) Krol.W., Scheller.S., Czuba.Z., Matsuno.T., Zydowicz.G., Shani.J. : Inhibition of neutrophil's chemiluminescence by ethanol extract of propolis and its phenolic components. *J. Ethnopharmacology* 55. 19-25. (1996).
- 47) Sakaguchi.S., Kanda. N., Hsu.C.C., Sakaguchi.O., *Microbiol. Immunol.*, 25, 229(1981)
- 48) Sakaguchi.O., Kanda.N., Sakaguchi. S., Hsu.C.C., Abe. H.,

Microbiol. Immunol., 25, 229(1981)

49) Regina Volpert and Erich F. Elstner. Biochemical Activities of Propolis Extracts I. Standardization and Antioxidative Properties of Ethanolic and Aqueous Derivatives. *Z. Naturforsch.* 48c, 851-857 (1993)

50) Shuhei Sakaguchi, Eiji Tsutsumi, and Katsushi Yokota. Preventive Effects of a Traditional Chinese Medicine against Oxygen Toxicity and Membrane Damage during Endotoxemia. *Biol. Pharm. Bull.* 16(8) 782-786 (1993).

51) Eun-Mee Choi, Jyung-Rewng Park and Jung-Sook Seo. : Effect of Beta-Carotene Supplementation on Lipid Metabolism and Related Enzyme Activities in Rats. *J. Korean Soc. Food Nutr.* 23(5), 743-749 (1994).

52) Tsutomu, N., Hiranitsu and Kawakishi, S : The protective role of gallic acid ester in bacterial cytotoxicity and SOS responses induced by hydrogen peroxide. *Mutation Res.*, 303, 29(1993)

53) Shuhei Sakaguchi, Shinobu Furusawa and Ken-ichi Sasaki : The enhancing effect of tumor necrosis factor on oxidative stress in endotoxemia. *Pharmacology and Toxicology* 1996, 79.

54) Rangel-Frausto, M.S., Pittet, D., and Costigan, M : The natural history of the systemic inflammatory response syndrome(SIRS). *JAMA*

273: 117-123, 1995.

55) Aral, K., Lee.F., Miyajina.A., Miyatake.S. : Cytokine, coordinators of immune and inflammatory responses. *Annual Review of immunology*, 1997.

56) Baggiolini, M., Dewald. B., and Moser. B. : Human chemokines an up date. *Annual Review of Immunology*, 1997.

5

가

1

가

1.

가.

가

1)

1950

가

5-10%

.

.

,

가

가

.

가

.

가

.

.

2)

1995 , 1996 3,939 , 4,574
16%
가 , 가
· ,
·
1996 1,052 ,
408 , 617 .
가 가
· ,
가 332 가 .
·
가 가 가
가 , 10%
105 , ,
·
·
·
가 , 1996
· , 41가
·

가

가

,
가

가

가

cleaning

가 가

가

가

1)

(absorbant)

composting

1970

steroid saponin

가

2)

10

cost formulation)

가

ICF(least

(carrier)

(Engelhard

)

가

10

.(가 100 /kg)

Sarsaponin

Urease inhibitor

glycocomponent가 2

2

5- 10%

10

1995 20

11

<Table 5-1>

	(peat)	
	sarsaponin chemical: , , , formalin	, 10

<Table 5-2>

	Biochem	DPI	Alltech
	Sarsa	Micro Aid	Deodorase
Sarsaponin	14%	8%	10%
가	US \$ 25/Kg(p)	US \$ 20/Kg	US \$ 35/Kg

: (), -50, (),
 ()

. Propolis

1)

20

가 , ,
 , .
 , 가
 , ,
 가 .

2)

가 , 가
 . 가

가 , 가 . 가 1996 39
 15.6 가 가 (, 1 4kg 가).
 가 10 20 .
 가 , 가 .
 가 3 /kg 30-60 .

<Table 5-3>

1995	43,258	764,243	27,781	369,825	15,477 394,418
1996	39,678	719,224	24,900	319,460	14,478 399,764

:

BC 3,000 1899
 2 . ,
 1) : CONAP 120
 ,
 2) : 가 200 , 300 가
 . 10 , 2
 . 80%가 , , , ,
 . 1987

3)

:

3-6

가

가

가

(10%, 20%, 30%)

가

(water soluble)

40%

5% 가

60

30

가

PG, mineral oil

(10%, 20%)

PG, mineral oil

가 가

가 (GRAS)

가 .

<Table 5-4>

가	SiO ₂	(10, 20, 30%) (5%) 10	SiO ₂
		(),	
	water Mineral oil Propylene glycol	(10, 20%)	

. Sarsaponin

1)

Steroid sarsaponin

2)

0.13g/l , 0.43g/l, 0.97g/l

, ()

500

가

가

2,000

. 가

가

Kg 300

1,000

가

(boiling)

가

30%

가

가

,

,

가 가

,

spray drying

가

,

가

.

가

.

가

가

nacerati on

, polysorbate

.

2.

가

가.

1970

(1%),

가 가 . 1996 UR

가

, ,

, 가

가

.

가

가 .

1) : ,

, DHA

2) : ,

,

3) 가 , : , , 가

,

,

가

가

UGF

가

가

<Table 5-5>

가

가

가

가

가

5-10%

가

()

1997

450 , 250

50% 가

가 가

<Table 5-5>

			가		
OK	1992	1,900 /10	200,000	가	
	1988	1,400 / 6	100,000		
	1993	1,800 /10	50,000		
	1995	1,900 /10	50,000		
	1994	1,950 /10	50,000		
dha	1991	1,700 /10	200,000		
	1994	1,400 / 6	50,000		DHA
	1991	1,700 /10	100,000		
300	1995	2,000 /10	100,000		DHA
	1981	1,200 / 6	150,000		
	1986	1,650 /10	50,000		

가가

1)

2) 가

2

1.

가.

1

가

가

가

가

.

.

가

가

가

가

가

가

가

가

가
가

가

가

가

가

2000

가

가

UR

가

,

가가

가 , ,

propolis 가 가가 가

<Table 5-6> 가

	TYPE	TYPE	TYPE	TYPE
	가	가	가	가

local

brand

national brand (NB)

NB

가

private brand (PB) 3 가 . 「 1996

」

가 4가

78

가

1kg

265

170

600

가

1.6

2 가

<Table 5-7> 1kg

type	1kg ()					
			가			
)	326 (100.0)	135 (41.4)	45 (13.8)	82 (25.2)	3 (0.9)	61 (18.7)
)	282 (100.0)	107 (38.0)	44 (15.6)	72 (25.5)	7 (2.5)	52 (18.4)
)	179 (100.0)	80 (44.7)	42 (23.5)	40 (22.3)	7 (3.9)	10 (5.6)
)	189 (100.0)	103 (54.5)	29 (15.3)	20 (10.6)	7 (3.7)	30 (15.9)
)	164 (100.0)	85 (51.8)	26 (15.8)	20 (12.2)	6 (3.7)	27 (16.5)
)	147 (100.0)	69 (46.9)	29 (19.7)	22 (15.0)	4 (2.7)	23 (15.7)

Type
) 「 」 : , ,
 「 」 : . , ,
 가 , 가
 type 454 가 type 184
 ()
 3 4
 가,
 ()

<Table 5-8> 1kg 가 :

type						
	(A)	(B)	(C)	(D)	(A)/(B)	(C)/(D)
(A)	326	282	179	189	1.64	1.47
가(B)	454	340	226	450	1.90	1.84
(C)	128	58	47	261	2.6	3.7
(C)=(A)-(B)						

<Table 5-9> 가 : /kg

type	가		가		(A)/(B)	(C)/(D)
	(A)	(B)	(C)	(D)		
	454	270	-	-	1.7	-
	340	297	310	226	1.2	1.4
	450	263	253	190	1.7	1.3
	-	-	255	184	-	1.4
	415	277	273	200	1.5	1.4

가 가 .
 가 PB , NB
 NB , PB ,
 가
 가
 IMF

가 1997

1 347 , , 310

가 가

2,000

<Table 5-10>

	()	()	GNP* (10)	가 * (/10)
1988	6,992	42,031	131,371.3	448.2
1989	6,701	42,449	147,941.6	609.5
1990	6,810	42,869	178,262.1	576.0
1991	7,370	43,286	194,458.8	427.9
1992	7,891	43,663	204,231.0	489.5
1993	7,824	44,056	216,162.4	392.6
1994	7,728	44,453	234,333.3	429.1
1995	7,990	44,851	254,704.9	486.4
1996	8,315	45,248	272,323.8	468.9
1997	8,469	45,657	285,621.2	484.8

: * 1990 가

: 가 ,

가 , GNP,

가 , 가 가 가 가

가 가 , 가 ,
 , GNP
 GNP 가 가

$$\log EQ_t = \alpha + \beta \log P_t + \lambda \log EP_t$$

$EQ_t = t$
 $P_t = t$
 $EP_t = t$ 가
 $\alpha =$
 $\beta, \lambda =$

1988
 1997 10 가
 1990 가
 (log-log)

$$\log EQ_t = -17.698 + 2.562^{***} \log P_t - 0.138^* \log EP_t$$

(7.45) (- 1.95)

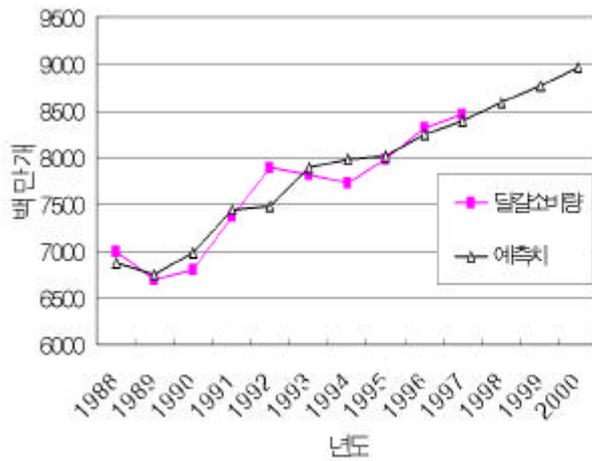
R2=0.89

() t

*** : p>1%

* : p>10%

가 1% 가 2.562% 가 가
 1% 0.138% . ,
 가 가가 가
 propolis가 가 가가
 가



<Fig. 5-1>

1997 85 가
 8,500 (100)
 가 가 2,000
 90 1

<Table 5-11>

				가
		59%	가	.
43%			1g	가
1,200				
	1,000		1g	5
				가가
	1,800			
1		\$	1,200	
1,423.5	가		1,200	
가				
	가	<Table 5-11>		1
		가		
가				60%

<Table 5-12> . <Table 5-12>

		1997		1
16,738		1300		
	가	1,447		.
10		10	53	
			1.5	2
			가	가가
	가	가	,	,
	가			

가

<Table 5-12> , 10

가								
					(A)			
		1200	1300	1400	1200	1300	1400	
가	3,933	3,933			3,933			3,933
	11,279	13,607	13,615	13,761	12,703	12,794	12,885	13,396
	600	600			600			600
	113	45			45			45
	813	813			813			813
	16,738	18,998	19,006	19,152	18,094	18,185	18,276	18,787
10								
	606.4	688	689	694	656	659	662	681

) : 「 1997
 가 ,
 (가) = + 가 1
 (가) : × 40%
 (가) : 60%)
 10 = / 276 × 10 (1
 : 276)

가
 가
 C 22,400 , propolis
 2,600 , 25,000 110

<Table 5-13> .

<Table 5-13>

				propolis		
			1			
1 1-3 7 8-9 10		25cc/1,000	5 0.7	3ppm/1	2.64	4.08
11-12 17 18-20 20		30cc/1,000	6 15	5ppm/1	4.4	6.8
21-23 27 28-29 30 31-33 40	SP	100g/1,000 100cc/1,000 500g/1,000	4.4 20 20 40	25ppm/1	44	68
50 60 70 80 90 110	SP BNE,	1 /1,000 200g/1,000 150g/1,000	13 8.8 80 150	50ppm/1	264	408
			362.9		337	520.88

: propolis
 가
 propolis 1ppm(1ng) 가:
 < 6> 1,400 0.088 ,
 0.136
 propolis =1 × × propolis 가 가
 120 287 , propolis 2

3 .
 propolis 가
 가 가
 propolis 가 .
 110
 1 362.9 propolis
 315.04 , 486.88
 1 47.86
 .
 110-130 가
 propolis
 가 가 . < 8>
 120 287 가
 propolis 2 propolis
 가 . propolis
 propolis 가 가 가
 가가
 가
 .
 가 가
 .
 Caucasians(*Apis mellifera caucasica*) .
 가

700-800

가

(crude

propolis)

가 ,

가 ,

,

가

.

(, , ,

가 ,

,

가 ,

,

,

,

,

,

,

,

,

,

,

,

,

,

,

,

,

,

)

,

가

가

.

가 가

.

20

US\$30/kg

US\$40/kg 가

, , ,

,

,

,

가

가

가

partiy()가 가

1

A

, B

C

A

B

A

100-200

가 가

가

(fresh royal jelly)

800 1,000

70%-80%

500g 1,000g

가

(省)

base

3

100 (A 30

/B 70)

network()

5

가

A

가

3) propolis

1 100g

300g

7-8

가

가

가

가

가 .
가

.

4)

가

, , 가
가 가

가

가

.

,

.

,

.

, , , ,
가

novable-frame hives

traditional log hive()

novable-frame hive()

top-bar hive가 .

가

.

.

.

가 , bee wax

가

· ·

가 base

· ,

Beeking & Develpment, Bee World, American Bee Journal, Apicultural Abstracts, Indian Bee Journal Chinese Bee Products Newspaper

·

·

가

·

2.

가.

· 가가

가

· 가
가

가

·
가가

가
 Sarsaponin yucca
 가 yucca saponin saponin
 80
 가 , 가

<Table 5-14> steroid kg 가

	가 (kg) (A)	(kg) (B)	(C)	1g 가 (A+B)/C
Sarsaponin	\$20	12,000	20g/kg	1,800 a
	8,000		15g/kg	1,900 b
	0		6g/kg	2,000 c
가	0		10g/kg	1,200
	0		5g/kg	2,400

- 1. Sarsaponin 1 1200 , 1300 , 1400
 a : 1200 , b : 1300 , c : 1400
- 2. 가
- 3. ethanol butanol
- 4. sarsaponin crude saponin
 16.9%

sarsaponin kg 20
 20 가 , 가 , kg 8,000
 가 . saponin
 kg 12,000 sarsaponin kg 20g,
 가 15g, 6g, 가 10g, 5g
 . 1g 가 1,300
 sarsaponin 1,900 (1,330), 가
 (1,200) sarsaponin .

<Table 5-15> 가 1

	Poultry		Swine			Cattle			
	Layer (20)	Broiler (6)	Stater (3-5kg)	Grower (20-50kg)	Finisher (50-80kg)	Beef (400kg)	Dairy (600kg, 20kg)		
(kg)	550g/	1070g/	250g/	1855g/	2575g/	7000g/	6400g/		
saponin 가 (ppm)	28.5	55.64	91.25	677.08	939.88	2555	2336		
가 (g)	50	100	120	100	60	200	200		
	1.43	5.56	10.95	67.7	56.4	511	467.2		
()	Yucca*	A	2,574	10,008	19,710	121,860	101,520	919,800	840,960
		B	2,717	10,564	20,805	128,630	107,160	970,900	887,680
		C	2,860	11,120	21,900	135,400	112,800	1,022,000	934,400
		1,902	7,395	14,564	90,041	75,012	679,630	621,376	
		2,860	11,120	21,900	135,400	112,800	1,022,000	934,400	
	가	1,716	6,672	13,140	81,240	67,680	613,200	560,640	
	3,432	13,344	26,280	162,480	135,360	1,226,400	1,121,280		

) Yucca A, B, C 1,200 , 1,300 , 1,400

saponin 가 Agro industrial

<Table 5-14>

1g 가

가

<Table 5-15> .

sarsaponin

Yucca 가 100

<Table 5-16> 가

	1200	1300	1400
Yucca	100	100	100
	73.9	70.0	66.5
	111.1	105.3	100.0
가	66.7	63.2	60.0
	133.3	126.3	120.0

<Table 5-16>

Yucca 가 100 1200 가
 73.9% 가 가 66.7%
 1,400
 가 가
 가 가
 가

가

1991

가

가
가
가

<Table 5-17>

	가 ()	(M/T)	(ha)
1988	35,807	14,834	11,698
1989	38,843	13,939	11,877
1990	36,404	13,889	12,184
1991	36,146	15,132	11,694
1992	30,809	13,508	11,364
1993	30,327	14,874	10,986
1994	30,500	14,292	10,123
1995	23,172	11,971	9,375
1996	23,304	10,147	8,940

:

가
가
가
가
가
가
가
가

가

가 가 . 가 가 . 가
가 가 .
가 가 가 .

1)

n 가 $I_1 \dots I_n$ 가 , 가 $V_1, V_2, \dots V_n$
 . $I_i \ I_j$ a_{ij}

$$a_{ij} = \frac{W_i}{W_j} \quad (1)$$

가 .

$A = [a_{ij}]$.

$$A = \begin{pmatrix} \frac{W_1}{W_1} & \frac{W_1}{W_2} & \dots & \frac{W_1}{W_n} \\ \frac{W_2}{W_1} & \frac{W_2}{W_2} & \dots & \frac{W_2}{W_n} \\ \vdots & \vdots & \ddots & \vdots \\ \frac{W_n}{W_1} & \frac{W_n}{W_2} & \dots & \frac{W_n}{W_n} \end{pmatrix} \quad (2)$$

A 가가 A
weight vector .

$$A = \begin{pmatrix} \frac{W_1}{W_1} & \frac{W_1}{W_2} & \dots & \frac{W_1}{W_n} \\ \frac{W_2}{W_1} & \frac{W_2}{W_2} & \dots & \frac{W_2}{W_n} \\ \vdots & \vdots & \ddots & \vdots \\ \frac{W_n}{W_1} & \frac{W_n}{W_2} & \dots & \frac{W_n}{W_n} \end{pmatrix} = \begin{pmatrix} W_1 \\ W_2 \\ \vdots \\ W_n \end{pmatrix} = n \begin{pmatrix} W_1 \\ W_2 \\ \vdots \\ W_n \end{pmatrix} \quad (3)$$

weight vector A vector n
 n A
 A (2)
 가 , A
 vector vector가 가 weight
 vector , A r2,

$$\lambda = \begin{pmatrix} \lambda_1 \\ \vdots \\ \lambda_n \end{pmatrix} \quad (4)$$

vector

$$A \lambda = \lambda_{\max} \lambda \quad (5)$$

A

$$(C.I.) = \frac{\lambda_{\max} - n}{n - 1}$$

$$(C.I.) = 0.1$$

$$(C.R.) = 0.1$$

n	1	2	3	4	5	6	7	8	9	10	11	12
M	0.00	0.00	0.58	0.90	1.12	1.24	1.32	1.41	1.45	1.49	1.51	1.53

$$(C.I.) = \frac{C.I.}{M}$$

가

1)

가

가

가 , (, ,)

(,) .

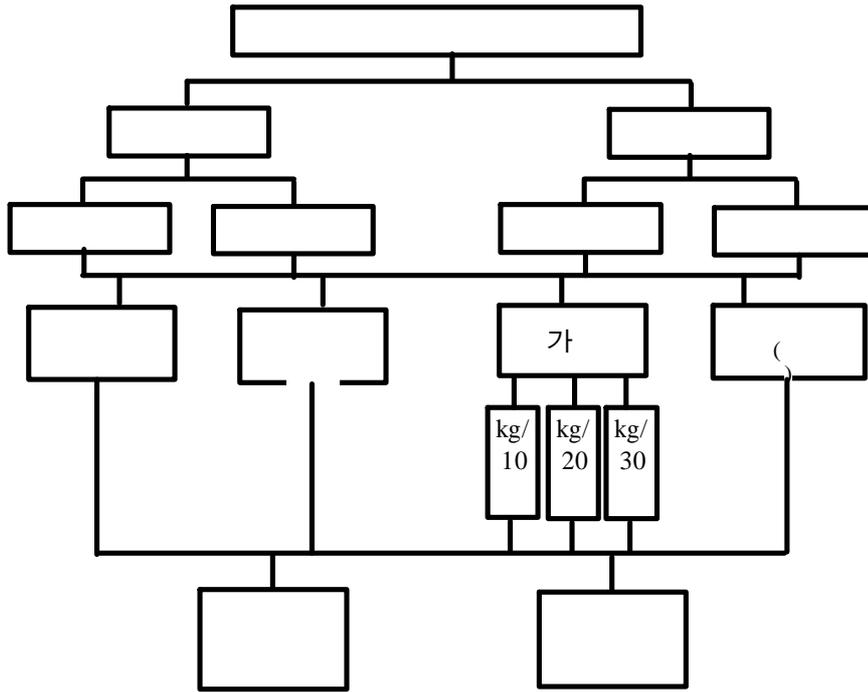
가 가

가

가

가 가

가



<Fig 5-2>

가 1,000 25%, 1000
 75% (가 ,
 , 1998) 0.25:0.75
 50:50
 . 가
 kg 3 , 5 , 10 .
 , () 가 ,

<Table 5-18>

	/	/	/	/	
	0.008	0.014	0.025	0.033	0.080
	0.034	0.031	0.091	0.146	0.302
가	0.025	0.070	0.060	0.098	0.253
	0.058	0.010	0.198	0.098	0.364

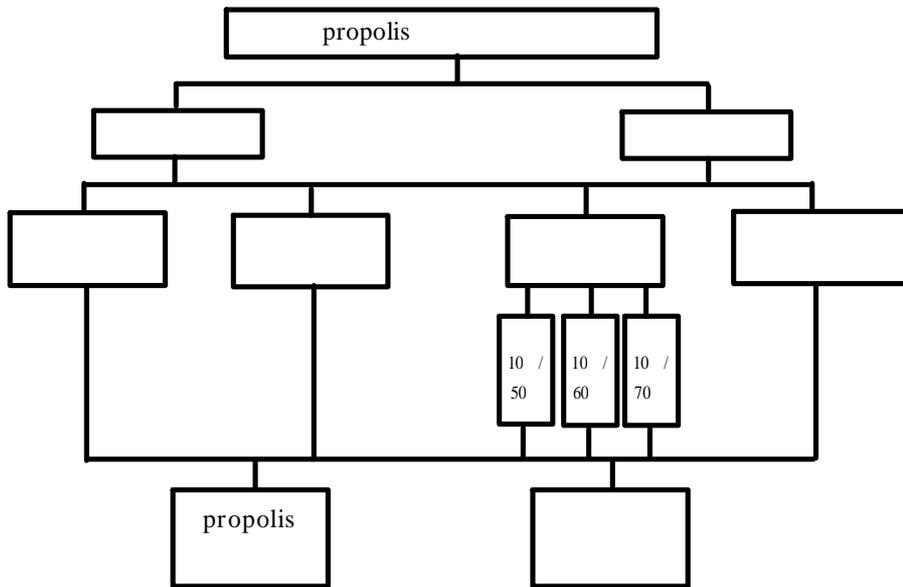
가
 (0.364), (0.302), 가 (0.253),
 (0.080) . 가
 가(0.198) 가
 가 가
 가 가
 (0.01) .
 , ,

<Table 5-18>

(0.626)
 (0.374) .
 가
 가 가
 .
 가

2) Propolis

propolis 가
propolis 가
propolis
propolis
가 가



<Fig. 3> propolis

가

가 가 1
 1 . 10%
 가 90% 1:9

<Table 5-20>

	0.010	0.059	0.069
	0.038	0.466	0.504
가	0.030	0.120	0.150
	0.022	0.255	0.278

propolis 가
 (0.504), (0.278), 가
 (0.150), (0.069) . 가
 가 가
 가 .

	0.010	0.059	0.069
	0.038	0.466	0.504
가	0.030	0.120	0.150
	0.022	0.255	0.278

3

가

가

1.

2

가

Saponin

Propolis

가

.

Propolis

가

가

.

saponin

eucca

sarsaponin

saponin

propolis

saponin propolis

가

saponin propolis

.

가

가

가

가

HACCP

가

.

.

2. 가 가

PSE

A, B

가

가

가

<Table 5-22>

		가		가			
		7,500	-	5,700	4,370	1,800	3,130
		8,000	-	6,400	5,160	1,600	2,840
		-	9,800	6,200	5,970	3,600	3,830
		-	9,800	6,800	6,730	3,000	3,070

: '98. 10

: 1,300: 1

, 가

kg 1800 , 2300

가

400 , 500

kg 1240 1330 (74 77%)

kg 70 230 (2 9%)

가 가
가 가

<Table 5-23> 110kg 가 가

()	90%	137, 979	80%	148, 968	50%	76, 655	50%	93, 107
()	10%	20, 596	20%	50, 507	50%	102, 949	50%	126, 257
	100%	158, 570	100%	199, 475	100%	179, 604	100%	219, 369
	177, 580							
	203, 893							
	19, 010		21, 895		2, 024		41, 789	
	45, 323		4, 410		24, 289		15, 476	

: , , 1999. 1

가 <Table 5-23> .
가 10%, 20% 가
가 158, 570 199, 475 .
177, 580 가 19, 010
21, 895 203, 893
가 45, 323 , 4, 410
. ,

가 가
 50% 2,024
 41,789 가 가
 가 가

<Table 5-24> 가

		: 3.0		: 2.7		
		A	B	A	B	
		196,077	215,685	235,292	215,685	235,292
	가	44,174	44,174		44,174	
		92,198	96,488*		78,945**	
	가	5,209	5,209		5,209	
		1,862	1,862		1,862	
		10,412	10,412		10,412	
		153,855	158,145		140,602	
	가	3,350	3,350		3,350	
		14,150	14,150		14,150	
		171,355	175,645		158,102	
			42,222	57,540	77,147	75,083
		24,722	40,040	59,647	57,583	77,190

: 110kg , 가 , 1999

A () 10% 가
 B () 20% 가
 kg 가 279.4

가 kg
 10 , 3 13 가

* /kg(279.4 +13 =292.4) × (3.0) × 110kg
 ** /kg(279.4 +13 =292.4) × (2.7) × 110kg

가
 가 < 3> .
 가 kg 가 10 , 3
 가
 10% 20% 가
 .
 가
 15,318 52,468 가가 .
 , (3.0) 가
 4,290 () 10% 20%
 19,608 39,215
 가 가 15,318 34,925
 . 2.7
 32,861 52,468 .

3.

가.

UR VIO 1994 1
 21
 . 1997 6 1
 가
 .
 1999 82 74

가 IMF 가
가

1997

1998

<Table 5-25>

		'91	'92	'93	'94	'95	'96	'97	'98	'99(P)
°		516.5	603.4	628.4	650.0	680.4	737.6	767.5	806.0	819.8
	“	-	2.1	10.0	3.9	6.7	4.4	4.0	17.6	17.0
	“	498.8	601.3	618.4	621.1	639.3	691.8	698.7	732.7	740.8
	“	17.7	-	-	25.0	34.4	41.4	64.8	55.7	62.0
(MMA)		-	-	-	-	17.5	23.3	14.6	-	-
°		516.5	603.4	628.4	650.0	680.4	737.6	767.5	806.0	819.8
	“	510.8	584.9	613.2	632.2	661.7	696.9	698.3	700.7	711.6
	“	3.6	8.5	11.3	11.1	14.3	36.9	51.6	88.3	90.0
	“	2.1	10.0	3.9	6.7	4.4	3.8	17.6	17.0	18.2
1	kg	11.8	13.4	13.9	14.2	14.8	15.3	16.0	15.1	15.2
	%	97.7	102.8	100.8	98.2	96.6	99.3	95.5	104.5	104.1

IMF 가 1 1998
15.1kg 0.9kg 1999 0.1kg 가
28.1kg 15.1kg 1998 1 54%

가 7.4kg(26%), 가 5.6kg(20%)

75.2 23.9

32% 가

가가

1990 23.3%

1999 6

54.4%

가

1997 7 1

1997 33.4%

2004 25%

1.2%

. <Table 5-24>

가

가 , 가

1999

1998

가

IMF

1997

1997

70% 가

1999

45%

가

15%

가

가

10%가

(,)

5%가

1998

90%

1999

7

48.8%

가

가

가

가

가

가가

가

가

가

,

가 가 .
 1999 가 10
 11%, 가 17 18% 가
 50 54%, 65%

90 10
 1996 37 1999 90 가
 . 가가 가
 . 가 가
 , 가
 , 1993

1991- 1997
 13,797 가 9,253 1 415
 540 , 500 가
 , 289 .
 1992 973% 가(1998)
) 가
 가 1997

2002 HACCP
 2001 HACCP
 가

HACCP

가

1992 () 가 ‘ ’

가

, , 가 , , ,

가 가

가 . 가

97 7

,

49 (10)가

가 10 , 8 , 6 .

16

가

<Table 5-26>

가, 가 , , ,

1998 3

40%

<Table 5-26>

가

(1999)

.					
(8)	가			,	
				"	
				"	
				"	
				"	
				"	
		가		-	
			A-	,	
(3)			DEA	"	
				"	
				"	
(1)			,		
(10)				-	가
	()		,	,	
	()			"	
	()			"	
				"	
				-	
		()		,	
				"	
				"	
			3	"	
(2)	가			-	
				,	
(3)	()			"	
	()			-	
				,	

.					
(6)				,	
	()			"	
				-	
	가			"	
				-	
				,	
(2)				"	
				-	
(1)				,	
(3)		가		"	
				"	
				-	
(5)				,	
				"	
				-	
	()			,	
				-	
(5)				,	
				-	
				,	
				-	
	()				

가

10-20% 가

가

OEM

1 가

가

가가

가가 가

1 가

kg 1

가가

가

가

1996

가3

가3

DHA

10%,

20%

가

가

가

,

.

.

HACCP

.

.

,

가

가 HACCP

.

가

가

,

가

,

,

,

.

.

<Table 5-27>

		(ng/kg)	
	0	0.006	:5 ()
	5	-	:
	9	-	, , (0.1ng/kg)
	0	0.025	:0 (,), 3 ()
	5	0.020	:
	9	0.025	, , (0.2ng/kg)
	0	0.011	:7 ()
	9	0.006	:
			(0.05), , (0.1)

: , 1992

30

가 가 가 가 가 FDA

. <Table 5-27>

, ,

가

. <Table 5-28>

가 54.4%

27.3% 가 가 80%

가

<Table 5-28>

	54.4%	27.3%	9.1%	9.1%	100%

.

가

12

가

.

가

가

. ,

가

가

가

.

가

.

가

HACCP

,

.

가

가 .
가 가

가 가 .
가 가

.

가 가 .
가 가 가
가 가
가

.

가

, ,

.

4.

IMF

가 .

2002

HACCP

2001

HACCP

가

.

, HACCP , , 1999. 9.
, , 1999. 4.
, , 1992
, 가 ,