

**Development of new products by  
reutilization of food waste**

( ) : 가

:  
:

“ ”

- ( “ 1.  
2.  
3.  
4.  
5.  
6.

“)

1998. . .

- :
- :
- :( )
- :( )
- :
- :
- :
- :
- :
- :
- :

•

•

가.

30%

,

가가

가 .

,

.

,

,

,

film

,

가

.

1)

가 가

15%

가 가

15% 가

가

( )

( ).

가

2)

가 10,000 - 20,000 ppm

, oligo , isoflavon, phytate

isoflavon 가

, cholesterol 가 .

,  
가 가 가 가

3)

가  
가 가 가  
가

4)

가

. 가

. 가

film

.

, film

.

.

5)

가

,

. 가

(

)

가

가

.

가

가

가 .

.

.

( 2 ) 6

가

3

.

,

( , , ) ,

,

( , , )

.

,

.

(

, 1997. 11. )

.

( 2 )

3 pilot plant

( paste , 2 )

---

---

o

o

o

(2 )

o

o

o

---

.

1)

,

.



2) 1

가 pilot

3) 2

thiosulfates

50% ethanol

가 가  
polyphenol flavonoid

4) 3

가

가

5) 4

가

ACE

가

6) 5

가

가

가

가

가 ( , , ) 가

가

( 40 6 p531-535 (1997),

30 5 p1197-1202 (1998)) 가

가

가

7) 6

가

.

.

# Summary

In order to utilize the food waste from food industry, several food products were developed in this research works. In addition to food products development, food wastes produced from industry was statistically surveyed for possible waste utilization as food products. The results are obtained as follow.

## Part 1. Development of natural spice seasoning.

In this work natural spice seasoning products were investigated for their preparation conditions and procedures for effective reutilization of red pepper seeds and garlic root portion, the wastes from spice food industry. For the red pepper seeds(RPS), the seeds were treated with steaming or roasting and then extracted in water with addition of phosphates, organic acid, salt and sugar at various pH and temperature. The results showed that RPS without heat treatment produced better hot taste than heat treated ones. In the temperature range of 70 - 100 °C, the extracts prepared at 80 °C for 30

minutes resulted a relatively high solid yield(27%) with mild color and the highest in hot taste. Among the natural materials added to RPS extracts to improve the harmonized hot taste, mussel was chosen because of its high palatability. The mussel extraction study showed that enzymatic hydrolysis with Protease (commercial) before extraction and addition of  $K_2P_2O_7$  or  $Na_3P_0_4$ (1.0%) followed by extraction at 90 for 40 minutes were optimal in terms of solid yields, color and sensory properties. The preferred mixing ratio of RPS and mussel was 10 : 90(w/w).

For extraction of root portion of garlic, several conditions were studied. It was found that hydrolysis of ground garlic with viscozyme. (a commercial polysaccharase) The ground garlic was ground and followed by extraction at 30 and pH 5.5 for 30 minutes with addition of  $K_3P_0_4$  or  $K_2P_2O_7$  were optimal condition in terms of high solid yields(34-36%) and retention of garlic flavor. The garlic oleoresin extracted with ethanol was stable under vacuum or nitrogen condition with addition of ascorbic acid and cysteine during storage. The discoloration such as greening and browning, which is the major problem in garlic products, was significantly reduced by addition of cysteine, ascorbic acid,  $Na_3P_0_4$  and  $Na_2P_2O_7$ , individually

or in mixture.

The spray dried RPS-mussel extracts and garlic paste processed by pilot plant with using the selected conditions and procedures, were evaluated high in their qualities by industrial personnel.

## Part 2. Conformation and separation of phytochemical compounds in red pepper seeds and garlic root portion

In order to extract the functional components of garlic effectively, solid yields, electron donating ability(EDA), nitrite-scavenging effects(NSE) and peroxide value of extracts from five kinds of organic solvents and four kinds of acids were determined. Methanol extracts were highest in the yield and functionality.

Concentration and pH of methanol and ethanol and the addition effects of citric acid and NaCl to alcohol were conducted to increase the functionality of extracts.

Considering the using to food 50% ethanol were evaluated to best

solvent.

The extracts prepared from boiled garlic for 10-30min and at 40-120 . The EDA, NSE and total thiosulfinates contents were investigated. The EDA and NSE was reduced extending the heating time and raising the heating temperature but reducing rate of EDA was large but that of NSE was little. The total thiosulfinates contents greatly reduced, especially they cannot be detectable when garlic was heated at 80 or more.

When the extracts from heated garlic were stored at 4 , 25 , 50 for 60 days, the EDA and total thiosulfinates contents were decreased more drastically at higher temperature storage, but there was a little change in NSE.

When garlic extracts(0.5-2%) were added to soy sause there were snot observed gas formation and growth of film yeast.

### Part 3. Statistical survey of the waste from food industry

A current status of general solidwastes from food industry sector of which production is up to 7% of total domestic manufacturing

production in 1995 was surveyed by based on the reported statistical natural food resources data.

The Amount of eatable foodstuff in 1995 was approxiually 34,446,000MT. 24%(8,481,000MT) of it were put into further food processing as a raw materials and remainder were used as a food, directly. 1,938,000MT of general solid wastes were produced at the pretreatment(1st processing) grain sand livestock and 3,668,000MT were generated from the food processing(2nd processing), which was approxiually 56% of raw materials. Volume of generated waste was firstly depend on food source type and these kinds of general solidwaste produced from processing of crops, fishery, and meats were produced 311,900MT, 395,300MT, and 12,100MT respectively.

The other type is process specific, such as extracting or conversion of a certain chief ingredient, and the ratio of waste verse raw material were 168%(fermentation), 84.7%(oil extraction), 55.1%(starch production), etc

Annual average of general solid waste generated from each food industry in 1995 was 371.8MT, which was different from manufacturing size, and Type of raw materials and process.



#### Part 4. Separation of phytochemical components from soybean curd whey

Soybean curd wheys (SCW) left after producing tofu from 10 tofu-producing plants were collected and analyzed for the contamination level (COD, BOD), physicochemical properties (viscosity, color, etc.) and functional components (total protein, oligosaccharides, amino acids etc.). For separation and concentration of oligosaccharides and proteins in SCW, ultrafiltration (UF) and nanofiltration (NF) methods were applied. By UF, 90% of total protein and 50% of ash in SCW were removed. NF concentrated oligosaccharides efficiently. The effects of soybean oligosaccharide concentrated from SCW on *Bifidobacteria* and pathogenic microorganism were measured. The contents of heavy metals i.e., Pb, Cd in oligosaccharide were below the threshold level of them by Calpis CO., in Japan. Refined oligosaccharides by ionic filtration were dried to the fine powders with bright white color, and showed no significant differences in sensory quality compared with Japanese products.

The characteristics of soybean whey protein (SWP) concentrated by

UF and NF were compared with isolated soybean protein(ISP). The solubility of SWP was greater than ISP, the emulsifying activity of SWP was less than that of ISP. SWP were fractionated by gel filtration and each fraction was evaluated for ACE inhibiting activity. The fraction superior in ACE inhibition effect was analyzed further by prep-FPLC, TLC and MALDI.

#### Part 5. Preparation and utilization of calcium components from eggshell

Eggshell crushed by a ball mill was separated from its membrane under the optimum conditions (30rpm, 30min, volume fraction 0.08) based on the difference in specific gravity in water. In order to increase the calcium ionization rate, calcium-citric acid complex(CC) and calcium-malic acid complex(CM) were prepared.

The calcium ion concentration of fresh noodle fortified with 1.0%(w/v) of CC and CM were 98 and 105ppm and those of loaf bread were 240 and 220ppm respectively. The sensory evaluation with fresh noodle showed that 0.6% of CC and 0.8% of CM had significant

difference( $p=0.05$ ) as compare to control. As for the loaf bread, both CC and CM at the level of 0.8% had significant difference( $p=0.01$ ).

Concentration of calcium ion eluted from the Ramen fortified with 1.0%(w/v) of CC and CM were 62.8 and 24.8 ppm, respectively. These concentrations were 198 and 78 times higher than control and 11.4 times higher compared to MEP(0.7%, w/v). Both sensory results and reology of Ramen fortified with up to 0.7%(w/v) CM were similar to control. Soy milk fortified with 0.2%(w/v) CC or 0.25%(w/v) CM represents the optimum quality.

In summary, a new method to efficiently separate calcium component and enhance *in vivo* calcium absorption rate was developed. The organic acid eggshell calcium(OAEC) was incorporated with various food including fresh noodle, loaf bread, Ramen, soy milk to determine the threshold concentrations of OAEC without sacrificing their original sensory attributes.

Plant design for producing OAEC on the medium scale was also carried out. Based on the annual production rate of 240 ton/year, mass and energy balances were made along with equipment selection, plant layout, and economic analysis in detail.

Part 6. Development of processing method for biodegradable  
film from potato starch

Potato starch extracted from potato wastes was modified by crosslinking(CL) or hydroxypropylation(HP) and polyethylene(PE) cast films were prepared that contained 5% or 10%(wt) of the modified potato starch. Physicochemical properties of the modified potato starches and mechanical properties of the films were examined and thermal and biodegradability of these films were determined using rapid and representative methods. Water binding capacity increased but blue value, iodine adsorption, solubility, and swelling power decreased with the increase of degree of substitution in CL and HP. CL increased the gelatinization temperature of the starch but HP lowered the gelatinization temperature. CL did not affect the crystal region of starch granule.

Relative crystallinity of the starch was decreased and starch granule was destroyed by HP. Mechanical properties of CL-starch/PE films and HP-starch/PE films except 10.0 HP-starch/PE film were stronger than those of native-starch/PE films. Thermal degradability of the films was evaluated by heat treatment at 70 for 12 weeks.

CL-starch/PE films and HP-starch/PE films were more sensitive on thermal degradability than native-starch/PE film. Biodegradability of the films was determined by the incubation with *Pseudomonas aeruginosa*. CL-starch/PE films and HP-starch/PE films were degraded easier than native-starch/PE film. Therefore, recycling of potato wastes was economically useful as well as mechanical properties of the films containing modified starches were improved and thermal and biodegradability of these films were fast.

가가

.

가

,

.

1.

.

**pH**

, ,

가

.

가

가

가

**80**

**30**

**(27%)**

가

.

가

가

가

.

protease( )

K<sub>4</sub>P<sub>2</sub>O<sub>7</sub> Na<sub>3</sub>P<sub>0</sub>4(1.0%) 가 90 40

10 : 90

가

K<sub>3</sub>P<sub>0</sub>4 K<sub>4</sub>P<sub>2</sub>O<sub>7</sub> 가 Viscozyme

pH 5.5 30 30 가 (34

36%)

가

Ethanol

oleoresin

ascorbic acid

cysteine 가

가

가

cysteine, ascorbic

acid, Na<sub>3</sub>P<sub>0</sub>4 Na<sub>4</sub>P<sub>2</sub>O<sub>7</sub> 가

pilot plant 가

-

가

가 , 가 4

가 methanol 가 . Ethanol methanol

, pH(3 8) , NaCl(1 5%), citric acid(1 5%), NaH<sub>2</sub>PO<sub>4</sub>(0.5 2%) NaH<sub>2</sub>PO<sub>4</sub>(0.5 2%) alcohol 가

가 ethanol

50% . 40 120 10 30

ethanol 50% (EDA) (NSE),

thiosulfates . EDA NSE 가 가 가

가 EDA NSE

. thiosulfates 가 80

가 .가 60 (4 ;

25 ; 50 ) thiosulfates

가 가

10% 20%

. (0.5 2%) 가 30



가

.

3

7%

가

가 . .

34,446 MT(

36,199 MT )

24%

8,481 MT 가

가 (1 가 )

1,938 MT

가 가

56% 3,668 MT

가 가

311.9 MT,

395.3 MT,

12.1 MT

2,949 MT

(168.0%),

(84.7%),

(55.1%)

371M/T(

587M/T)

4

10

BOD COD 17,177 ppm 19,980 ppm

2.7%, 0.434%, 1.214% 0.682

1.139% daidzein 2.505mg/g, genistein 1.224

mg/g 94.3 147.0mg/g

(RCM) polysulphone (PSM) pH 3.5 가

PSM 10 COD 79.25%,

98.42%가 가 , RCM

90%, 50%

sucrose, raffi nose,

stachyose가 3.58%, 0.69%, 3.26% 10

*bi fi dobacteri um*

Pb, Cd

가 (SWP)

(ISP)

SWP가

SWP가

. ANS

SWP S-S

Lysine/argi ni ne 1.71 mole

35% . gel chromatography

ACE

prep- FPLC,

TLC MALDI .

5 .

ball mill

( 30 rpm, 30 ,

volume fraction 0.08) .

(Calcium citrate(CC), Calcium malate(CM))

가 .

CC CM 1.0%(w/v) 가 98, 105ppm

, 240, 220ppm .

CC, CM 가 , 0.6, 0.8% 5%

가 CC, CM 0.8%

1.0% .

CM CC 1.0%(w/v) 가

62.8, 24.8ppm 198 , 78 , MEP(0.7%w/v) 11, 4

. CM 0.7%(w/v) 가

. CC 가 가 0.2%(w/v)

. CM 가 CM 가

0.25% (w/v) .

,  
( ,

, , ) 가

240

.

6 .

,  
가

, 5% 10% polyethylene

,  
가

가 가 가 , 가

가 가 . 가

가

. 가

.

가 . 10%

가

. 70 12 가

. *Pseudomonas*

*aeruginosa*

가

.

,

.

# Content

Part 1. Development of natural spice seasoning.

Chapter 1. Introduction .....	56
Chapter 2. Materials and Methods.....	60
Section 1. Red pepper seeds(RPS) extracts.....	60
1. Materials.....	60
2. RPS extracts preparation.....	60
3. Analysis of physicochemical properties.....	64
Section 3. Garlic and garlic roots.....	65
1. Materials.....	65
2. Control of garlic discoloration.....	66
3. Garlic aqueous extracts and oleoresin preparation.....	68
4. Analysis of physicochemical properties.....	70
5. Sensory evaluation.....	71
Chapter 3. Results and Discussion.....	73
Section 1. Red pepper seeds(RPS) extracts.....	73
1. RPS extracts.....	73

2. RPS-Mussel extracts.....	81
3. Produced of RPS-Mussel seasoning products in pilot plant ....	97
Section 2. Garlic and garlic roots.....	103
1. Control of discoloration.....	103
2. Garlic roots extracts preparation.....	118
3. Garlic oleoresin preparation.....	122
4. Changes during storage.....	126
5. Produced of garlic - paste products in pilot plant.....	131
Chapter 4. References .....	137

Part 2. Conformation and separation of phytochemical compounds  
in red pepper seeds and garlic root portion

Chapter 1 Introduction.....	141
Chapter 2 Materials and Methods .....	143
Section 1 Materials.....	143
Section 2 Methods.....	143
1. Extraction methods.....	143
2. Extraction of heated garlic .....	144
3. Changes in functional properties of extracts by heated garlic during storage at different temperature.....	145



4. Effect of garlic on formation of gas and film in film yeast..	145
5. Effects of garlic on pork lipids.....	147
6. Experiments on physico-chemical methods.....	147
Chapter 3. Results and discussions .....	151
Section 1. Extraction of functional components.....	151
1. Yields and functionality of extracts by organic solvents....	151
2. Yields and functionality of extracts by acids.....	153
3. Yields and functionality of extracts by different concentration of methanol.....	155
4. Yields and functionality of extracts by different concentration of ethanol.....	158
5. Yields and functionality of extracts by different pH of alcohols .....	160
6. Yields and functionality of extracts by 50% ethanol containing different citric acid contents.....	162
7. Yields and functionality of extracts by 50% ethanol containing different NaCl contents.....	165
Section 2. Functionality of extracts by 50% ethanol of heated garlic .....	167
Section 3. Changes in functionality of extracts during storage .....	168

1. Changes in functionality of extracts from fresh garlic during storage.....	168
2. Changes in functionality of extracts from heated garlic during storage .....	171
Section 4. Effects of garlic on lipids of ground pork.....	176
1. Changes of peroxide value.....	176
2. Changes of thiobarbituric acid.....	178
Section 5. Effects of garlic on gas and film formation of film yeasts.....	179
Chapter 4. References.....	182

Part 3. Statistical survey of the waste from food industry

Chapter 1. Introduction.....	186
Chapter 2. Methods and Scopes.....	190
Section 1. Survey methods.....	190
1. Classification of food industry.....	190
2. Survey Target.....	191
3. Literature survey.....	192
Section 2. Case study.....	193

Chapter 3. Results .....	195
Section 1. Current status of food industry.....	195
1. Manufacturing status.....	195
2. Material usage in food processing.....	198
Section 2. Current status of food waste production.....	201
1. Mass of Waste.....	201
Section 3. Waste generation process.....	206
1. Pattern of waste at processing step.....	206
2. Waste production on processing Type.....	212
Section 4. Characteristics of waste for reuse.....	224
1. Characteristics of waste.....	224
2. Reuse and recycling.....	226
Chapter 4. References.....	228

Part 4. Separation of phytochemical components from soybean curd whey

Chapter 1. Introduction.....	229
Chapter 2. Material and methods.....	233
Section 1. Characteristics of soybean-curd whey and soybean soaking water.....	233
1. Materials.....	233

2. Analysis of contamination.....	233
3. Proximate analysis.....	234
4. Quantitative analysis of functional components.....	235
Section 2 . Recovery and concentration of functional components	
.....	240
1. Materials.....	240
2. Recovery and concentration of functional components using	
ultrafiltration.....	240
3. Recovery and concentration of functional components	
using nanofiltration.....	240
Section 3. Purification of soybean oligosaccharide from UF/NF	
.....	241
1. Quantitative analysis of soybean oligosaccharide.....	241
2. Measurement of other components.....	241
3. Effects of soybean oligosaccharide on <i>bi f i dobacterium</i> activity	
.....	241
4. Analysis of physical characteristics of purified soybean	
oligosaccharide.....	243
Section 4. Recovery of soybean oligosaccharide using other methods	
.....	243
1. Purification of oligosaccharide using ion exchange resin....	243
2. Recovery of soybean oligosaccharide by Steffen method.....	244

3. Recovery of soybean oligosaccharide using activated charcoal .....	244
Section 5. Separation of soybean-curd whey protein and functional property.....	245
1. Measurement of physicochemical characteristics of freeze-dried soybean-curd whey protein.....	245
2. Measurement of functional properties of freeze-dried soybean-curd whey protein.....	246
3. Structural characteristics of freeze-dried soybean-curd whey .....	248
4. Nutritional analysis of freeze-dried soybean-curd whey .....	251
Section 6. Isolation of functional peptide from concentrated with UF/NF and analysis of functionality .....	253
1. Purification and recovery of low molecular peptides using UF/NF .....	253
2. Purification and isolation of peptide .....	253
3. Analysis of ACE inhibitory activity.....	254
4. Purification and isolation of glycoprotein using thin layer chromatography.....	255
5. Measurement of molecular weight using MALDI.....	256

Chapter 3. Results and discussion .....	257
Section 1. Characteristics of soybean-curd whey and soybean soaking water.....	257
1. Survey of soybean-curd whey factory.....	257
2. Analysis of contamination.....	258
3. Proximate analysis.....	262
4. Quantitative analysis of functional components1.....	264
Section 2. Recovery and concentration of functional components .....	268
1. Recovery and concentration of functional components using UF .....	268
Section 3. Recovery of functional components and soybean oligosaccharides using UF/NF .....	275
1. Soybean oligosaccharides.....	275
2. Ionic calcium .....	276
3. Proximate components of concentrate using UF/NF .....	277
4. Physicochemical characteristics of concentrate using UF/NF .....	280
5. Effects of concentrated soybean oligosaccharides on <i>bifidobacterium</i> activity.....	286
6. Characteristics of purified soybean oligosaccharides .....	287

Section 4. Recovery of soybean oligosaccharide using other methods	
.....	289
1. Concentration and recovery of functional components.....	289
Section 5. Separation of soybean-curd whey protein and functional	
characteristics.....	292
1. Physicochemical characteristics of concentrated soybean-curd	
whey protein.....	292
2. Functional properties of concentrated soybean-curd whey protein	
.....	295
3. Structural characteristics of concentrated soybean-curd	
whey protein .....	298
4. Nutritional characteristics of concentrated soybean-curd whey	
protein.....	302
Section 6. Separation of functional peptide from soybean-curd whey	
concentrated by UF/NF and functional properties.....	303
1. Recovery and concentration of low molecular weight peptide	
by UF/NF.....	303
2. Fractionation of ACE inhibitory peptide and characterization	
.....	305
3. Separation and purification of glycoprotein by Thin layer	
chromatography.....	307

4. Molecular weight analysis by MALDI mass spectrometry	308
.....	
Chapter 4. References.....	310

Part 5. Preparation and utilization of calcium  
components from eggshell

Chapter 1. Introduction.....	315
Chapter 2. Methods.....	317
section 1. Separation and utilization of natural inorganic compounds from eggshell.....	317
1. Materials and reagent.....	317
2. Removal of egg membrane.....	317
3. Ash of eggshell.....	317
4. Measurement of Hausner ratio.....	318
5. Measurement of concentration of calcium ion.....	318
6. Preparation of organic acid eggshell calcium .....	319
7. Agglomeration process of organic acid eggshell calcium .....	319
8. Measurement of WSI and WAI.....	319
Section 2. Physical properties of various food fortified with organic acid eggshell calcium.....	319



1. Fresh noodle.....	319
2. Preparation of loaf bread.....	323
3. Preparation of Ramen.....	325
4. Soy milk.....	326
Section 3. Organic acid eggshell calcium plant design.....	327
1. Process flow-diagram.....	327
2. Fixed capital cost estimation.....	327
3. Estimate of production cost.....	328
Chapter 3. Results.....	329
Section 1. Separation & utilization of natural inorganic compounds from eggshell.....	218
1. Removal of egg membrane.....	329
2. Decoloration of eggshell.....	330
3. Degree of ionization of eggshell calcium powder.....	333
4. WSI and WAI of organic acid eggshell calcium powder.....	336
5. SEM of organic acid eggshell calcium powder.....	337
Section 2. Physical properties of food fortified with organic acid calcium.....	337
1. Fresh noodle.....	337
2. Loaf bread.....	346
3. Ramen.....	349
4. Soy milk.....	353

Section 3. Plant design for organic acid eggshell calcium .....	356
1. Process flow diagram of producing organic eggshell calcium .....	356
2. Analysis of fixed capital cost estimation.....	360
3. Estimation of production cost for production of organic acid eggshell calcium .....	360
Chapter 4. References.....	363

Part 6. Development of processing method for biodegradable film from potato starch

Chapter 1. Introduction.....	365
Section 1. Background of study.....	365
1. Studies for biodegradable plastics.....	368
2. Objectives.....	368
Chapter 2. Materials and methods.....	368
Section 1. Preparation and characterization of modified starches .....	368
1. Preparation of potato starch.....	368
2. Preparation of modified starches.....	368

3. Degree of substitution in starch.....	369
4. Physicochemical properties of starch.....	370
Section 2. Preparation of evaluation of modified starch-polyethylene films.....	372
1. Preparation of starch-polyethylene films.....	372
2. Color of starch-polyethylene films.....	372
3. Mechanical properties of starch-polyethylene films.....	373
Section 3. Thermal and Biodegradability of starch-polyethylene films.....	376
1. Thermal degradability.....	376
2. Biodegradability.....	377
Chapter 3. Results and Discussion.....	379
Section 1. Physicochemical properties of starch.....	379
1. Degree of substitution in starch.....	379
2. Physicochemical properties of starch.....	381
Section 2. Characterization of starch-polyethylene films.....	393
1. Color of starch-polyethylene films.....	393
2. Mechanical properties of starch-polyethylene films.....	394
Section 3. Thermal and biodegradability of starch-polyethylene films.....	398

1. Thermal degradability of starch-polyethylene films.....	398
2. Biodegradability of starch-polyethylene films.....	408
Section 4. Economic evaluation.....	417
Chapter 4. References.....	418

1 ( ) ( )

1	.....	56
2	.....	60
1	.....	60
1.	.....	60
2.	.....	60
3.	.....	64
2	.....	65
1.	.....	65
2.	.....	66
3.	<b>Oleoresin</b> .....	68
4.	.....	70
5.	.....	71
3	.....	73
1	.....	73
1.	.....	73
2.	- .....	81

3. Pilot	.....	97
2	.....	103
1.	.....	103
2.	.....	118
3.    oleoresin	.....	122
4. oleoresin	.....	126
5. Pilot	.....	131
4	.....	137

2 ( )

( )

1	.....	141
2	.....	143
1	.....	143
2	.....	143
1.	.....	143
2. 가	.....	144

3.	.....	145
4.	가	..... 145
5.	.....	147
6.	.....	149
3	.....	151
1	.....	151
1.	.....	151
2.	.....	153
3.	Methanol	..... 155
4.	Ethanol	..... 158
5.	pH	..... 160
6.	Citric acid 가	..... 162
7.	NaCl 가	..... 163
2	가	..... 167
3	.....	168
1.	.....	168
2.	.....	171
4	.....	176
1.	가(POV)	..... 176
2.	thiobarbituric acid value(TBA)	..... 178

5	가	... 179
4	.....	182
3	( ) ( )	
1	.....	186
2	.....	190
1	.....	190
1.	.....	190
2.	.....	191
3.	.....	192
2	.....	193
3	.....	195
1	.....	195
1.	.....	195
2. 가	.....	198
2	.....	201
1.	.....	201
3	.....	206



1. 가	.....	206
2. 가	.....	212
4	.....	224
1.	.....	224
2.	.....	226
4	.....	228
4	( ) ( )	
1	.....	229
2	.....	233
1	.....	233
1.	.....	233
2.	.....	234
3.	.....	235
4.	.....	236
2	, .....	240
1.	.....	240
2.	.....	240

3. Nanofiltration	.....	240
3    UF/NF	,	
.....		241
1.        .....		241
2.        .....		241
3.        Bifidobacterium	.....	241
4.        .....		243
4        가	.....	243
1.        .....		243
2. Steffen	.....	244
3.        .....		244
5        .....		245
1.        .....		245
2.        .....		246
3.        .....		248
4.        .....		251
6    UF/NF		
.....		253
1.        .....		253
2.        .....		253

3. ACE	.....	254
4. Thin layer chromatography	.....	255
5. MALDI	.....	256
3	.....	257
1	.....	257
1.	.....	257
2.	.....	258
3.	.....	262
4.	.....	264
2	, .....	268
1.	.....	268
3	UF/NF .....	275
1.	.....	275
2.	.....	276
3.	UF/NF .....	277
4.	UF/NF .....	280
5.	UF/NF <i>bi fi do bacteri um</i>	
.....	.....	286
6.	.....	287
4	가 .....	289

1.	.....	289
5	.....	292
1.	.....	292
2.	.....	295
3.	.....	298
4.	.....	302
6	UF/NF	
	.....	303
1.	UF/NF	303
2.	ACE	305
3.	Thin layer chromatography	307
4.	MALDI mass spectrometry	308
4	.....	310
	5 ( ) ( )	
1	.....	315
2	.....	317
1	.....	317
1.	.....	317

2.	.....	317
3.	.....	317
4. Hausner ratio	.....	318
5.	.....	318
6.	.....	319
7.	.....	319
8.	(WSI) (WAI) .....	319
2	.....	319
1.	.....	319
2.	.....	323
3.	.....	325
4.	.....	326
3	.....	327
1.	.....	327
2.	.....	327
3.	가 .....	328
3	.....	329
1	.....	329
1.	.....	329

2.	.....	330
3.	.....	333
4.	WSI WAI.....	336
5.	(SEM).....	337
2	.....	337
1.	.....	337
2.	.....	346
3.	.....	349
4.	.....	353
3	.....	356
1.	.....	356
2.	.....	360
3.	가 .....	360
4	.....	363

6 ( ) ( )

1	.....	365
1	.....	365
1.	.....	365
2.	.....	367
2	.....	368
1	.....	368
1.	.....	368
2.	.....	368
3.	.....	369
4.	.....	370
2	.....	372
	<b>Starch- pol yethyl ene</b>	
1. Starch- pol yethyl ene	.....	372
2. Starch- pol yethyl ene	.....	373
3. Starch- pol yethyl ene	.....	373
3	.....	376
1.	.....	376
2.	.....	377
3	.....	379
1	.....	379

1.	.....	379
2.	.....	381
2	Starch-polyethylene .....	393
1.	Starch-polyethylene .....	393
2.	Starch-polyethylene .....	394
3	Starch-polyethylene .....	398
1.	Starch-polyethylene .....	398
2.	Starch-polyethylene .....	408
4	.....	417
4	.....	418

**1.**

**1**



가 .

가 가 (1989 7 ha) (1)

1 1 8 9g( )

가 20 . 60% 가

가 40% 가 (2).

Thresh(3)가 capsai ci n

capsai noi d

(47) capsai ci n(8-methyl - N-vani llyl - 6-nonenami de) di hydro-

capsai ci n . capsai ci n 가

100 di hydrocapsai ci n 63, nordi hydrocapsai ci n

11, homcapsai ci n 5, homodi hydrocapsai ci n 3

(8-11).

, , Vit C (12) ,

, (13) 7

, (14) GC

.

, ,

, ,

가 . 가

, 15%

가 가 (15) 가

가  
가  
가

가 (16),

가 (17-19)

(20) polyphenol oxi dase가 Singh (21)

6%

(22) 65 0.1%, 75 0.5%, 85

1.0%

Lukes(23)가

S- (1-propenyl) cysteine sul foxi de (24)

가 soybean oil

가  
(25)

14% 가 가

oleoresin

(26)

methyl alcohol

oleoresin 가 가

(27)

ethanol

extract

가

, , anti forming agent 가

oleoresin

, paste,

가

drink

가

가

가

가

가

가

가

10 15%가

가

가 ,

.

, pilot

plant

. pilot plant

.

2

1

1.

( ) 가  
 , , , , ,  
 , ,  
 . 가 NaCl,  
 sucrose, citric acid, ethanol, (NaH<sub>2</sub>PO<sub>4</sub>, Na<sub>2</sub>HPO<sub>4</sub>, Na<sub>3</sub>PO<sub>4</sub>, sodium  
 citrate Na<sub>4</sub>P<sub>2</sub>O<sub>7</sub>, Na<sub>5</sub>P<sub>3</sub>O<sub>10</sub>, (NaPO<sub>3</sub>)<sub>n</sub>) .  
 Novozyme 89, Alcalase, Protamex 3 ( ) Novo Nordisk  
 ( ) Acid Protease A, Acid Protease , Protease  
 M ( ) Amano Enzyme ( U. S. A ) .

2.

가

1)

stirrer/hot plate(PC-320) Fig. 1 .  
 hot plate  
 5cm

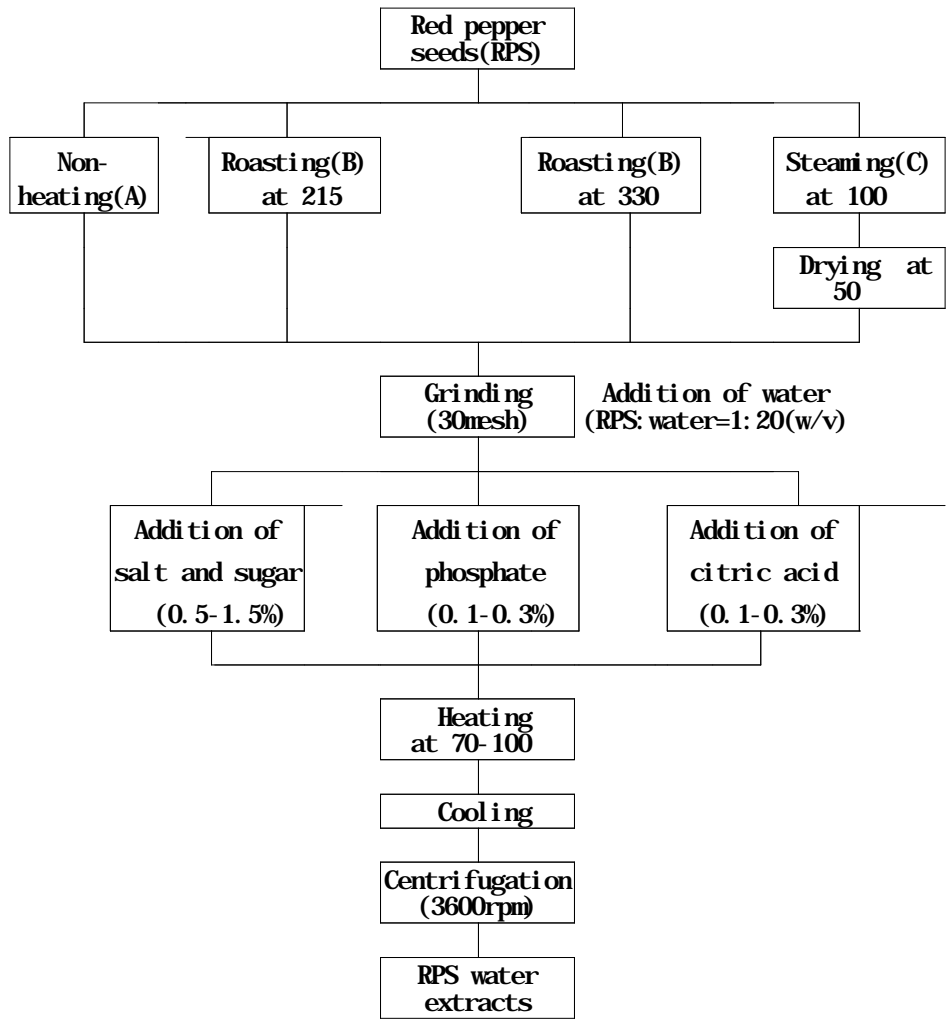


Fig. 1. Schematic diagram for preparation of several aqueous extraction of red pepper seeds.

215 330 10, 20, 30 .  
 100 10, 20, 30 50 4  
 . 가 가 30mesh

-20

1: 20(w/v) , 0.5 0.3%  
 (70-100 ) (10-60 ) 3600rpm  
 20 (HMR-220IV, ) .

2)

ethanol (2) 0, 10, 20%  
 ethanol 1: 20(w/v) 60 1  
 3600rpm 20  
 3 10  
 2

waring blander 1 .

Fig. 2 2 가

80 100 20 120 , 0.3%, 0.6%,  
 0.9%, (NaH<sub>2</sub>PO<sub>4</sub>, NaHPO<sub>4</sub>, Na<sub>3</sub>PO<sub>4</sub>, Na<sub>4</sub>P<sub>2</sub>O<sub>7</sub>, Na<sub>5</sub>P<sub>3</sub>O<sub>10</sub>, (NaPO<sub>3</sub>)<sub>n</sub>)  
 sodium citrate 0. 2.0% 가 .  
 pH 6N NaOH HCl 가 2 10 .  
 0.01 1.0% 가  
 가 pH  
 30 120 . 90 40  
 shaking 30 120 20 3600rpm  
 15 (Whatman No. 41) .

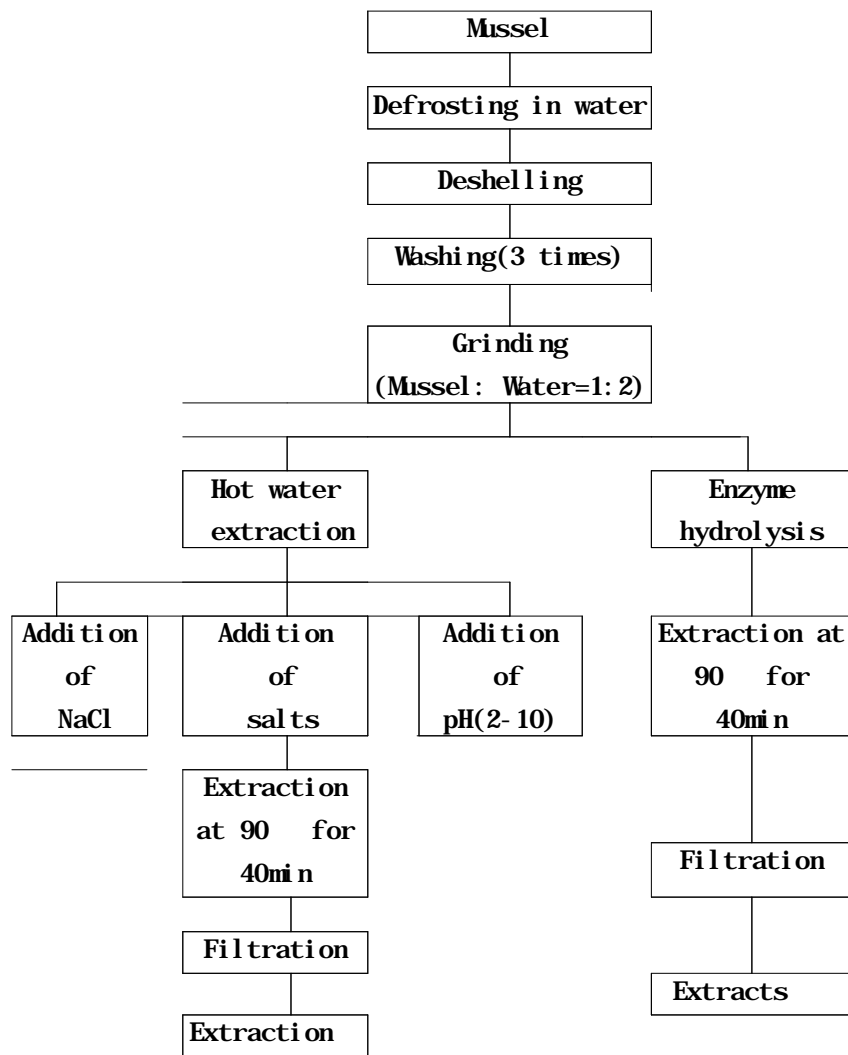


Fig 2. Schematic diagram for preparation of mussel extracts



40 가 (5, 10, 15, 20%)  
80 30 3 cheese clothes  
3600rpm 15  
(Whatman No. 41)

3.

가.

105 (29)

가 가

Brookfield viscometer(Model DV  
Brookfield Engineering, U. S. A) 8ml 2.0cm  
6.5cm 30 spindle No. 21  
100rpm

(Chroma meter CR-200, Minolta) Hunter  
L, a, b

Wätman filter paper(#4)  
spectrophotometer(Beckman. DU. 650 made in USA) 600nm  
3

가

10 (hot oder), (roasted oder),  
 (hot taste), (roasted taste) 9 가

80 30 . 가

45-50 20ml .

가 가 (savory odor), (fish odor),  
 (umami taste), (fish taste)

9 . 가 45

10ml .

가 (savory odor), (fish odor),  
 (hot odor) (off odor) (umami taste), (fish taste),  
 (hot taste), (off taste) .  
 Duncan .

## 2

1.

(Allium sativum L.) 1995  
 가 . -20

15%

Cell

uclast 1.5L, Extrazym, Viscozym L 3 ( ) Novo Nordisk( )

7 (methanol, ethanol, acetone, isopropyl alcohol, n-hexane, methylene chloride, ethylene chloride)

2.

가.

(

) 가 20mesh

가 .

. 가 가 pH

Table 1

: 1:1(w/w)

25 24

. pH HCl pH 4.0, 4.5, 5.5

**Table 1. Chemicals and their concentrations used for the study of effects on color changes of gound garlic.**

Additives	Concentration	Additives	Concentration
Salts		Antioxidants	
NaCl	0.5 2%	BHA	0.02%
NaH <sub>2</sub> PO <sub>4</sub>	0.1%	BHT	0.02%
Na <sub>2</sub> HPO <sub>4</sub>	0.1%	PG3)	0.02%
Na <sub>3</sub> PO <sub>4</sub>	0.1%	Glutathione	0.1%
SPP1)	0.1%	Others	
SMP2)	0.1%	Cysteine	1mM
Na <sub>2</sub> S <sub>2</sub> O <sub>5</sub>	0.5%	L - a s c o r b i c acid	10mM
Na <sub>2</sub> SO <sub>4</sub>	0.3%		
Organic acids			
Acetic acid	0.5 2%		
Citric acid	0.5 2%		
Lactic acid	0.5 2%		

1) Sodium polyphosphate, 2) Sodium metaphosphate, 3) Propyl gallate

. 가

2

50 100 60

가

25 24

,

. 가

pH

0.05M citrate buffer

pH 3.0 5.0

80

0 10

가

.

.

가

ascorbic acid(A), cysteine(C), sodium metabisulfite(Na<sub>2</sub>S<sub>2</sub>O<sub>5</sub>) (S),

sodium triphosphate( $\text{Na}_3\text{P}_04$ ) (P) ,

25 7

3. oleoresin

가.

1)

가 가  
 가 waring blendor 가 20mesh  
 가 3 30 100rpm  
 10 .  
 7 2 가  
 100rpm 30 50 , 1 4 .  
 (HMR-220IV, )  
 15000rpm 15 (Whatman No. 4)

2) pH 가

pH 1N HCl 가 pH 2.5  
 5.5 , 가 0.1M (NaCl, KCl,  $\text{CaCl}_2$   
 $\text{NaH}_2\text{P}_04$ ,  $\text{Na}_2\text{H}_2\text{P}_04$ ,  $\text{Na}_3\text{P}_04$ ,  $\text{Na}_4\text{P}_207$ ,  $\text{KH}_2\text{P}_04$ ,  $\text{K}_2\text{H}_2\text{P}_04$ ,  $\text{K}_3\text{P}_04$ ,  $\text{K}_4\text{P}_207$ ) 0.1M  
 (0.1M acetic acid, citric acid, lactic acid, tartaric  
 acid, phosphoric acid) 가

30 10 .  
 가 (3) K<sub>2</sub>P<sub>2</sub>O<sub>7</sub> phosphoric acid,  
 tartaric acid, sodium oxalate, CaCl<sub>2</sub> 가 30  
 10 .  
 3)  
 2 3mm : =1:5(w/w)  
 가 waring blendor 20mesh 가 3 1N  
 HCl 가 pH 5.0 100 .  
 0.04 0.12%가 가 50 30  
 60 . büchner funnel aspirator  
 Celluclast,  
 Extrazym, Viscozym Table 2 .

Table 2. Properties of commercial enzymes used for extraction of garlic juice

Commercial name	Optimum conditions	
	pH	Temp( )
Celluclast	4.5 6.0	50 60
Extrazym	4.5	50
Viscozym	3.5 5.5	40 50

. Oleoresin

oleoresin 8가  
 ethanol , 30 2 3600rpm 15

(Whatman No. 4)

Rotary vacuum evaporator (Eyela N-N Series, Japan) 30 ° Brix

oleoresin . oleoresin

0.1% Na<sub>2</sub>S<sub>2</sub>O<sub>5</sub> Na<sub>3</sub>PO<sub>4</sub> 0.1% ascorbic acid,

cysteine 1% 가 .

4.

가.

3000rpm 15

(Whatman No. 4) 105 가

가

가

. pH, ,

pH pH meter(DP-215M, Dongwoo Co.) , AOAC

10ml 0.1N NaOH

(%) .

(Chroma meter CR-200, Minolta Co., Japan)

Hunter L, a, b , 3

spectrophotometer(DU 650, Beckman, USA)

600nm

thiosulfinate

oleoresin 0.45µm membrane filter

Sep-pak C18

HPLC

HPLC(Waters Associate, USA)

RI (Waters 486)

UV detector(Waters 410)

column Aminex HPX-87A stainless

column(7.8 × 300mm, Bio Rad)

0.005M H2SO4

1.5ml/min

20µl

thiosulfinate

Freeman

oleoresin 1ml 0

10ml 가

5ml 10ml

hexane

가

5

Thermolyne(37600

Mixer)

hexane

262nm

thiosulfinate

5.

oleoresin

( , , ) (

, )

가 3 가

1 , 5 ,

9

9



9 가 .

SAS

(ANOVA) Duncan

### 3

#### 1

1.

가. 가

1)

, 가 가 ,

.

(100 ) (215 , 330 ) 80

30 , ,

Table 3 .

가 가

(27.03%) 100 10, 30 33.20, 36.12%

215 30

37.37%, 330 20 40.10% 가

가 , .

가 600nm

10

가 가 . L b

가 L b 가

. a 가 .

2)

100

80

30

Table 4

가

215

330

가

Table 3. Effects of roasting or steaming on characteristics of aqueous extracts of red pepper seeds extracted at 80

Temperature ( )	Time (min)	Solid yield (%)	Viscosity (cps)	Absorbance (600nm)	Hunter		
					L	a	b
100)	0	27.03	1.50	1.01	38.27	0.32	14.57
	10	33.20	1.50	0.64	44.74	-0.12	19.50
	20	28.72	1.50	0.77	47.55	0.03	21.86
	30	36.12	1.50	0.64	47.73	0.13	23.44
215)	0	27.03	1.50	1.01	38.27	0.32	14.57
	10	31.23	1.75	1.92	27.66	0.27	14.38
	20	27.40	1.50	1.36	30.78	0.14	14.74
	30	37.37	2.00	1.11	31.26	0.03	15.75
330)	0	27.03	1.50	1.01	38.27	0.32	14.57
	10	33.21	2.00	1.51	31.34	-0.22	14.60
	20	40.10	2.25	1.01	32.35	-0.06	18.89
	30	29.82	1.75	0.54	42.96	3.14	35.56

1) : Steamed red pepper seeds at 100

2) : Roasted red pepper seeds at 215 or 330

10 , 215

30

330

20

가

(Table 5)

가

(A)가 가

215 30 , 330 20 , 10 .  
 330 가 가 .

**Table 4. Comparison of odor and taste of aqueous extracts of red pepper seeds extracted at 80 for 30min after heating treatments**

Heat treatment	Description	Heating time(min)			F- value
		10	20	30	
steaming (100 )	hot odor	4.80	4.20	3.80	0.40
	roasted odor	4.60	3.80	4.80	0.32
	hot taste	6.40a	5.20a	3.20b	6.32**
	roasted taste	3.00	3.40	4.20	0.41
Roasting (215 )	hot odor	4.40	6.00	6.60	2.13
	roasted odor	3.00	3.40	3.80	0.14
	hot taste	4.00b	5.20ab	6.60a	3.01*
	roasted taste	2.20	2.60	3.80	1.07
Roasting (330 )	hot odor	6.60	5.60	5.00	1.21
	roasted odor	2.60b	4.60a	6.20a	10.17**
	hot taste	5.20	6.20	5.00	0.54
	roasted taste	3.00b	4.60b	6.60a	10.41**

\*\* significant p<0.01

a b Mean scores within row by the same letter are not significantly different at the 5% level

**Table 5. Comparison of odor and taste of aqueous extracts of red pepper seeds extracted at 80 for 30min after roasting and steaming.**

Description	A	B	C		F- value
			215 /30min	330 /20min	
hot odor	7.33a	4.67b	6.83a	5.50ab	3.38*
roasted odor	4.33	4.33	5.17	6.17	0.64
hot taste	7.50a	4.50b	7.17a	4.67b	8.21***
roasted taste	3.67b	4.67ab	5.17ab	6.67a	3.19*

\* significant p<0.05

\*\*\* significant p<0.001

a b Mean scores within row by the same letter are not significantly different at the 5% level

A: Red pepper seeds without heat treatment

B: Steamed red pepper seeds at 100 for 10min

C: Roasted red pepper seeds

1)

가

(

)

가 가

, 가

Table 6, 7

(Table 6)

가

70 80

30

27-29%

가

70

80

1.50cps

가

90

100

1.50cps

가

가 1.25cps

70-90



2)

Table 7 70- 100

가 . 5% 80 가

Table 8 80

10 30 가 10 50 80 30

Table 7. Effects of temperature on odor and taste of aqueous extracts of red pepper seeds after extraction for 30 min.

Description	Heating temperature( )				F- value
	70	80	90	100	
hot odor	5.50	7.25	7.00	4.75	3.07
roasted odor	4.75	4.75	5.75	6.75	2.59
hot taste	6.25ab	7.75a	7.25ab	5.00b	4.35*
roasted taste	4.75	4.75	5.25	5.50	0.35

\* significant p<0.05

a b Mean scores within row by the same letter are not significantly different at the 5% level

**Table 8. Effects of extraction time at 80 on odor and taste of aqueous extracts of red pepper seeds.**

Description	extraction time(min)						F- value
	10	20	30	40	50	60	
hot odor	6.25a	5.75ab	5.50ab	4.75ab	3.50ab	2.75b	3.68*
roasted odor	3.50	4.50	5.25	5.25	5.00	5.25	0.72
hot taste	6.00a	6.25a	6.50a	5.75ab	4.25ab	3.00b	4.08*
roasted taste	2.75	3.50	4.25	5.00	4.50	5.50	1.87

\* significant p<0.05

ac Mean scores within row by the same letter are not significantly different at the 5% level

. , , 가

1)

, , , 가가

80 , 30

가 Table 9 .

80 30 (27.03%)

citric acid 가 가

18-22% 0.2% 가 29.21%

.

가 가가 가 1.5% 가

3.50cps citric acid (Na<sub>2</sub>HPO<sub>4</sub>) 가 0.2

0.3% , 0.5%

가 가 . 가 가



citric acid 가 가  
citric acid 가 L 가  
(Na<sub>2</sub>HPO<sub>4</sub>) L citric acid a  
-0.25, -0.20  
(Na<sub>2</sub>PO<sub>4</sub>) a 가

**Table 9.** Effects of concentration of NaCl, sucrose, Na<sub>2</sub>HPO<sub>4</sub> and citric acid on characteristics of aqueous extracts of red pepper seeds extracted at 80 °C for 30min.

Concentration (%)	Solid yield (%)	Viscosity (cps)	Absorbance (600nm)	Hunter			
				L	a	b	
	0	27.03	1.50	1.01	38.27	0.32	14.57
NaCl	0.5	27.21	2.00	1.44	44.59	-0.35	13.11
	1.0	23.42	2.00	0.47	41.06	-0.33	12.85
	1.5	24.61	2.50	0.40	42.58	-0.25	12.93
	0.5	17.59	2.00	0.55	39.64	0.36	13.70
Sucrose	1.0	18.42	2.50	2.87	37.15	0.42	14.48
	1.5	22.23	3.00	4.31	30.86	1.03	15.95
	0.1	24.38	1.50	0.50	26.59	0.52	15.45
Na <sub>2</sub> HPO <sub>4</sub>	0.2	29.21	1.50	1.60	19.84	0.87	16.64
	0.3	28.22	2.00	0.75	21.72	0.98	16.53
	0.1	26.00	1.75	0.68	35.44	-0.32	12.96
Citric acid	0.2	27.21	2.00	0.66	40.70	-0.80	12.81
	0.3	25.60	2.00	0.56	48.55	-1.20	12.36

2)

Table 10 11 , , , citric acid

가 80 30

(NaCl) 0.5 1.5% 가 (Table 10)

0.5% 가

0.5% 가

가 masking

(Na<sub>2</sub>HPO<sub>4</sub>) 가

citric acid 가

가 가가

masking

2. -

가.

가

가 , . 가

5% 가 10 가 1

Table 12, 13 .

**Table 10. Comparison of odor and taste of aqueous extracts of red pepper seeds extracted at 80 °C for 30min after addition of various concentration of NaCl.**

Additives	Description	concentration(%)				F- value
		0	0.5	1.0	1.5	
NaCl	hot odor	5.75ab	6.75a	6.00ab	5.00b	4.55*
	roasted odor	5.00	5.50	4.50	4.50	1.57
	hot taste	6.00ab	7.00a	7.00a	5.00b	3.67*
	roasted taste	5.25	5.75	6.00	3.75	2.19
Sucrose	hot odor	6.75ab	4.25b	5.25ab	4.25b	5.15*
	roasted odor	5.75a	4.50ab	4.75ab	3.75b	2.15*
	hot taste	6.00ab	7.00a	7.00a	5.00b	5.43**
	roasted taste	5.75	4.00	5.25	3.25	1.95

\* significant p<0.05

\*\* significant p<0.01

a b Mean scores within row by the same letter are not significantly different at the 5% level

**Table 11. Comparison of odor and taste of aqueous extracts of red pepper seeds extracted at 80 °C for 30min after addition of various concentration of Na<sub>2</sub>HPO<sub>4</sub> and citric acid.**

Additives	Description	Na <sub>2</sub> HPO <sub>4</sub> concentration(%)				F- value
		0	0.1	0.2	0.3	
Na <sub>2</sub> HPO <sub>4</sub>	hot odor	7.00	6.75	6.25	5.25	1.77
	roasted odor	5.75a	5.25ab	5.00ab	4.50b	2.89*
	hot taste	7.00a	6.75ab	6.75ab	5.00b	3.04*
	roasted taste	5.75a	4.75b	3.75c	3.25d	11.80***
Citric acid	hot odor	7.00	6.75	6.00	5.50	1.94
	roasted odor	5.75a	4.50b	3.50c	2.50d	24.73***
	hot taste	7.00	7.00	7.00	6.50	0.20
	roasted taste	5.75a	4.75b	3.50c	2.50d	27.71***

\*\*\* significant p<0.001  
 ad Mean scores within row by the same letter are not significantly different at the 5% level

**(Table12)**

26.81% 가 가 (24.11%), (23.43%) 가  
 , , , .  
 가 가 , ,  
 .  
 L , , a  
 (-) , 1.22  
 .  
 가 가

**Table 13**

가 , - 가  
 가 가  
 . 가 가  
 . 가  
 - - - . - -  
 . 가 가  
 .

**Table 12. Characteristics of aqueous extracts of seasoning materials with addition of 5% red pepper seed**

Natural seasoning materials	Solid yield(%)	Absorbance (600nm)	Hunter		
			L	a	b
Beef	23.02	1.64	41.03	1.22	13.51
Anchovy	23.43	1.70	44.38	0.05	15.65
Pollack	26.81	2.09	33.16	0.47	12.98
Mussel	22.59	1.38	55.20	- 0.76	13.83
Shrimps	24.11	1.73	38.40	0.08	11.58
Sea mustard	21.65	0.96	50.28	- 0.36	14.14
Sea tangle	22.47	1.21	52.75	0.39	16.87

**Table 13. Sensory characteristics of natural seasoning extracts with addition of 5% red pepper seed**

Description	Natural seasoning materials						
	beef	anchovy	pollack	mussel	shrimps	sea mustard	sea tangle
hot odor	5.33	5.00	5.00	5.67	5.00	4.67	4.33
savory odor	4.00	3.67	5.00	6.00	4.67	3.33	4.33
fish odor	1.67	3.00b	3.67ab	2.88b	4.67ab	4.33ab	6.53a
off odor	1.33	4.33	1.67	2.13	1.00	1.00	1.00
hot taste	4.33	4.33	4.67	4.25	5.67	4.33	4.00
savory taste	4.33abc	4.67abc	5.00abc	5.88a	5.01a	3.33bc	5.31a
fish taste	3.67b	3.00b	3.33b	2.63b	4.33ab	3.33ab	6.33a
off taste	1.33	2.67	1.33	1.88	1.00	1.33	1.33
total acceptability	68ab	79bc	86cd	51a	64ab	97d	74bc

Total acceptability was tested ranking test.

Mean scores within row followed by the same letter are not significantly different at 5% level.

\* P < 0.05

1)

**Table 14**

80-100

80



pH

가 가      Na<sub>3</sub>P<sub>04</sub>

L a      a      L

a b      .      1.0% 가

가      가      Na<sub>3</sub>P<sub>04</sub> 1.0% 가

sodi um

phosphate tribasic, sodi um polyphosphate, sodi um pyrophosphate가

(34) ,      (35)

sodi um citrate 가

**Table 14. Changes in the characteristics of mussel extracts during aqueous extraction at 80 100**

Temp ( )	Time (min)	Solid yield (%)	Absorbance (600nm)	Color			Sensory properties	
				L	a	b	umami taste	fish taste
80	20	15.71	0.19	72.21	-0.97	+18.37	+	++++
	40	18.17	0.27	75.81	-0.75	+20.83	+	++++
	80	17.02	0.29	79.91	-1.53	+21.08	+	+++
90	20	17.41	0.23	79.86	-1.65	+19.22	+	++++
	40	18.97	0.37	83.96	-1.04	+21.52	+++	++
	80	16.45	0.18	79.11	-1.48	+22.51	++	+++
100	20	17.90	0.14	83.06	-1.55	+22.18	+	+++
	40	18.48	0.12	83.65	-1.47	+22.55	++	+++
	80	16.65	0.17	85.83	-2.08	+25.33	++	+++



3) pH

pH 6.25 , NaOH HCl 가 pH 2-10

Table 16

pH 18.97% pH 2  
 28.04% pH가 가  
 pH 8 가 pH 10 27.07% .  
 pH 10 가  
 L (79.9)  
 pH 2 pH 10 L a b  
 pH 7 가

**Table 15. Effect of phosphates addition on the characteristics of aqueous extracts of mussel extracted at 90 °C for 40 min**

Phosphates	Concentration (%)	pH	Solid yield (%)	Absorbance (600nm)	Color			Sensory properties	
					L	a	b	umami taste	fish taste
Control	0	6.25	18.97	0.37	83.96	-1.04	+21.52	+++	++
NaH <sub>2</sub> PO <sub>4</sub>	0.5	6.05	17.51	0.42	73.17	-1.00	+20.19	++	+++
	1.0	5.90	16.80	0.52	68.26	-0.84	+17.83	+++	++
	1.5	5.80	16.05	0.59	69.53	-1.29	+17.36	+	++
Na <sub>2</sub> HPO <sub>4</sub>	0.5	7.66	20.67	1.33	13.52	+1.88	+ 8.04	++	+++
	1.0	7.96	20.99	2.29	16.36	+2.20	+ 9.28	+++	+
	1.5	8.11	22.54	3.68	25.30	+4.15	+13.48	+++	+
Na <sub>3</sub> PO <sub>4</sub>	0.5	8.44	22.18	0.60	14.61	+2.51	+8.74	++	++
	1.0	9.53	24.53	1.64	22.11	+2.42	+11.74	++++	+
	1.5	10.23	25.52	4.49	29.37	+3.42	+15.39	+++	+
Na <sub>4</sub> P <sub>2</sub> O <sub>7</sub>	0.5	7.50	19.85	0.76	15.45	+2.04	+8.95	+++	++
	1.0	8.32	22.32	2.04	17.20	+2.21	+ 9.85	++++	+
	1.5	8.64	23.45	3.61	25.34	+3.10	+13.20	+++	+
sodium hexameta phosphate	0.5	6.35	19.00	0.78	57.68	+2.14	+22.72	++	+++
	1.0	6.28	18.57	0.82	51.41	+3.24	+21.80	++	++
	1.5	6.24	17.81	0.94	51.91	+3.05	+21.53	++	++++
sodium citrate	0.5	6.96	19.37	0.84	24.33	+3.44	+12.32	++	+++
	1.0	7.14	21.65	0.98	31.41	+4.46	+15.76	+	++
	1.5	7.25	24.00	1.41	41.89	+4.50	+19.45	+	++

pH 2 10 가  
 pH 10 가 .  
 pH 12 가  
 pH 12 가  
 (34)가  
 (35) .

**Table 16. Effect of pH on the characteristics of aqueous extracts of mussel extracted at 90 for 40min**

pH	Solid yield (%)	Absorbance (600nm)	Color			Sensory properties	
			L	a	b	umami taste	fish taste
2.00	25.07	5.81	10.57	+1.91	+6.19	++++	++
4.00	18.53	0.40	80.52	- 1.13	+21.50	+++	+
6.00	17.26	1.77	27.51	+4.02	+12.84	++	+++
8.00	23.68	5.99	11.93	+2.41	+7.07	++	+
10.00	28.04	9.06	8.63	+3.07	+5.74	++++	+

4)

56%

가

Novowyme 89, Alcalase, Protamex, Papain

Acid Protease A, Acid Protease , Protease M

0.01- 1% 가 (Table 17) 가

. 가 가 Protease A, Protease , Papain

가 0.05% 가 . 가

Protamex 0.05% Acid Protease 0.1% .

10 L 40-90

pH . Protamex a 0.26

Protease A -2.8 b Protease A가 23

Protease A

0.1% 가

가

Protease A

Acid Protease

A 0.01%

Protamex 0.05%

Table 18 2가

Protamex 0.05% 90 , Protease 0.01% 90 가

5)

, , pH

Table 19 .

Protamex(0.05%, 60 )가,

Protease (0.01%, 90 )가 가 Na<sub>3</sub>P<sub>04</sub> 1.0%,

Na<sub>4</sub>P<sub>2</sub>O<sub>7</sub> 1.0%, 90 40 , pH 2 10 .

가 pH 2

10 90 40

100 30 (18.97%)

Protamex Na<sub>3</sub>P<sub>04</sub> 가

pH 10 30.53% 60%

가 (Table 19) Na<sub>3</sub>P<sub>04</sub>

가 Na<sub>4</sub>P<sub>2</sub>O<sub>7</sub> 가 pH 10

가 pH

. pH 2 pH 10

. a pH 10

. pH 10 a maillard

**Table 17. Effect of enzyme addition on characteristics of aqueous extract of mussel extracted at 90 for 40min after enzymatic hydrolysis for 30 min**

Enzyme	Enzyme conc(%)	Solid yield (%)	Absorbance (600nm)	Color			Sensory properties	
				L	a	b	umami taste	fish taste
Novozyme 89	0.01	20.52	0.11	57.41	+0.80	+21.00	+++	++
	0.05	21.03	0.18	68.00	+0.60	+21.19	+++	+
	0.1	20.68	0.22	73.72	+0.06	+21.00	+++	+
Protamex	0.01	23.73	0.45	47.17	+0.26	+14.84	++	+
	0.05	24.06	0.53	54.08	+0.46	+17.91	+++	+
	0.1	25.58	0.61	51.17	+0.65	+17.25	+	+
Alcalase	0.01	22.34	0.42	50.82	+0.27	+14.91	++	++
	0.05	23.42	0.47	37.65	+1.14	+12.57	++	+
	0.1	24.04	0.52	49.56	+0.77	+15.48	++	+
Protease A	0.01	22.21	0.17	87.59	+1.31	+20.47	+++	++
	0.05	24.29	0.07	84.57	- 2.84	+23.02	+++	++
	0.1	25.52	0.03	83.33	- 2.80	+25.41	+++	+
Protease	0.01	20.22	0.43	85.35	- 2.03	+18.43	++++	++
	0.05	21.99	0.39	88.80	- 2.91	+19.33	+++	++
	0.1	23.40	0.41	82.92	- 2.17	+21.67	++	+
Protease M	0.01	20.34	0.29	88.54	- 2.58	+18.29	+++	++
	0.05	21.45	0.34	89.32	- 3.06	+20.85	++	+
	0.1	22.68	0.29	84.11	- 2.43	+23.05	+	+
Papain	0.01	20.70	0.45	88.95	- 3.42	+23.56	++	++
	0.05	20.18	0.43	84.66	+3.26	+27.65	++	++
	0.1	24.34	0.52	87.10	+3.43	+26.16	+	+

. **Protamex(0.05%)**  
**Na<sub>3</sub>P<sub>0</sub>4(1.0%)** 가 **pH 10** **90**  
.

**Table 18. Changes in the characteristics of mussel extracts as affected by time of enzyme hydrolysis time extracted at 90 for 40 min**

Enzyme	Hydrolysis Time (min)	Solid yield (%)	Absorbance (600nm)	Color			Sensory properties	
				L	a	b	umami taste	fish taste
Protamex 0.05%	30	20.77	0.24	52.35	+1.04	+19.94	+++	+++
	60	24.06	0.33	49.62	+0.99	+19.66	++	++
	90	24.28	0.35	49.03	+0.76	+19.81	++	++
Protease 0.01%	30	19.87	0.38	88.04	-2.81	+19.49	+++	++
	60	21.11	0.45	87.95	-2.05	+19.12	++++	+
	90	22.87	0.49	89.51	-2.47	+19.81	+++	+

**Table 19. Changes in the characteristics of enzyme-treated mussel extracts as affected by phosphates and pH adjusted extracted at 90 for 40 min**

Enzyme	Phosphates	pH	Solid yield (%)	Absorbance (600nm)	Color			
					L	a	b	
Protamex (0.05%), 60min)	Na <sub>3</sub> PO <sub>4</sub> (0.5%)	2	27.20	0.41	83.28	-2.01	+26.35	
		9.53	27.86	0.57	59.55	+2.25	+26.17	
		10	30.53	0.99	25.70	+12.14	+17.34	
	Na <sub>4</sub> P <sub>2</sub> O <sub>7</sub> (1.0%)	2	23.92	0.20	86.64	-2.29	+21.64	
		8.32	25.74	0.32	70.36	+0.48	+28.16	
		10	27.98	2.24	23.10	+3.04	+12.77	
	Protease (0.01%), 90min)	Na <sub>3</sub> PO <sub>4</sub> (0.5%)	2	25.20	0.05	92.26	-3.16	+17.17
			9.53	24.86	0.03	77.79	-1.96	+28.02
			10	29.62	0.43	37.04	+10.03	+23.97
Na <sub>4</sub> P <sub>2</sub> O <sub>7</sub> (1.0%)		2	24.17	0.03	93.75	-3.26	+16.51	
		8.32	21.99	0.28	93.92	-3.87	+19.48	
		10	29.92	1.17	24.37	+8.05	+16.11	

가

Table

20

Protease

pH 2

pH 10

가

Protamex

pH10

가

Na<sub>3</sub>P<sub>04</sub>

Na<sub>4</sub>P<sub>2</sub>O<sub>7</sub>

가

가

Na<sub>3</sub>P<sub>04</sub>가 Na<sub>4</sub>P<sub>2</sub>O<sub>7</sub>

가

pH

10 (Table 19 )

Protamex

Na<sub>3</sub>P<sub>04</sub>, Na<sub>4</sub>P<sub>2</sub>O<sub>7</sub> 1%

가

가

가

30

21

Protamex 0.05%

Na<sub>3</sub>P<sub>04</sub>

1.0% 가

pH 10

가

(protamex 0.05%, Na<sub>3</sub>P<sub>04</sub> 1.0%, pH10)

가

80

(Table 21)

5%

가

18.97%

31.57%

가가

가

가

가

가

L

b

b

**Table 20. Sensory scores of mussel extracts extracted at 90 for 40 min as affected by phosphates and pH.**

Enzyme	Sensory attribute	Na3PO4(1.0%)			Na4P2O7(1.0%)			F- value	
		pH 2	pH 9.53	pH 10	pH 2	pH 8.32	pH 10		
Protamax 0.05%	Odor	Savory	3.00c	5.00ab	4.88ab	3.88bc	3.63c	5.75a	7.16*
		Fish	5.38a	3.25c	3.88bc	4.63ab	2.63c	3.63bc	5.60*
	Taste	Umami	5.88a	3.88b	6.88a	6.25a	6.00a	6.50a	4.43*
		Fish	5.38	3.38	3.75	3.63	3.25	3.38	2.79
Protease 0.01%	Odor	Savory	4.75ab	3.63b	5.00a	5.13a	3.63b	5.88a	4.67*
		Fish	3.25b	6.50a	6.13a	3.00b	2.63b	5.13a	11.32**
	Taste	Umami	5.38a	3.50c	6.38a	5.13ab	4.00bc	6.38a	6.58*
		Fish	3.75	6.25	4.88	4.38	4.13	3.63	3.64

a-c Mean scores follow by the same letter are not significantly

\* P < 0.05

\*\* P < 0.01

가

**Table 22**

가

가

5%

가

가

가

10%

가

가

가

가

5%

10%

가 가

가

5 10%

가

가



**Table 21. Effect of red pepper seeds addition ratio on properties of mussel extracts prepared by combined method<sup>a</sup> at 90 for 40 minutes**

Addition of pepper seed(%)	Solid yield(%)	Absorbance 600nm	Color		
			L	a	b
control	27.97	2.24	23.10	+3.04	+12.77
5	31.57	5.32	14.07	+3.50	+9.01
10	32.09	5.13	9.43	+3.63	+6.25
15	33.31	5.88	8.98	+4.45	+6.02
20	33.56	10.91	7.03	+4.24	+4.76

<sup>a</sup> Mussel extract was prepared by addition of Na<sub>2</sub>P<sub>2</sub>O<sub>7</sub>(0.5%), hydrolysis with Protanex(0.05%, 60min) and at pH 10 and 90 for 40 min

**Table 22. Sensory properties of RPS-mussel extracts as affected by addition ratio of red pepper seeds**

Description	Seed concentration(%)				F- value
	5	10	15	20	
Hot odor	3.25c	6.13b	7.25ab	8.13a	29.04**
Savory odor	6.00a	4.50a	2.75b	2.25b	10.12*
Fish odor	2.88b	4.50a	2.00bc	1.25c	12.72**
Hot taste	4.25c	5.63b	6.75b	8.63a	22.06**
Savory taste	5.88a	4.38b	2.89c	1.75d	26.61**
Fish taste	2.63b	3.88a	2.50b	1.63b	6.71*
Total acceptability	56a	57a	82bd	92cd	

Total acceptability was tested ranking test. Mean scores within row followed by the same letter are not significantly different at 5% level.  
\* P < 0.05 , \*\* P < 0.01

### 3. Pilot

가.

Pilot

가 . 가  
80 30 가  
가 , 가 ( 가  
Protamax Na<sub>3</sub>P<sub>0</sub>4 1% 가 pH 10  
90 40 ) .  
5-10% 가 2

. pilot

1) Fig. 3

Pilot

2)

Protamex Protease

Protamex

가 ( )

5% 10% 5%가 5% 가

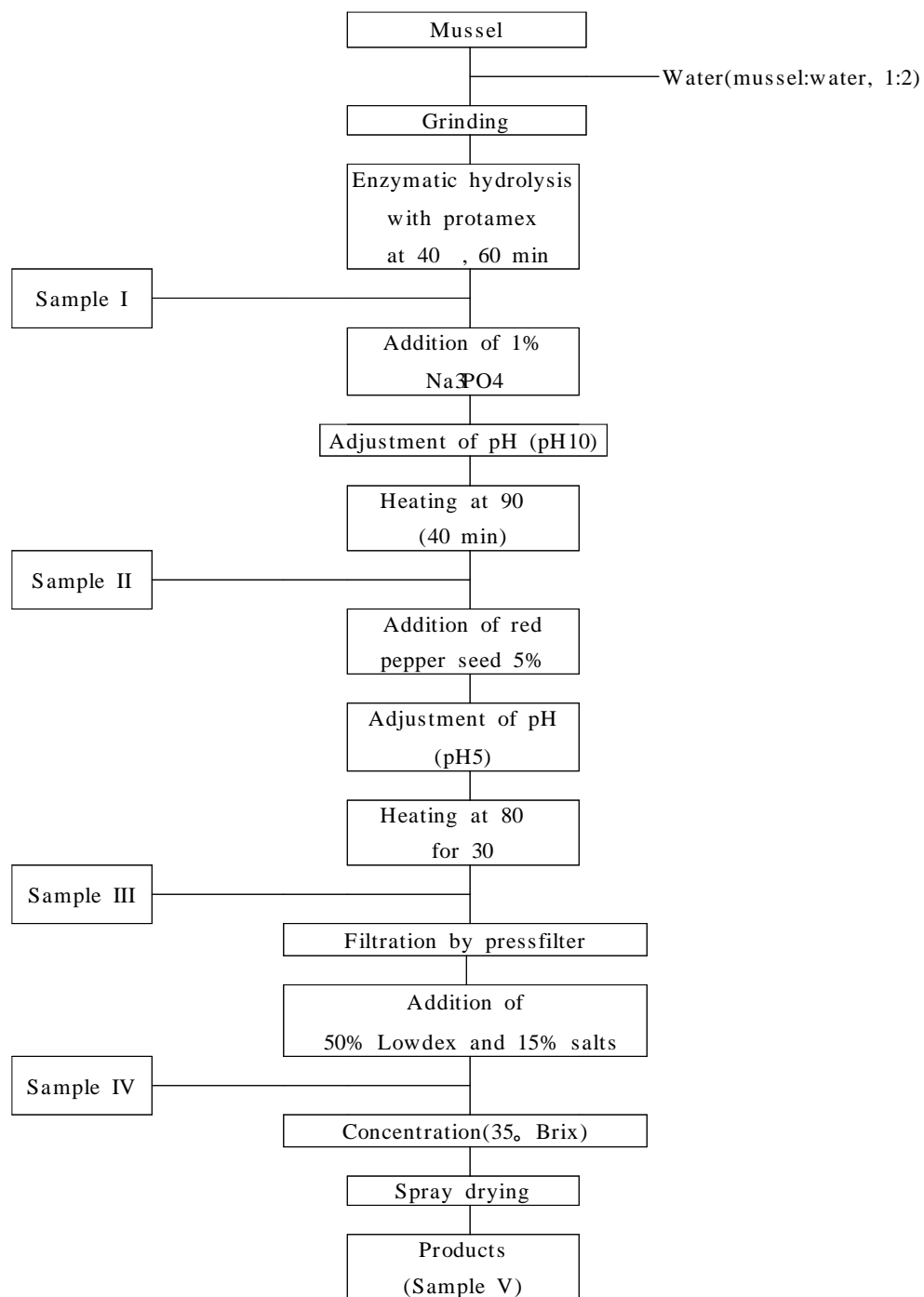
. pH NaOH HCl 가

5N 5N

3)

(1 , )

(2 , )



**Fig. 3. Schematic diagram for pilot plant scale preparation of red pepperseed-mussel spice.**

4) Pilot

(working volum 100kg)

(150 200mesh Filter Press)

50% Lowdex( ) 15% 가 .

35 ° brix가

Pilot

(sample ,

, , , )

(Hunter Lab),

, 가

.

. pilot

( )

1)

가)

( , 가 ) 75%

가

가 가

)

Filter press(180mesh)

pore

Decanter

Decanter

)

(100Kg)

가

)

30% , Pilot scale ( 1000M<sup>3</sup>/hr) . S/D(spray drying)

S/D

)

Pilot scale test , Decanter (Plan scale Production) 가

2)

type

가

가

가

가

3

가

가

( - ) , ,

(Table 23, 24)

( - ) (600nm) (420nm) 가  
(L ) (b )

가 Lowdex .

**Table 23. Physicochemical data of red repper seed - mussel spice produced from pilot plant scale preparation**

	Samples				
Moisture(%)	94.72	94.33	91.62	71.45	4.36
Protein(%)	2.03	2.63	2.87	9.8	23.62
Lipids(%)	1.05	0.58	1.26	0.57	2.11
Carbohydrate(%)	1.91	1.01	2.02	6.60	36.39
Reducing sugar(%)	0.4	-	-	2.30	9.45
Ash(%)	0.29	2.63	2.87	9.8	23.62
Viscosity(cps)	10.0	10.0	10.0	20.0	10.0
Absorbance(420nm)	23.93	28.46	26.84	18.13	14.48
Turbidity(600nm)	16.03	12.80	16.80	9.09	6.48
Color L	4.89	5.61	2.73	12.94	17.60
(1/2dilution) a	3.57	4.17	3.58	6.48	3.69
b	3.37	3.90	1.89	8.78	25.03

( )

가

**Table 24. Sensory properties of red pepper seed - mussel spice produced from pilot plant scale preparation**

		Samples					F- value
		I	II	III	IV	V	
Odor	Hot	3.60a	3.20a	6.80b	3.40a	2.10a	9.94
	Savory	3.10	5.20	4.40	4.10	3.80	1.20
	Fish	8.60a	7.50ab	6.30b	7.40ab	6.50b	4.04
	Sweet	1.80	3.00	2.00	2.04	2.70	1.05
Taste	Hot	2.00ab	1.20b	6.90c	2.70a	2.10ab	33.91
	Fish	8.30a	5.90b	5.50b	5.50b	4.30b	7.45
	Umami	2.00a	2.20a	4.50b	4.90b	3.70b	5.03
	Salty	2.60a	1.60a	5.10b	6.70b	4.30cb	12.46
	Sweet	1.60	1.70	2.20	2.40	1.90	0.67

**ac Mean scores follow by the same letter are not significantly at 5% level**

2

1.

가. NaCl 가

가 가

NaCl , , ,  
가, pH , 가 가

NaCl 가 25  
24 Table 25 .  
가 (0%) L 38.12, a  
-3.25 가 L 38.42 39.66, a -3.16  
-3.40 가 , b 가 가  
가 .

가 acetic acid 0.5% 가 L  
38.12 31.71 (-)a -3.25  
-10.49 가 (-)b 3.12 -0.64  
. Acetic acid 0.5% 가 L  
가 a (-)b 가 0.5% 가



acid acetic acid citric acid lactic acid

Table 25. Effects of NaCl and organic acids on color changes of ground garlic after storage at 25 for 24 hours.

	Hunter values	Concentration				
		0%	0.5%	1.0%	1.5%	2.0%
NaCl	L	38.12	38.42	38.47	39.56	39.66
	a	- 3.25	- 3.16	- 3.25	- 3.32	- 3.40
	b	3.12	3.66	3.87	3.98	4.01
Acetic acid	L	38.12	31.71	34.84	36.33	38.45
	a	- 3.25	- 10.49	- 10.85	- 10.17	- 10.76
	b	3.12	- 0.64	- 1.81	- 1.81	- 1.08
Citric acid	L	38.12	27.68	35.54	39.60	44.38
	a	- 3.25	- 8.22	- 10.72	- 9.99	- 10.47
	b	3.12	- 0.68	- 1.54	- 0.71	- 0.88
Lactic acid	L	38.25	29.00	34.54	38.69	42.11
	a	- 3.25	- 9.22	- 10.45	- 10.22	- 10.36
	b	3.12	- 0.96	- 1.60	- 0.48	0.16

pH,

가

가

polyphenol oxidase

pH

pH

Table 26

pH 6.4 6.6

pH 4.0

가 . pH 5.5 a  
 -5.95 .  
 pH pH 4.0 .  
 pH ,  
 (27) 가 , (NaH<sub>2</sub>PO<sub>4</sub> Na<sub>2</sub>HPO<sub>4</sub> Na<sub>3</sub>PO<sub>4</sub>)

Table 26. Effects of pH, salts, antioxidants and others on color changes of ground garlic after storage at 25 for 24 hours.

	Hunter values		
	L	a	b
Control	39.28	-3.55	3.82
pH 4.0	41.86	-0.29	-0.53
pH 4.5	39.10	-4.52	-0.70
pH 5.5	38.18	-5.95	-0.60
NaH <sub>2</sub> PO <sub>4</sub>	39.01	-3.51	2.56
Na <sub>2</sub> HPO <sub>4</sub>	38.94	-2.87	2.36
Na <sub>3</sub> PO <sub>4</sub>	38.33	-2.62	2.31
SPP	40.19	-3.01	4.19
SMP	39.51	-3.38	3.84
Na <sub>2</sub> S <sub>2</sub> O <sub>5</sub>	40.00	-2.07	3.81
Na <sub>2</sub> SO <sub>4</sub>	36.28	-2.23	1.94
BHA	43.30	-4.86	2.01
BHT	45.82	-5.47	2.04
PG	45.12	-5.41	1.98
glutathione	41.24	-3.72	4.37
Cysteine	40.17	-2.87	4.59
Ascorbic acid	41.70	-3.35	4.10

가

(-)a b

Na<sub>3</sub>PO<sub>4</sub>가

allergy-like

FDA

(37) sodi um metabi sulfite

(22) sodi um metabi sul fi te가

(38)

cysteine ascorbic acid

가

. 가

가

가

(50-100 )

(0-60 ) 가

25

(Fig. 4)

, L

가

가

가

가

a

10

가

(-)a

가

가

가 10

가

.

10

0

20

, 90

30

가

가

. 70 가 b 가 가  
. 가  
. (39)  
50 30 가 80 가  
가 . 가  
가 .

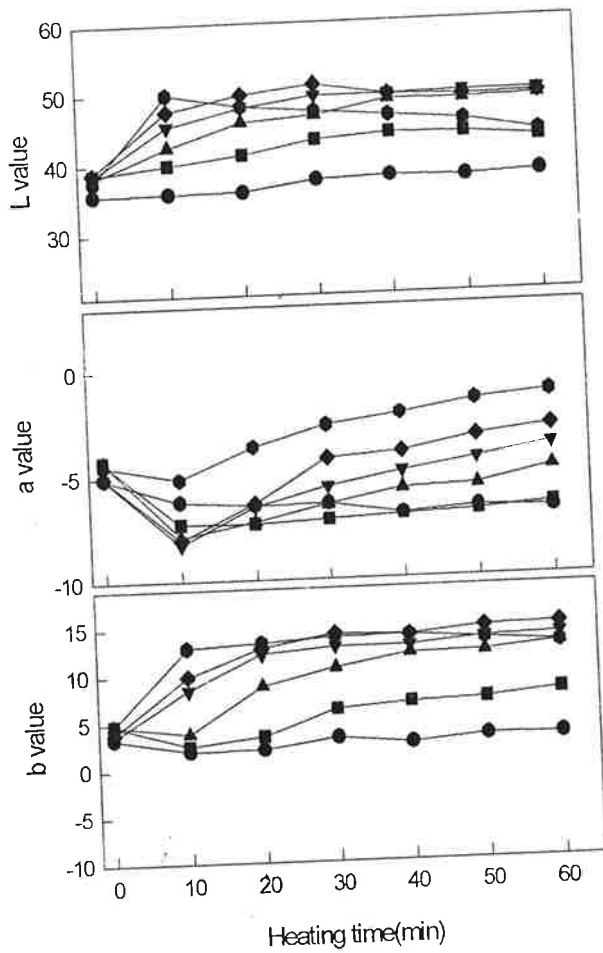


Fig. 4. Effects of heating temperature and time on color values of ground garlic after 24 hours of storage at 25°C

(—●— 50°C, —■— 60°C, —▲— 70°C, —▼— 80°C,  
—◆— 90°C, —●— 100°C)

Table 27 가

80 pH

가 가 (-) a 가 pH 3

-9.94가 b (-) 10 가

가 가 . 10

pH 4.0 5.0 . Fig. 2

10 가 a b

pH .

NaCl 가

acetic, citric, lactic

pH . 가 80

30 , 90 20 가

80-90 10 가 .

Table 27. Effect of pH and heating at 80 on color changes of ground garlic after storage at 25 for 24 hours.

pH	Heating time (min)	Hunter values		
		L	a	b
3.0	0	44.85	-4.71	2.41
	2	44.06	-7.73	-3.81
	6	43.96	-9.80	-3.65
	10	45.22	-9.94	-2.12
4.0	0	40.67	-4.13	2.26
	2	39.12	-7.58	-3.35
	6	42.86	-8.48	-2.85
	10	46.11	-10.20	-2.27
5.0	0	38.40	-3.37	2.47
	2	39.42	-6.65	-1.80
	6	41.22	-7.34	-0.85
	10	45.75	-8.87	6.59

가  
 ascorbic acid(A), cysteine(C), sodium metabisulfite(S), trisodium phosphate(P)

25  
 가 80 90 가  
 가

1)  
 0.1M A, C, S, P 25 7  
 L, a, b 44.28, -1.93,  
 11.27, Fig. 5 L 가

가

L

. a S

7

a A 가 가

4가

가

A가

가

(2)

. b

S

가

가

S

가

2) 가가

0. 1M A, C, S, P 가

6가

bi nary mi xture

Fig. 6

CP(C P ) 가 가

L

L

가 . , a, b

AP

가

. Binary mi xtures

A, C, P S

가 가

sul fi tes가

가

Soto (4)

0 a, b

가



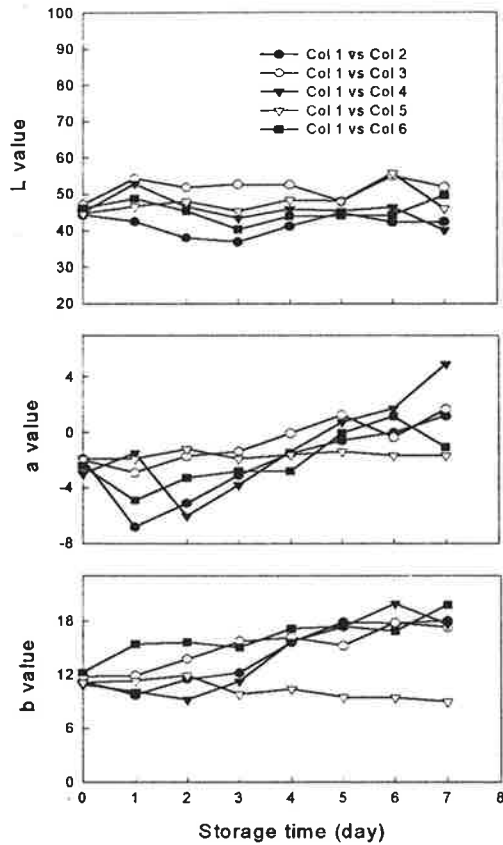


Fig. 5. Effect of 0.1M cysteine, ascorbic acid,  $\text{Na}_2\text{S}_2\text{O}_5$  and  $\text{Na}_3\text{PO}_4$  on Hunter values of ground garlic during storage at  $25^\circ\text{C}$

수 있었다.

0.1M의 A, C, S, P를 셋 혹은 네가지씩 혼합하여 처리한 경우(CAS, CAP, ASP, CSP, CASP)의 변색억제 효과는 Fig. 7과 같다. 3가지 이상의 물질로 처리한 경우 대조구에 비해 높은 L값을 계속 유지하는 것으로 나타났으며 CAP처리구를 제외한 모든 처리구의 a, b값은 거의 일정하게 유지되었다. 3가지 이상의 혼합물질 처리구의 a값은 저장 0일의 수치와 비교시 거의 일정하게 유지되어 2가지 물질을 혼합한 것보다도 갈변 및 녹변억제에 매우 효

과적임을 알 수 있었다. 그러나 b값의 경우에는 두가지 물질을 혼합하였을 경우와 그 경향이 매우 유사하였다.

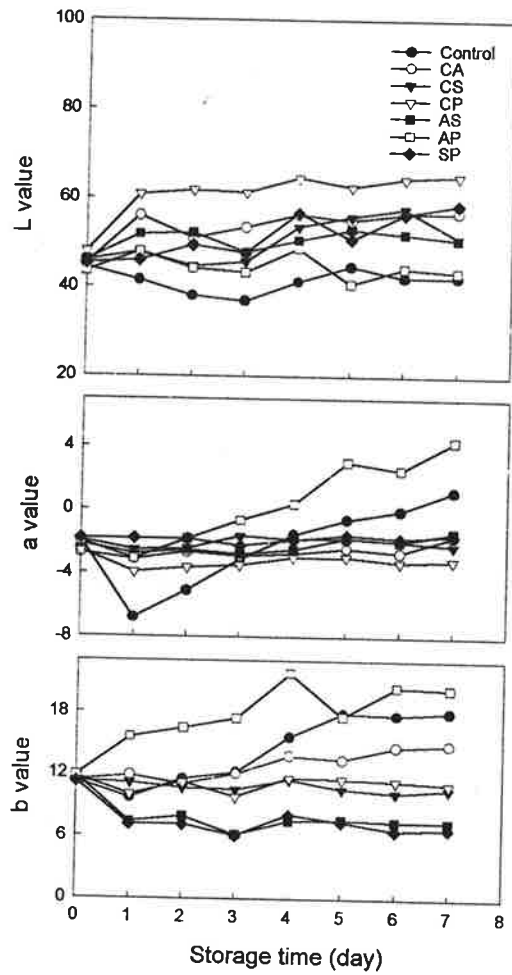


Fig. 6. Effect of binary mixtures of cysteine, ascorbic acid and  $\text{Na}_2\text{S}_2\text{O}_5$  at 0.1M each on Hunter values of ground garlic during storage at 25°C

3)

가

binary/ternary/quaternary

가

0.03M

0.01M

. 0.03M

L, a, b

(Fig. 8).

0.01M

(Fig. 9)

. ,

0.01M

a

.

가

L

, b

가

,

0.03M CS, CAS, CSP, CASP

가

.

가

.

,

A, C, S, P binary, ternary,

quaternary

.

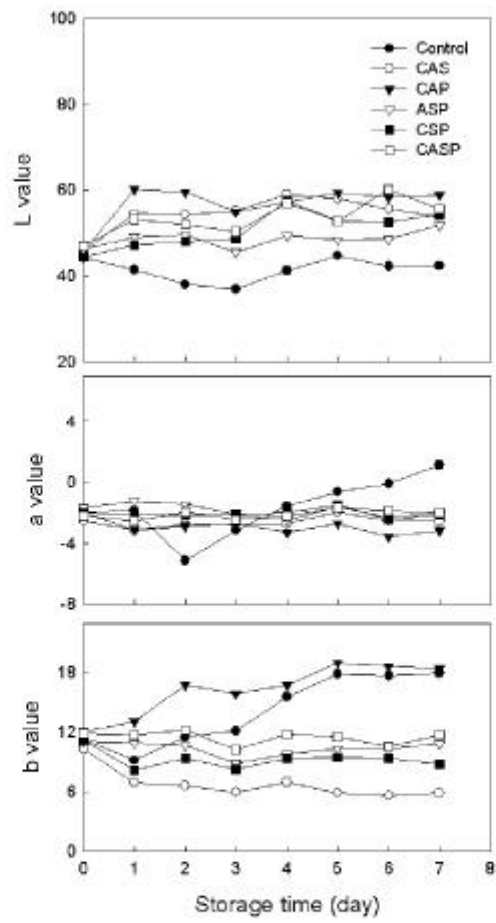


Fig. 7. Effect of ternary mixtures of cysteine, ascorbic acid,  $\text{Na}_2\text{S}_2\text{O}_5$  and  $\text{Na}_3\text{PO}_4$  at 0.1M each on Hunter values of ground garlic during storage at  $25^\circ\text{C}$

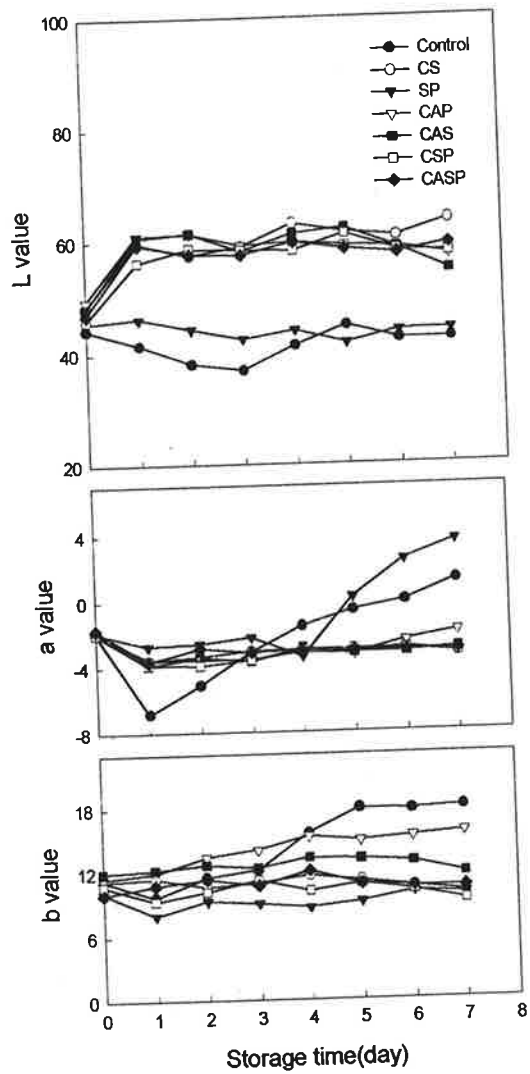


Fig. 8. Effect of binary and ternary mixtures of cysteine, ascorbic acid,  $\text{Na}_2\text{S}_2\text{O}_5$  and  $\text{Na}_2\text{PO}_3$  at 0.03M each on Hunter values of ground garlic during storage at  $25^\circ\text{C}$

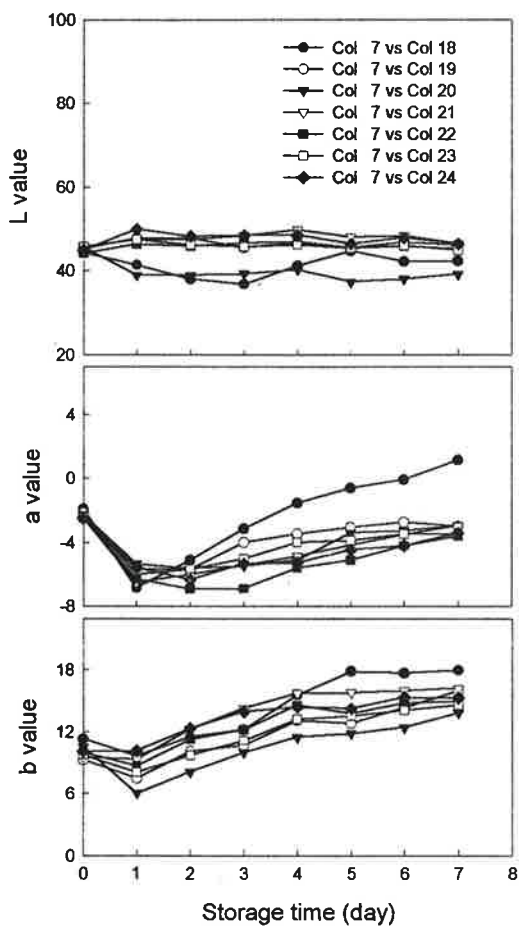


Fig. 9. Effect of binary and ternary mixtures of cysteine, ascorbic acid,  $\text{Na}_2\text{S}_2\text{O}_5$  and  $\text{Na}_3\text{PO}_4$  at 0.01M each on Hunter values of ground garlic during storage at  $25^\circ\text{C}$

2.

12%

5% .

pH, 가,

가. pH

pH가

Table 28 . ,

pH 2.5 5.5

pH가

가 pH 5.5

29.32% .

pH가

L

(-)a pH 4.5 5.5

, pH 3.5

pH 4.5

(4) pH가

L

가 b

가

Table 28. Effects of pH on solid yields and color of garlic extract prepared at 30 for 30minutes.

pH	Solid yield (%)	Hunter values		
		L	a	b
control	27.30	86.96	- 3.15	18.85
2.5	26.59	94.87	- 1.16	5.31
3.5	26.87	94.65	- 3.20	6.32
4.5	27.30	94.48	- 4.40	9.22
5.5	29.32	93.77	- 5.68	16.48

acidulants 가

가

(Table 29).

27.30% 가 가

K<sub>4</sub>P<sub>2</sub>O<sub>7</sub> 가 36.06%

가 , NaCl , KCl, CaCl<sub>2</sub>

31 33%

sodium oxalate 가 36.22%

33%

K<sub>4</sub>P<sub>2</sub>O<sub>7</sub>,

CaCl<sub>2</sub>, tartaric acid, sodium oxalate

Table 30 가

K<sub>4</sub>P<sub>2</sub>O<sub>7</sub> 가 35.82 38.28%

, CaCl<sub>2</sub> sodium oxalate



가  
 tartaric acid  
 phosphoric acid 가  
 가 가 가  
 tartaric acid  
 L 93.31, a -3.04, b 19.23 가  
 CaCl<sub>2</sub>, phosphoric acid 가 L 89.11 96.21  
 K<sub>2</sub>P<sub>2</sub>O<sub>7</sub>, sodium oxalate L  
 48.51 62.36, 72.27 81.01  
 K<sub>2</sub>P<sub>2</sub>O<sub>7</sub>, sodium oxalate 가  
 가 , tartaric  
 acid 가 가

Table 29. Effects of salts and acidulants on solid yields of garlic extract prepared at 30 for 30minutes.

Salts	Solid yield (%)	Acidulants	Solid yield (%)
Control	27.30	Acetic acid	33.86
NaCl	35.26	Citric acid	33.46
KCl	34.40	Lactic acid	33.48
CaCl <sub>2</sub>	34.17	Phosphoric acid	33.99
NaH <sub>2</sub> PO <sub>4</sub>	32.03	Tartaric acid	32.79
Na <sub>2</sub> HPO <sub>4</sub>	33.84	Oxalic acid	33.09
Na <sub>3</sub> PO <sub>4</sub>	33.30	Sodium oxalate	36.22
Na <sub>4</sub> P <sub>2</sub> O <sub>7</sub>	31.79		
KH <sub>2</sub> PO <sub>4</sub>	31.48		
K <sub>2</sub> HPO <sub>4</sub>	33.87		
K <sub>3</sub> PO <sub>4</sub>	33.72		
K <sub>4</sub> P <sub>2</sub> O <sub>7</sub>	36.06		

**Table 30. Effects of some additives on solid yields and color of garlic extract prepared at 30 for 30minutes.**

	Conc. (M)	Solid yield(%)	Hunter values		
			L	a	b
Control		27.30	80.77	- 1.32	24.74
K <sub>4</sub> P <sub>2</sub> O <sub>7</sub>	0.03	35.82	62.36	2.26	24.15
	0.06	36.24	52.60	3.36	21.74
	0.09	38.28	48.51	3.46	19.93
CaCl <sub>2</sub>	0.03	33.98	89.11	- 3.57	21.00
	0.06	34.20	93.27	- 3.22	19.36
	0.09	35.94	94.30	- 3.37	19.91
Tartaric acid	0.03	35.30	93.31	- 3.04	19.23
	0.06	34.02	91.58	- 2.23	14.48
	0.09	32.66	94.08	- 2.23	11.01
Phosphoric acid	0.03	33.02	90.83	- 3.02	22.73
	0.06	32.44	96.21	- 2.67	21.43
	0.09	32.41	94.95	- 3.43	20.24
Sodium oxalate	0.03	34.40	81.01	- 2.78	21.56
	0.06	34.72	75.19	- 0.59	23.45
	0.09	35.40	72.27	- 0.05	23.71

**Celluclast, Extrazyn, Viscozyn**  
**0.04 0.08%** 가  
**Table 31** .  
**(pH , acidulants 가)**

가 가 , Celluclast  
 Extrazym 90 0.04% 가  
 32.48%, 32.56% 가 0.08% 가 0.04% 가  
 , 가 가  
 Celluclast 0.04%-90 , Extrazym 0.08%-90 , Viscozym  
 0.08%-60 , 가 가 Extrazym  
 26.7% 가 .  
 (٪)  
 0.08% 0.12% 가 60 90  
 가 14% .  
 , 가  
 L , (-)a  
 가

3. oleoresin

가.

12%  
 8 가 30 50 1

Table 32

30 (43.4%) methanol (35.9%)

ethanol (27.1%), isopropanol (14.2%), acetone(14.2%)  
 diethyl ether ethylene chloride, methylene chloride,  
 n-hexane 1-2%

Table 31. Effects of enzyme concentration and reaction time on solid yields and color of garlic extracts at 30 .

Enzyme	Enzyme conc.(%)	Reaction time(min)	Solid yield (%)	Hunter values		
				L	a	b
Control	0	30	24.60	85.15	- 5.96	12.68
		60	26.75	83.13	- 7.17	13.04
		90	26.25	80.28	- 8.89	14.31
Celluclast	0.04	30	31.06	78.43	- 8.30	4.93
		60	31.82	69.37	- 10.26	- 1.60
		90	32.48	66.56	- 11.87	4.80
	0.08	30	31.21	77.81	- 9.11	4.07
		60	31.06	74.40	- 9.03	0.76
		90	31.88	68.69	- 10.22	1.05
Extrazym	0.04	30	31.26	74.86	- 8.06	4.38
		60	31.43	74.30	- 9.12	1.31
		90	32.56	69.74	- 10.22	2.67
	0.08	30	31.55	79.87	- 8.18	3.61
		60	31.51	72.46	- 9.24	0.19
		90	33.25	69.04	- 9.86	2.42
Viscozym	0.04	30	30.92	82.54	- 8.11	7.80
		60	31.32	70.48	- 9.27	- 0.80
		90	31.16	65.43	- 10.14	- 0.28
	0.08	30	30.97	80.41	- 8.29	6.25
		60	31.46	71.25	- 9.06	- 0.24
		90	31.09	68.00	- 9.74	0.12

가 0.177 가

가 30 50 가  
 43.4% 53.8% 10% 가가  
 methanol, ethanol, isopropanol, acetone  
 가 5% ,  
 diethyl ether, n-hexane, ethylene chloride, methylene chloride  
 가가 가 .  
 가 가  
 30 .

methanol, ethanol, isopropanol, acetone  
 30 4  
 Table 33 . 가 가  
 , 3 가 . L  
 methanol ethanol, acetone, isopropanol 가  
 가 . a - 가가  
 2 .  
 가  
 65%  
 . methanol  
 methanol  
 ethanol oleoresin .

**Table 32. Effects of various solvents on solid yield and turbidity after extraction of garlic root portion at 30 -50 for 1hr**

	Solid yield(%)			Absorbance(600nm)		
	30	40	50	30	40	50
Water	43.4	49.7	53.8	0.177	0.146	0.135
Methanol	35.9	38.6	39.7	0.020	0.051	0.040
Ethanol	27.1	28.3	29.0	0.019	0.030	0.039
Acetone	11.1	11.2	11.6	0.009	0.054	0.024
Isopropanol	14.2	14.7	15.2	0.008	0.017	0.022
Diethyl ether	2.1	2.2	3.0	0.006	0.007	0.018
n- hexane	0.5	0.6	0.4	0.004	0.011	0.009
Ethylene chloride	0.6	0.7	0.8	0.032	0.007	0.001
Methylene chloride	0.9	1.1	1.1	0.055	0.011	0.006

**Table 33. Effects of extraction time on solid yield, turbidity and color of extracts of garlic root portion extracted at 30 for 2-4hrs**

Extraction time		Solid yield(%)	Absorbance (600nm)	Hunter		
				L	a	b
2hrs	Water	54.1	0.185	83.00	- 2.84	22.75
	Methanol	41.1	0.026	96.08	- 6.42	21.78
	Ethylanol	28.2	0.021	96.70	- 5.78	19.62
	Isopropanol	14.9	0.014	97.19	- 4.64	16.66
	Acetone	10.9	0.015	97.00	- 4.16	15.12
3hrs	Water	56.1	0.184	83.70	- 2.88	23.17
	Methanol	41.3	0.024	96.01	- 6.63	22.01
	Ethylanol	28.7	0.017	96.55	- 6.03	20.62
	Isopropanol	15.7	0.008	97.47	- 4.95	17.38
	Acetone	11.1	0.013	96.93	- 4.35	15.96
4hrs	Water	58.5	0.148	85.62	- 3.17	23.17
	Methanol	41.6	0.035	95.85	- 6.34	20.10
	Ethylanol	29.1	0.027	96.21	- 6.58	20.99
	Isopropanol	16.0	0.017	97.37	- 5.47	17.87
	Acetone	11.2	0.018	97.34	- 5.33	17.50

4. Oleoresin

가. , 가

Ethanol ( , , )  
 34 . pH 가 2 pH 5.86  
 가 a 가 .  
 thiosulfate 가  
 가 .

Table 34. Changes in some characteristics of garlic oleoresin during storage in N<sub>2</sub>, O<sub>2</sub> and vaccum conditions at 25

Storage time (weeks)	Storage conditions	pH	TA (%)	Absorbance		Hunter color		
				thiosulfinate	turbidity	L	a	b
0		5.96	0.52	2.29	0.22	56.38	5.82	19.54
	N <sub>2</sub>	5.88	0.54	1.53	1.04	34.00	2.55	19.33
1	Vaccum	5.89	0.53	1.47	1.32	30.86	4.99	17.47
	O <sub>2</sub>	5.82	0.62	1.39	1.60	29.74	6.43	15.94
	N <sub>2</sub>	5.90	0.56	1.32	1.53	29.60	7.32	18.24
2	Vaccum	5.88	0.56	1.31	1.41	30.95	10.97	15.76
	O <sub>2</sub>	5.86	0.65	1.25	1.58	29.04	14.34	15.86

가 25 14 , 0.1% Na<sub>2</sub>S<sub>2</sub>O<sub>5</sub> Na<sub>2</sub>PO<sub>4</sub> 1% (Table  
 35) pH , 14  
 가 Na<sub>2</sub>PO<sub>4</sub> 가 pH 5.44 pH 6 가  
 . pH 14

Table 35. Effects of N<sub>2</sub>, vacuum, Na<sub>2</sub>S<sub>2</sub>O<sub>5</sub> and Na<sub>2</sub>PO<sub>4</sub> on some characteristics of garlic oleoresin during storage at 25

	day	pH	TA(%)	Absorbance		Hunter		
				turbidity	thiosulfinate	L	a	b
Control (Air)	0	5.44	0.58	1.09	2.68	46.28	0.80	27.75
	14	5.46	0.55	0.40	0.25	60.72	21.17	40.92
N <sub>2</sub>	0	5.44	0.58	1.09	2.68	46.28	0.80	27.75
	14	5.45	0.54	0.43	0.26	64.67	16.51	42.21
Vacuum	0	5.44	0.58	1.09	2.68	46.28	0.80	27.75
	14	5.44	0.56	0.50	0.28	62.80	15.57	41.25
Na <sub>2</sub> S <sub>2</sub> O <sub>5</sub>	0		0.58	1.09	2.68	46.28	0.80	27.75
	14	5.41	0.49	0.33	0.22	66.63	15.14	43.88
Na <sub>2</sub> PO <sub>4</sub>	0		0.58	1.09	2.68	46.28	0.80	27.75
	14	5.92	0.40	0.39	0.19	49.99	25.42	34.15

가 . L  
 가 , a



. , Na<sub>2</sub>S<sub>2</sub>O<sub>5</sub> 가 a  
 가 Na<sub>3</sub>PO<sub>4</sub> 가  
 가  
 . Na<sub>3</sub>PO<sub>4</sub> 가 pH pH 6.5  
 PPO 가 a .  
 alliin thi osul fi nate  
 14 10  
 . Na<sub>2</sub>S<sub>2</sub>O<sub>5</sub>, Na<sub>3</sub>PO<sub>4</sub>  
 .  
 . ascorbic acid cysteine 가  
 oleoresin 가 pH  
 가 ascorbic  
 acid(0.1%) cysteine(1.0%) 가 4 28  
 (Table 36). L  
 28 ascorbic acid cysteine 가 가  
 L . a 가 28 5.72  
 ascorbic acid cysteine 가 가 4.12, 5.44  
 가가 가 가 .  
 가  
 . b a 가  
 L .

**Table 36. Changes in free sugar, organic acids color and turbidity of oleoresin as affected by addition of ascorbic acid and cysteine during storage at 4 under nitrogen stuffing**

		Control		Ascorbic acid		Cysteine	
		0 day	28 day	0 day	28 day	0 day	28 day
Sugars (ppm)	Sucrose	325.68	328.23	325.68	340.09	325.68	342.91
	Glucose	29.59	47.01	29.59	47.23	29.59	49.96
	Fructose	124.49	119.19	124.49	130.06	124.49	124.73
Acids (ppm)	Citric acid	78.13	63.58	78.13	45.88	78.13	53.54
	Malic acid	131.18	82.64	131.18	87.49	131.18	85.66
	Lactic acid	103.40	75.43	103.40	56.21	103.40	67.36
	Acetic acid	9.86	10.21	9.86	9.64	9.86	9.66
	Succinic acid	25.94	21.26	25.94	16.76	25.94	17.92
Color	L	63.28	43.77	63.28	53.43	63.28	50.36
	a	- 0.65	5.72	- 0.65	4.12	- 0.65	5.44
	b	37.91	27.73	37.91	33.27	37.91	31.62
Turbidity		0.6213	0.9864	0.6213	0.7190	0.6213	0.8066

sucrose 가 , fructose, glucose .  
 Sucrose, glucose, fructose 28  
 가 sucrose  
 ascorbic acid cysteine 가 .  
 Fructose 28 ,  
 glucose 가 28  
 2 가 가 . oleoresin  
 sucrose가 65% .  
 , ascorbic acid, cysteine 가



**Table 37. Sensory properties of garlic oleoresin as affected by addition of ascorbic acid and cysteine during storage at 4 under nitrogen stuffing**

Treatment	Sensory attribute	Storage time(days)					F- value	
		0	7	14	21	28		
Control	Odor	Fresh	5.3a	4.6b	4.6b	4.6b	4.1b	4.79*
		Hot	5.9a	5.0b	5.0b	4.4b	4.7b	6.00*
		Acidic	3.9	3.9	4.1	3.9	3.7	0.47
	Taste	Hot	6.3a	5.7b	5.4b	5.4b	4.6c	10.07**
		Acidic	1.9ab	2.1a	1.7ab	1.3b	1.4b	2.38
		Fresh	7.4a	7.3a	7.3a	6.6b	6.5b	5.62*
Ascorbic acid	Odor	Hot	6.3a	6.3a	6.3a	5.7ab	5.6b	2.82
		Acidic	4.4ab	3.9b	4.6a	4.1ab	4.0ab	2.02
		Fresh	7.4a	7.3a	7.3a	6.6b	6.5b	5.62*
	Taste	Hot	8.4	8.3	8.3	8.0	7.9	1.23
		Acidic	2.4	2.4	2.3	2.6	2.7	0.70
		Fresh	7.9a	7.6ab	7.5abc	7.0bc	6.9c	3.89
Cysteine	Odor	Hot	6.7	6.4	6.4	6.4	6.4	0.41
		Acidic	3.1a	2.7ab	2.7ab	3.0a	2.3b	2.48
		Fresh	7.9a	7.6ab	7.5abc	7.0bc	6.9c	3.89
	Taste	Hot	8.7a	8.3a	8.3a	8.3a	7.6b	4.79*
		Acidic	2.6a	2.0ab	1.9b	1.9b	2.1ab	2.22
		Fresh	7.9a	7.6ab	7.5abc	7.0bc	6.9c	3.89

±-c Means in a column with same letters are not significantly different at 1.0%(\*\*) or 5.0%(\*) level

5. Pilot

가. paste

1)

( )

2

paste

2) paste Fig. 10

Super mill (nascoroid mill) 300mesh

Viscozym L (Novo co.) 0.2% 가

pH 5 50 60 . Paste

xanthan gum 0.5% 가 Na<sub>3</sub>P<sub>0</sub>4,

K<sub>2</sub>S<sub>2</sub>O<sub>5</sub> Cystein 0.05%, 0.3%, 0.02% 가

3) paste 95%

1) Paste

2)

3) 가 xanthan gum  
(0.5% 0.2% 가 ) (xanthan gum +  
guar gum)

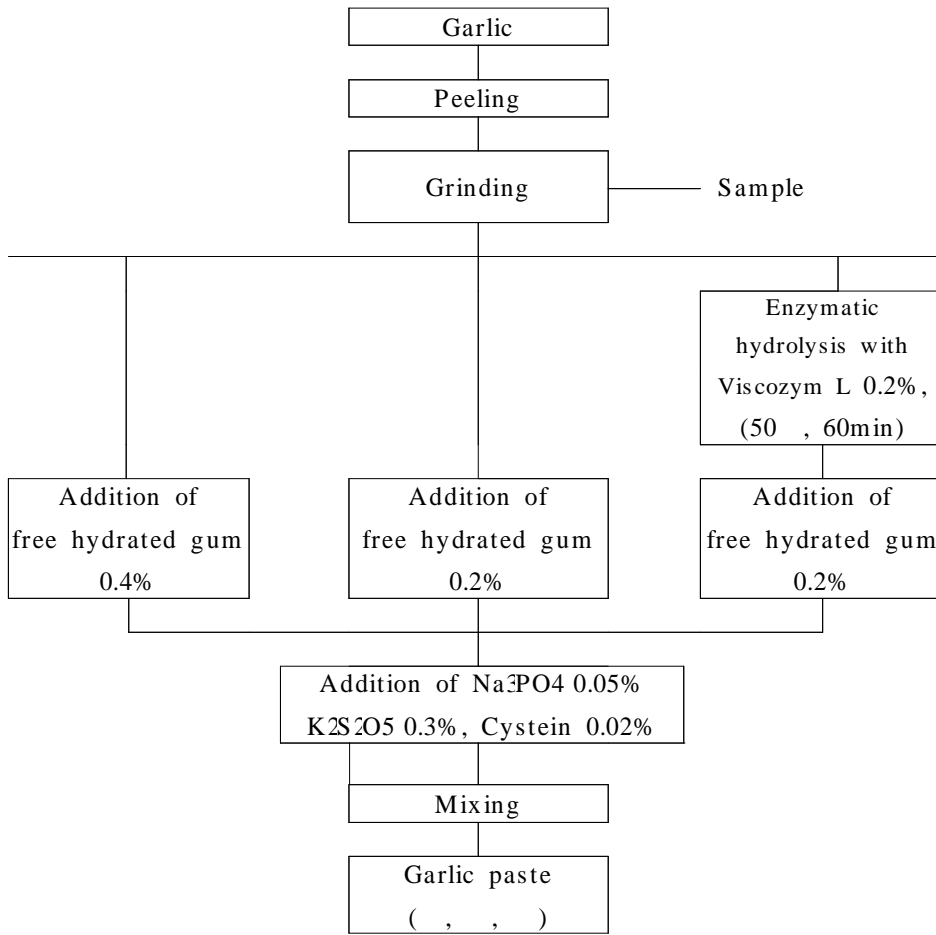


Fig. 10. Schematic diagram for pilot plant scale preparation of garlic paste.

paste ( ) 3가 ( , , )  
 , , 41 .  
 75.3-76.2%  
 ( )가 3.06% paste ( , )

free hydrated gum 0.4% 가  
 (19.5%)  
 1.2%  
 1.05%  
 가 free hydrated gum  
 가  
 가 L, a, b 가

Table 41. Proximate composition and some properties of samples obtained from pilot plant scale preparation of garlic paste

	Samples			
	I	II	III	IV
Moisture(%)	76.2	75.3	75.6	76.1
Protein(%)	2.63	2.63	2.39	3.06
Lipids(%)	1.23	1.04	1.25	1.26
Carbohydrate(%)	18.89	19.74	19.54	18.26
Reducingsugar(%)	4.67	3.92	4.12	5.03
Ash(%)	1.05	1.29	1.22	1.32
Viscosity(cps)	2.00 × 10 <sup>2</sup>	2.32 × 10 <sup>3</sup>	4.34 × 10 <sup>3</sup>	1.43 × 10 <sup>2</sup>
color L	41.36	43.29	40.10	40.06
a	+2.31	+1.01	+1.65	+0.81
b	+14.00	+13.05	+0.81	10.95

- . ground garlic
- . garlic paste added with 0.2% free hydrated gum
- . garlic paste added with 0.4% free hydrated gum
- . sample hydrolyzed by 0.2% Viscozym L

가

2

paste

가

Table 37, 38

(
· (1.5 3ton/H) : 3,000
· (1.5ton) : 5,500
· (2ton/batch) : 2,000

Table 38. - paste

( : )

		105,000	
	43% 1)	45,150	
	14% 1)	14,700	
	15% 1)	15,750	
	35% 1)	36,750	
	13% 1)	13,650	
	20% 1)	21,000	<b>252,000</b>
	30% 1)	31,500	<b>283,500( )</b>
	7% 2)	19,845	
	15% 2)	42,525	
			<b>345,870</b>

1) : %, 2) : %



Table 39. - paste

( : )

	-	paste
가	4,936.22	4,391.75
(kg )	32.80	80.80
	117.67	72.09
( )	38.96	6.54
( )	294.21	6.30
	5.74	3.49
	4.80	0.37
	331.00	
	470.00	470.00
	6,231.39	5,031.34
	810.08	654.07
	7,041.47	5,685.41

Table 40. - paste 가 ( : )

	-	paste
(%)	90.60	84.20
(kg/hr)	500.00	459.00

•

## 4

1. : (1990)
2. , : 가 capsaicin .  
 , 14, 409(1985)
3. Thresh : *Pharm. J. Trans.*, 3, 473(1876)
4. Nelsn, E. K. : *J. Ind. Eng. Chem.*, 2, 419(1910)
5. Nelsn, E. K. : *J. Chem. Soci.*, 41, 1115(1919)
6. Huffnan, V. L., Schalde, E. R., Villalon, B. and Burns, E. E. :  
Volatile components and pungency in fresh and processed Jalapens  
peppers. *J. Food Sci.*, 43, 1809(1978)
7. Rahway, N. J. , U.S.A : The Merck Index. An Encyclopedia of Chemicals,  
Drugs, and Biologicals. 11th ed, Merck & CO., Ins. 1767, 266(1989)
8. , , : oleoresin .  
 , 16(3), 85(1987)
9. Bennet, D. J. and Kirby, G. W. : Constitution biosynthesis of  
capsaisin. *J. Chem. Soc.*, 442(1968)
10. Crombie, L., Dandegaonker, S. H. and Simpson, K.B. : Amide of  
vegetable origin. Part . Synthesis of capsaicin. *J. Chem. Soc.*,  
1025(1955)
11. Lee, K., Suzuki, T., Kobash, M., Hasegawa, K. and Iwai, K. :  
Quantitative microanalysis of capsaicin, dihydrocapsaicin using  
mass fragmentography. *J. Chromatogr.*, 123, 119(1976)
12. , , :  
 , 24, 3, 266(1992)

13. : . (1984)
14. : Cas liquid chromatography .  
 , 11, 278(1979)
15. : .  
(1996)
16. Cruess, W. V. : Experiments on garlic and onion extracts. *Fruit products J.*, 23, 305-313 (1944)
17. , : ( 1 ).  
가 , 24(1), 43(1986)
18. , : ( 2 ).  
가 , 24(1), 54(1986)
19. , : ( 3 ).  
가 , 24(1), 87(1986)
20. Kin, D. Y., Rhee, C. O. and Kim, Y. B. : Characteristics of polyphenol oxidase from garlic(*Allium sativum* L.) (in Korean). *J. Korean Agric. Chem. Soc.*, 24, 167-173 (1981)
21. Singh. L. J., Pruthi. J. S., Sankaran. A. N., Indiranna, K. and Girdhari L. : Effect of type of packaging and storage temperature on flavour and colour of garlic powder. *Food Science.* 8, 457-460 (1959)
22. Kin, H. K., Jo, K. S., Shin, D. H. and Kim, I. H. : Effects of phosphate complex treatment on the quality of dried garlic flakes (in Korean). *Korean J. Food Sci. Technol.*, 19, 75-80 (1987)
23. Lukes, T. M. : Factors governing in greening of garlic puree. *J. Food Sci.*, 51, 1577-1578 (1986)

24. Bae, R. N., Lee, S. K. : Factors affecting browning and its control methods in chopped garlic. *J. Kor. Soc. Hort. Sci.*, 31, 213-218 (1990)
25. Shin, D. B. : Effect of extraction and dehydration methods on flavor compounds of garlic powder (in Korean). Ph. D. Thesis, Chung-Ang Univ., Korea (1995)
26. Bae, T. J., Kang, H. I., Kim, H. J., Choi, O. S. and Ha, B. S. : Studies on oleoresin product from spices 3. Rapid processing of garlic oleoresin (in Korean). *J. Korean Soc. Food Nutr.*, 22, 73-77 (1993)
27. Jo K. S., Kim, H. K., Kwon, D. J., Park, M. H. and Shin, H. S. : Preparation and keeping quality of garlic oleoresin (in Korean). *Korean J. Food Sci. Technol.*, 22, 846-851 (1990)
28. 김, 동진, 박, 명희, 조, 김숙, 이, 호진, 이, 호진 : Oleoresin  
가 . , 24(2), 137(1992)
29. A.O.A.C. : *Official Methods of Analysis*, 15th ed., Association of Official Analytical Chemists, Washington D. C., p. 912(1990)
30. Hartwig, P. and McDaniel M. R. : Flavor characteristics of lactic, malic, citric, and acetic acids at various pH levels. *J. Food Sci.*, 60, 384-388 (1995)
31. 김, 동진 : 가 . , 64, 47(1982)
32. 김, 동진 : . , p. 306(1978)
33. Sofos, J.N. : Use of phosphates in low-sodium meat products, *Food. Technol.*, 40(9), 52(1986)

34. , , :  
 , 17(2), 65(1985)
35. , , , : 가가  
 , 17(4), 257(1985)
36. Montecalvo, J. Jr., Constantinides, S. M. and Yang, C. S. T :  
Optimization of processing parameters of the preparation of  
flounder frame protein product. *J. Food Sci*, 49, 172(1984)
37. Browing of foods : Control by sulfites, antioxidants and other means,  
*Food Technol.*, 40, 52(1986)
38. Dziezak, J. D. : Preservatives : Antioxidants. *Food Technol.*, 40,  
94-102(1986)
39. Kin, D. M. and Kin, K. H. : On the development of flesh greening of  
the stored garlic. (in Korean) *Korean J. Food Sci. Technol.*, 22,  
50-55(1990)
40. Sayavedra-Soto, L. A. and Montgomery, M. W. : Inhibition of polyphenol  
-oxidase by sulfite. *J. Food Sci.* 51, 1531-1536(1986)
41. Cho, J. S. : Control of color changes and improvement of extraction  
yield of ground garlic. M. S. Thesis, Sejong Univ., Korea(1996)

2 .

1

가

가

, 가 , , 가

1997 1 6.6kg

(1,2). alliin

. alliin 가

alliinase thiosulfinate alliin

pyruvic acid가 가 thiosulfinate가

thiosulfinate diallyl-disulfide sulfide mercaptan

(3,5).

가 (6,7)

allicin thiosulfinate -SH 가

(8,9) 가

allicin thiosulfinate, garlic oil,

scordinin ethanol peroxidase

di mutase 가 가 . (10,11) 가

가

가

resin                      oleoresin, allicin  
alliinase

,                      (1: 1).

가

가

salad dressing

가    가    .

가

## 2

### 1

가

. Cysteine, 5,5'-di-thio-bis(2-nitrobenzoic acid)  
(DTNB), 1,1-Diphenyl-2-picryl-hydrazyl (DPPH), HEPES(N-[2-Hydroxy  
ethyl]piperazine-N'-[2-ethanesulfonic acid]) 2-thiobarbituric  
acid(4,6-Dihydroxy-2-thiopyrimidine) Sigma Chem. Co. (St. Louis,  
MO, U.S.A.)

### 2

#### 1.

4 homogenizer(Nissei, MA  
-10, Ace homogenizer, Japan) 가 14,000rpm  
5 30 4  
30 5,000rpm( Beckman, Model J2-21 centrifuge)

- 64



가.

가 ,

, petroleum ether, benzene, chloroform, ethyl acetate , methanol  
acetic, citric, tartaric acid

5 가

. Methanol ethanol

Methanol ethanol

. 50% Ethanol pH citric acid, NaCl 가

Methanol 50% ethanol 50% 1N, 0.1N HCl NaOH 가 pH3  
8 , citric acid 1 5%, NaCl 1 5%, NaH<sub>2</sub>PO<sub>4</sub>  
0.5 2.0%, Na<sub>2</sub>HPO<sub>4</sub> 0.5 2.0% 가

5

2. 가

10g polyethylene film silicon  
oil 가 40 120 10 30 가

ethanol 50% 가 5 4 honogenizer  
 14.000rpm 30  
 4 30 5,000rpm

3.

(40 120 ), (10 30min) 가 , ethanol 50% 가  
 5 가 (4. , 25 ° , 50 )  
 30 5

4.

가

가.

가 가  
 SS-1, SS-2, SP-1, SP-2, SP-3  
 . (10)

yeast extract 0.3%, malt  
 extract 0.3%, peptone 0.5%, glucose 1%, salt 18%(YMPGSB)

pH(5.0)

. YMPGSB

가 가

PDA (potato dextrose agar, NaCl 5%)

30 96 가 . Screw cap

tube Durham YMPGSB(NaCl 18%) 20ml 121

15 colony 1 30 가

, 가 PDA(NaCl 5%) 30

96 colony 1 YMPGSB(NaCl 18%) 30

48 . Screw cap tube Durham

121 15 (0.5 2%)

(Christ Alpha 1-4, B, Braun Biotech. International)

(0.5 1.5%) 1% 30 가

. YMPGSB

가

YMPGSB(NaCl 18%) screw cap tube 20ml 121 15

, screw cap tube (0.5 1.5%) (

0.5 2%) 1% 30

5.

100kg

1-2 가

2 (7mm plate) ,

10% 가 . 5%,

10%, 20% 10% ethanol 50%

가 ,

polyethylene film 1 2

가 가 (100 autoclave 10 가 )

0, 2, 4, 6, 8 10 (4 ± 1 )

6.

가.

가 (10Mℓ) 105 ,

(17) . ,

DPPH 8mg ethanol 300Mℓ . 0.5Mℓ

DPPH 5Mℓ 가 vortex mixer 10



[ 5,5'-di thio -bis(2-nitrobenzoi c acid) ] 1Mℓ 가

27 10 412nm

standard curve

가(peroxide value)

가 AOAC Cd 8-53 (2)

5g petroleum ether 10Mℓ

celite 545 10g

BHA, tocopherol 0.05% 가

(0.8) 가

가 1g 250Mℓ flask

(acetic acid chloroform 3:2 ) 30Mℓ 가

KI 0.5Mℓ 1

30Mℓ 가 0.01N 0.1N sodi um

thi osul fi nate 1% 가

. Thiobarbituric acid value(TBA가)

Witte (2)

10g 20% trichloroacetic acid 2M phosphoric acid

25Mℓ 가 homogenizer 10,000rpm 1.5

50Mℓ volumetric flask 20Mℓ

가 shaki ng 50Mℓ

5Mℓ test tube(15 × 200mm) 0.005M  
2-thiobarbituric acid 5Mℓ 가 ,  
, 15 530nm  
10g OD .

### 3

#### 1

1.

가 Table  
1 methanol, chloroform, petroleum ether, benzene, ethyl  
acetate 가 methanol 8.1%  
가 1%

DPPH (Table 1)  
methanol > benzene >  
petroleum ether > chloroform > ethyl acetate  
methanol 36.8%  
1.8 2.7 .

pH (Table 1)  
methanol pH  
pH 1.2 64%, pH 3.0 38%  
pH 4.2 pH 6.0  
, pH pH 1.2 pH  
3.0 pH 4.2 pH 6.0

. Chloroform, petroleum ether, benzene

pH .



**Table 1. Solid yields, electron donating abilities (EDA) and nitrite-scavenging effects of garlic extracts as affected by various solvents**

Solvents (polarity index)	Yields (%)	EDA (%)	Nitrite scavenging (%)			
			pH1.2	pH3.0	pH4.2	pH6.0
Petroleum ether (0.1)	0.58 ± 0.04	19.63 ± 3.85	1.06 ± 0.09	1.64 ± 0.32	0.10 ± 0.03	0
Benzene (2.7)	0.50 ± 0.00	20.22 ± 0.10	0.43 ± 0.07	0	0	0
Chloroform (4.1)	0.78 ± 0.04	13.14 ± 2.26	0.54 ± 0.06	0.02 ± 0.02	0	0
Ethyl acetate (4.4)	0.50 ± 0.00	19.22 ± 2.51	23.35 ± 1.85	2.70 ± 0.37	0.56 ± 0.13	0
Methanol (5.1)	8.13 ± 0.81	36.33 ± 2.50	63.19 ± 1.39	38.81 ± 1.73	22.53 ± 1.55	0

Fox Ackerman(2) ascorbic acid, cysteine, nicotinamide adenine  
dinucleotide

nitric oxide

pH protonate form nitrous acid(pKa=3.4)

, 2, 3

5g 가 BHA tocopherol 0.05%  
가 30  
가 Fig. 1  
가 가 BHA

tocopherol 가 가 가  
 BHA tocopherol  
 가가 가  
 가가 . 6~7 가가  
 2  
 . (24)  
 가 BHA < tocopherol < petroleum  
 ethermethanol < benzene < ethyl acetate < chloroform < control

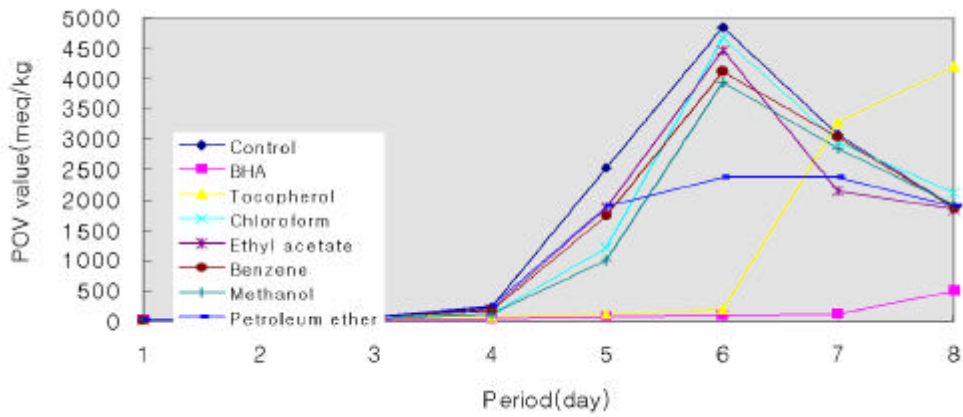


Fig.1. Changes in peroxide value of garlic extracts as affected by various solvents.

2.

acetic, citric, tartaric acid

, Table 2 citric, acetic, tartaric,  
 phosphoric acid citric acid 28.7%

, phosphoric acid 23.5% 가

가 (Table 2)

citric < tartaric < phosphoric < acetic acid

acetic acid citric acid 5% citric,  
tartaric, phosphoric acid pH

pH , pH

pH 1.2 pH 3.0 pH 4.2 pH

6.0 pH

1.2 pH 3.0 . pH 가

(Table2). Tartaric acid

pH1.2 50%,

pH3.0 14% acetic acid

citric acid citric acid pH1.2

tartaric acid 60%, pH3.0% 50% .

가 가 가

Fig. 2 . BHA tocopherol 가가

가

가가 . 가 가

가 가 6~7 가가

. 가 BHA < tocopherol < tartaric  
< acetic < phosphoric < citric acid < control .

가 , methanol

1/2,

1/2-1/3 methanol

### 3. Methanol

가                   methanol  
methanol

**Table 2. Solid yields, electron donating abilities (EDA) and nitrite-scavenging effects of garlic extracts as affected by various acids**

Acids	Yields(%)	EDA(%)	Nitrite scavenging effects(%)	
			pH1.2	pH3.0
Acetic acid	26.90 ± 0.18	19.18 ± 0.51	38.76 ± 8.85	15.83 ± 5.56
Citric acid	28.72 ± 1.06	12.78 ± 1.44	29.87 ± 3.00	7.14 ± 3.15
Phosphoric acid	23.47 ± 1.77	14.52 ± 2.23	34.33 ± 0.95	14.41 ± 3.10
Tartaric acid	25.97 ± 2.39	13.37 ± 0.60	50.08 ± 10.50	14.29 ± 4.18

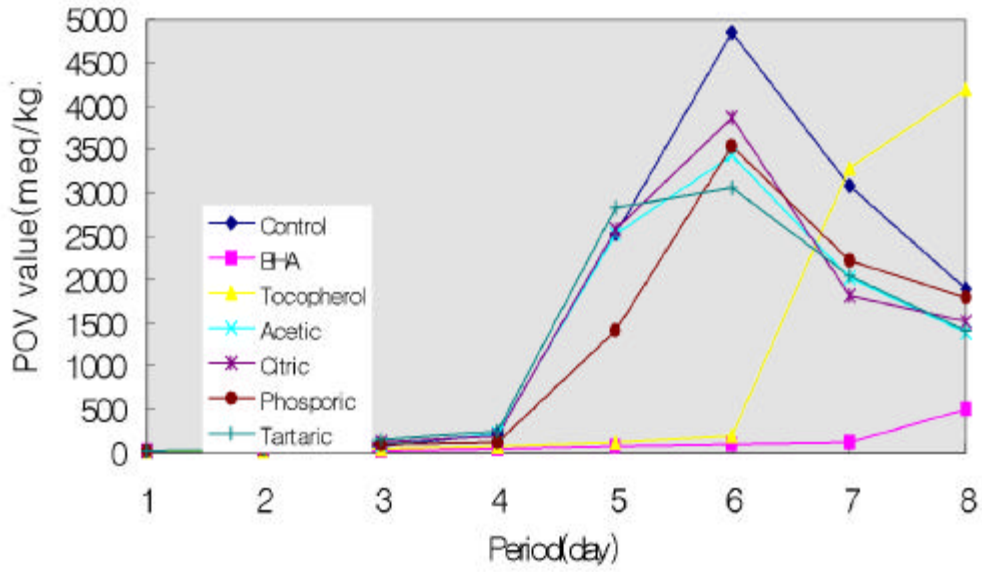


Fig. 2. Changes in peroxide value of garlic extracts as affected by various acids.

Table 3

methanol 50% 가 methanol  
 3.5 26.43  
 28.17%  
 Methanol DPPH (Table  
 3) methanol 가 50% 38.58% 가  
 methanol 100가 26.06% methanol 0, 25, 75%  
 30.9 32.0% 가  
 Methanol (Table 3)  
 pH 1.2 pH 3.0 methanol

pH

. Methanol pH 1.2 methanol  
100% methanol 25%, 75%가 가  
. pH 3.0 methanol 100% 가  
methanol 가 가 .  
thiosulfinate allicin  
(di allyl thiosulfinate) ,  
가  
. , allicin ,  
allinase  
alliin sulfides diallyl mono-,  
di-, oligosulfides; vinyl dithiols; ajoenes (5).  
thiosulfates , methyl allyl, 1-propenyl  
allyl, allyl 1-propenyl, 1-propenyl methyl, dimethyl thiosulfinate  
. (2) Han (2) thiosulfinate cysteine 가  
S-allyl mercaptocysteine thio-  
sulfinate  
allicin alliin allyl  
thiosulfates . Lawson (2)  
allicin thiosulfates 60-80% , Yu (2)  
30 allicin  
diallyl disulfide diallyl trisulfide 75%  
, Han (2) thiosulfates 0.7 allicin  
, Han

**Table 3. Solid yields, electron donating abilities (EDA), nitrite-scavenging effects (NSE) and contents of total thiosulfinates of garlic extracts as affected by various methanol concentrations**

Concentrations (%)	Yields (%)	EDA (%)	NSE (%)		Thiosulfinates (mM)
			pH 1.2	pH 3.0	
MeOH 0	27.44 ± 0.77	30.90 ± 1.15	60.94 ± 3.19	27.13 ± 3.18	4.31 ± 0.16
MeOH 25	28.17 ± 1.64	68.13 ± 1.78	36.92 ± 4.77	32.00 ± 1.46	4.16 ± 0.07
MeOH 50	27.20 ± 0.55	38.58 ± 2.49	66.70 ± 2.53	37.98 ± 1.37	4.26 ± 0.23
MeOH 75	26.43 ± 2.16	30.96 ± 2.10	67.47 ± 4.26	37.98 ± 3.44	2.32 ± 0.21
MeOH 100	8.35 ± 1.08	26.06 ± 0.21	63.19 ± 1.20	39.15 ± 1.39	1.00 ± 0.4

methanol thiosulfinates (Table 3)  
, methanol 50% 4.1  
6 4.31mM 가 methanol  
thiosulfinates HPLC allicin  
dialk(en)yl thiosulfinates Larry (28,29)  
allicin .

4. Ethanol

가

가

methanol ethanol (0 100 %)

Table 4 .

methanol 가 50% 가

. Ethanol 0%, 25% 0.5 1.7% 50%

ethanol 3.5

ethanol

.

Ethanol DPPH (Table 4)

methanol ethanol 가 50% 가

, 100% ethanol 1.5

,

(Table 4) ethanol 100%

methanol

pH1.2 pH3.0 2

pH 1.2 0 75%

ethanol 50% . pH 3.0 75% ethanol 가

가 가 .

Ethanol thiosulfates (Table 4) 50%

ethanol 0%, 25%, 50% 4.3 4.4nM

가 ethanol 가 thiosulfates

methanol ethanol

. Allicin



**Table 4. Solid yields, electron donating abilities (EDA), nitrite-scavenging effects (NSE) and contents of total thiosulfinates of garlic extracts as affected by various ethanol concentrations**

Concentrations (%)	Yields (%)	EDA (%)	NSE(%)		Thiosulfinates (mM)
			pH 1.2	pH 3.0	
EtOH 0	27.44 ± 0.77	30.90 ± 1.15	60.94 ± 3.19	27.13 ± 3.18	4.31 ± 0.16
EtOH 25	26.39 ± 0.70	38.62 ± 0.42	62.47 ± 0.65	28.70 ± 1.39	4.30 ± 0.17
EtOH 50	28.11 ± 1.00	40.81 ± 1.93	64.24 ± 0.27	34.06 ± 2.11	4.40 ± 0.62
EtOH 75	18.13 ± 1.59	31.78 ± 1.79	61.30 ± 3.26	34.62 ± 1.39	2.20 ± 0.56
EtOH 100	8.01 ± 0.30	26.58 ± 0.11	37.94 ± 5.38	26.22 ± 1.53	0.60 ± 0.21

(28,29) ethanol 가  
 , methanol, ethanol  
 50%  
 , ethanol 50%가 가

5. pH

Ethanol ethanol 50%  
 methanol 50% pH3 8

Table 5 . pH가 ethanol 50% methanol 50%

DPPH

(Table 5) pH가

ethanol 50% methanol 50%

가

가

pH

thiosulfinates

(Table 5)

ethanol methanol

ethanol 50%

methanol 50%

pH6

pH 7

allicin

pH가 6.5

Yu (27)

Stoll Seebeck(30)

allinase

pH 4~8

, Mazelis

Crews(4)

allinase

pH가 6.5

Ethanol

methanol

pH

Table 6

, pH

pH 1.2

, pH

가

pH 1.2

pH3.0

pH가

가

**Table 5. Solid yields, electron donating abilities (EDA) and contents of total thiosulfinates of garlic extracted as affected by 50% methanol and 50% ethanol at different pH**

pH	MeOH 50%			EtOH 50%		
	Yields (%)	EDA (%)	Thiosulfinates (mM)	Yields (%)	EDA (%)	Thiosulfinates (mM)
3	21.69 ± 0.69	33.37 ± 2.84	4.41 ± 0.02	22.39 ± 0.66	33.87 ± 3.30	4.24 ± 0.07
4	23.21 ± 2.31	33.11 ± 1.76	4.44 ± 0.01	23.60 ± 0.58	34.78 ± 2.26	4.17 ± 0.16
5	24.90 ± 1.58	35.84 ± 2.13	4.51 ± 0.04	23.97 ± 0.68	35.62 ± 2.85	4.33 ± 0.14
6	27.55 ± 0.52	37.48 ± 0.59	4.69 ± 0.00	24.78 ± 0.05	36.10 ± 1.87	4.65 ± 0.09
7	27.39 ± 1.76	38.31 ± 0.61	4.68 ± 0.03	26.33 ± 1.20	38.02 ± 2.44	4.64 ± 0.11
8	28.40 ± 4.08	38.17 ± 1.25	4.66 ± 0.01	25.51 ± 1.07	38.44 ± 3.49	4.48 ± 0.02

6. Citric acid 가

ethanol 50%

methanol 50% citric acid 1 5% 가

Table 7 . Citric acid 1% 가 가

methanol 1/2, ethanol 1/3 citric acid 가 , alcohol methanol ethanol .

**Table 6. Nitrite-scavenging effects of garlic extracts as affected by 50% methanol and 50% ethanol at different pH**

unit : %			
pH of alcohol	PH of buffer	MeOH 50%	EtOH 50%
3	1.2	65.95 ± 0.76	65.14 ± 1.58
	3.0	32.28 ± 2.97	32.00 ± 2.57
4	1.2	65.32 ± 2.73	65.79 ± 1.64
	3.0	33.56 ± 3.70	33.04 ± 3.51
5	1.2	65.47 ± 0.69	67.01 ± 1.42
	3.0	32.28 ± 1.04	35.24 ± 2.38
6	1.2	66.54 ± 2.17	67.18 ± 2.75
	3.0	33.31 ± 0.03	34.37 ± 2.41
7	1.2	66.75 ± 3.56	66.19 ± 1.07
	3.0	33.69 ± 2.62	36.98 ± 2.62
8	1.2	67.10 ± 1.78	68.27 ± 1.51
	3.0	34.05 ± 0.66	37.04 ± 2.47

(Table 7) citric acid 가 가  
 ethanol 50% methanol 50%  
 , 가  
 citric acid 5% 가 가 1.5 ,  
 alcohol methanol ethanol  
 .  
 thiosulfates (Table 7) ethanol 50% methanol 50%  
 citric acid 가 가  
 citric acid 5% 가



**Table 8. Nitrite-scavenging effects of garlic extracts as affected by different concentrations of the citric acid in 50% methanol and 50% ethanol**

**unit : %**

Conc of citric acid(%)	pH	MeOH 50%	EtOH 50%
0	1.2	66.70 ± 0.76	64.24 ± 0.27
	3.0	37.98 ± 2.97	34.06 ± 2.11
1	1.2	64.24 ± 1.22	62.63 ± 3.56
	3.0	32.21 ± 1.83	32.19 ± 1.72
2	1.2	64.18 ± 2.61	63.52 ± 3.98
	3.0	32.35 ± 2.58	34.84 ± 2.00
3	1.2	63.24 ± 2.36	63.07 ± 2.45
	3.0	33.07 ± 1.42	32.04 ± 0.66
4	1.2	64.17 ± 2.48	63.34 ± 2.17
	3.0	32.70 ± 1.82	32.12 ± 1.28
5	1.2	62.04 ± 2.13	60.10 ± 1.64
	3.0	32.54 ± 1.06	30.33 ± 2.49

7. NaCl 가

ethanol 50% NaCl 1 5% 가

**Table 9**

. Citric acid NaCl 가 ethanol  
50%

(Table 9) NaCl 가  
citric acid

(Table 9) pH  
1.2 , NaCl  
가 .  
thiosulfates (Table 9)  
NaCl 1% 가 가 , 1% 가  
NaCl 가 가 citric acid NaCl 가  
가 thiosulfates 가 4 5%  
가 가 2% 가  
thiosulfates NaCl 가 .  
ethanol  
ethanol 50% methanol 50% pH (3 8), NaCl, citric  
acid 1 5%  $\text{NaH}_2\text{PO}_4$   $\text{Na}_2\text{HPO}_4$  0.5 2% ( )  
가 , , , thiosulfates  
 ,  
ethanol 50%가 가

**Table 9. Solid yields, electron donating abilities (EDA), nitrite-scavenging effects(NSE) and contents of total thiosulfinates of garlic extracted as affected by different concentration of NaCl in 50% ethanol**

Conc. of NaCl(%)	Yields (%)	EDA (%)	NSE(%)		Thiosulfinates (mM)
			pH 1.2	pH 3.0	
0	28.11 ± 1.00	40.81 ± 1.93	64.24 ± 0.27	34.06 ± 2.11	4.40 ± 0.64
1	27.48 ± 2.05	37.76 ± 1.16	66.90 ± 4.19	31.40 ± 0.84	4.21 ± 0.01
2	24.83 ± 1.44	37.49 ± 1.16	67.68 ± 3.46	30.09 ± 0.67	4.45 ± 0.02
3	23.15 ± 0.63	33.94 ± 0.51	67.68 ± 0.18	34.13 ± 0.33	4.35 ± 0.11
4	24.29 ± 4.20	31.76 ± 0.26	67.94 ± 4.91	35.20 ± 0.50	4.52 ± 0.03
5	20.55 ± 2.61	32.12 ± 0.25	66.91 ± 2.37	36.51 ± 1.01	4.52 ± 0.02

## 2 가

가 , 가  
 . 40 120 10 30 가 ethanol 50%  
 .  
 가 DPPH Table  
 10 . 가 10 가 가  
 가 , 20 가 60  
 80 1/3  
 가 . 30 가 40 가 20%  
 80 20 가 1/3 8



0 . 가  
 pH (Table 10) pH  
 pH1.2 가 가 , 가

가 thiosulfates (Table 10) 가  
 가 , 가 가 80  
 가 thiosulfates  
 . HPLC allicin dialk(en)yl  
 thiosulfates Larry (22,23) allicin, diallyl  
 disulfide .

### 3

1.

ethanol 50% 4 ° , 25 ° , 5  
 0 30 5  
 .  
 Table 11 4  
 30  
 72% 25  
 15 15 42%  
 . 50 5 50%가 15

**Table 10. Changes in electron donating abilities (EDA), nitrite-scavenging effects(NSE) and contents of total thiosulfinates of garlic extracts as affected by 50% ethanol with different heating temperature and heating times**

Heating time (min)	Heating temp.( )							
	fresh	40	60	80	100	120		
EDA (%)	10	40.81 ± 1.93	39.19 ± 0.10	35.34 ± 2.81	25.00 ± 1.46	19.34 ± 2.39	18.43 ± 0.73	
	20	40.81 ± 1.93	38.06 ± 1.94	37.10 ± 2.41	13.10 ± 0.11	13.86 ± 0.52	12.49 ± 0.19	
	30	40.81 ± 1.93	32.77 ± 3.86	31.37 ± 3.44	13.94 ± 0.33	14.58 ± 0.32	16.28 ± 1.94	
NSE(%)	10	pH1.2	72.28 ± 2.36	72.14 ± 8.99	71.74 ± 0.32	65.38 ± 1.41	65.28 ± 0.63	68.60 ± 0.58
		pH3.0	39.68 ± 1.98	32.00 ± 1.70	28.67 ± 1.98	27.86 ± 2.41	27.04 ± 1.57	28.57 ± 1.56
	20	pH1.2	72.28 ± 2.36	71.29 ± 2.38	70.59 ± 0.17	64.03 ± 1.57	72.64 ± 1.74	73.62 ± 1.17
		pH3.0	39.68 ± 1.98	33.68 ± 1.63	28.75 ± 1.83	28.63 ± 0.26	29.25 ± 1.85	26.82 ± 0.53
	30	pH1.2	72.28 ± 2.36	70.82 ± 1.93	68.25 ± 0.15	64.98 ± 0.60	66.87 ± 1.14	69.80 ± 1.10
		pH3.0	39.68 ± 1.98	32.19 ± 1.89	29.80 ± 2.01	29.23 ± 1.18	28.81 ± 0.59	30.11 ± 1.53
Thiosulfinates (mM)	10	4.52 ± 0.11	4.55 ± 0.00	4.16 ± 0.31	0.06 ± 0.01	0	0	
	20	4.52 ± 0.11	3.52 ± 0.05	3.42 ± 0.13	0.01 ± 0.00	0	0	
	30	4.52 ± 0.11	3.38 ± 0.07	2.84 ± 0.14	0	0	0	

33%가 가 .  
 가 .  
 (Table 11) 4 pH1.2  
 30 85%  
 , pH3.0 5  
 30 65% . 25  
 pH 1.2 4 25  
 25 30 80% . pH3.0 5

**Table 11. Changes in electron donating abilities (EDA) and nitrite-scavenging effects and of garlic extracts prepared by ethanol 50% extraction as affected by during storage time at different temperature**

Storage temp.	Storage (days)								
	0	5	10	15	20	25	30		
EDA (%)									
4	40.81 ± 1.93	37.76 ± 0.91	34.25 ± 5.94	32.56 ± 1.32	32.89 ± 1.86	32.54 ± 1.82	29.23 ± 2.90		
25	40.81 ± 1.93	34.43 ± 1.68	24.62 ± 0.42	16.49 ± 2.49	16.08 ± 0.91	16.96 ± 3.75	17.87 ± 1.22		
50	40.81 ± 1.93	20.74 ± 3.67	17.22 ± 1.80	13.73 ± 2.19	13.01 ± 3.35	13.90 ± 0.75	13.36 ± 0.41		
NSE (%)	4	pH1.2	72.28 ± 2.36	70.24 ± 2.43	67.14 ± 0.49	67.81 ± 1.20	67.02 ± 0.99	65.26 ± 1.49	63.05 ± 1.52
		pH3.0	39.68 ± 1.98	32.64 ± 0.00	29.74 ± 3.38	28.63 ± 2.67	27.97 ± 2.59	27.29 ± 1.73	26.12 ± 1.15
	25	pH1.2	72.28 ± 2.36	68.76 ± 0.76	66.59 ± 1.03	65.41 ± 0.99	65.41 ± 1.15	65.41 ± 1.97	63.10 ± 1.34
		pH3.0	39.68 ± 1.98	30.67 ± 2.23	27.71 ± 0.81	27.11 ± 1.12	27.04 ± 0.40	26.07 ± 1.29	21.41 ± 2.07
	50	pH1.2	72.28 ± 2.36	65.58 ± 0.71	67.39 ± 1.42	65.80 ± 1.51	64.24 ± 1.33	64.84 ± 0.19	62.98 ± 0.20
		pH3.0	39.68 ± 1.98	29.14 ± 0.40	26.25 ± 1.14	27.79 ± 0.92	27.28 ± 2.60	27.02 ± 0.55	22.60 ± 1.38

가 25 25  
 30 55% 50  
 pH1.2 5  
 pH 3.0 25  
 가 가

thiosulfates

Table 12

thiosulfates가

25 11 , 50 1 thiosulfates가  
 . 4 thiosulfates  
 30 1/2 . Larry (28,29)

allicin

Table 12. Changes in contents of thiosulfinates of garlic extracts as affected by 50% ethanol extraction as affected by during storage time at different temperature

( unit : nM )

Storage temp. ( )	Storage (day)								
	0	1	2	5	10	15	20	25	30
4	4.52±0.11	4.15±0.09	4.13±0.13	3.76±0.32	3.47±0.00	3.39±0.13	2.58±0.13	2.17±0.14	2.03±0.0
25	4.52±0.11	3.29±0.10	3.42±0.31	1.35±0.11	0.09±0.14	ND	ND	ND	ND
50	4.52±0.11	0.49±0	ND	ND	ND	ND	ND	ND	ND

ND : Not Detected

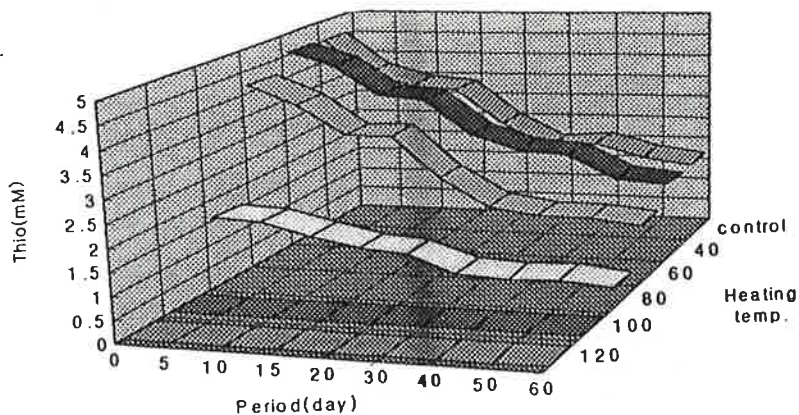
2.

(40 120 ), (10 30min) 가  
4°, 25°, 50 30 5

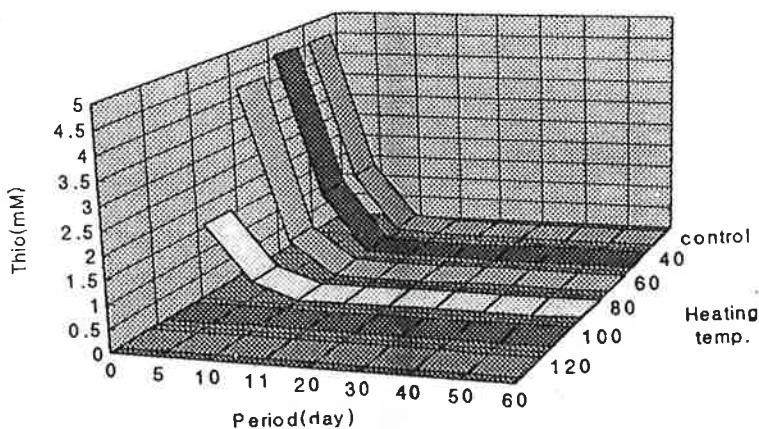
Fig. 3 5 .

10 가  
가  
40.81% 40  
60 12%가 80 32% 100  
50% 4 4

10 min  
4°C 저장



10 min  
25°C 저장



10 min  
50°C 저장

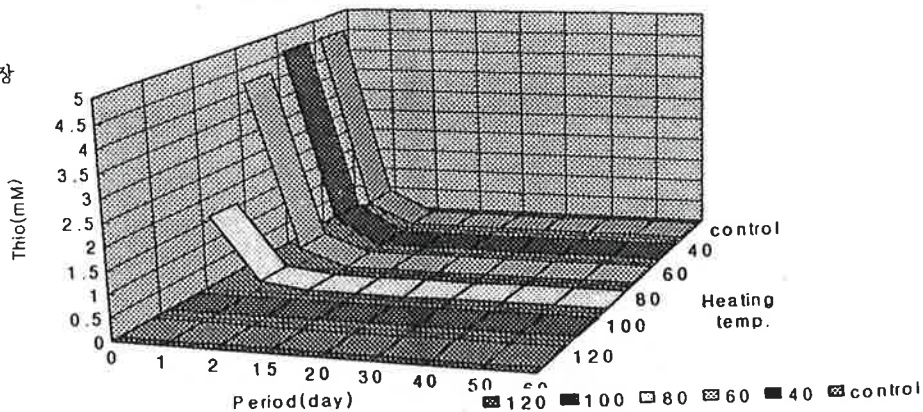


Fig. 5. Changes in contents of thiosulfinates of garlic extracts as affected by 50% ethanol during storage time at different temperature after different heating time for 10 min.

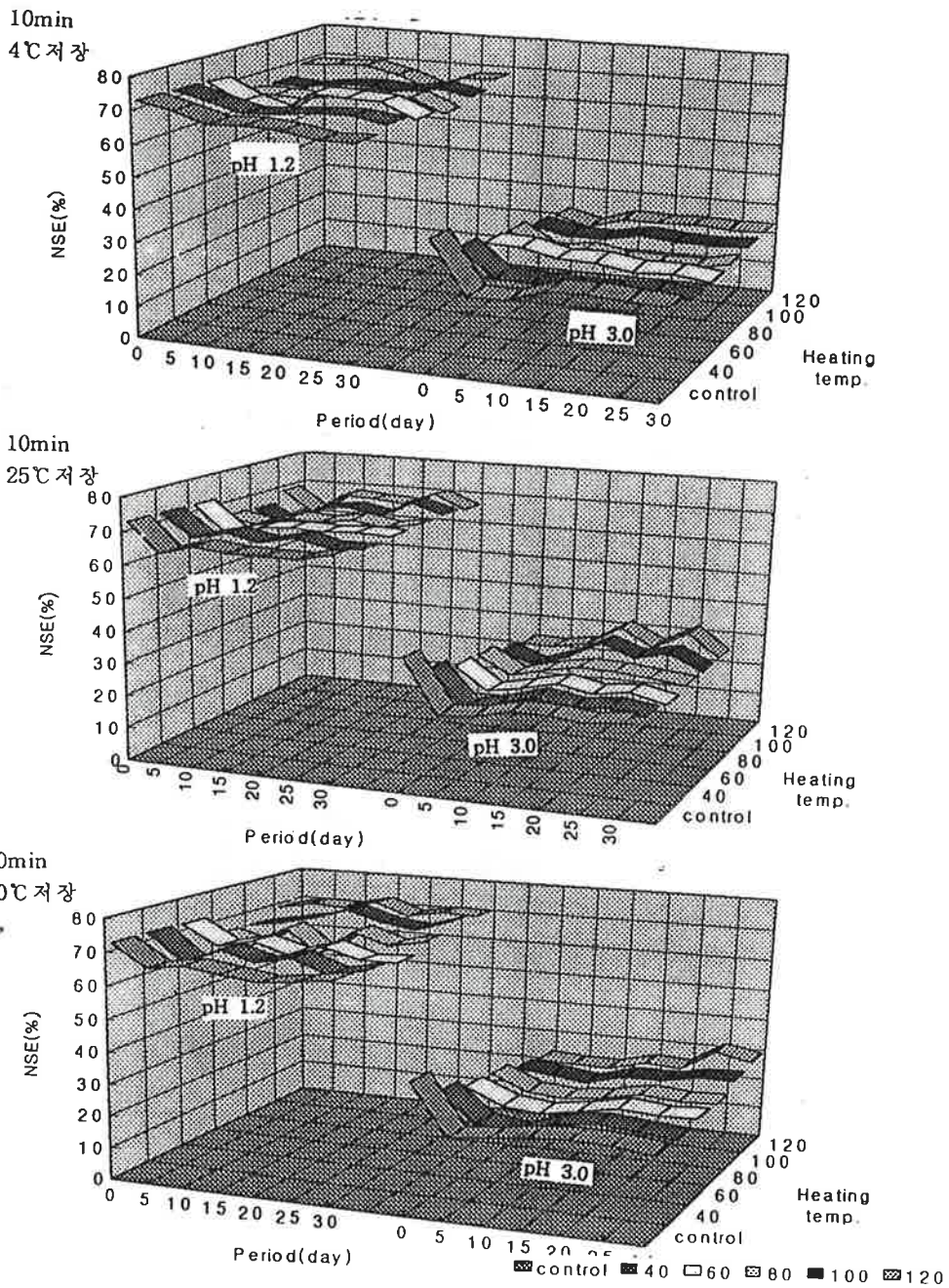
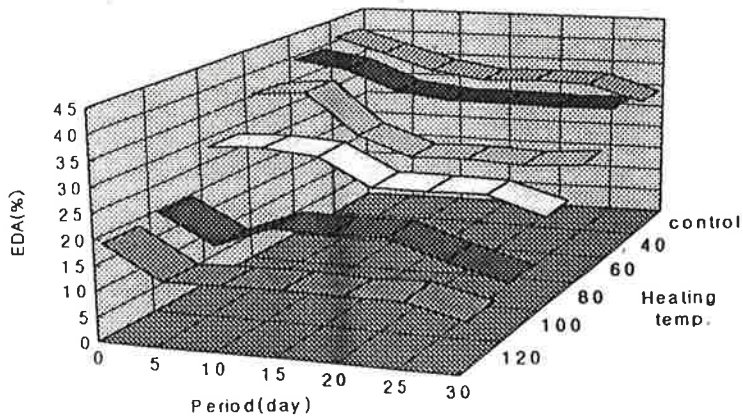
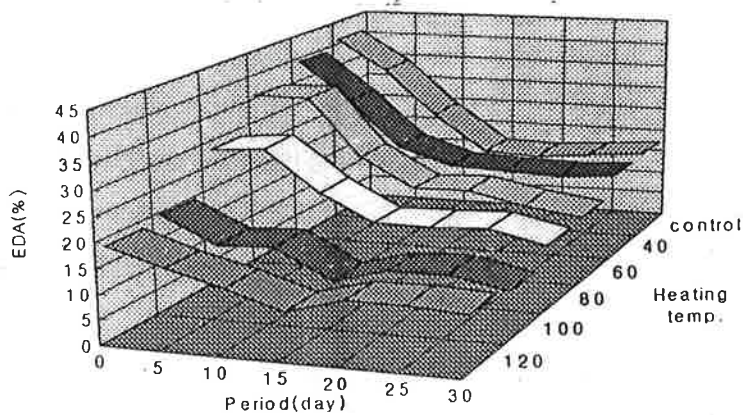


Fig. 4. Changes in nitrite-scavenging effects(NSE) of garlic extracts as affected by 50% ethanol during storage time at different temperature after different heating time for 10min.

10min  
4℃ 저장



10 min  
25℃ 저장



10min  
50℃ 저장

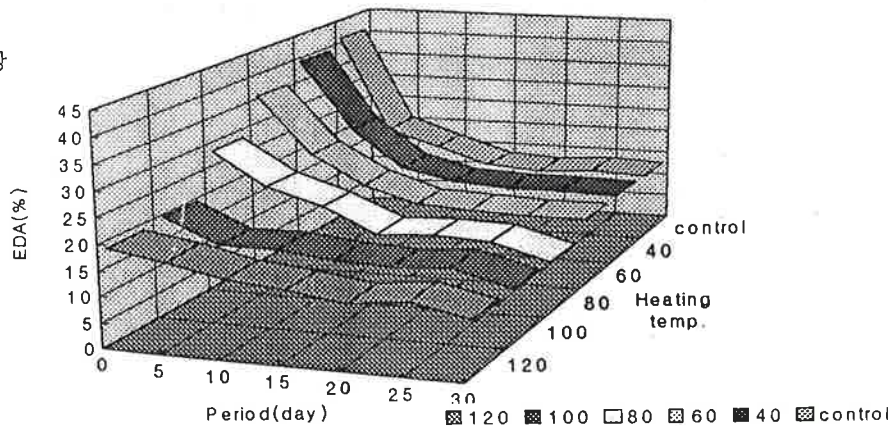


Fig. 3. Changes in electron donating abilities (EDA) of garlic extracts as affected by 50% ethanol during storage time at different temperature after different heating time for 10 min.

0 60 30  
 50% . 80 50%  
 4 25 , 25 10 , 50  
 5 . 100 가 5  
 2/3 . 20  
 10 .  
 가 가 40 60 가 4 10 가  
 2/3가 80 가  
 5 2/3 가 .  
 30 20 10  
 0 120 가 10 20  
 . 30

Maillard

가 Fig. 6 8

. 가  
 , 가

pH1.2 5

0 30 73.3% 65%  
 4 25 가 .

thi osul fi nates Fig. 9 11

. 가 thi osul fi nates

가 가 가  
 .10 가 100 , 30 가 80

thi osul fi nates , 가



thi osul fi nates가 , 25 4 60 11 , 50  
 1 thi osul fi nates

4

4  
 10% ethanol 50% 5, 10, 20% 가 가 , 가  
 10 가, TBA가 .

1. 가 (POV)

가 가  
 Table 13 .  
 가 2 가 가  
 8 가가 10 .  
 가 6 20% 가 가  
 . 10% 50% ethanol 가 20% 가  
 6 가가 가 . 가  
 가가 8 가

2

Gustone Norris(25)

. 가

POV 가

POV

,

가

here

가 , here

(31),

,

가

가(32)

.

가

가

5%

20%

가

가

50% ethanol

20%

가

.

ethanol

.

**Table 13. Changes in peroxide value of ground pork treated with garlics and garlic extracts during 10 days storage at 4 unit: meq/kg**

Treatment 1)	Storage days						
	0	2	4	6	8	10	
uncooked	Control	31 ± 4	42 ± 7	287 ± 63	390 ± 13	725 ± 25	605 ± 32
	G- 5	30 ± 1	40 ± 14	255 ± 20	323 ± 15	600 ± 20	550 ± 21
	G- 10	26 ± 1	30 ± 0	215 ± 7	318 ± 35	600 ± 20	425 ± 33
	G- 20	24 ± 1	30 ± 14	150 ± 28	195 ± 26	575 ± 10	450 ± 50
	G- ex.	27 ± 3	25 ± 7	113 ± 9	172 ± 37	550 ± 12	480 ± 10
cooked	Control	49 ± 2	118 ± 40	340 ± 42	483 ± 39	1048 ± 28	827 ± 30
	G- 5	43 ± 6	78 ± 4	260 ± 12	361 ± 26	941 ± 53	768 ± 42
	G- 10	35 ± 6	55 ± 0	150 ± 10	295 ± 37	924 ± 66	712 ± 45
	G- 20	25 ± 5	39 ± 3	140 ± 18	258 ± 26	848 ± 41	736 ± 15
	G- ex.	19 ± 5	30 ± 7	148 ± 24	276 ± 40	856 ± 49	727 ± 38

1)Control: no garlic ; G-5: 5% garlic added on ground pork weight basis; G-10: 10% garlic added on ground pork weight basis; G-20: 20% garlic added on ground pork weight basis; G-ex.: 10% garlic extracts added on ground pork weight basis.

2. Thiobarbituric acid value (TBA가)

Table 14 . TBA가

10가 . 가

가

가 TBA가

가 .

가 가 TBA가 8

가 .

TBA가 10 가 20%  
 가 6  
 가 TBA 10% 가  
 20% 가 8  
 TBA 가 peroxi de 2  
 ral onaldehydes .

가

가

50%ethanol

가

가

5

가

가

Table 15 .

가 (0.5 2%)

30

SP-1

가

. SP-1

0.5%

가

8

가 , 19

, 1%

가

가

.

(0.5 1.5%)

가

.

1%

(10)

.

**Table 14. Changes in thiobarbituric acid value(TBAvalue) of ground pork treated with garlics and garlic extracts during 10days storage at 4**

unit : OD/10g

Treatment 1)	Storage days						
	0	2	4	6	8	10	
uncooked	Control	0.08 ± 0.00	0.18 ± 0.00	0.30 ± 0.00	0.39 ± 0.00	0.66 ± 0.00	0.81 ± 0.00
	G- 5	0.09 ± 0.00	0.20 ± 0.00	0.28 ± 0.00	0.35 ± 0.00	0.56 ± 0.00	0.77 ± 0.00
	G- 10	0.09 ± 0.00	0.18 ± 0.00	0.25 ± 0.00	0.30 ± 0.00	0.42 ± 0.00	0.702 ± 0.00
	G- 20	0.09 ± 0.00	0.11 ± 0.00	0.14 ± 0.00	0.13 ± 0.00	0.31 ± 0.00	0.51 ± 0.00
	G- ex.	0.05 ± 0.00	0.07 ± 0.00	0.08 ± 0.00	0.08 ± 0.00	0.40 ± 0.00	0.63 ± 0.00
cooked	Control	0.07 ± 0.00	0.20 ± 0.01	0.34 ± 0.00	0.45 ± 0.02	0.75 ± 0.01	0.92 ± 0.00
	G- 5	0.08 ± 0.00	0.12 ± 0.00	0.38 ± 0.00	0.43 ± 0.00	0.69 ± 0.01	0.86 ± 0.00
	G- 10	0.08 ± 0.00	0.09 ± 0.00	0.34 ± 0.01	0.39 ± 0.00	0.44 ± 0.00	0.73 ± 0.00
	G- 20	0.10 ± 0.00	0.10 ± 0.00	0.16 ± 0.00	0.16 ± 0.01	0.34 ± 0.00	0.61 ± 0.00
	G- ex.	0.05 ± 0.00	0.08 ± 0.00	0.09 ± 0.00	0.15 ± 0.00	0.41 ± 0.00	0.75 ± 0.00

1)Control: no garlic ; G-5: 5% garlic added on ground pork weight basis; G-10: 10% garlic added on ground pork weight basis; G-20: 20% garlic added on ground pork weight basis; G-ex.: 10% garlic extracts added on ground pork weight basis.

, 가  
 가 가  
 (2) 가 가 가 가  
 p-hydroxy benzoic acid ester , benzoic acid,  
 sodium benzonate, sorbic acid 가  
 (2).  
 가 가  
 가

**Table 15. The first day when gas formation and film was observed on YMPCSB incubated at 30 during 30 days**

(unit: day)

Conc.	Strains	SS-1	SS-2	SP-1	SP-2	SP-3	
Control	Gas	1	1	1	1	1	
	Film	3	2	2	-	3	
Garlic extract	0.5%	Gas	-	-	7	-	-
		Film	-	-	19	-	-
	1%	Gas	-	-	-	-	-
		Film	-	-	-	-	-
	2%	Gas	-	-	-	-	-
		Film	-	-	-	-	-
Freeze-dry garlic	0.5%	Gas	-	-	-	-	-
		Film	-	-	-	-	-
	1%	Gas	-	-	-	-	-
		Film	-	-	-	-	-
	1.5%	Gas	-	-	-	-	-
		Film	-	-	-	-	-

- : Gas and Film was not observed during 30 days of storage

가

## 4

1. : . p296(1998)
2. : '97 가 . p46(1997)
3. Carson J. F. : Chemistry and biological properties of onions and garlic, *Food review Interactions*, 13(1&2), 71(1987)
4. Mazelis, M and Crews, L. : Purification of the alliin lyase of garlic, *allium sativum L. Biochen. J.*, 108, 725(1968)
5. Bronitz, M.H. and Pascale, J.V. : Flavor components of garlic extract. *J. Agr. Food Chem.*, 19(2), 273(1971)
6. Cavallito, C. J. and Bailey, J. H. : Alliin, the antibacterial principle of *Allium sativum*. I. Isolation, physical properties, and antibacterialaction. *J. An. Chen. Sco.*, 66, 1950(1944)
7. Cavallito, C. J. and Bailey, J. H. : Alliin, the antibacterial principle of *Allium sativum*. . Isolation, physical properties, and antibacterialaction. *J. An. Chen. Sco.*, 66, 1952(1944)
8. Small, L.D., Bailey, J.H. and Cavallito, C.J. : Alkyl thiosulfinates, *J. An. Chen.*, 69, 1710(1947)
9. Bogin, E. and Abrams, M. : The effect of garlic extract on the activity of some enzymes, *Fd. Cosnent. Toxicol.*, 417(1976)
10. , : , 가 ,24(1), 43(1986)
11. 内藤茂三, 山口直彦, 横尾良夫: ネギ類植物からの 抗酸化物の 検 索, *日本食品工業學會誌*, 28, 6(1981)

12. : DNA .  
 , 20(3), 287(1988)
13. , , , : oreoresin  
 . , 22, 846(1990)
14. Sakai, I : Efficiency of garlic ingredients and methods for  
 deodorized effect. *Shokuhim to Kaihatsu*, 27, 19(1992)
15. , , , 野口明德 :  
 allinase 가 .  
 , 28(3), 593(1976)
16. , : .  
 , 10(1), 97(1997)
17. , : .  
 , 17, 506(1985)
18. Kato, H., Lee, I.E., Cheyen, N.V., Kin, S.B. and Hayase, F. :  
 Inhibitory of nitrosamine of formation by nondialyzable  
 melanoidins. *Agric. Biol. Chem.*, 51(5), 1333(1987)
19. , , , , :  
 . 1.  
 , 20(5), 463(1987)
20. Joan Han, J., Lawson, L., Grace Han, G. and Peter Han, P. : A  
 spectrophoto-netric method for quantitative determination of  
 allicin and total garlic thiosulfinates. *Analytical Biochemistry*,  
 225, 157(1995)
21. AOCS 3rd. ed. An. Oil Chem. Soc. Cd 8-53 (1978)



22. Witte, V.C., Krause, G.F., and Bailey, M.E. : A new extraction method for determining 2-thiobarbituric acid values of pork and beef during storage. *J. of Food Sci.*, 35, 582(1970)
23. Fox, J.B. and Ackerman, S.A. : Formation of nitric oxide myoglobin: mechanism of the reaction with various reductants. *J. of Food Sci.*, 33, 364(1987)
24. Tung-Hsi Yu, Chung-May Wu, and Yoh-Cherng Liou : Volatile compounds from Garlic. *J. Agric. Food Chem.*, 37, 725(1989)
25. Gunston, F.D. and Norris, F.A. : Lipids in foods chemistry, biochemistry, technology. *Ferganon Press Inc.*, p58(1983)
26. Lawson, L.D., Wood, S.G. and Hughes, B.G. : *Planta Med.*, 57, 263(1991)
27. Yu, T.H., Wu, C.M., and Liou, Y.C. : Effects of pH adjustment and subsequent heat treatment on the formation of volatile compounds of garlic, *J. of Food Sci.*, 54(3), 632(1989)
28. Larry D.L., Zhen-Yu J.W. and Bronwyn, G.H. : Identification and HPLC quantitation of the sulfides and dialk(en)yl thio sulfinates in commercial garlic products, *Planta Med.*, 58, 263(1992)
29. Larry I.D., and Bronwyn G.H. : Characterization of the formation of allicin and other thiosulfinates from garlic. *Planta Med.*, 58, 263(1992)
30. Stoll, A., and Seebach, E. : Chemical investigation on alliin, the specific principle of garlic. *Advan. Enzynol.*, 11, 377(1951)

31. Pikul, J., Leszczynski, D.E., Bechtel, P.Z. and Kummerow F.A. :  
Effect of frozen storage and cooking on lipid oxidation in  
chicken meat. *J. of Food Sci.*, 49, 838(1984)
32. Igene, J.O., King, J.A., Pearson, A.M. and Fray, J. I. :  
Influence of heme pigments, nitrite and non-heme iron in  
development of warmed-over flavor (WOF) in cooked meat. *J.  
Agric. Food Chem.*, 27, 838(1979)
33. 大西 博 : 醤油醸造と微生物( ). *J. Jpn. Soy Sauce Res. Inst.*,  
5(3), 129(1979)
34. 竹内弘明 : Studies on decrease of p-oxybutyl benzoate in  
laminated can with vinylchloride. 日本醤油技術會 秋季 研究 發表  
會, 16(1966)

3 .

1

가

가

가

가 ( )

'95 가 ( ) (

) 6.77% 가가 ( 가

6.83%) , 6.92%,

7% . [ , , 1997(1995 )]

23,620,070M/T

70% 16,433,980M/T

6,871,206M/T(41.8%), 5,392,082M/T(32.8%), 1,929,610M/T(11.7%)

3,412,357M/T

가 50.8% , 3,773,733M/T

51.4% 가 36.6% 가 11.7%

가

.(Table. 1.1)

1,536,781M/T( 6.5%)

60% 14,116,656M/T

93.6% .(Table. 1.2)

39,273,507M/T 76.7% 30,112,088M/T

. . . 1 가  
 가  
 ,  
 가  
 ( ) 가  
 , 가  
 가  
 .

Table. 1.1 . . .

		(M/T)	(%)
		5,392,082(32.8%)	22.8
		736,471(4.5%)	3.1
		194,671(1.2%)	0.8
		6,871,206(41.8%)	29.1
		1,929,610(11.7%)	8.2
		1,148,992(7.0%)	4.9
		106,102(0.7%)	0.4
		54,846(0.3%)	0.2
		16,433,980(100%)	69.5
		1,721,042(50.4%)	7.3
		914,213(26.8%)	3.9
		777,102(22.8%)	3.3
		3,412,357(100%)	14.5
		1,955,939(51.8%)	8.3
		1,377,481(36.6%)	5.8
		440,313(11.7%)	1.9
		3,773,733(100%)	16.0
		23,620,070	100

: 1994 , , 1995

Table. 1.2 . . . . . ('94)

( : M/D)

					/ (%)
	<b>16,433,980</b>	<b>13,214,023</b>	<b>464,085(30.2%)</b>	<b>29,183,918</b>	<b>45.3</b>
	<b>3,412,357</b>	<b>747,427</b>	<b>1,061,424(69.1%)</b>	<b>3,098,360</b>	<b>24.1</b>
	<b>3,773,733</b>	<b>155,206</b>	<b>11,272(0.7%)</b>	<b>3,917,667</b>	<b>4.0</b>
	<b>23,620,070</b>	<b>14,116,656</b>	<b>1,536,781(100%)</b>	<b>36,199,945</b>	<b>39.0</b>

**2**

# 1

1.

11

, , ( 2.1

1, 2, 3 ) 가 . 1 가

가

Table. 2.1

Table. 2.1

1.	가
2.	가
3.	, , 가
4.	
5.	
6.	,
7.	
8.	
9.	가
10.	
11.	( , , , , , )

2.

1994

가

'96 1

(Fig. 2. 1)

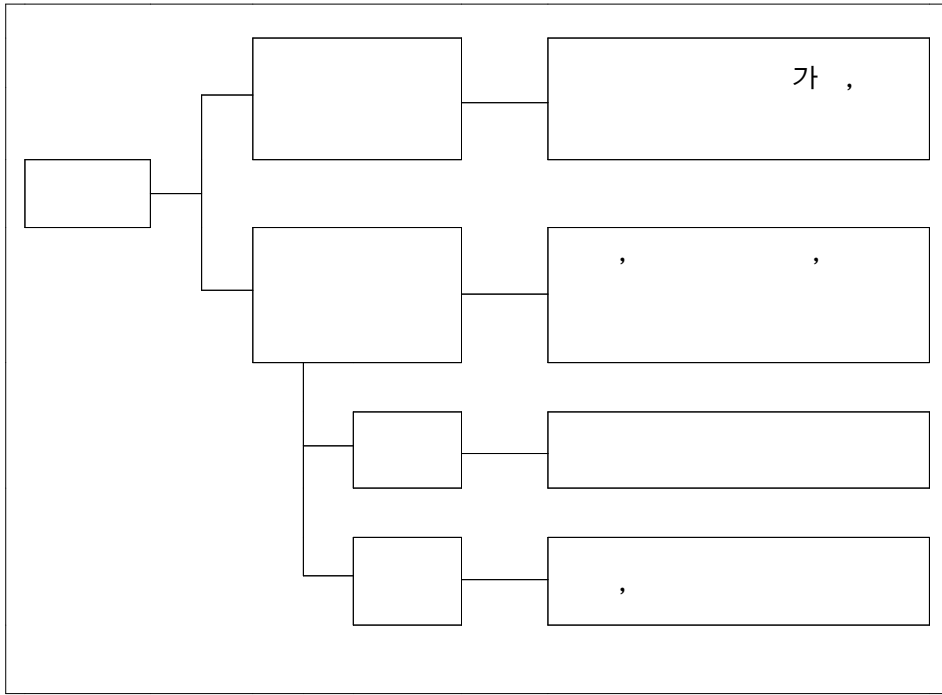
120

가

Fig. 2. 1

(1996. 1.)





3.

가 .  
 .  
 , 가 ,  
 .  
 : 1994 , ,  
 가 ,  
 .  
 :

가

- -

1.

가. (kg, ton, kl)

. (kg, ton, kl)

: / ,

: "

. (kg, ton, kl)

. (kg, ton, kl)

. 가

2. (%)

가. 가 \_\_\_\_\_(%)

. \_\_\_\_\_(%)

. \_\_\_\_\_ (%)

. \_\_\_\_\_ (%)

. \_\_\_\_\_ (%)

. (       ) \_\_\_\_\_ (%)

3.

### 3

#### 1

##### 1.

11

. (Table. 3. 1)

28. 6% 19, 470, 346M/I 1996 , ,  
가 , .

가

3, 306M/I .

, , , 가

, , .

가 50 .

가 87. 4% . Table. 3. 1

가 가

Table. 3. 1 가 Table. 3. 2

. 7. 2%

Table. 3.1

('96)

	( )	(%)	(M/T)	(%)	/ (MI/ )
가	901	7.3	2,775,258	4.1	3,080.2
가	573	4.6	7,814,883	11.5	13,638.5
, , 가	1,539	12.4	5,050,736	7.4	3,281.8
	498	4.0	997,873	1.5	2,003.8
	156	1.3	19,470,346	28.6	124,809.9
,	1,830	14.7	2,607,726	3.8	1,425.0
	789	6.4	1,624,614	2.4	2,059.1
	96	0.8	1,937,433	2.8	20,181.6
가	1,062	8.6	3,543,263	5.2	3,336.4
	297	2.4	6,817,282	10.0	22,953.8
( , , , , , , )	4,671	37.6	15,445,742	22.7	3,306.7
	12,412	100.0	68,085,156	100.0	5,485.4

: 1

( :

3 )

:

(<http://www.foodnet.co.kr>)

Table. 3.2

		( )	( )	( )	( )	( )	( )	가가			
		( )	( )	( )	( )	( )	( )	( )	( )	( )	( )
		96,202	100	2,951,885	100	364,821	100	358,887	100	159,448	100
		87,931	91.4	1,173,461	39.8	74,772	20.5	73,914	20.6	33,681	21.1
		9,353	8.6	1,778,424	60.2	290,049	79.5	284,937	79.4	125,767	78.9
		6,248	100	206,451	100	26,430	100	26,234	100	10,885	100
		5,460	87.4	75,425	36.5	5,239	19.8	5,177	19.7	1,891	17.4
		788	12.6	131,026	63.5	21,191	80.2	21,058	80.3	8,994	82.6

: 2 (2Digits)  
 , ( )  
 500 (5-49 )  
 : 1995 , 1997

2. 가

1 . 1 ( )가 ( )

가 가

2 가 가 .

( , , , ) 34.4%,

3.9%, 16.7%, 23.7% 가 가

.

34,446 M/T(

36,200 M/T. Table. 1.1) 24% 8,481 M/T 가

( 가 , 가 , )

. (Table. 3.3)

Table. 3.3 . 가

( : M/T)

	(A)	가 (B)	B/A (%)	가
	5,059.8	174.0	3.4	, , , ,
	312.5	307.0	98.2	가 ,
	6,052.2	44.0	0.7	, ,
	5,410.6	2,423.2	44.8	, , ,
	489.4	12.8	2.6	,
	247.1	123.3	49.9	, ,
	1,453.4	1,509.0	72.9	, , , ,
	66.1	9.5	14.3	
	63.1	35.7	56.6	
	26.9	23.9	88.8	
	19,181.1	4,662.4	24.3	
	2,673.7	153.7	5.7	, , ,
	1,602.6	88.2	5.5	, , , , , ,
	172.6	3.6	2.1	, , ,
	493.8	0.3	0.1	,
	599.4	1.2	0.2	, , ,
	408.5	1.1	0.3	, , , ,
	307.3	3.1	1.0	, , , , ,
	169.2	0.2	0.1	,
	175.5	4.6	2.6	가 , , , ,
	149.5	6.3	4.2	, , ,
	150.7	3.9	2.6	, , ,
	6,902.8	266.2	3.9	

, 가 [( + )- ] 가 가

Table. 3.3





## 2

1.

가.

가  
1994 ( , 1995) 가  
가  
가 가  
1 , 가  
가 (1 가 )  
1,795.3 M/T, 142.9 M/T 1,938.2 M/T  
92.6% . Table. 3.4

1994  
23,620 M/T/ 34,446 M/T , 24%  
8,481 M/T 가 ( 가 , 가 ,  
) , 가 8.5% 719.3 M/T  
가  
( , 1996)  
33.3%, 25.1%,  
25.0%, 15.9%, 14.6%  
395.3 M/T, 106.1 M/T, 88.7 M/T,

75.1 M/T . 가  
 3.9% 266.3 M/T 206.9

M/T ( 77.7% 가 ), 22.3%

59.3 M/T

( )

.

, , , ,

가 299.2 M/T

75.1 M/T . 가

3,234.8 M/T 767.6 M/T(23.7%) 가

, 12.1 M/T

가 . 가

가 , 3,098.4 M/T 가

2,486.0 M/T 가 , 395.3 M/T

.

.

'93 1,203

653,923M/T '94 가

Table. 3.5

가 121 2.0%(  
 12,412 1%)

. 가 7,420M/T

가 0.13% 가  
 가 4,261M/I( 57%)  
 가 94  
 . (Table. 3.6)

가

. (Table. 3.7)

Table. 3.4

( )

( : M/I/ )

	가		(%)
	2,948.2	106.1	3.6
	136.1	20.0	14.7
	0.1	1	0.7
	1,509.0	18.1	1.2
	9.5	2.4	25.0
	59.6	0.5	0.9
	266.3	88.7	33.3
	299.2	75.1	25.1
	186.7	8.0	4.3
	580.9	4.1	0.7
	2,486.0	395.3	15.9
	8481.0	719.3	8.5

Table. 3.5

	1993 ( )				1994 ( )			
				/				/
	(M/I/ )	(%)	( )	(M/I/ . )	(M/I/ )	(%)	( )	(M/I/ . )
	653, 923	8.0	1, 203	543.6	7, 420	0.5	128	58.0

: , , 1997

Table. 3.6

('94)

( : M/I/ )

	(M/I/ )	* (%)	(%)	( )	/ (M/I/ . )
	136.69	0.03	1.84	5	27.34
	4, 261.86	1.26	57.44	7	608.84
	916.50	0.50	12.35	94	9.75
	13.90	0.01	0.19	4	3.48
	17.70	0.03	0.24	2	8.85
	1.68	0.04	0.02	2	0.84
	15.87	0.01	0.21	4	3.97
	8.00	0.22	0.11	1	8.00
	25.55	0.27	0.34	7	12.81
	39.43	0.04	0.53	1	60.00
	2, 008.13	11.50	27.06	1	2, 008.13
	7, 419.76		99.99	128	2, 739.2

Table. 3.7

('94)

( : / )

			93						93
,		10	99			8			117
			16			14			29
			92						92
			1						1
가	7		59	2		1			68
	40	4,252	334		0.16	1		2,008	6,635.16
	47	4,262	694	2	0.16	24		2,008	7,035.16

: , , 1996

### 3

#### 1. 가

##### 가. 1 가

1 가 2 가  
1 가 가  
가 , 가 . 2 가  
,  
가 . Table. 3.8 . 1 가  
. 1 가  
23.5% 가 25.3%가  
55.0% 20.3% .  
가  
. 가  
2.4% 가 1  
2 가 가 가 1 가 .

Fig. 3.1 가 가

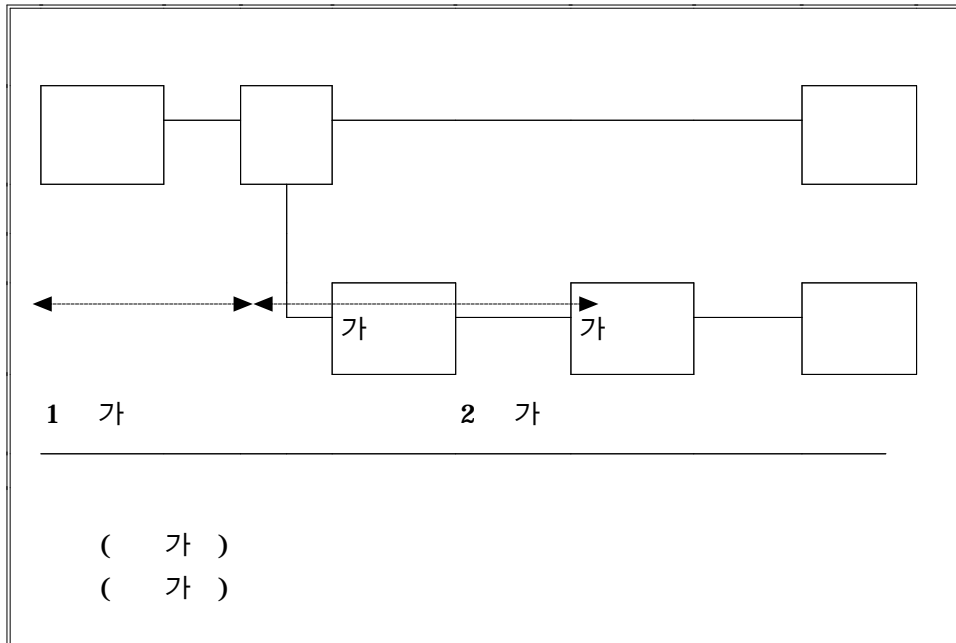




Table. 3.8 1 가

		(M/D)	(M/D)	(%)
가	* *	7, 088, 751	1, 795, 284	25. 3
	* *	6, 571, 122 514, 540 3, 080	1, 511, 358 283, 002 924	
가	*	1, 153, 992	142, 941	12. 4
		169, 280	22, 006	
		705, 886	84, 706	
		278, 826	36, 229	
		8, 242, 743	1, 938, 225	23. 5

(3)

: '95 , . . , 1995  
1994 , , 1995

. 2 가  
 2 가 가 가 가  
 가 ( )  
 2 가 290 68,348,103(M/T)  
 (1996 ).  
 가 , , , ,  
 , , .  
 , , .  
 ,

. Table. 3.9 20 가 2

가  
 . 5,291 M/T( 가 8,481 M/T  
 )  
 2,950 M/T 55.8% .  
 , 가 .  
 가 875,211(M/T)  
 620,898(M/T) ( )가 322,751(M/T)  
 . 가 168%  
 가 . 84.7%,  
 55.1%, 40.0%가 . 2 가  
 1 가 .

Table. 3.9 2 가

		(M/T)	(M/T)	(%)
가	* * * * *	299, 153	75, 211	25. 1
		160, 000	40, 000	
		63, 000	13, 860	
		36, 000	8, 280	
		40, 153	13, 071	
	*	1, 583, 129	875, 211	55. 1
		78, 128	14, 720	
		1, 492, 160	559, 560	
		12, 841	11, 525	
	*	125, 005	50, 002	40. 0
	*	211, 873	22, 579	10. 7
		67, 110		
		67, 110	22, 514	
		2, 523	65	
		75, 130		
	( ) ( ) )	747, 518	620, 896	83. 1
			617, 170	
			3, 726	
	* * * * *	225, 146	190, 774	84. 7
		4, 183	2, 510	
		189, 161	172, 137	
		13, 638	6, 137	
		18, 164	9, 990	
	*	266, 295	88, 676	33. 3
		153, 736	55, 146	
		88, 162	23, 372	
		24, 397	10, 158	

(\*)

Table. 3.9

		(M/T)	(M/T)	(%)
( ) 가	*	192, 113. 4	322, 750. 5	168. 0
		86, 451. 0 105, 662. 4		
가	*	1, 637, 800	703, 500	43. 0
			255, 596 326, 151 121, 753	
		5, 290, 818	2, 949, 600	55. 8

(\*)

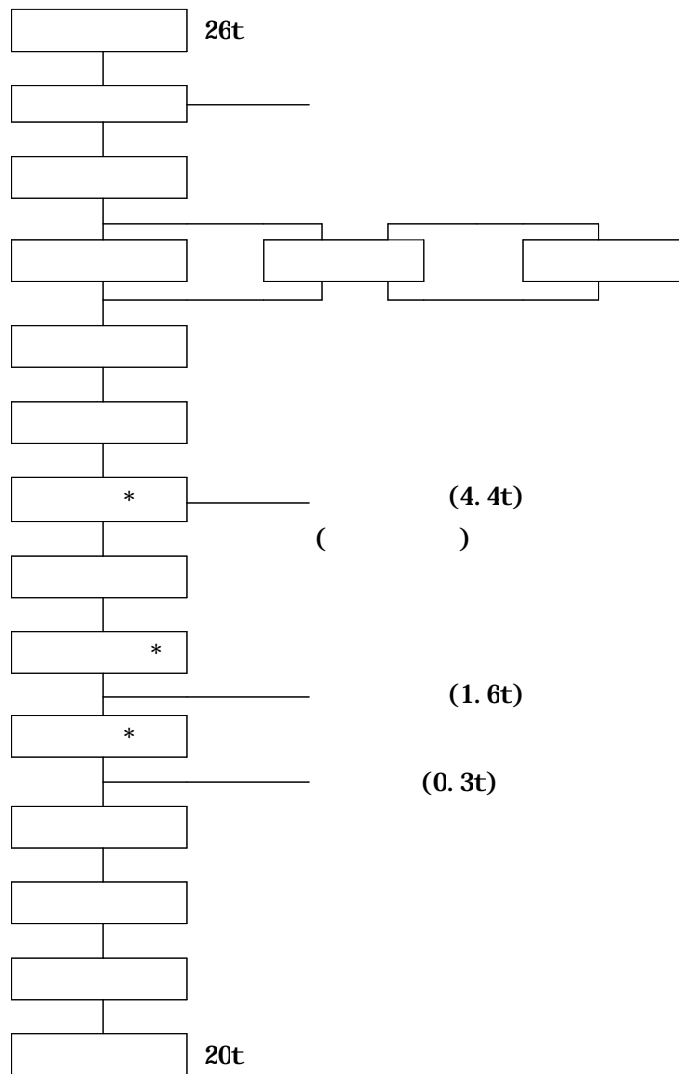
2. 가

가. 1

가

80%, 20% . 가  
(92%) , , 7 , 5 .  
가 ( 8%) , 가 .  
4,560(M/I) ( )  
( 13% )  
17 20%가 6 8%가  
가 1%가 .

Fig. 3.2



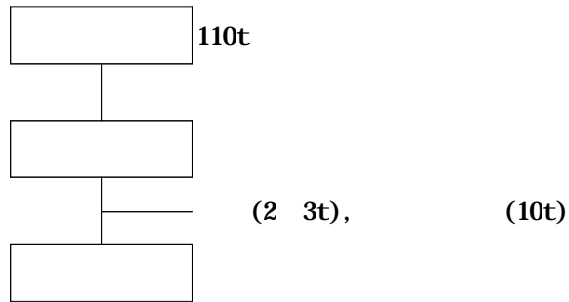
(\*)

1 가

가

10%

Fig. 3.3



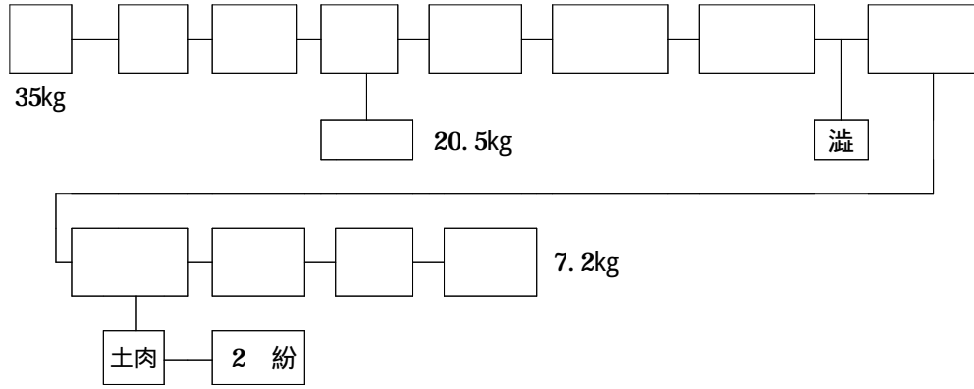
2 가

1) 가

(CCH100) 가

( )

Fig. 3.4 ( )



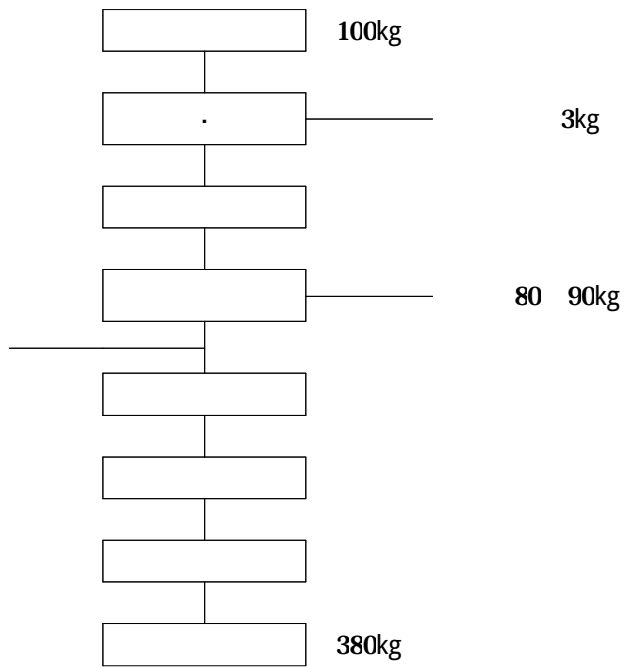
， ，  
 40%가  
 ，  
 18%가

2)

가  
 Fig. 3.5



Fig. 3.5



100kg

380kg

가

( )

가 ,

3) 가

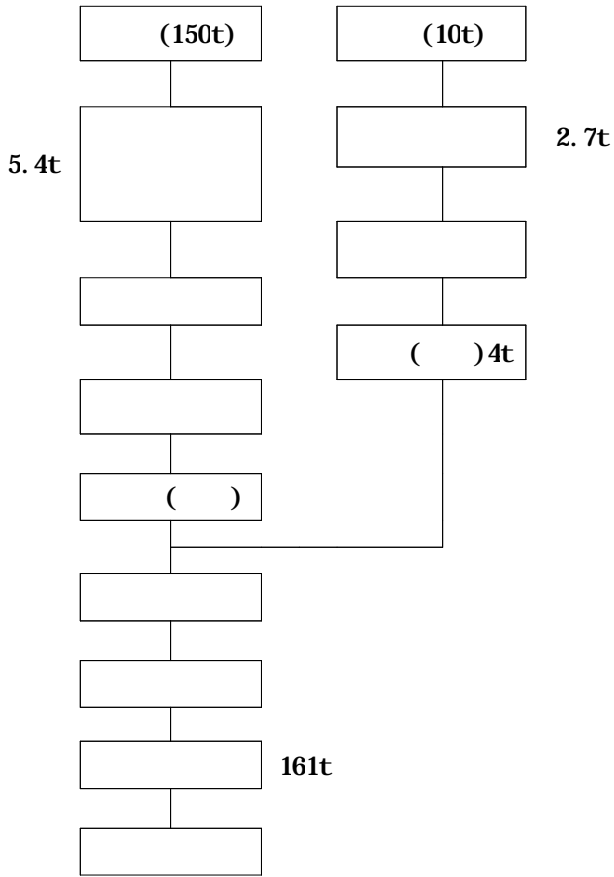
(70 90%),

가

가

가 ( , ) , Fig 3.5 .

Fig. 3.6



( ) ,

가

. 가

가

, 가 2 3가

4) 가

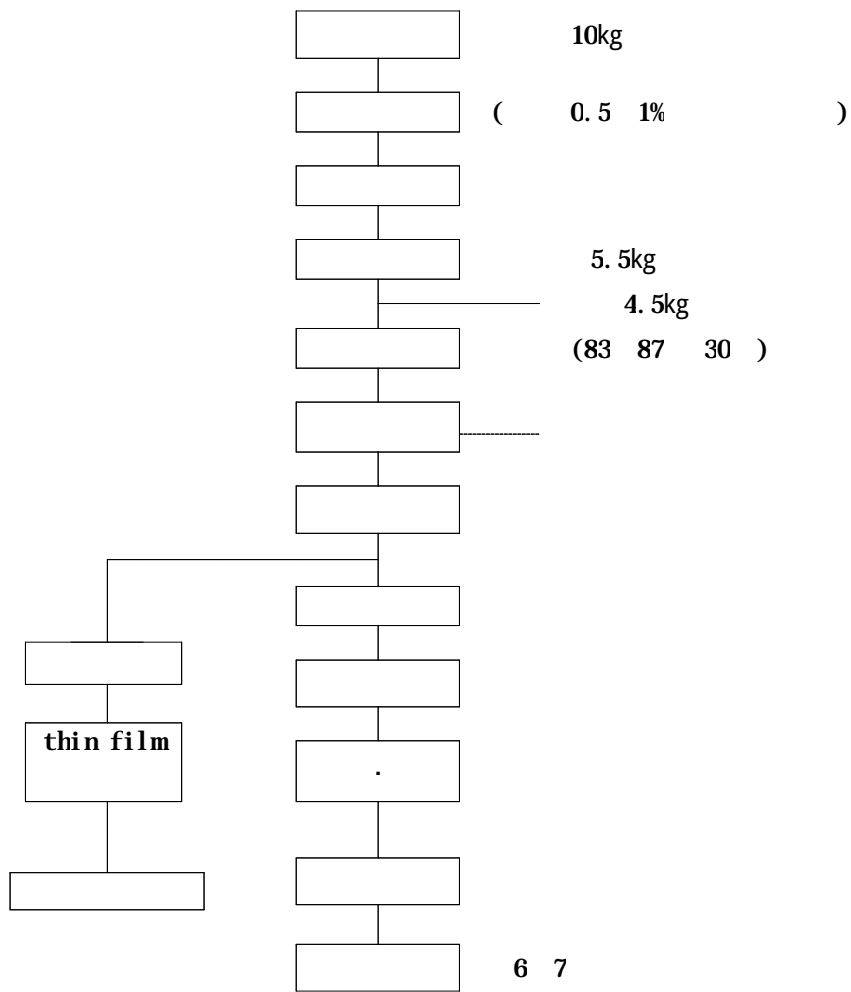
80 90%

가

가

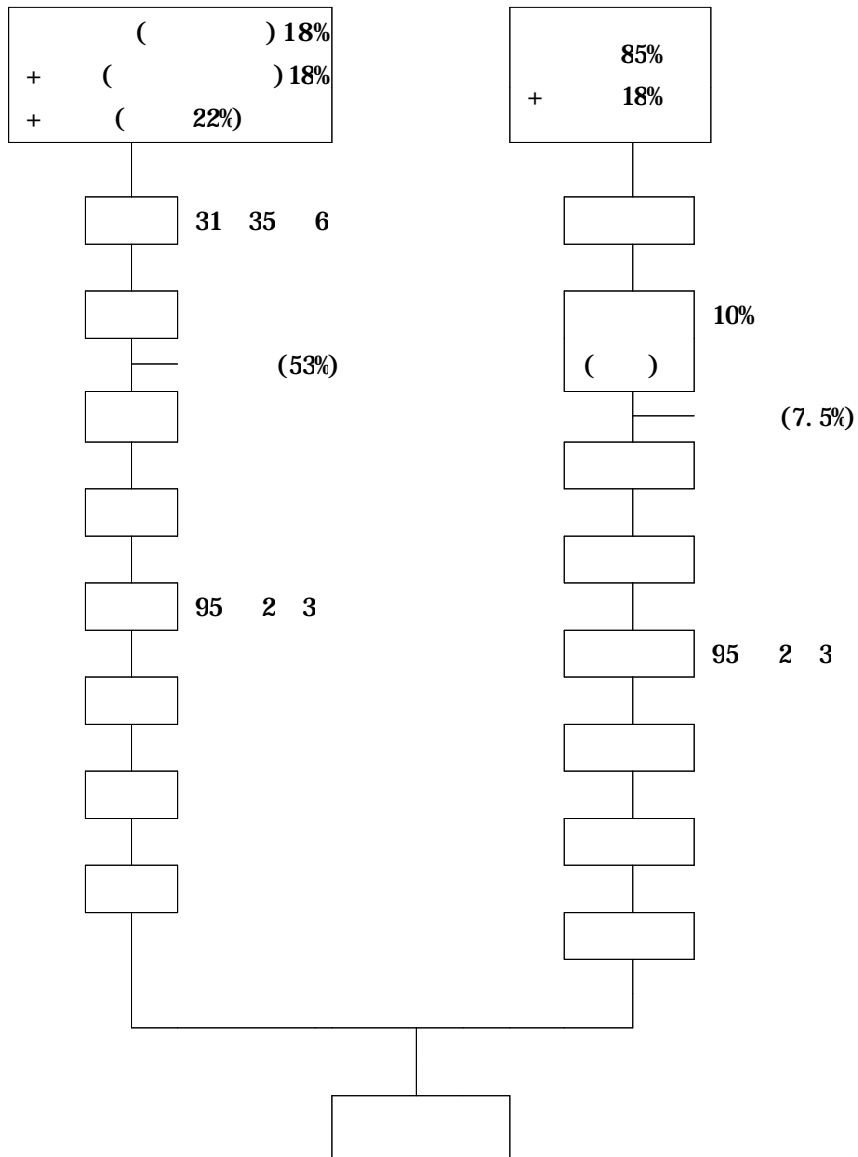
, 가

Fig. 3.7 ( )



가  
가  
5) 가  
( ) ( )  
가 , 가  
가 , 가

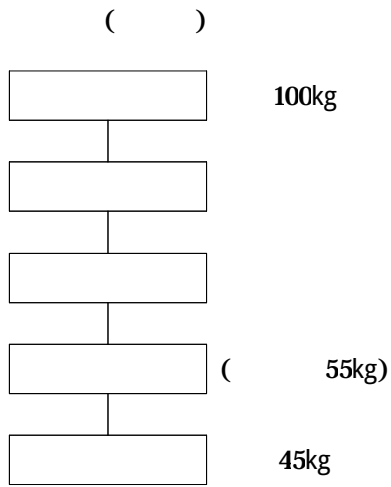
Fig. 3.8



6) 가

, , , .  
, ( , salad oil,  
) 가 (butter, margrin, shortening )  
가 . ,  
.

Fig. 3.9

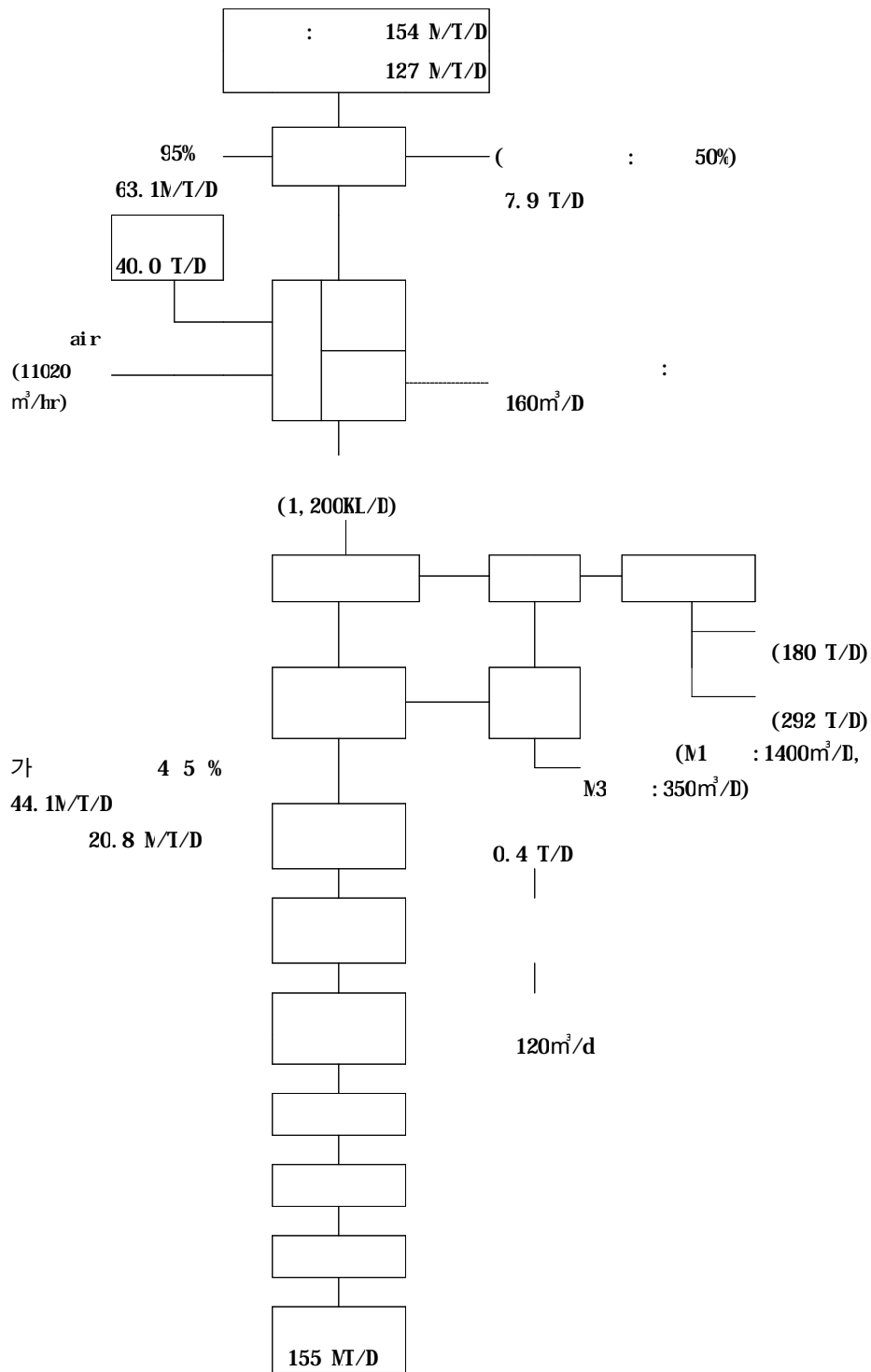


7) ( )가

, 가 ,  
, , , (MSG),  
,

Fig. 3.10

Fig. 3.10

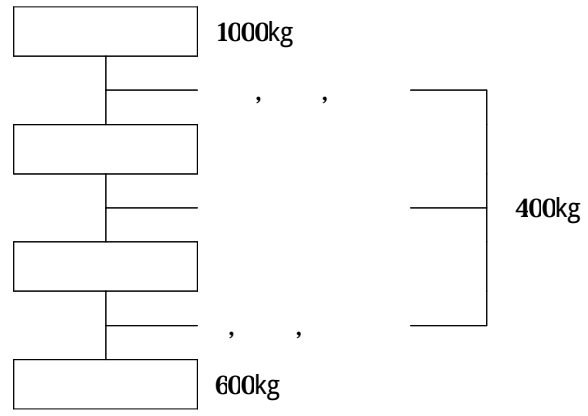


.            2 가  
           가                            가 ,    가 ,    가  
           가                            (    ,    ,    ,    ),    ,  
           ,            ,            ,            ,    가    ,                            .  
 가

Fig. 3.9

가                            30 50%

Fig. 3.11 가





# 4

1.

## 1 가

,  
(15 20% ) 가 ,  
, 70 95%  
가 , , ,  
. 가 1 가  
, 가  
가

## 2 가

60 90%가  
. 가 ,  
가 ( ) 70 85%  
. , 가 .  
가  
. ( , , ) ,  
( ). Table. 3.11 가

Table. 3.11 가

		(%)	(100g )
1 가	가	15 20	11.7g, 79.6g, 6.8g , 110.4ng( )
	가	70 90	80.3g, 17.5g, 1.5g , 110.4ng( )
	가	70 95	72.4g, 20.7g, 4.6g , 231.1ng( )
2 가	가	70 85	86.4g, 10.3g, 1.4g , 19.0ng( )
		85 90	73.6g, 23.0g, 1.1g , 595.2ng( )
		80 90	9.2g, 41.3g, 22.6g , 638.6ng( )
		80 85	9.2g, 41.3g, 22.6g , 638.6ng( )
		60 70	14.8g, 68.4g, 10.3g , 411.5ng( )
		15 20	3.9g, 39.5g, 20.2g , 817.2ng( )
		60 90	90.3g, 6.1g, 2.0g , 139.8ng( )

2.

가  
 ,  
 .  
 가 ( ), ( )  
 ), ( , ) 가  
 .  
 가  
 ( , ) ( )

Table. 3.11 가

가  
 .  
 1 가 ( )  
 , , 가 , ,  
 , , , , 가 .  
 2 가 가 ,  
 , , , , , ,  
 , 가  
 가 . , 가  
 ,  
 , 가  
 , , 가

Table. 3.11 가

				가
1	가			, ( ): ,
	가 가	,	, ,	bone meal, ( ), , ( ) feather meal
2	가			, ( ) ( : pH ) fish meal, (Ca ), ( ), Chitin , Chito : , , ,
		( ):		, ( ): , , lint( , ) furfural
	가			( ), , gluteen, ( )
	가			, ( ), , ( ): , , ( , ), , ( , ), ,
				, , ( , 가 ), ,
				, , , , , pectine , ( ), , ,
가			, ( ) , ,	

: , 1993

## 4

1. 1994 , , 1995
2. '94 가 , , 1995. 7
3. , ' , 1995. 9
4. , , , 1997
5. , , 1997
6. , , , 가 , , 1996
7. 1995 , , 1996
8. 1995 , , 1997. 4
9. , , 1993. 12
10. <http://www.nfrda.re.kr>, 「 」
11. <http://www.suhyup.co.kr>, 「 」
12. <http://www.nori.go.kr>,
13. <http://www.afnc.co.kr>, 「 」
14. <http://www.agr.or.kr>, 「 」
15. <http://www.nlcf.co.kr>, 「 」
16. <http://www.kfri.re.kr>,
17. <http://www.foodnet.re.kr>, 「 」
18. <http://www.rda.go.kr>, 「 가 」
19. <http://www.naf.go.kr>, 「 」
20. <http://www.affis.or.kr>,
21. <http://www.rdc.or.kr>, ,
22. <http://provin.kyonggi.kr>, ,
23. <http://www.aginfo.snu.ac.kr>, 「 가 가 」

4 .

1

가 가

가 .

550 ,

가

10 ( ,

1992) . 1,500 4,000

가

“probiotic

material”

*bifidobacterium*

(Hoover, 1993a : Hoover, 1993b).

sucrose, raffinose,

stachyose ,

raffinose stachyose

가

70% 가  
 ( , 1995). 70% 가  
 , 가  
 .

### Isoflavone

(Barnes, 1995), (Shi no  
 et al., 1988), (Fotsis et al., 1995), (Naim  
 et al., 1976), aldehyde dehydrogenase (Keung et al., 1993)  
 . isoflavone  
 dai dzei n geni stei n . isoflavone ,  
 , isoflavone 가 ,  
 isoflavone

fraction , glyci ni n(11S) - conglyci ni n(7S)  
 . 2S, 7S, 11S, 15S  
 . 3

( 1996).  
 가 (1996), (1992)  
 Wang (1989) ,

, 가

peptide , protease  
 가 가 ,  
 ( , 1995).  
 peptides  
 ( , 1995),  
 ( , 1996), ( , 1995), ( ,  
 1996) .  
 가

3 ( , 1994).  
 peptide  
 peptides .

, , ,  
 가 .  
 , ,  
 ,



가

(ultrafiltration)

가 10 100psi

NaCl 95-99%

200-100psi

(Nanofiltration, NF)

1

peptide

, Isoflavone, peptide

## 2

### 1

1.

가.

10

2

Table 1

Table 1. Area and scales of soybean-curd processing companies

Companies	Area	Scale(output per year)
JA	Kyungi - do, Guri	18,750 ton
KD	Seoul, Kangdong	2,592 ton
PC	Kyungi - do, Pucheon	365 ton
DY	Kyungi - do, Shi heung	4,562 ton
VI	Kyungi - do, Shi heung	1,800 ton
DS	Seoul, Sungdong	2,203 ton
HY	Seoul, Sungdong	1,478 ton
YH	Seoul, Youngdeungpo	3,500 ton
KA	Seoul, Kwanak	4,000 ton
PM	Kangwon - do, Chuncheon	19,000 ton

2.

가.

(Biochemical Oxygen Demand

: BOD) 5-Day BOD test

( , 1988;

, 1992a). 가

aerator , phosphate ( pH 7.2, KH<sub>2</sub>PO<sub>4</sub> 48.5g,

K<sub>2</sub>HPO<sub>4</sub> 21.75g, Na<sub>2</sub>HPO<sub>4</sub>•7H<sub>2</sub>O 33.04g, NH<sub>4</sub>Cl 1.7g 1

), 2.25% MgSO<sub>4</sub>•7H<sub>2</sub>O , 2.75% CaCl<sub>2</sub> , 0.025%

FeCl<sub>3</sub>•6H<sub>2</sub>O 1 1Mℓ 가 .

BOD (300Mℓ )

15

20

5

(Dissolved Oxygen Meter: YSI model 58)

(Chemical Oxygen Demand : COD)

( , 1988; , 1992a).

7.5Mℓ digestion vessel 2.5Mℓ, digestion (103

2 K<sub>2</sub>Cr<sub>2</sub>O<sub>7</sub> 10.216g 167Mℓ, HgSO<sub>4</sub> 33.3g

500Mℓ 1 ) 1.5Mℓ, -

( 1kg 5.5g 1 2 )

3.5Mℓ , 150 block digester 2

, 가 COD meter(DR/2000, HACH)  
 600nm COD .  
 . pH , , , Brix  
 pH meter( , model DP-215M, digital pH/ion meter)  
 pH .  
 (Physica, standard measuring drive system SM-LM, Z4  
 Din) .  
 $20.5 \pm 0.5$  , (shear rate) 40.3 4030 1/s  
 (shear stress)  
 .  
 (Conductivity meter: YSI model 35)  
 .  
 (Abbe, Ermacat A-302) brix  
 .  
 .  
 (Minolta Spectrophotometer CM-3500d)  
 Hunter scale (L), (a), (b)  
 .  
 3.  
 . dry-ice acetone

-80 (deep-freezer) 24  
 (freeze-dryer: , FD 5508) 24 .  
 . microkj el dahl , phenol -  
 sulfuric acid .  
 4.  
 가.  
 (HPLC)  
 . , 10Mℓ 10% lead acetate 2Mℓ  
 가 12,000 × g 15  
 . Sep-Pak C18 cartridge(Waters) 10  
 μℓ (HPLC, Waters model 200 series)  
 . fructose, sucrose, raffinose, stachyose Sigma  
 (Luis, MO) , HPLC Table 2 .

Table 2. Conditions of HPLC for oligosaccharides analysis of soybean soaking water and soybean-curd whey

Instrument	Waters model 200 series
Column	Waters carbohydrate analysis column (3.9 × 300mm, Waters)
Guard column	μ-bondapak insert C18
Mobile phase	75% acetonitrile in water
Flow rate	1.0Mℓ/min
Temperature	40.0
Detector	RI detector (Differential refractometer R401)
Injection volume	10μℓ

isoflavone                      Table 3

(HPLC, Hewlett Packard 1090 series II)

0.35g                      1N HCl 3Mℓ                      가                      100                      (HY-1000, Dae  
han sci. co.)                      1                      가                      가                      MeOH 3Mℓ                      가

12                      isoflavone                      0.5 μ m syringe  
filter(Millipore Co.)                      20μℓ                      Si gra  
(Si gra chemical co., USA)

Table 3 . Conditions of HPLC for isoflavones analysis of soybean soaking water and soybean-curd whey

Instrument	HP 1090 series II Hewlett Packard Co.
Column	YMC-PACK ODS-QM 303 S-5 (120 , 250 × 4.6 mm I. D., YMC Inc. Wilmington, NC)
Guard column	YMC C18 guard column
Moblie phase	0.1% Acetic acid in Acetonitrile : H <sub>2</sub> O = 35 : 65 (v/v)
Flow rate	1.0Mℓ/min
Detector	UV detector (254nm)
Injection volume	20μℓ

Waters Pi co-Tag                      Table 4

20ng                      6N HCl

15Mℓ                      가                      150                      1                      가                      가

3                      50Mℓ가                      0.45μm micro  
filter                      50μℓ                      Pi co-Tag Workstation  
(H<sub>2</sub>O: MeOH: triethyl amine  
(TEA=2: 2: 1, v/v))                      10μℓ                      ,                      vortex mixer

Pi co-Tag Workstation                      Phenyl isothi ocyanate  
(PITC)                      (H<sub>2</sub>O : MeOH : TEA  
: PITC = 1 : 7 : 1 : 1)                      20μℓ                      vortex mixer                      20



5ml 가 420 .

10ml

(Atomic Absorption Spectrophotometer, GBS model 904)

. calibration 1, 2, 3, 4 ppm

(Junsei Chemical Co., Calcium Standard

Solution) .



2

,

1.

2.

45µm

(Bus-200, SK Industry, Korea)

(MCO : molecular weight cut off) 10,000 spiral- wound

(regenerated cellulose) polysul phone

(Millipore Co.)

25 , 2kgf/cm<sup>2</sup>

가

pH

calcium sequestering

EDTA 0.01M 가

가

(VCR: Volume Concentration Ratio) 2 10

(permeate)

(retentate)

### 3. Nanofiltration

Nanofiltration Helicon RO 4 Cartridges가

Nanonax-50

(Milipore, CDRN 50004)

5

5 /min, 130psi

5

30

3 /min, 200psi

500psi

100ml/min 가

### 3 UF/NF

,

1.

2.

, , (COD, pH, ,  
, Brix)

3. *Bifidobacterium*

가.

(Korean Collection for Type Culture,  
KCTC) from 4 (*Bifidobacterium infantis*,  
*Bifidobacterium longum*, *Salmonella typhimurium*, *Staphylococcus aureus*)

UF/NF (DOWEX 50W)  
0.5% 0.45µm filter 10MØ  
glucose

0.5% glucose 가 BHI broth  
 ( , Bifidobacterium longum, bifidobacterium infantis 0.05%  
 cysteine 가) 가 .

**Bifidobacterium**  
 .  
 0.05% cysteine BHI broth  
 anaerobic jar(DIFCO) 37 , 48 1  
 2 . 0.5Mℓ  
 (10,000 × g)  
 0.5Mℓ vortex suspensi on  
 10Mℓ 0.1% peptone  
 0.1Mℓ  
 BL agar 37 48  
 . 0.9Mℓ cryogenic vial  
 10%(v/v) glycerol 0.1 Mℓ 가 -20  
 .

**Salmonella typhinurium Staphylococcus aureus**  
 .  
 10Mℓ BHI broth 37 24  
 1 BHI 10Mℓ 1Mℓ  
 2 .  
 slope agar 37 24 broth 1 loop  
 0.1% peptone 9.9Mℓ tube

0. 1M petri dishes

37 24

4.

가. ICP

(ICP- AES: ICPS- 1000IV)

60%

60%

panel

Duncan's multiple range test

4 가

1.

가.

1 (Sigma Co., DOWEX 50W),

1 (Sigma Co., DOWEX 1X2-200)

1N NaOH 1: 5(w/v) 가

20 1

1N HCl 1: 5(w/v) 가

, 20 1  
 1N HCl  
 1N NaOH  
 2. Steffen  
 (1995)  
 (CaCl<sub>2</sub> · 2H<sub>2</sub>O) 8-10% 가 , ,  
 100-120% (CaO) 가 5 20  
 . 425 1 cold saccharate  
 , 75-85 가 2 hot saccharate  
 . 2  
 1 , 1 30 , 2  
 saccharate 가  
 . 30 , 1 , 1 30  
 , cake 400Mℓ 가  
 325Mℓ 75Mℓ 가  
 HPLC .

3.

100 400 (Sigma Co. , )  
 0.6g, 0.8g, 1.0g, 1.2g  
 3  
 (Sigma Co. , DOWEX 50W) 10 10,000 × g

10M $\varnothing$

3

(rotary evaporator)

가

10M $\varnothing$

HPLC

5

].

가.

10%

pI

50 , 30

pH

10, 900  $\times$  g

15

pH 7.5

30

pH

(soybean-curd whey protein

; SMP)

pH 4.1, 6.0, 8.0

20

0.0-2270 1/s

2.

가.

Frazen (1976)	Sathe (1981)		
(SWP)	(ISP)	pH	.
0. 1g	10Mℓ	1N HCl	1N NaOH
pH 2, 4, 1, 6, 8, 10, 12		, 10	pH
10, 900 × g	15	(Vi si on Co.)	.
1Mℓ	3Mℓ	4Mℓ	30
540nm		pH 12	
100%	.		

1)

	(SWP)	(ISP)	pH	Pearce
(1978)				.
	0. 2g	50Mℓ	1N NaOH	1N HCl
	pH 2, 4, 1, 6, 8, 10, 12		10	pH
	( ) 2Mℓ	Osterizer Mixer	' Mix '	
node 1	.	0. 1Mℓ	0. 1% SDS	
1: 150	500nm		(Hi tachi	
spectrophotometer)				

$$T = 2.303 A/L$$

T : Turbidity , A : Absorbance at 500nm,  
L : Pathlength of Cuvette(1cm)

2)

30 , 60 0.1Mℓ  
0.1% SDS 15Mℓ vortex mixer 500nm

$$\text{Emulsion stability(\%)} = \frac{\%T \text{ of emulsion after } t \text{ min.}}{\%T \text{ of initial emulsion}} \times 100 \quad (\%T: \text{ turbidity})$$

(t = 30 min, 60 min)

1)

Yasunatsu (1972) (SWP),  
(ISP) 0.2g 50Mℓ  
1N NaOH 1N HCl pH 2, 4.1, 6, 8, 10, 12  
, 10 pH Osterizer mixer  
' Whip ' mode 5 30

2)

30, 60

$$\text{Foam stability(\%)} = \frac{\text{Vol. of foam after } t \text{ min}}{\text{Vol. of initial foam}} \times 100 \quad (t = 30 \text{ min, } 60 \text{ min})$$



3.

가.

Davis Orstein Lamli (Bollag and Edelstein, 1991)

. Vertical slab instrument (Hoffer Scientific Instrumets, SE 600 series) 12.5%

sodium dodecyl sulfate(SDS)-polyacrylamide gel (16cm × 18cm) 5% stacking gel . (SWP),

(ISP) Eppendorf 1:5

vortex mixer . 5

가 650 × g 10 25μℓ

stacking gel well SDS tris-glycine buffer

(pH 8.3) SWP 90V, 15 ISP 200V 5

. 0.6Mℓ 1M Tris-HCl (pH 6.8), 5Mℓ 50% glycerol, 2Mℓ 10% SDS, 0.5Mℓ 2-mercaptoethanol, 1Mℓ 1% bromophenol blue 0.9Mℓ H<sub>2</sub>O .

coomassie blue G-250 0.1% MeOH: glacial acetic acid: water (4.5: 1: 4.5) 가 30

MeOH: glacial acetic acid: water(1: 1: 8) .

nyosin -

galactosidase, phosphorylase, albumin(from bovine), albumin(from egg),

carbonic anhydrase, trypsin inhibitor .

1) ANS

Hettiarachcy (1995) Wagner (1990)

가 0.06%-0.001%

0.01M phosphate buffer(pH7.0)

. 80 $\mu$ l

1-anilino-8-naphthalene sulfonate(ANS: 0.01M phosphate buffer 8.0 mM

) 4Ml 가 .

ANS- 390nm(excitation wavelength),

470nm(emission wavelength) spectrofluorometer(Kontron, SFM25)

ANS 80% full

scale

(ANS 가 )

ANS가 가

sample blank

2) SDS

Kato (1984)

0.07 mM SDS

가 0.1 %

30 . SDS-

(Spectra/Por Membranes, MWC: 12-14,000)

0.02M

phosphate buffer(pH 7.0) 25Ml 48

500 $\mu$ l

5Ml chloroform

25Ml screw-capped test

tube vortex mixer

2.5mM methyleneblue

1.25Ml vortex mixer

2,500rpm(

) 10

(chloroform

SDS-methylene mixture)

655nm

. SDS

SDS

SDS 500µg SDS µg

3) thiol disulfide bond

가) thiol

Beveridge (1974) (SWP),

(ISP) thiol 0.06g

tris-glycine buffer(10.4g tris, 6.9g glycine, 1.2g EDTA/liter, pH8.0) 1M 4.7g guanidine hydrochloride 가 10M

1M urea-guHCl (tris-glycine buffer

5M guHCl가 8M urea) 4M 4mg/M buffer

Ellman's reagent(5,5'-dithiobis-2-nitrobenzoic acid; DNTB) 0.05M

15 412nm

thiol

$\mu\text{M SH/g} = \frac{73.53 A_{412} D}{C}$	<p>A<sub>412</sub>: Absorbance at 412 nm  C: Sample concentration(mg/M)  D: Dilution factor  73.53 = 106 / (1.36 x 10<sup>4</sup>)  1.36 x 10<sup>4</sup>: molar absorptivity</p>
--	---

) Disulfide bond

10M 1M mercaptoethanol 0.05

M, urea-guHCl 4M 1 12%

trichloroacetic acid 10M 가 1

6700 x g 10 (Vision Co.)

2-mercaptoethanol 2 12% trichloroacetic acid 10

Mℓ . 8M urea in  
 tris- glycine 10Mℓ 3Mℓ 3Mℓ 8M urea in tris-glycine  
 , 4mg /Mℓ buffer Ellman's reagent 0.04Mℓ 15  
 412nm thiol

thiol disul fide group thiol  
 thiol thiol thiol  
 2 di sul fide groups .

4)

4.

가.

Miller (1981) Knuckles(1985) pepsin  
 pancreatin .  
 0.2g 4.8Mℓ 0.1N HCl (pH 2) 19Mℓ,  
 pepsin 1Mℓ .  
 (Spectra/Por Membrane MCCO: 12-14, 000) pH 2 HCl 200Mℓ  
 가 37 2

2 0.1 NaHCO<sub>3</sub> 200Mℓ가

1N NaHCO<sub>3</sub> 2.5Mℓ 가, 15  
 pH 7 pancreatin-bile 2.5 Mℓ 가 .  
 37 2 ,

pepsin, pancreatin

.

Lowry

bovine serum albumin

가

가

.

2

가

, 20

.



1g 2ml  
 1.5ml loading 0.4ml/min  
 3 fraction collector(Buchler Co. U.S)  
 3ml 280nm Lowry (Lowry et al.,  
 1951) ACE 가

. Prep-FPLC chromatography

GPC ACE 가  
 prep-FPLC(Model AKTA Pharmacia Co.) Table 5

Table 5. Operating condition of gel permeation chromatography

Column	Superdex 75 HR 10/30
Pressure	0.50MPa
Flow rate	0.5ml/min
Loading volume	100 $\mu$ l(20ng/ml)
Eluate FracSize	1ml
UV	280nm

3. ACE

Angiotensin-I (ACE) (Sigma Co.) 1g  
 (pH 8.3) 10ml 가 5 24 ,  
 (10,000rpm, 30 ) ,  
 hippuryl-His-Leu(Sigma Co.) ACE  
 Cushman (30) ACE ACE 50%

peptide-nitrogen

Gel chromatography

(0.1M sodium borate (pH8.3) containing 300mM NaCl)

hippuryl-His-Leu (25mM) 100μl 가 37 5

preincubation ACE 100μl 가 37

30 1.0N HCl 500μl 가 .

0.1ml 가

가 . ethyl acetate 3.0ml 가 10

, 3000rpm 5 1ml .

100 4 3ml 가 228nm

ACE .

---

% Inhibition

= [ 1 - { (As - Ac) / (Ab - Ac) } ] × 100

Ab =

As =

Ac =

Asc =

---

#### 4. Thin layer chromatography

ACE

가 silica gel plate(silica gel 60, layer thickness 0.22mm) spot . BuOH: Acetic acid: H<sub>2</sub>O(4:1:

1) spot TLC 3 .

12cm TLC 110 5

, UV

ninhydrin(ninhydrin : Acetic acid )



, 50% H<sub>2</sub>SO<sub>4</sub> .

### 5. MALDI

Matrix CHCA ( -cyano-4-hydroxycinnamic acid ) 0.01g H<sub>2</sub>O 500  
μℓ 30% acetonitrile 500μℓ 1 vortex CHCA  
1μℓ 0.3% TFA 4,000rpm, 9  
matrix . component insulin  
(bovine), Thioredoxin (E. Coli), Apomyoglobin (horse)  
0.5pmol/μℓ, 2.75pmol/μℓ, 4.0pmol/μℓ mixture  
. Standard mixture 30% acetonitril in 0.01% TFA 100μℓ  
vortex microcentrifuge tube 24μℓ  
standard 1μℓ 가 . ACE  
10pmole , 5μℓ  
laser plate loading vaccune 5  
. plate voyager Table 6

Table 6. Operating condition of MALDI mass spectrometry

Accelerating Volatage	25000
Grid Voltage	75.0%
Guid wire Voltage	0.05%
Laer	2540 - 2100
Pressure+	3.01e-07

### 3

#### 1

1.

4,000

, 1,500 4,000 , 1,500

Table 7 . 가 PM, JA, DY, KA , YH, KD, DS,  
WI , HY, PC . 50kg 124kg 165kg  
PM>JA, DY>YH>KA, KD>WI>HY>PC> DS

. , , ,  
가 , , ( , 1992)

PC, DY, HY, YH, KA 5

.  
, , 가  
( , 1992) KA>JA,  
WI, DY, HY, YH>PV>DS>PC>KD 가

가

( , 1992).

CaSO<sub>4</sub>

CaSO<sub>4</sub>

MgCl<sub>2</sub>

CaCl<sub>2</sub>

GDL(glucono- -lactone) 가

. CaSO<sub>4</sub>

가

가 GDL MgCl<sub>2</sub>

가 .

Table 7

2%

JA

1.8%, PM

2.1%

JA, KD, DY, PM

50kg 205.9 , KD 144

300 , DY 75 , PM 1,342 .

JA, PC, DY, WI, YH, KA, PM

└

3

3 가 1 , 가 2 가

2.

가.

BOD COD

Table 7. Contents of the questionnaires on soybean-curd processing companies

Company Contents	JA	KD	PC	DY	VI	DS	HY	YH	KA	PM
Company Scale (ton output per year)	18,750	2,592	365	4,562	1,800	2,203	1,478	3,500	4,000	19,000
Yield (kg) of soybean curd (50kg soybean)	158.4	144	134.4	158.4	140	122.4	136	150	144	165
Water to soak	tap water	tap water	tap water	underground water	tap water	underground water	tap water	underground water	tap water	sterilized industrial water
Soaking Time (hr)	8	3	4	8	8	5-6	8	8	10	6-6.5
Water Addition for curd-making	octuple	sextuple	octuple						octuple	
Curd Filtration Method	centrifuge	centrifuge	centrifuge	centrifuge	pressing		centrifuge	centrifuge	centrifuge	drum
Coagulants	CaCl <sub>2</sub> +CaSO <sub>4</sub> +MgCl <sub>2</sub> +GDL	CaCl <sub>2</sub> +CaSO <sub>4</sub> +MgCl <sub>2</sub> +GDL	CaSO <sub>4</sub>	CaCl <sub>2</sub> +CaSO <sub>4</sub> +MgCl <sub>2</sub> +GDL	CaSO <sub>4</sub>	GDL	CaSO <sub>4</sub>	CaSO <sub>4</sub>	CaSO <sub>4</sub>	CaSO <sub>4</sub> +MgCl <sub>2</sub> +GDL
Coagulants Contents	CaSO <sub>4</sub> 12g MgCl <sub>2</sub> 2g GDL 2g/ 1kg soybean	undrilled bean curd (70) 130g/100 soymilk	2% of soybeans (W/W)		2% of soybeans (W/W)			2% of soybeans (w/w)	2% of soybeans (w/w)	2% of soybeans (w/w)
Coagulation Time (min)	20-30	1-5	15-20	12	10		5-10	8	10	15
Treatment of Soybean curd Whey										
Treatment of Soybean Soaking Water										
			soymilk heating after filtration	soymilk heating after filtration			soymilk heating after filtration	soymilk heating after filtration	soymilk heating after filtration	

Table 8

BOD COD 12,920

22,760 ppn 13,000 37,100 ppn BOD COD

164 7,530 ppn 213 9,000 ppn BOD COD

(1992a) BOD가 20,821 ppn COD

19,258 ppn

BOD

60 80 ppn COD 70 90 ppn ( ,

1993), BOD COD

Table 8. BOD and COD of the soybean-curd whey and the soybean soaking water

(unit : ppn)

Companies	soybean curd whey		soybean soaking water	
	BOD*	COD**	BOD*	COD**
JA	19,640	21,300	1,270	1,275
KD	22,760	37,100	7,080	9,000
PC	13,820	14,100	183	511
DY	16,880	17,600	651	750
VI	12,920	13,000	1,290	1,175
DS	13,170	17,200	7,530	7,925
HY	16,950	19,800	7,515	8,800
YH	16,400	19,300	7,125	7,725
KA	18,150	17,200	1,701	1,775
PM	21,080	23,200	164	213
Mean	17,177	19,980	3,451	3,915

\* : Biochemical Oxygen Demand  
 \*\* : Chemical Oxygen Demand

pH, , brix,

pH, , brix,

Table 9

pH가 , 가

pH가 ( , 1987; Powers et al., 1959) . 가 ( , 1992b; , 1992) newtonian . 1.372 2.066 cp 1.125 1.346 cp 가 . 1.55 6.79 mhos 0.25 2.74 mhos . Brix 가 ( , 1992a) brix , Brix 1.2 4.8 brix . 1.335 1.340, 1.331 1.334 .

Table 9. pH, viscosity, conductivity, refractive index, Brix of the soybean-curd whey and the soybean soaking water

Comp-anies	soybean-curd whey					soybean soaking water			
	pH	viscosity (cp)	conductivity (mhos)	refractive index	Brix	pH	viscosity (cp)	conductivity (mhos)	refractive index
JA	5.29	1.585	3.50	1.337	3	4.51	1.224	0.62	1.331
KD	4.62	2.066	6.79	1.340	4.8	5.72	1.234	2.74	1.333
PC	5.73	1.733	4.66	1.337	3	5.40	1.200	0.65	1.334
DY	5.57	1.598	1.55	1.337	3	4.11	1.275	0.63	1.331
VI	5.72	1.629	2.44	1.335	1.2	4.41	1.152	0.46	1.332
DS	5.43	1.780	3.82	1.335	1.2	4.05	1.318	0.25	1.333
HY	4.68	1.372	3.77	1.337	2.8	4.04	1.235	0.51	1.333
YH	4.97	1.582	2.40	1.337	2.8	4.35	1.346	1.67	1.333
KA	5.32	1.575	2.94	1.336	2	4.22	1.241	0.44	1.333
PM	4.53	1.871	3.40	1.337	2.8	4.27	1.125	0.47	1.333
Mean	5.19	1.679	3.53	1.337	2.7	4.51	1.235	0.84	1.333

isoflavone

Table 10

L 34.10 89.97, a -4.50 -0.41, b 10.50 14.63 ,  
 L 66.10 94.86, a -1.42 0.26, b 1.51 14.59 .

Table 10. Hunter's color values of the soybean-curd whey and the soybean soaking water

Con-panies	L	a	b	L	a	b
JA	85.55	-3.28	12.87	91.71	-0.21	3.16
KD	34.10	-1.43	14.40	79.05	-0.84	14.59
PC	89.97	-2.99	10.50	94.86	-0.34	1.51
DY	89.39	-4.50	14.63	78.80	0.08	6.92
VI	85.86	-3.32	13.36	90.13	-0.21	3.45
DS	87.86	-3.06	11.86	66.10	0.26	12.04
HY	68.83	-0.77	13.23	82.72	-1.24	9.84
YH	66.32	-0.41	13.65	74.91	-1.10	13.56
KA	52.06	-0.88	14.46	78.93	0.11	8.41
PM	67.19	-1.35	14.41	94.26	-0.33	1.95
Mean	72.71	-2.20	13.34	83.15	-0.38	7.54

L : lightness, a : redness, b : yellowness

3.

가. ,

Table 11

2.0 3.8%, 0.11 1.01%  
 0.318 0.670%, 0.002 0.123% .

0.3 0.4%, 0.1% Wang(1989)  
 12.1 20.4%, 0.6 15.5%가  
 (1987)  
 12 25%가  
 10  
 KA 0.036%, 8 YH 0.019%, 6 PM 0.017%, 4  
 PC 0.009% ,  
 가 .  
 가, ,  
 Table 11 , 0.98 1.64%,  
 0.011 0.316% .  
 33.3 60.2% , 10.0 53.0% .

**Table 11. Contents of total solid, total protein, total sugars in the soybean-curd whey and the soybean soaking water**

Companies	soybean-curd whey			soybean soaking water		
	total solid (%)	total protein(%)	total sugars (g/ )	total solid (%)	total protein(%)	total sugars (g/ )
JA	2.1	0.350	12.636	0.11	0.008	0.399
KD	3.8	0.670	16.414	1.01	0.123	3.163
PC	2.3	0.355	11.645	0.11	0.009	0.110
DY	2.9	0.505	12.116	0.21	0.014	0.212
WI	2.0	0.318	10.050	0.11	0.002	0.241
DS	2.5	0.510	11.180	0.70	0.004	0.955
HY	2.4	0.380	9.828	0.75	0.014	1.336
YH	2.9	0.435	11.513	0.77	0.019	1.153
KA	3.2	0.387	12.983	0.26	0.036	0.287
PM	3.1	0.450	13.012	0.11	0.017	0.583
Mean	2.7	0.434	12.138	0.41	0.025	0.844



4.

가.

BOD COD 가  
( , 1994)

HPLC Table 12

sucrose: raffinose : stachyose가 5: 1: 4

(Kennedy et al., 1985; Kuo et al., 1988; , 1995)

HY, YH, PM sucrose :

raffinose : stachyose 5. 2: 1: 4. 4

Table 12. Contents of oligosaccharides in the soybean-curd whey

(unit : g/ )

Companies	sucrose	raffinose	stachyose	total
JA	3.829 ± 0.120	0.823 ± 0.062	3.619 ± 0.146	8.271
KD	4.444 ± 0.018	0.965 ± 0.057	4.380 ± 0.021	9.789
PC	3.852 ± 0.055	0.721 ± 0.082	3.774 ± 0.020	8.347
DY	5.052 ± 0.044	0.845 ± 0.023	4.246 ± 0.070	10.143
WI	4.353 ± 0.041	0.769 ± 0.063	3.523 ± 0.091	8.645
DS	4.077 ± 0.084	0.776 ± 0.056	3.427 ± 0.027	8.280
HY	2.788 ± 0.024	0.739 ± 0.045	3.293 ± 0.153	6.820
YH	3.267 ± 0.226	0.752 ± 0.038	3.886 ± 0.113	7.905
KA	5.326 ± 0.071	1.142 ± 0.015	4.922 ± 0.024	11.390
PM	3.741 ± 0.048	0.786 ± 0.023	4.283 ± 0.061	8.810
Mean	4.073 ± 0.765	0.832 ± 0.129	3.935 ± 0.512	8.840 ± 1.290

isoflavone

Table 13      daidzein      genistein      ,      daidzein      2.08  
 2.88ng/g, genistein      0.976      1.437ng/g(      )      .      whole  
 bean      1744μg/g(Vinton      )      isoflavone      가  
 (Wang, Murphy, 1994)      10.3      39.9%가 recovery      .  
 isoflavone      L      (r=0.7826)      .

Table 13. Contents of isoflavones in the soybean-curd whey solids

(unit : ng/g)			
Companies	Daidzein	Genistein	Total
JA	2.880 ± 0.009	1.437 ± 0.002	4.317
KD	2.079 ± 0.026	0.976 ± 0.011	3.054
PC	2.367 ± 0.011	1.115 ± 0.006	3.482
DY	2.782 ± 0.011	1.344 ± 0.006	4.126
WI	2.662 ± 0.015	1.179 ± 0.005	3.842
DS	2.645 ± 0.013	1.413 ± 0.005	4.058
HY	2.611 ± 0.017	1.398 ± 0.005	4.009
YH	2.461 ± 0.010	1.216 ± 0.014	3.677
KA	2.134 ± 0.033	1.029 ± 0.008	3.163
PM	2.433 ± 0.029	1.134 ± 0.016	3.567
Mean	2.505 ± 0.262	1.224 ± 0.166	3.730 ± 0.417

(Wang, Cavins, 1989).

가 가 (Hackler, Stillings, 1967a).

(Wang, Cavins, 1989).

amino acid analyzer

가 0.002 0.12%

가 .

Table 14 .

17 94.333 147.035 ng/g

, 0.198 0.422 % .

56.57 96.38 % 78.58% .

가 HY, PC, DS, WI

. JA, KD ,

GDL 가 GDL

가 (1996) .

, lysine 가 glutamic acid glutamine,

aspartic acid asparagine . Wang (1989)

lysine isoleucine, leucine,

valine, phenylalanine

, cysteine methionine

trypsin inhibitor (Wang,

Cavins, 1989). Wang (1989)

0.33

0.49

Table 14. Contents of amino acids in the soybean-curd whey solids

(unit : ng/g)

	JA	KD	PC	DY	VI	DS	FY	YH	KA	PM
Asx*	7.232	11.258	13.319	12.273	15.106	14.054	17.008	11.732	11.134	11.190
Glx**	11.612	17.212	19.064	18.934	22.009	22.325	25.695	17.463	17.294	18.567
Ser	2.902	3.547	5.153	3.796	5.598	5.192	5.576	5.513	3.805	5.232
Gly	2.675	3.517	4.158	4.408	4.503	4.649	5.219	4.605	3.595	4.502
His	5.254	4.379	6.379	5.128	5.600	5.546	5.689	4.951	3.139	5.065
Arg	5.501	6.490	7.778	7.407	8.211	8.029	8.897	7.126	5.745	7.881
Thr	4.017	5.308	4.605	4.398	6.222	6.128	7.276	4.184	4.887	5.999
Ala	2.716	3.215	3.531	3.522	3.680	3.546	4.637	3.303	2.970	3.663
Pro	2.541	4.423	4.467	4.869	4.975	5.126	5.873	5.306	3.550	4.704
Tyr	7.178	6.965	10.908	8.593	7.611	9.280	7.657	8.505	6.611	7.235
Val	2.312	1.995	2.229	2.371	2.411	2.523	2.125	2.657	2.348	2.399
Met	2.570	3.720	2.821	3.259	3.611	3.571	3.488	2.946	2.979	2.902
Cys	0.456	0.453	1.485	0.548	0.624	0.602	0.700	0.456	0.800	0.760
Ile	2.345	3.302	2.963	5.900	6.987	3.365	6.126	2.680	5.796	5.560
Leu	4.778	4.254	5.833	6.096	6.497	5.787	6.027	4.938	5.480	5.718
Phe	3.884	1.457	4.073	4.763	5.427	4.009	4.272	3.762	5.082	4.511
Lys	26.362	29.582	36.547	34.472	35.844	39.884	30.776	32.996	31.391	29.509
Total	94.333	111.078	135.313	130.737	144.912	143.618	147.035	123.123	116.605	125.399

\* : Asx = Aspartic acid + Asparagine

\*\* : Glx = Glutamic acid + Glutamine

DS

CaSO<sub>4</sub>

CaSO<sub>4</sub>가

가

Table 15

193.5 678.1 ppm

, JA DY

(r=0.7317)

Table 15. Ionic calcium and Total calcium concentrations of the soybean-curd whey

Companies	ionic calcium	total calcium
JA	134.0	641.3
KD	37.3	225.2
PC	120.0	640.5
DY	188.0	678.1
VI	41.3	281.9
DS	69.2	478.9
HY	106.0	553.5
YH	168.0	498.1
KA	93.0	193.5
Mean	106.3 ± 52.4	465.7 ± 187.5

2

1.

가. pH EDTA 가

pH EDTA

Fig. 2

polysulphone

pH가 가

, pH 3.5

가

pH 7.0

pH

Kuo Cheryan(1983)

EDTA 0.01M

가

, calcium

sequestering

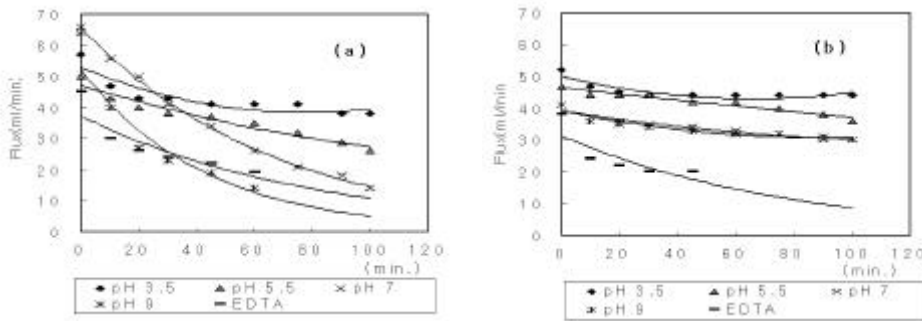


Fig. 2. Effect of membrane type and chemical treatment on flux of soybean curd whey during ultrafiltration.

(a) regenerated cellulose membrane (b) polysulphone membrane

. pH

pH

가

pH

Fig. 3

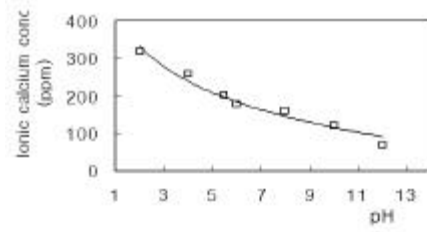


Fig. 3. Changes of ionic calcium concentration in soybean curd whey at

. Kroll (1984)

various pH.  
pH가 가

pH가 가

가

가 pH

가

1)

(COD)

COD

Table 16

COD가 23,900, 23,800ppm

polysulphone

COD

56.58%, 79.25%

COD

COD

(MRV)

가

가

, polysulphone

COD

**Table 16. Effect of membrane type and volume concentration ratio (VCR) on the reduction of COD of soybean curd whey during ultrafiltration**

Fraction	VCR2	Regenerated cellulose (unit: ppm)	Polysulphone (unit: ppm)
SCW(pH 3.5) 1		23,900	23,800
Perneate	2	13,600(27.66) 3	11,800(54.62)
	3	18,700(29.17)	14,100(63.28)
	4	19,350(37.58)	18,400(60.17)
	5	19,350(45.03)	20,200(65.17)
	10	19,450(56.58)	20,000(79.25)
Retentate	2	18,800	26,000
	3	26,400	38,400
	4	31,000	46,200
	5	35,200	58,000
	10	44,800	96,400

1. Soybean curd whey
2. VCR : volume concentration ratio
3. The value in parenthesis is MRV(membrane rejection value) of COD, %.

$$MRV(\%) = [1 - (C_{perneate}/C_{retentate})] \times 100$$

2) , ,

(VCR)

**Table 17**

polysul phone

가

5 가

10

polysul phone

가 가

가

, polysul phone

polysul phone



4

5

가 가

4가

, polysulphone

5

가

10

가 가

polysulphone

5 가

Table 17. Proximate analysis of soybean curd whey, permeate and retentate

Fraction	VCR (%)	Regenerated cellulose				Polysulphone			
		Slid (%)	Ash (%)	Protein (%)	Total sugar (g/l)	Slid (%)	Ash (%)	Protein (%)	Total sugar (g/l)
SCW1 (pH3.5)		2.58	0.64	0.43	20.10	2.94	0.56	0.462	17.05
Permeate	2	1.58	0.42	0.064(86.75)	8.99(27.91)	1.77	0.35	0.062(90.61)	8.55(42.31)
	3	2.05	0.54	0.061(91.21)	12.73(19.02)	2.12	0.51	0.053(95.43)	10.74(43.08)
	4	2.16	0.55	0.032(95.92)	12.59(16.95)	2.24	0.51	0.047(96.18)	12.72(43.21)
	5	1.70	0.51	0.021(98.30)	9.17(32.77)	2.12	0.53	0.037(98.36)	14.01(40.20)
	10	1.85	0.50	0.029(97.72)	9.43(48.22)	2.40	0.50	0.050(98.42)	13.25(61.76)
Retentate	2	2.35	0.48	0.483	12.47	3.25	0.57	0.66	14.82
	3	3.13	0.59	0.694	15.72	4.11	0.43	1.16	18.87
	4	3.51	0.57	0.785	15.16	4.77	0.64	1.23	22.40
	5	3.53	0.61	1.237	13.64	5.12	0.61	2.25	23.43
	10	4.36	0.60	1.270	18.21	8.39	0.60	3.16	34.65

1) SCW : soybean curd whey

2) VCR : volume concentration ratio

3) The value in parenthesis is MRV(membrane rejection value) of protein, %.

4) The value in parenthesis is MRV(membrane rejection value) of total sugar, %.

L( ), a( ), b( ) Hunter

Table 18

L

가 . ,  
 polysul phone 가 .  
 a ,  
 b . b  
 b .  
 1. 10 1. 70cP,  
 0. 96 1. 13cP, 10 2. 61 2. 88cP  
 , Newton .

Table 18. Hunter's color value of soybean curd whey, permeate and retentate

Fraction	VCR2	Regenerated cellulose			Polysul phone		
		L	a	b	L	a	b
SCW1		57. 71	0. 05	12. 38	60. 48	0. 27	12. 76
Perneate	2	96. 69	-1. 53	5. 27	98. 27	-1. 78	3. 54
	3	94. 91	-2. 02	7. 08	98. 60	-2. 40	6. 05
	4	96. 18	-1. 90	7. 46	98. 44	-2. 49	6. 65
	5	94. 74	-1. 61	6. 15	98. 42	-2. 46	5. 66
	10	95. 79	-1. 68	6. 70	98. 32	-2. 68	7. 47
Retentate	2	61. 46	0. 20	11. 15	61. 03	0. 36	12. 84
	3	59. 63	-0. 07	12. 10	48. 80	-0. 21	13. 02
	4	60. 67	-0. 08	12. 55	41. 34	-0. 77	13. 22
	5	53. 32	-0. 22	12. 21	32. 33	-0. 66	12. 91
	10	40. 02	-0. 99	12. 95	19. 80	2. 76	11. 11

L = lightness (0 100), a=green red(-60 +60), b=blue yellow(-60 +60)

1) SCW : soybean curd whey

2) VCR : volume concentration ratio

3)

Table 19 .

Table 20 .

4

가 , 4  
 95.9%  
 4 가 polysul phone  
 가 가 가 가  
 가 , 2  
 90.61%

Table 19. Oligosaccharide contents of soybean curd whey, permeate and retentate

(unit : g/l)

Fraction	VCR2	Regenerated cellulose			Polysul phone		
		sucrose	raffinose	stachyose	sucrose	raffinose	stachyose
SCW1 (pH3.5)		4.67	0.45	3.43	4.95	0.54	3.49
Perneate	2	1.93	0.22	1.23	2.80	0.26	1.43
	3	3.80	0.29	2.08	3.03	0.26	1.96
	4	4.21	0.30	2.43	3.52	0.26	2.45
	5	2.64	0.24	1.72	3.86	0.28	2.61
	10	2.97	0.29	2.12	4.66	0.42	2.58
Retentate	2	2.86	0.31	2.23	2.82	0.31	2.46
	3	4.18	0.43	2.90	3.78	0.44	3.72
	4	3.73	0.35	2.76	4.75	0.54	5.21
	5	3.74	0.26	2.53	4.57	0.67	5.50
	10	3.50	0.43	3.15	4.35	0.77	9.21

1) SCW : soybean curd whey

2) VCR : volume concentration ratio

Table 20. Membrane Rejection Value of sum of raffinose and stachyose content

(unit : %)

VCR1	Regenerated cellulose	Polysulphone
2	42.91	38.99
3	28.83	46.63
4	12.22	52.87
5	29.75	53.16
10	32.68	69.94

1) VCR : volume concentration ratio

### 3 UF/NF

1.

: : = 4 : 1 : 5

(Kennedy et al., 1985 ; Kuo et al., 1988 ; , 1995).

HPLC

Table 21

Table 21. Soybean oligosaccharides contents of the soybean-curd whey by UF/NF

(unit : g/ )

Samples	Sucrose	Raffinose	Stachyose
Sumul	5.55	0.43	3.58
UF-P	1.31	n. d.	1.91
UF-R	3.27	0.34	1.85
NF-P	1.35	n. d.	n. d.
NF-R	35.8	6.80	32.6
NF-RIE	20.9	2.30	20.6

UF-P : permeate by ultrafiltration n. d. : not detected

UF-R : retentate by ultrafiltration NF-P : permeate by nanofiltration of UF-P

NF-R : retentate by nanofiltration of UF-P

NF-RIE : solution passed into mixed ion exchange (+ : - = 2 : 1) of NF-Rc

5.5g/ , 0.43g/  
 3.58g/  
 90%  
 nano 10  
 20  
 Nano nano  
 Table 22 20  
 Table 24  
 가 , 42%  
 2.

CaSO4

, Table 22 .

Table 22. Ionic calcium concentrations of the soybean-curd whey by UF/NF processing

Samples	Concentration
Sunmul	83.6
UF-P	58.5
UF-R	122.0
NF-P	34.2
NF-R	1670.0
NF-RIE	990.0

F-P : permeate by ultrafiltration n. d. : not detected  
 UF-R : retentate by ultrafiltration NF-P : permeate by nanofiltration of UF-P  
 NF-R : retentate by nanofiltration of UF-P  
 NF-RIE : solution passed into mixed ion exchange (+ : - = 2 : 1) of NF-R

83.6ppm

(1997)

가

UF/NF

.

(r=0.9899),

(r=0.9844), COD(r=0.9913)

.

가

가

nano

Donnan effect

. CaSO<sub>4</sub>

가

(diafiltration)

.

3. UF/NF

가.

**Table 23. Total sugar contents of the soybean-curd whey by UF/NF processing**

Samples	Sugar contents
Sunnul	12.21
UF-P	8.71
UF-R	12.78
NF-P	2.84
NF-R	151.09
NF-RIE	120.83

UF-P : permeate by ultrafiltration

UF-R : retentate by ultrafiltration

NF-P : permeate by nanofiltration of UF-P

NF-R : retentate by nanofiltration of UF-P

NF-RIE : solution passed into mixed ion exchange (+ : - = 2 : 1) of NF-R

2

**Table 23**

**Table 24**

0.49%

0.05%

90%

0.5%

0.28%

57%

nano

Nano

12

10

50%

가

UF/NF

Table 26

가 .  
 0.05% 90%  
 0.5%  
 0.48%, 0.68%  
 58% .  
 가 fouling 가

Table 24. Contents of total protein and total ash in the soybean-curd whey by UF/NF processing

Samples	Unit : %(w/w)	
	Protein	Ash
Sunnul	0.49	0.50
UF-P	0.05	0.48
UF-R	0.89	0.68
NF-P	0.04	0.30
NF-R	1.31	2.89
NF-RIE	0.66	1.32

UF-P : permeate by ultrafiltration

UF-R : retentate by ultrafiltration

NF-P : permeate by nanofiltration of UF-P

NF-R : retentate by nanofiltration of UF-P

NF-RIE : solution passed into mixed ion exchange (+ : - = 2 : 1) of NF-R



UF NF

2

**Table 25. Isoflavone contents in the soybean-curd whey by UF/NF processing**

	Isoflavone( $\mu\text{g/g}$ )		
	Dai dzei n	Geni stei n	Ge+De
summul	996. 416	285. 4686	1281. 885
NF- R	1027. 04	308. 98	1336. 02
UF- P	621. 96	179. 50	801. 45
UF- R	633. 01	201. 18	834. 20

4. UF/NF

가.

COD

70- 90ppm ( , 1993).

COD Table 26 26, 500

Fig. 4 COD 13, 400 50%

nano 5, 500 80%

가

**Table 26. COD of permeate and retentate of the soybean-curd whey by UF/NF**

(unit : ppm)	
Samples	COD value
Sunmul	26,500
UF-P	13,400
UF-R	34,000
NF-P	5,500
NF-R	466,000
NF-RIE	213,000

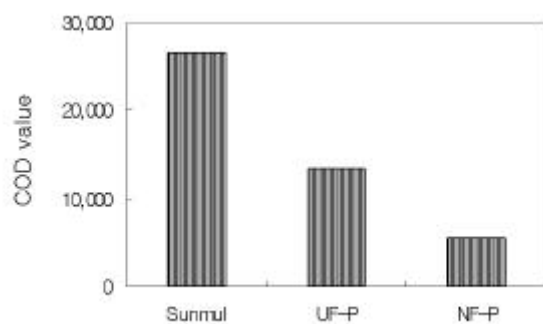
UF-P : permeate by ultrafiltration

UF-R : retentate by ultrafiltration

NF-P : permeate by nanofiltration of UF-P

NF-R : retentate by nanofiltration of UF-P

NF-RIE : solution passed into mixed ion exchange (+ : - = 2 : 1) of NF-R



UF-P : permeate by ultrafiltration

NF-P : permeate by nanofiltration of UF-P

**Fig. 4. Comparison of COD value from UF/NF permeate**

. pH

pH

Table 27

pH

(1997)

. UF/NF

pH가

fouling

pH

pI 3.7-4.0

(Wang H. L., Cavins J. F., 1989)

가

pH

가

pH

UF/NF

Table 27. pH change of the soybean-curd whey passed into UF/NF processing

Samples	pH
Sunnul	5.26
UF-P	5.62
UF-R	4.65
NF-P	5.56
NF-R	4.54
NF-RIE	4.07

UF-P : permeate by ultrafiltration

UF-R : retentate by ultrafiltration

NF-P : permeate by nanofiltration of UF-P

NF-R : retentate by nanofiltration of UF-P

NF-RIE : solution passed into mixed ion exchange (+ : - = 2 : 1) of NF-R

fouling 가

가

Table 28      20      0. 1823cp

0. 1224cp

0. 6486cp

가

diafiltration

diafiltration

Table 28. Viscosity of the soybean-curd whey passed into UF/NF processing

Samples	Viscosity(cp)
Sunnul	0. 1823
UF-P	0. 1275
UF-R	0. 1524
NF-P	0. 1224
NF-R	0. 6486
NF-RIE	0. 2914

UF-P : permeate by ultrafiltration

UF-R : retentate by ultrafiltration

NF-P : permeate by nanofiltration of UF-P

NF-R : retentate by nanofiltration of UF-P

NF-RIE : solution passed into mixed ion exchange (+ : - = 2 : 1) of NF-R

Table 29

가

가

Table 29. Conductivity of the soybean-curd whey passed into UF/NF processing

Samples	Conductivity (mMhos/cm)
Summul	5.4
UF-P	4.8
UF-R	5.3
NF-P	3.3
NF-R	14.6
NF-RIE	10.5

UF-P : permeate by ultrafiltration

UF-R : retentate by ultrafiltration

NF-P : permeate by nanofiltration of UF-P

NF-R : retentate by nanofiltration of UF-P

NF-RIE : solution passed into mixed ion exchange (+ : - = 2 : 1) of NF-R

, Brix

Table 30

(1992b)

2.2 brix

3.7brix

1. 3380

. Brix

(r=0.9771)                      nano

24.0 brix                      6                      .

14.3 brix

60 brix

가                      가                      .

**Table 30. Refractive index and brix of the soybean-curd whey passed into UF/ NF processing**

Samples	Refractive index	Brix
Sunnul	1.3380	3.7
UF-P	1.3365	2.5
UF-R	1.3390	4.3
NF-P	1.3355	1.9
NF-R	1.3700	24.0
NF-RIE	1.3545	14.3

UF-P : permeate by ultrafiltration  
 UF-R : retentate by ultrafiltration  
 NF-P : permeate by nanofiltration of UF-P  
 NF-R : retentate by nanofiltration of UF-P  
 NF-RIE : solution passed into mixed ion exchange (+ : - = 2 : 1) of NF-R

Hunter's values L(lightness), a(redness), b(yellowness)

Table 31                      UF/NF                      가

가                      .

Table 31. Hunter's color values of the soybean-curd whey by UF/NF processing

Samples	La	ab	bc
Sunnul	60.21	0.55	14.10
UF-P	92.84	-0.98	5.55
UF-R	54.41	0.82	10.39
NF-P	93.75	0.04	2.50
NF-R	36.85	5.20	20.13
NF-RIE	42.32	-0.89	14.35

a : Lightness    b : redness    c : yellowness

UF-P : permeate by ultrafiltration

UF-R : retentate by ultrafiltration

NF-P : permeate by nanofiltration of UF-P

NF-R : retentate by nanofiltration of UF-P

NF-RIE : solution passed into mixed ion exchange (+ : - = 2 : 1) of NF-R

5. UF/NF

bi fi do bacteri um

*B. infantis*                    1                     $1.4 \times 10^5$ cfu/Mℓ,                     $3.8 \times 10^5$ cfu/Mℓ  
, *B. longum*                    1  
 $3.3 \times 10^5$ cfu/Mℓ,                     $1.6 \times 10^5$ cfu/Mℓ

*S. typhinurium*, *S. aureus*                    5%

가                    1,                    broth  
2                    .                    2

*S. typhinurium*  $2.1 \times 10^8$ cfu/Mℓ, *S. aureus*  $9.5 \times 10^8$ cfu/Mℓ  
 $5.7 \times 10^4$ ,  $4.2 \times 10^6$

Table 32

Table 32. Viable cell counts of the cultures of concentrated solutions from UF/NF processing

	control 1a	control 2b	samples
<i>S. typhinurium</i>	$8.5 \times 10^4$	$2.1 \times 10^8$	$5.7 \times 10^4$
<i>S. aureus</i>	$2.4 \times 10^6$	$9.5 \times 10^8$	$4.2 \times 10^6$
<i>E. infantis</i>	$1.4 \times 10^6$	$1.8 \times 10^5$	$3.8 \times 10^5$
<i>E. longum</i>	$3.3 \times 10^5$	$1.4 \times 10^5$	$1.6 \times 10^5$

a : samples + 0.5% glucose

b : BHI broth incubation without samples(0.05% cysteine addition in case of *E.*

*infantis*, *B. longum*)

6.

가.

.

Table 33

Pb, Cd 2ppm

Pb 10ppm

Pb

As

8ppm

1ppm

DDTC-Ag

Ca, Na



Table 33. Mineral analysis of purified soybean oligosaccharides

(unit : ppm)					
	Ca	As	Cd	Pb	Na
	252	8	< 2	< 2	410
	6.8	8	<2	2	22

가

3%

(Minolta ; Japan)

L=92.04, a=-0.50, b=1.52

98%

(p<0.05).

# 4 가

1.

가.

UF/NF

가

Table 34

99.5%,

97.3%

가

Table 34

86%

, Table 35

yellowness

가

lightness가 가

99%

가

2 : 1

98.18 %

Table 34. Proximate common contents of soybean-curd whey treated with different ion exchange resin

	(unit : g/ )		
	Total sugar contents	Protein	Ash
Sunnul	15.332	0.410	0.857
Cation exchange resin	15.261(99.54)*	0.287(69.92)*	0.007(0.92)*
Anion exchange resin	14.920(97.31)*	0.057(13.82)*	0.016(1.83)*
Mixed resin(+ : - = 2 : 1)	15.053(98.18)*	0.143(34.96)*	0.008(0.94)*

( )\* : % recovery

Table 35. Hunter's color values of the soybean-curd whey treated with different ion exchange resin

	La	ab	bc
Sunnul	60.21	0.55	14.10
Cation exchange resin	66.30	1.86	11.81
Anion exchange resin	85.13	0.89	5.58
Mixed resin(+ : - = 2 : 1)	86.24	0.84	5.24

La : lightness , ab : redness, bc : yellowness

. Steffen

120 , 150 , 90 ,  
 30 , 60 , 90  
 2 , 1  
 51.6%, 63.5%, 53.1%  
 Table 36 89.4%,  
 89.91%, 78.84%

**Table 36. Recovery of soybean oligosaccharide in steffen method**

		(unit : g/ )	
	Summul	Recovery a	Recovery b
Sucrose	5.00	2.58(51.6)*	4.47(89.4)*
Raffinose	0.52	0.33(63.5)*	0.47(89.91)*
Stachyose	4.35	2.31(53.1)*	3.43(78.84)*

( )\* : Recovery

a : Using distilled water in cleaning cake after second filtration

b : Re-using permeated solution in cleaning cake after second filtration

**Table 37**

**Table 37**

50%

0.6g, 0.8g, 1.0g, 1.2g

Fig. 5

가 10% 1.0g

89.7%,

81%

가 가

Table 37. Change of oligosaccharides recovery by charcoal concentration

Charcoal contents(g)	(unit : %)		
	Sucrose	Raffinose	Stachyose
0.6	16.1	29.9	46.2
0.8	22.0	62	88.7
1.0	32.8	81	89.7
1.2	48.4	33.6	47.6

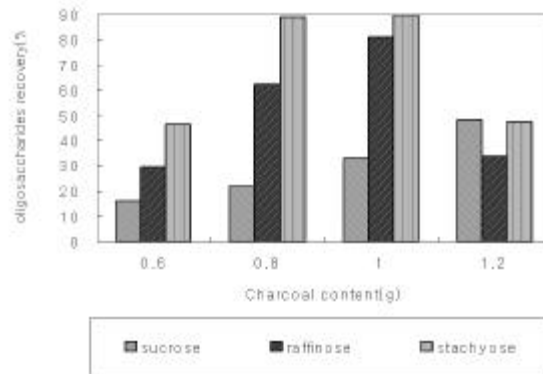


Fig. 5. Adsorption characteristics of oligosaccharides at various amounts of activated charcoal

5

1.

가.

2

Table 38

		0.41%
	16.19%	2
	23.82%	
	43.15%	
91.47%		0.875%
	18.81%	2
	17.78%	12.99%
2.55%		
	1.53%	41.7%
		가
28.1%		
	4.2%	

Table 38. Proximate analysis of aFDW, bFDCW, cSWP and dISP

	unit: w/w(%)		
	Crude protein	Ash	Total sugars
aFDW	16.19	18.81	41.7
bFDCW	23.82	17.78	28.1
cSWP	43.15	12.99	4.2
dISP	91.47	2.55	-

aFreeze-dried soybean-curd whey

bFreeze-dried concentrated soybean-curd whey by ultrafiltration

cSoybean-curd whey protein dIsolated soybean protein

3%

pH 4.1, 6, 8

(Physical)

20, 0-2270/sec

3% SWP ISP  
 가 가 가  
 가 . SWP  
 pH 가가  
 , ISP pH 4.1, 6, 8 가

Hunter's values L( ), a( ), b( )

Table 39 . , L  
 , a -1.91, -1.46,  
 86.35, 83.40, 80.68, 78.31 , a  
 0.09, 1.59, b 15.59, 11.66, 12.22, 13.99 .  
 가 L , a  
 가 L  
 -0.91626 , a 0.91939 ,

Table 39. Hunter's color values of  $\alpha$ FDW,  $\beta$ FDCW,  $\gamma$ SWP and  $\delta$ ISP

	L	a	b
$\alpha$ FDW	86.35	-1.91	15.59
$\beta$ FDCW	83.40	-1.46	11.66
$\gamma$ SWP	80.68	0.09	12.22
$\delta$ ISP	78.31	1.59	13.99

L : lightness, a : redness, b : yellowness

$\alpha$ Freeze-dried soybean-curd whey

$\beta$ Freeze-dried concentrated soybean-curd whey by ultrafiltration

$\gamma$ Soybean-curd whey protein

2.

가.

pH가

가

pH 4.1

,

가

가

.

(SWP)

ISP

ISP

(1992)

.

SWP

.

SWP

가

가

.

.

1)

SWP, ISP pH

pH 4.1

가

.

ISP

.

SWP

ISP



가

(Quaglia et al., 1990 ; , 1993 ) .

2)

pH SWP ISP 30 , 60

ISP 가

가 , SWP pH 6 가

. SWP ISP 가

가 가 . SWP가 ISP

SWP 가

가 ISP

가

1)

(SWP), ISP pH

Table 40 .

pH 2, 4.1, 6, 8, 10, 12

6Mℓ, 6Mℓ, 4Mℓ, 1Mℓ, 4Mℓ, 4Mℓ ISP

. ISP pH 4.1 pH 8 가 4Mℓ, 5Mℓ

pH 12 80Mℓ

Table 40 .

Batch-type

가

가



pH 8  
 가 1M  
 , pH 4.1  
 가 가  
 가 ( , 1993 ;  
 Pomeranz, 1985)

whipping, toppings, frozen dessert  
 albumin  
 ISP

3.

가.

200V ISP  
 basic subunit 20 kDa acidic  
 subunit 38 kDa . 66  
 90kDa , , ' . 90V  
 SWP subunit 가 subunit가 200kDa  
 29 49  
 kDa 7S subunit 11S  
 acidic subunit . 7S  
 subunit 66kDa .  
 20kDa band .  
 subunit



protein . di sul fi de bond  
 가  
 S-S .

(SWP) (ISP)  
 Table 42 . glutamic acid aspartic acid  
 tryptophan  
 mole % SWP 0.36 ISP 0.37

Table 42. Amino acid composition of soybean-curd whey protein (SWP) and isolated soybean protein (ISP)

unit :

	SWP	ISP
Asx*	9.65	10.77
Glx*	18.58	17.85
Ser	5.15	5.38
Gly	9.46	7.43
His	2.82	2.59
Arg	4.43	5.91
Thr	5.19	3.83
Ala	9.60	6.46
Pro	5.94	6.79
Tyr	0.78	2.34
Val	5.47	5.96
Met	1.14	1.08
Ile	4.74	5.42
Leu	6.65	8.28
Phe	2.77	4.24
Lys	7.58	5.67
∑E/T	0.36	0.37
Lys/Arg	1.71	0.96
∑MRH	35	37

\* : Asx = Aspartic acid + Asparagine

\*\* : Glx = Glutamic acid + Glutamine

∑E/T : the sum of essential amino acids/ the sum of total amino acids ratio

∑MRH : Molar ratio of hydrophobic amino acids (mol/mol, %)

cholesterol 가  
 cholesterol 가 lysine/  
 arginine ( , 1990 ;Tanaka & Sugno, 1989) .  
 Lysine/arginine SWP 1.71 ISP 0.96  
 casein  
 . proline, leucine, valine, phenylalanine, alanine  
 , mole SWP가 35%, ISP가 37% .

Table 43. Average hydrophobicity of soybean-curd whey protein (SWP) and isolated soybean protein (ISP)

unit : cal · mole<sup>-1</sup>

	SWP		ISP
	Ft	Ft × n/100	Ft × n/100
Ile	2,970	140.778	160.974
Tyr	2,870	22.386	67.158
Phe	2,650	73.405	112.36
Pro	2,600	154.44	176.54
Leu	2,420	160.93	200.376
Val	1,690	92.443	100.724
Lys	1,500	113.7	85.05
Met	1,300	14.82	14.04
Ala	730	70.08	47.158
Arg	730	32.339	43.143
His	500	14.1	12.95
Glu	550	102.465	98.175
Asp	540	52.11	58.158
Thr	440	22.836	16.896
Ser	40	2.06	2.152
Gly	0	0	0
Sum		1068.892	1195.854

Table 43

Bigelow Ney  
( , 1985).

$$Q = \frac{Ft \cdot n}{100} \text{ (cal} \cdot \text{mole}^{-1}\text{)}$$

Q : average hydrophobicity  
Ft : transfer free energy of an amino acid from an aqueous solution(at a certain concentration) to an ethanolic solution (at the same concentration)  
n : mole % of each amino acid

SWP가 1068.892 cal · mole<sup>-1</sup>, ISP가 1195.854 cal ·

mole<sup>-1</sup> ISP가 . SWP  
가 1068.892 cal · mole<sup>-1</sup> , 가 가  
가 , 가  
가 .

4.

가.

(SWP)

(ISP)

(*in vitro* digestibility)

가

7.83%,

0.522% .

가

59.3%

52.9%

, 가

. ACE

0.2%

crude extracts

ACE

25%, 21% , pepsin-pancreatin

SWP ISP

54%, 41%

crude extract

pepsin-pancreatin

가 ACE

## 6 UF/NF

1. UF/NF

가.

ACE

MWC가 10,000

polysulphone

15

50

ACE

table 44



NF-R IC<sub>50</sub> 53μg/ml

(NF-R)

Table 44. Purification of ACE inhibitory peptides from the nanofiltrated soybean-curd whey

Purification step	ACE Inhibition ratio(%)
Sumul	54
UF-perneatea	19
UF-Retentateb	40
NF-Retentatec	80

a UF-Perneate, permeate by ultrafiltration

b UF-Retentate, retentate by ultrafiltration

c NF-Retentate, retentate by nanofiltration of UF-perneate

Lowry

peptide-nitrogen

Table 45

0.49%

0.05%

10.000

90% 가

Table 45. Concentrations of total protein in the soybean-curd whey by UF/NF

Purification step	Protein(ng/ml)
Sunrul	86
UF- Pa	10.2
NF- Rb	17.7
F2 Fraction(FD)c	0.47

a UF-Permeate, permeate by ultrafiltration  
 b NF-Retentate, retentate by nanofiltration of UF-permeate  
 c Freeze-dried sample of F5 fraction.

2. ACE

open column chromatography , .  
 1g 3ml 1.5ml 4 peak가

chromatogram 280nm

Fig. 6 . chromatogram 4

ACE Table 46 . ,

F2 ACE

, F2 IC50 20µg/ml .

Gel permeation chromatography Fig.

7 , A -globulin

(bovine, M.W 158,000) , B ovalbumin(chicken, M.W 44,000),

C myoglobin(horse, M.W 17,000) , C peak 가

F2 10,000 dalton .

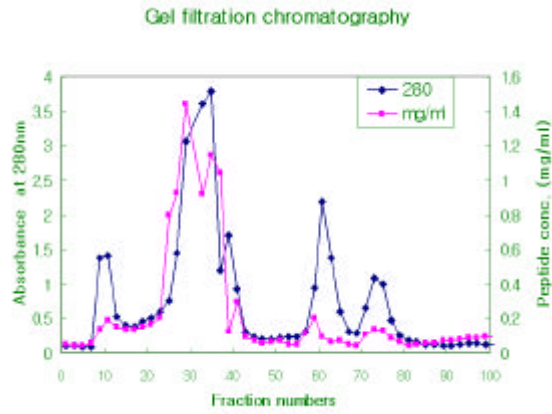


Fig. 6. elution profile of nanofiltrated soybean curd whey

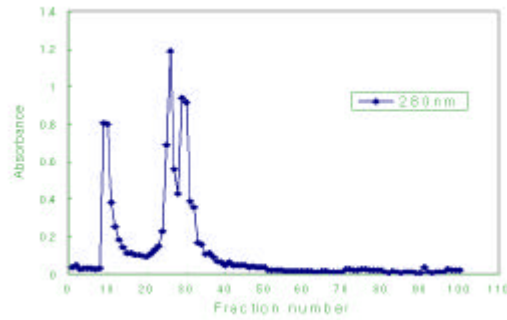


Fig. 7. Elution profile of molecular weight standards

Table 46. ACE inhibitory activity of each fraction obtained from the first gel filtration chromatography of the peptide fractions from NF-Ra

Fractions	ACE Inhibition ratio(%)
F1 (Fraction No. 10 - 29)	17.5
F2 (Fraction No. 30 - 49)	80
F3 (Fraction No. 50 - 69)	8.4
F4 (Fraction No. 70 - 89)	20.2
F5 (Fraction No. 90 - 100)	4.3

a NF-R, retentate by nanofiltration of UF-perneate

b IC<sub>50</sub>, defined as the concentration which inhibits 50% of the angiotensin-I converting enzyme activity.

N.D. : not detected

ACE (Ariyoshi, Y. 1993). gel filtration  
 2,000  
 10,000 가 .  
 ACE 가 F2 ,  
 prep-FPLC(AKTA, Superdex 75 HR)  
 F2 20ng/ml 100μl ,  
 3 , Fig. 8 .  
 chromatogram 2 peak F21, F22  
 ACE Table 47

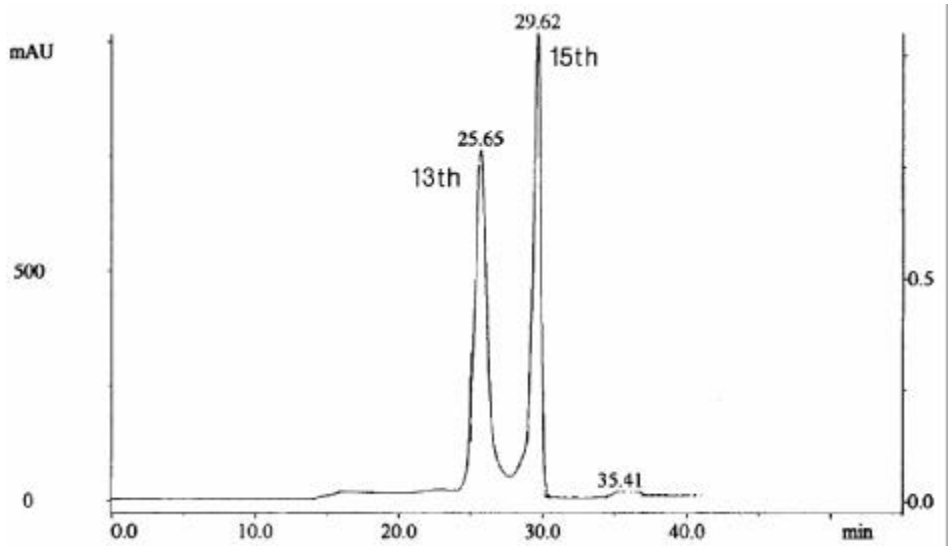


Fig. 8. Preparative FPLC chromatogram of the F2 fraction

Table 47. Purification of ACE inhibitory peptides from nanofiltrated soybean curd whey

Purification step	IC <sub>50</sub> ( $\mu\text{g}/\text{ml}$ )
Sunmul	123
NF-Rb	53
F2 fraction(FD)c	20
F2.2 fraction	13

a IC<sub>50</sub>, defined as the concentration which inhibits 50% of the angiotensin-I converting enzyme activity.

bNF-R, retentate by nanofiltrated

cFreeze-dried sample of F5 fraction

#### 4. MALDI mass spectrometry

MALDI (Matrix-assisted laser desorption/ionization)

mass spectrometry

bioanalytical ( Cheung, H. S., et al., 1980 ;

Hillenkamp, F., et al., 1991). ACE F2

MALDI mass spectrometry

10,000dalton .

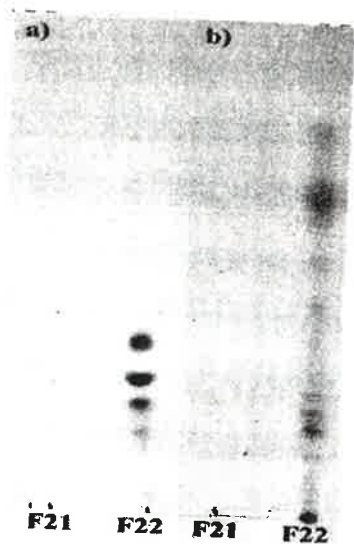


Fig. 9 Thin layer chromatogram on silica gel of compounds isolated from F21, F22

Developing solvent: BuOH : Acetic acid: H<sub>2</sub>O = 4:1:1

a) Ninhydrin reaction b) Sulfuric reaction

# 4

. . . (1992)  
, , , , .  
가 . 24(4):  
306. (1992a)  
, , , , . ,  
. 24(4): 311.  
(1992b)  
  
(1994)  
, .  
17: 26 (1983)  
. . .  
27(1): 43. (1994)  
가  
. (1993)  
  
(1994)  
, , , , ,  
Angiotensin Converting Enzyre(ACE) Peptide .  
Korean J. Food Sci. Technol., 27(2): 230 (1995)  
. . . (1988)  
. . (1993)

. (1996)

. 19(6): 492. (1987)

. 24(3): 199. (1992)

(1993)

(1997)

( , , )

: 3 (1996)

. 27(2): 225. (1995)

(1990)

가

25(5): 552 (1993)

Ariyoshi, Y Angiotensin converting enzyme inhibitors derived from food proteins. *Trend in Food Science and Technol.*, May, 139 (1993)

Beveridge, T., Tona, S.J., and Nakai, S. Determination of SH- and SS-groups in some food proteins using Ellman's Reagent. *J. Food Sci.*, 39: 49 (1974)



- Bollag, D.M. and Edelstein, S.J. Protein Method. Chapter 5. Gel Electrophoresis under denaturing conditions *Wiley-Liss* (1991)
- Cheung, H.S., Wang, F.L., Ondetti, M.A., Sabo, E.F. and Cushman, D.W. Binding of peptides substrates and inhibitors of angiotensin converting enzyme. *J. Biol. Chem.*, 255(2): 401(1980)
- Fotsis, T., Pepper, M., Adlercreutz, H., Fleischmann, G., Hase, T., Montesano, R. and Schweigerer, L. Genistein, a Dietary-driven Inhibitor of in vitro Angiogenesis. *Proc. Natl. Acad. Sci. USA.* 90: 2690. (1993)
- Franzen, K.L. and Kinsella, J. E. Functional Properties of Succinylated and Acetylated Soy Protein. *J. Agric. Food Chem.* 24(4): 788(1976)
- Hackler, L. R. and Stillings, B.R. Amino Acid Composition of Heat-processed Soymilk and Its Correlation with Nutritive Values. *Cereal Chem.* 44: 70 (1967)
- Hettiarachchy, N.S., Kalapathy U., and Myers D.J. Alkali-Modified Soy Protein with Improved Adhesive and Hydrophobic Properties. *JAOCS.*, 72(12): 1461 (1995)
- Hillenkamp, F., Karas, M., Beavis, R.C., and Chait, B.T. *Anal. Chem.* 63, 1193A (1991)
- Kato, A., Tsutsumi, N., Matsudori, N., and Kobayashi, K. and Nakai, S. *Agric. Biol. Chem.*, 45: 2755 (1981)
- Kaufmann, R. J. *Biotechnol.* 41(2-3): 155 (1995)

- Kennedy, I. R., Mwandenele, O. D. and McWhirter, K. S. Estimation of Sucrose, Raffinose and Stachyose in Soybean Seeds. *Food Chem.* 17: 85. (1985)
- Keung, W. M. and Vallee, B. L. Daidzein: A Potent, Selective Inhibitor of Human Mitochondrial Aldehyde Dehydrogenase. *Proc. Natl. Acad. Sci. USA.* 90: 1247. (1993)
- Knuclcs, B. E., Kuznicky, D. D., and Betschart, A. A. Effect of phytate and partially hydrolyzed phytate on in vitro protein digestibility. *J. Food Sci.*, 50: 1080 (1985)
- Kuo, T. M., Van Middlesworth, J. F. and Wolf, W. J. Content of Raffinose Oligosaccharides and Sucrose in Various Plant Seeds. *J. Agric. Food Chem.* 36: 32. (1988)
- Lowry, O. H., Rosebrough, N. J., A. Lewis Farr, and Rose, J. Randall Protein measurement with the folin phenol reagent. *J. Bio. Chem.*, 193: 265 (1951)
- Miller, C. D., Denning, H., and Bauer, A. Relation of nutrients in commercially prepared soybean curd. *Food Res.*, 17: 26 (1952)
- Nain, M., Gestetner, B., Bondi, A. and Birk, Y. Antioxidative and Antihemolytic Activities of Soybean Isoflavones. *J. Agric. Food Chem.* 24(6): 1174. (1976)
- Pomeranz, Y. Functional Properties of Food Components. AP Inc.,: 200(1985)
- Powers, J. J., Dratt, D. E. and Joiner, J. E. *Food Technol.* 15: 41. (1959)

- Quaglia, G. B., Orban, E. Influence of enzymatic hydrolysis on structure and emulsifying properties of Sardine (*Sardinops melanocephalus*) protein hydrolysates. *J. Food Sci.*, 55: 1571 (1990)
- Sathe, S. K., Salunke, D. K. Solubilization and electrophoretic characterization of great northern bean proteins. *J. Food Sci.*, 46: 82 (1981)
- Shino, A. Tsukuda, R., Odaka, H., Kitazaki, T., Tsuda, M. and Matsuo, T. Suppressive Effect of Ipriflavone on Bone Depletion in the Experimental Diabetic Rat. *Life Sci.* 42(11): 1123. (1988)
- Wagner, J. R. and Anon, M. C. Influence of Denaturation, Hydrophobicity and Sulfhydryl Content on Solubility and Water Absorbing Capacity of Soy Protein Isolates. *J. Food Sci.*, 55(3): 765 (1990)
- Wang, H. L. and Cavins, J. F. Yield and Amino Acid Composition of Fractions Obtained during Tofu Production. *Cereal Chem.* 66(4): 359. (1989)

5 .

1

가 , 96

43 가 ,

47 가

.

,

가 , 가

.

『

』

( ) “

가 .

1

75%

1, 500 2, 500mg

.

, , ,

”

, 『

』

( ) “ , 가 53% 가

38%, 25%, 18% ” .

(Ca<sup>2+</sup>) .

. 가 , 가

.

,

( , , , ) 가

240

.

## 2

### 1

1.

( )

, Acetic acid, citric acid, malic acid

Sigma(USA)

CC(calcium-citrate complex),

CM(calcium-malate complex)

.

2.

6 (ball

0.6 ) ball mill (Model 41-BN-200,

Kukje Science, Korea)

,

. ball mill ,

, ball mill

.

,

.

3.

250ml

가

200g

(Model FU-7063, Kukje Science, Korea)

,

(Model CR-300, MINOLTA, Japan) I( ), a( ),  
 b( ) E . 100mesh  
 , 3

#### 4. Hausner ratio

Bulk density 200Mℓ  
 (g/cm<sup>3</sup>) . , Hausner ratio  
 100 , bulk density tapped bulk density

$$HR = \frac{T}{O}$$

HR : Hausner ratio

T : Tapped bulk density

O : Bulk density of ashed eggshell

#### 5.

250Mℓ 50Mℓ 가  
 가 가

calcium probe ion meter(Model 720A pH/ISE meter, Orion, USA)

10Mℓ calcium ionic strength adjuster(4M KCl) 0.2Mℓ  
 가 .

6.

10%

5.6g : 50ml

CO2

CC(calciun-citrate complex), CM(calciun-malate complex)

7.

(UNI, Germany)

chamber

70

8.

(VSI)

(WAI)

2.5g

30ml

10

30

3000 x g

10

VSI

WAI

2

1.

가.



( )  
 (Calcium-citrate complex, calcium-malate complex)  
 , ,  
 14%, 0.45%, 13%, Farinogram  
 63%, 20  
 0.1% , 99.9%  
 , ( ) 98%  
 1 gas  
 280Ml  
 200  
 , Table 1  
 Otake (Model RC60B, 尾又葉鐵工所, Japan) , 가  
 , , 가 , (Model MG8, 尾又葉鐵工所, Japan)  
 80rpm 20 30  
 10 - 8 - 6 - 4 - 2.5mm  
 No. 10 3.0mm  
 ,  
 가 가 0.2 1.0%

Table 1. Composition of raw noodle

Ingredients	Flour basis(%)
Flour	100
Salt	3
Water	40
Organic calciums	0.2 1.0

가 Farinograph(Model  
 Sew, Brabender, Germany) , Farinogram 가  
 (absorption), (developing time), (stability),  
 (extensibility), (weakening of dough), (Dough consistency)

3 : 2.5 mm 6.5 cm  
 20 g 300 ml 5 25  
 1.5 . 130  
 ml 250 ml 가 .

가 .

$$\frac{Wt - Wo}{Wo} = k_w \sqrt{t}$$

$$\frac{Vt - Vo}{Vo} = k_v \sqrt{t}$$

W , V , o , t t

,  $k_w$  가 (min-1/2),  $k_v$  가

(min-1/2) .

13 6 , 1.5  
 Rheometer(Model CR-200D, Sun Scien. Co., Japan) Table 2

Table 2. Operating conditions for rheometer

Items	Conditions
Item selection	Mastication
Table speed	60mm/min
Chart speed	30No/sec
Load cell	1kg
Critical area	314mm <sup>2</sup>
Sample height	5mm
Sample width	10mm

10 g (Model GJM-496AU, IG Co., Korea) 1  
 90 ml 가 60 ,  
 icn meter(Model pH/ISE 720A, Orion, USA) .

13  
1 4 g 2 가 . 8  
2  
가 .

2.

가.

, Table 3 . 가

1)

• 1

, , 25 가  
mixer(Model A200I, Hobart, USA) 3 , 2  
가 24 가 . 27 , 75%  
4 1 .

Table 3. Formulas for loaf bread

Ingredients	Flour basis(%)	
	1st mixing	2nd mixing
Flour	70	30
Water	42	18
Salt	0.2	1.64
Yeast	26	
Emulsifier(SG-300)	2	
Food No25	0.2	
Sugar		10
Dried skim milk		2.7
Butter		15
-amylase		1
Organic acid eggshell calciums		0.2 1.0

2)

• 2

, , , 가  
 , 1 , 3 , 2  
 , 3 , 6 ,  
 가 27 가 15 , , bench time 15 , ,  
 panning . 2 38 , 85% 45 50

3)

2 가 oven(Model MF SS-2, Sungil Mech., Korea)

200 , 220 25 ,

, .

CR-200D, Sun Scien. Co., Japan)

Table 4

, plunger

Table 4. Measurement conditions of rheometer for loaf bread

Items	Conditions
Table speed	100mm/min
Chart speed	60No/sec
Critical area	1256mm <sup>2</sup>
Sample height	52mm
Sample width	130mm

10g 90MØ 가 , 60

(triangle test)

3.

가.

가 0.4, 0.7, 1.0%  
가 .

100 4 Texture Analyzer(TA-XI2 Stable  
Systems, U.K.) springiness, hardness, chewiness 40

가 calcium probe  
ion neter (720A pH/ISE neter, Orion, USA) .

blender 30g 300ml  
sanpling calcium ion neter

4.

가.

bile (B-3883), pepsin (P-7000), HCl (H-7020), pancreatin (P-1750) .

Signa

0.05, 0.1, 0.15, 0.2, 0.25, 0.3%(v/w) 가

가 5ml 20ml 0.1N HCl, 0.1N HCl/pepsin(0.1%), 0.05N HCl/pepsin(0.1%) 37 1 , 가 가 10ml 0.5% pancreatin 5% bile 가 0.1N HCl/pepsin/pancreatin/bile 0.05N HCl/pepsin/pancreatin/bile

3

1.





### 3

#### 1

1.

,  
.  
ball mill  
.  
,  
volume fraction( /ball mill  
) 0.08, ball mill 30 , 30rpm  
가 , 77%(w/w) .  
가 ,  
가  
.  
,  
,  
.  
, ball mill  
68%(w/w) .  
10 : 2 가 가  
, .

가 ,  
 가 ,  
 가 .

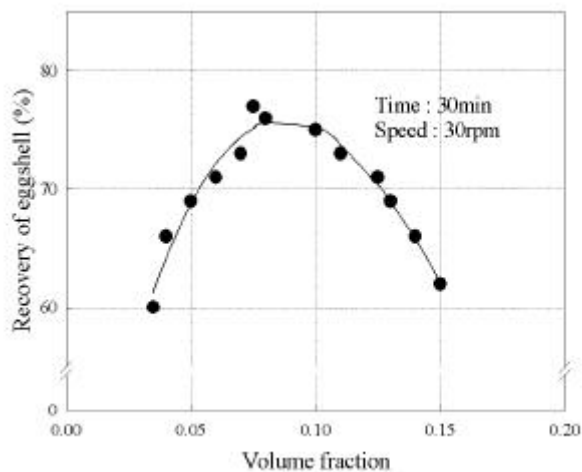


Fig. 1. Effect of volume fractions in a ball mill on the recovery of eggshell.

2.

가  
 가

850, 900, 1000

Hunter's colorimeter

. 850, 900, 1000

14, 9, 3

L

90

가

200g

. 120g(±5) 가

CaCO<sub>3</sub> + heat CaO + CO<sub>2</sub> 가 ,

가 . •

100, 56 120g

가 , ,

• tapped bulk density Hausner ratio

, 가 1.43

가 .

Table 4. Effect of ashing temperature on the color(lightness) and color difference of eggshell powder

Time(h)	900				1000			
	L*	a	b	E**	L	a	b	E
1	53.37	-0.15	+0.82	38.60	69.31	+0.10	+0.73	22.99
2	68.55	+0.09	+0.52	23.75	85.35	+0.17	+0.58	6.96
3	78.55	+0.12	+0.35	13.75	93.21	+0.22	+0.47	0.93
4	79.75	+0.07	+0.23	12.80				
5	83.53	+0.09	+0.22	6.77				
6	85.62	+0.19	+0.37	6.68				
7	87.75	+0.18	+0.35	4.55				
8	89.25	+0.18	+0.38	3.05				
9	91.78	+0.20	+0.40	0.53				

\* L : lightness(100=white, 0=black)

a : redness(-=green, +=red)

b : yellowness(-=blue, +=yellow)

\*\* E : color difference(= $[(L - L_0)^2 + (a - a_0)^2 + (b - b_0)^2]^{1/2}$ )

NBS( E) : unit classification : 0 0.5=trace, 0.5 1.5=slight,  
1.5 3.0=noticeable, 3.0 6.0=appreciable,  
6.0 12.0=nuch, over 12.0=very nuch.

Table 5. Change in eggshell weight with time at various ashing temperature (unit : g)

T (°C)	850	900	1000
Time(h)			
0	200	200	200
1	191	164	141
2	185	154	123
3	178	140	116
4	174	135	
5	170	130	
6	168	125	
7	164	122	
8	155	119	
9	153	118	
10	145		
11	140		
12	132		
13	129		
14	124		

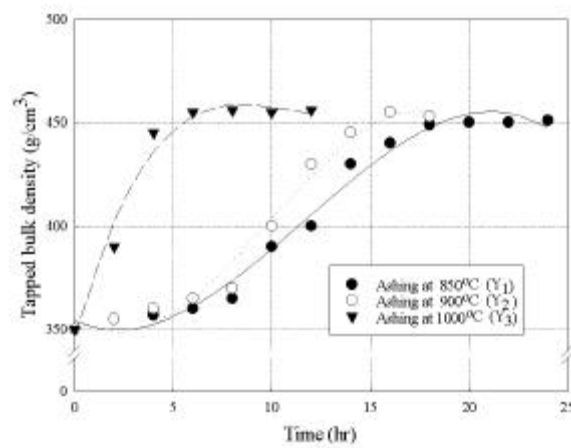


Fig. 2. Change in tapped bulk density with ashing time at various ashing temperatures.

3.

가 가 .

가 20ppm

가 L

가 가 L 70, 80, 90

600, 700, 1000ppm

50

가 가 .

(0 30 )

pH 가 L 90

pH 6.5

20

pH 14

가

(20)

가

가

가 (solubility : 0.002per 100parts by weight of the water at 100 ) ,

가 (solubility : 0.077per 100parts by weight of the water at 100 ) 가

(0.1 2%)

가

. citric acid

50Mℓ                      10g                      가 60  
, 1%                      3600ppm  
, pH 2.3                      20                      pH 9.5  
가                      pH                      ,                      가  
가 가                      .                      acetic acid  
1%                      3400ppm                      ,  
lactic acid                      0.5%                      4100ppm                      .  
pH가  
가                      가 가 .

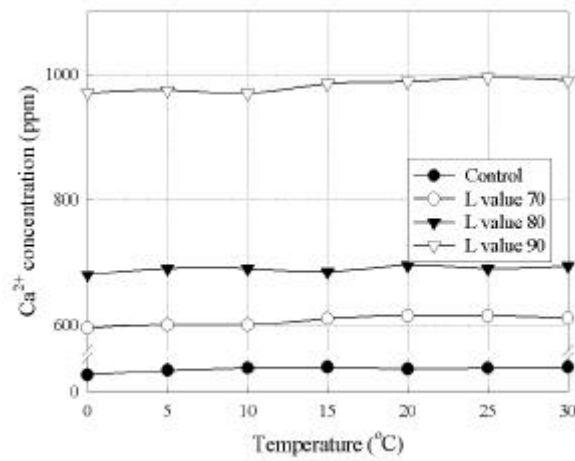


Fig. 3. Effect of temperatures on the water solubility of calcium of eggshell powder.

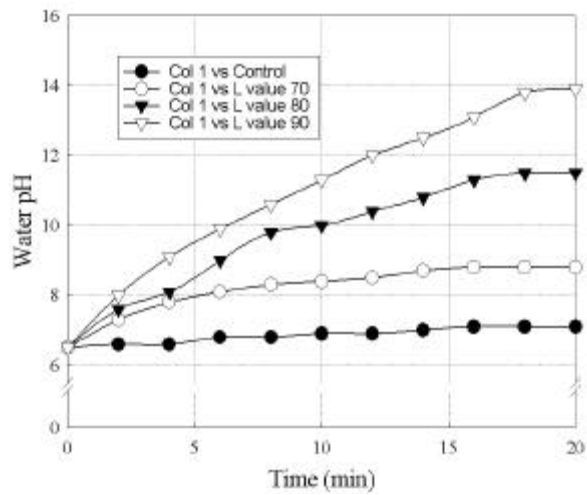


Fig. 4. Time course change of pH in water during eggshell powder solubilization.

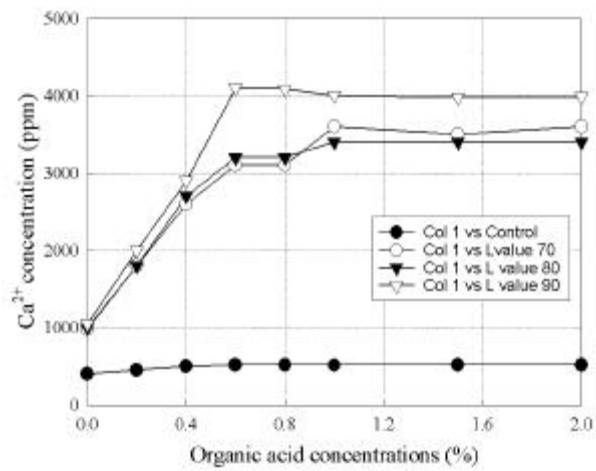


Fig. 5. Effect of organic acid concentrations on the solubility of eggshell powder.



4.

WSI WAI

1)

(WSI)

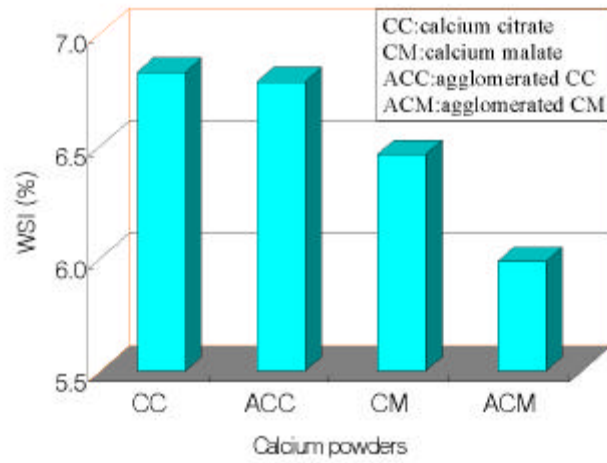


Fig. 6. WSI of some calcium powders

2)

(WAI)

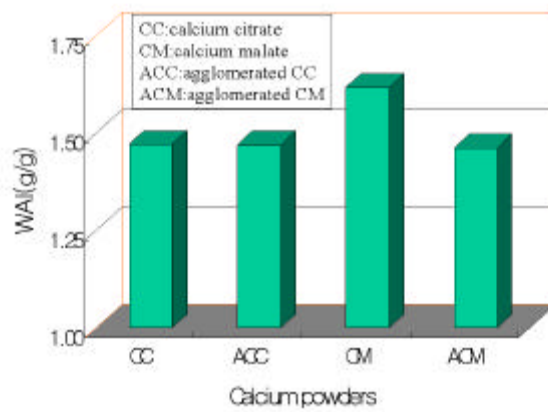


Fig. 7. WAI of some calcium powders

## 5. 유기산 칼슘 분말의 전자사진(SEM)

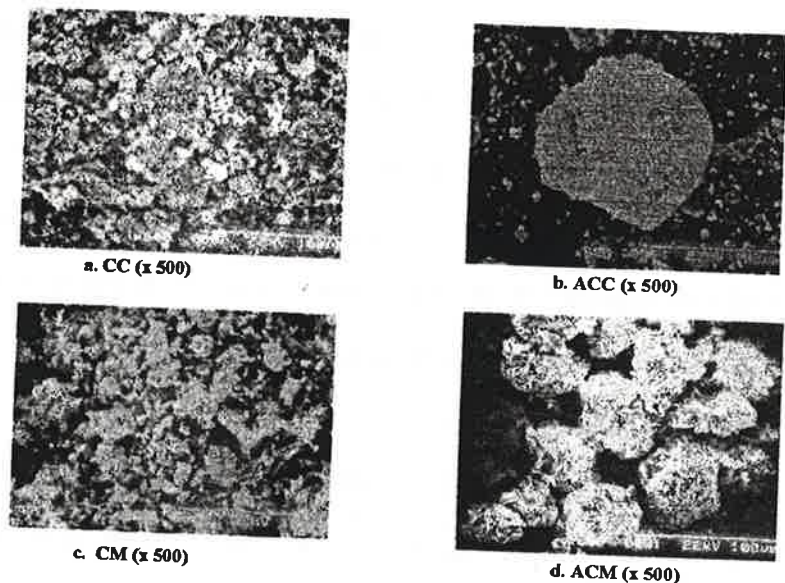


Fig. 8. SEM of some calcium powders

( CC:calcium citrate                      CM:calcium malate  
 ACC:agglomerated CC                  ACM:agglomerated CM )

## 제 2 절 유기산 칼슘이 강화된 식품의 물성

### 1. 숙면

#### 가. 반죽 성질

난각 칼슘분을 그대로 첨가할 경우 알칼리제의 첨가시와 유사하게 반죽의 경도를 높이는 것은 물론 반죽 성질에 많은 변화를 유발한다. 결과적으로 면의 절단력과 압착력의 증대 등 생면 특유의 식감에 나쁜 영향을 미치는 것으로 알려져 있다.

이와 같은 단점을 보완하기 위하여 본 연구에서는 제조한 난각 칼슘을 유기산

가 가 .

가 63% calcium citrate (CC)

calcium malate (CM) 가 CC 가

, CM 가 0.8% 가 가

1.0% 가 66% 가 (Fig. 9). 가

가 . 가

가 .

29 가

가 가 (Fig. 10). , CM 가

0.2% 가 9 . CC 가

가 가 가 가

. gluten

gluten 가

. 가 가

가 ,

. 가

가 .

가가 , CC 가

가 가 0.4% 가 8 12 가 .

가 가 1% 가 4

(Fig. 11). CM 가 가 0.2%

2  
 가  
 3가 CM 가 CC  
 가  
 가 CM 0.1% , CC  
 0.4%

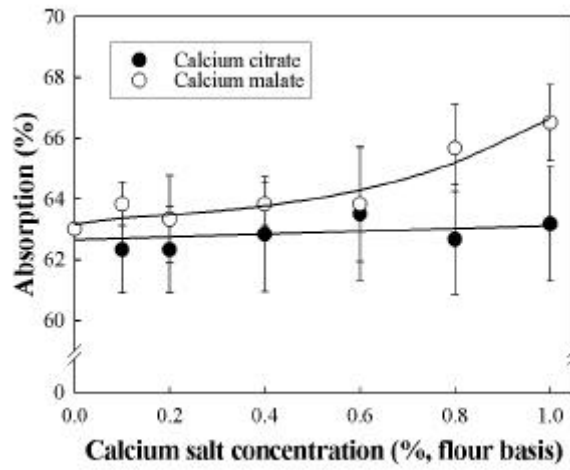


Fig 9. Effect of calcium salt concentration on the water absorption of wheat flour (Error bars indicate 95% confidence intervals).

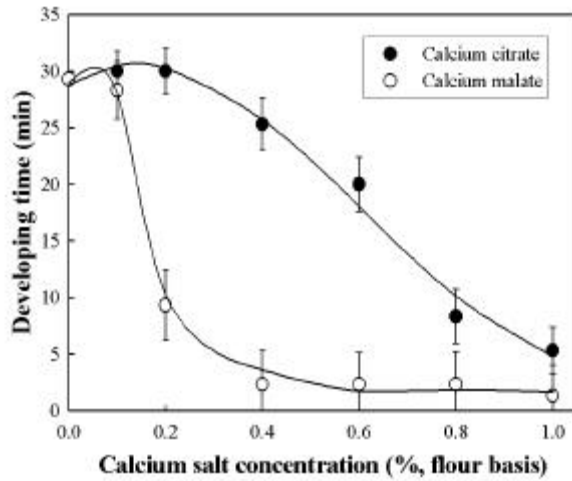


Fig. 10. Effect of calcium salt concentration on the development time of wheat flour (Error bars indicate 95% confidence intervals).

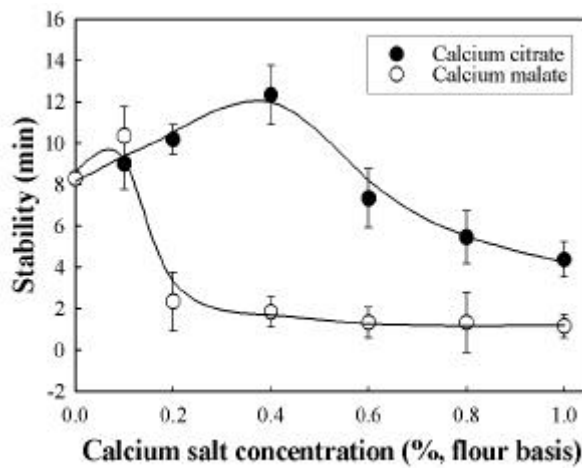


Fig. 11. Effect of calcium salt concentration on the stability time of wheat flour (Error bars indicate 95% confidence intervals).

가  
 20 g 5 25 가  
 가 가  
 가 가 CM 가 가  
 CC 가 (Table 6). CC 가 0.8%  
 , 가 가 0.072 min<sup>-1/2</sup> 0.067 min<sup>-1/2</sup>  
 . CM 0.2% 가 0.063 min<sup>-1/2</sup> CC 0.8% 가  
 CM 0.8% 가 0.058  
 min<sup>-1/2</sup> 가가 가  
 가 가 가 가

Table 6. Weight gain and volume expansion rate constants of cooked noodle by salt addition level

Cooked noodle		Weight gain rate constant (min <sup>-1/2</sup> )	Volume gain rate constant (min <sup>-1/2</sup> )
	Control	0.057	0.072
CC*	0.2%	0.057	0.070
	0.4%	0.056	0.069
	0.6%	0.055	0.069
	0.8%	0.054	0.067
CM**	0.2%	0.054	0.063
	0.4%	0.054	0.061
	0.6%	0.054	0.060
	0.8%	0.054	0.058

\*CC ; calcium citrate, \*\*CM ; calcium malate

Rheometer , hardness  
 가 , CC 가 가  
 가 . 가 가 CC 가  
 springiness 0.6% 가 , cohesiveness CC 가  
 0.1 . Adhesiveness CC가  
 가 . 가  
 가 가  
 , 가 가  
 가 .  
 가 ,  
 가 0.4%, 0.6%  
 , Nagao  
 가 , 가  
 가 .

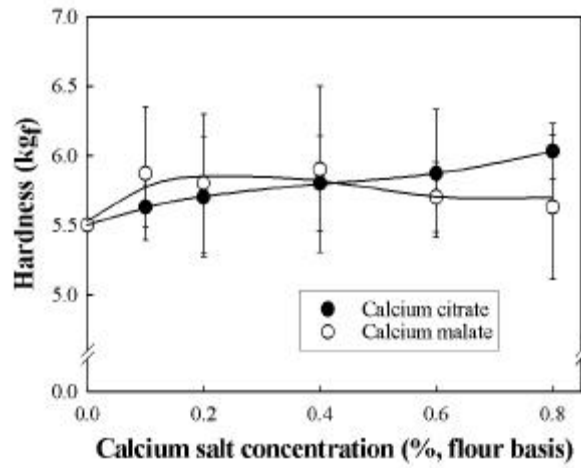


Fig. 12. Effect of calcium salt concentration on the hardness of wet noodle (Error bars indicate 95% confidence intervals).

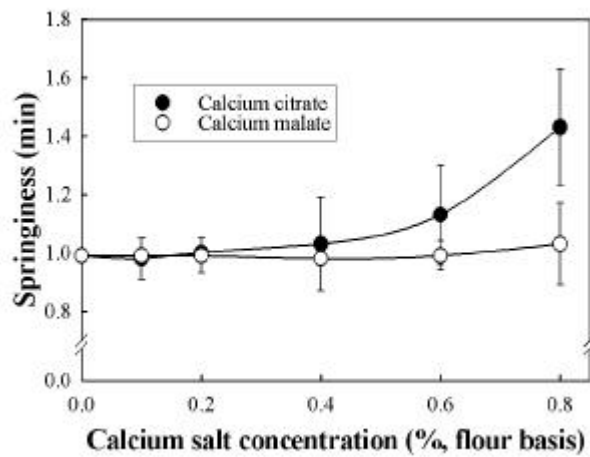


Fig. 13. Effect of calcium salt concentration on the springiness of wet noodle (Error bars indicate 95% confidence intervals).

가 13mg



가 가 , 18ppm .  
 가 CC, CM 가  
 1.0% 98, 105ppm .  
 0.6% 가 200g 1300ppm  
 360ppm 3.6 가 ,  
 pH 가 가 .  
 , 가  
 , 가  
 가 가 .

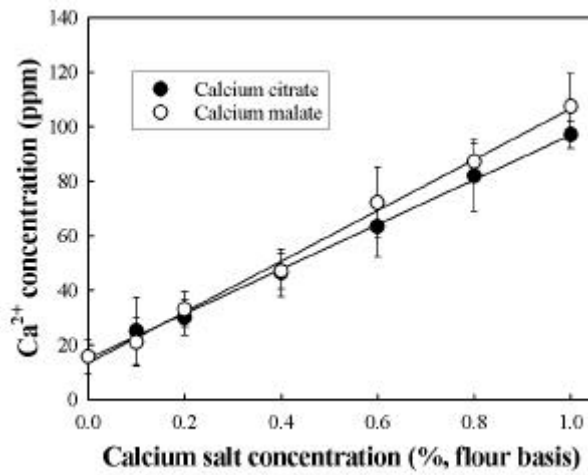


Fig. 14. Relationship between calcium salt concentration of raw noodle and calcium ion concentration of cooked noodle dispersed in distilled water with 1:9 weight ratio (Error bars indicate 95% confidence intervals).

.  
 가  
 CC, CM 0.8% 가 , 13  
 8 2 ,  
 chi-square( 2) test .  
 Table 7 CC 가 0.6% 5%  
 가 가 , CM  
 0.8% 가 5% 가 .  
 , 가 CM  
 가가 가 .  
 가 0.2%가 가  
 . 가  
 가

Table 7. Difference analysis for cooked noodle fortified with CC1) and CM2)

Organic acids-eggshell calcium salts	Salt Concentration (% , flour basis)	Number of correct answers out of 16	2
CC	0.2	5	0.008
	0.4	7	0.38
	0.6	11	7.51*
	0.8	16	29.07**
CM	0.2	6	0.08
	0.4	7	0.38
	0.6	9	2.82
	0.8	14	18.76**

1)CC ; calcium citrate, 2)CM ; calcium malate  
 \*Significant at p 0.05, \*\*Significant at p 0.01

2.

가.

Rheometer , hardness 1.0% CM 가  
 1.0 0.1 가가 hardness  
 . Springiness CC 가 , 가  
 . 가  
 . ,  
 가  
 , 가 가  
 가 .

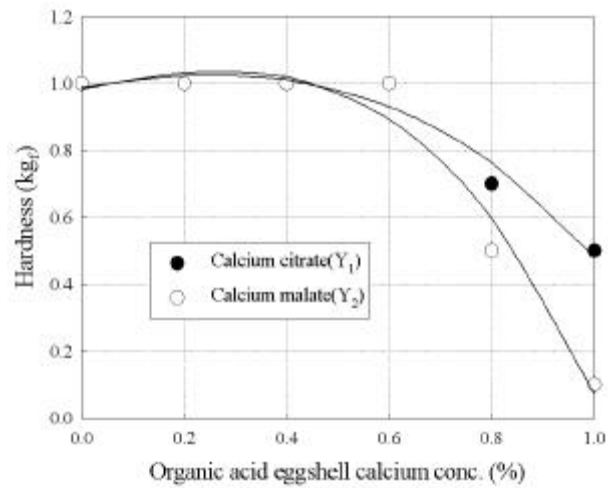


Fig. 15. Effect of organic acid eggshell calcium concentrations on the hardness of loaf bread.

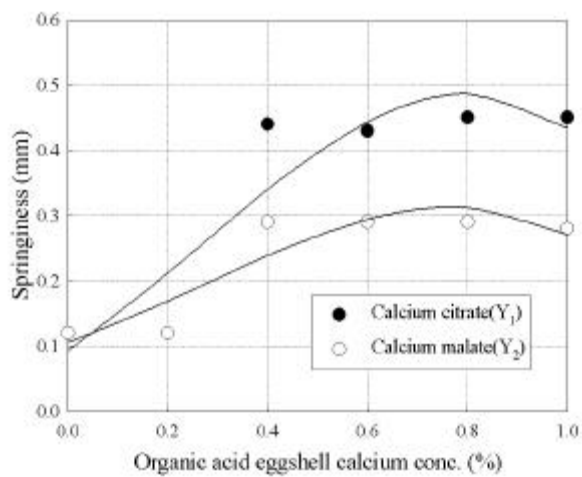


Fig. 16. Effect of organic acid eggshell calcium concentrations on the springiness of loaf bread.

1.0% CC 가

220ppm, CM 240ppm 가 , 200 가

. 1.0% 가

, 가

.

.

0.8% 가 8 2

, chi-square( 2) test .

0.2 0.8% 가 CC, CM 가

0.8% 가 1% .

가 CC, CM 0.6%

가

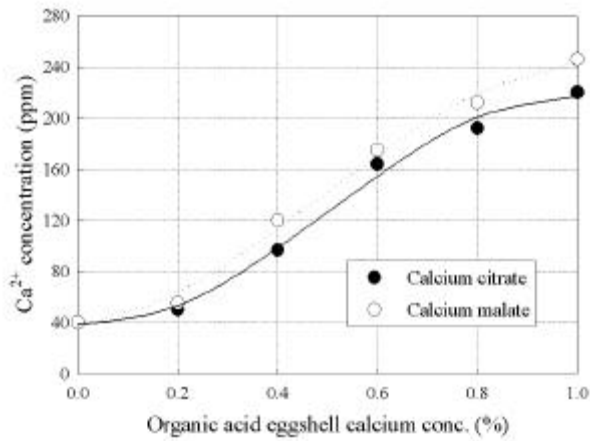


Fig. 17. Calcium ion concentrations of loaf bread with the amount of addition of organic acid eggshell calcium.

Table 8. Difference analysis for loaf breads from CC, CM fortified and concentrations

Organic acid eggshell calci ums	Number of correct answers out of 16	2	
CC	0.2%	9	0.38
	0.4%	9	0.38
	0.6%	7	3.78
	0.8%	2	13.13*
CM	0.2%	11	0.01
	0.4%	10	0.01
	0.6%	7	2.82
	0.8%	3	14.45*

3.

가.

1 CM CC 1.0%(w/v) 가

62.8, 24.8ppm

198 , 78 MEP(0.7% w/v) 11 , 4 .

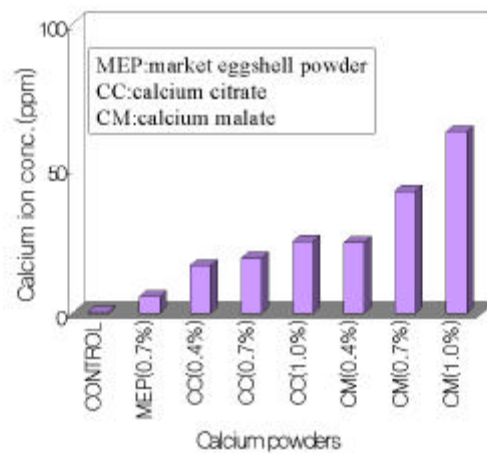


Fig. 18. Calcium ion conc. after 1 hour soaking

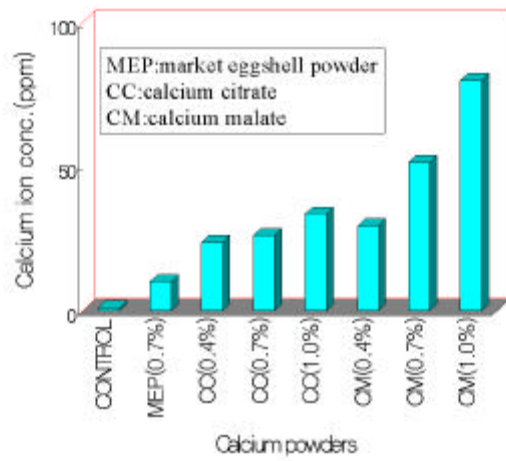


Fig. 19. Calcium ion conc. after 6 hour soaking

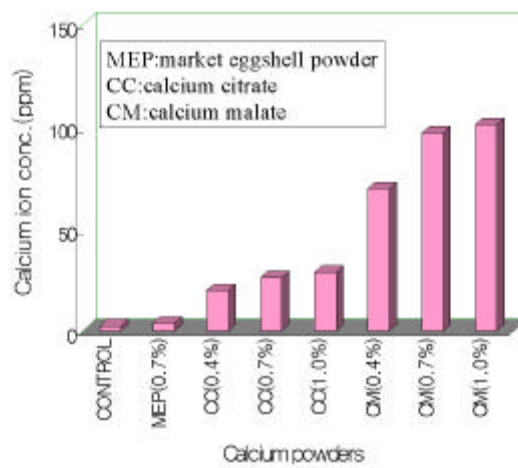


Fig. 20. Residual calcium ion conc. in Raren soup

### 1) Springiness

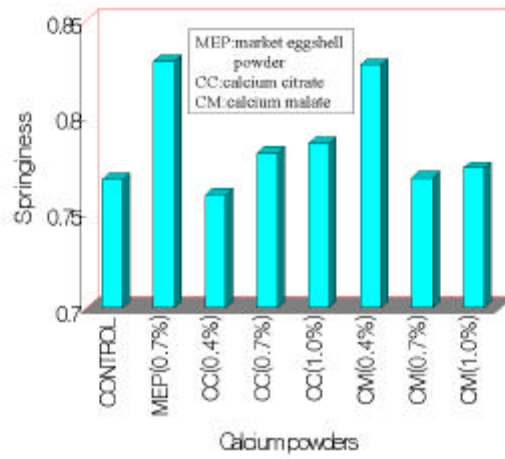


Fig. 21. Effect of calcium on springiness of Raren

### 2) Hardness

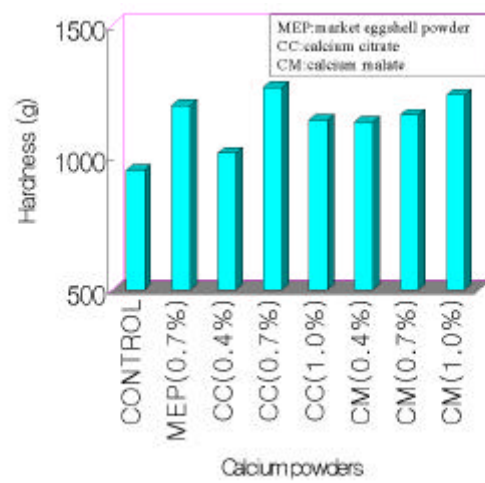


Fig. 22. Effect of calcium on hardness of Raren



### 3) Chewiness

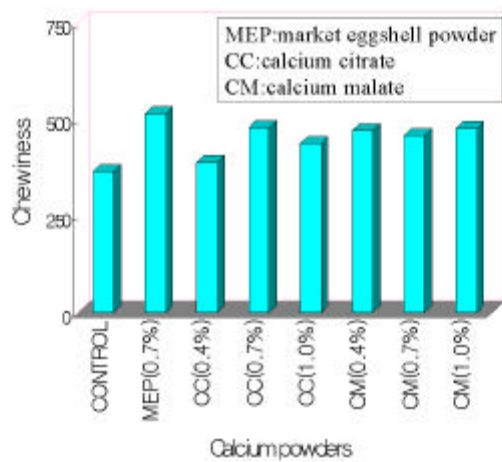


Fig. 23. Effect of calcium on chewiness of Ramen

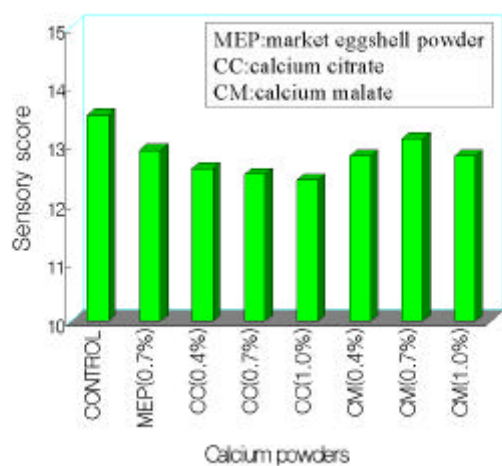


Fig. 24. Sensory results of Ramen fortified with calcium powders

4.

가.

가

calcium citrate 가 가 0.2%(w/v)  
 calcium citrate  
 calcium citrate 0.2%(w/v) 가 가  
 Calcium nalate 가 CM 가 0.25%(w/v)  
 CM CM 0.25%(w/v) 가  
 Calcium citrate nalate 가  
 5% CCM 0.15%(w/v)  
 가 가

Table 9. Calcium ion concentration in soy milk fortified with various forms of calcium additives (ppm)

	control	0.05%	0.1%	0.15%	0.2%	0.25%	0.3%
CC	4.60	6.73	10.54	22.66	30.84	39.26	50.60
CM	4.60	7.42	18.72	39.30	55.08	90.12	123.20
CCM	4.60	10.97	16.96	24.96	34.40	47.80	58.70

CC: calcium citrate CM: calcium nalate CCM: calcium citrate nalate

가

(0.1N HCl) 2.3 6.1 가

0.1HCl/pepsin 0.1N HCl

Table 10. Calcium ion concentration in soy milk fortified with various forms of calcium additives in 0.1N HCl (ppm)

	control	0.01%	0.02%	0.03%	0.04%	0.05%	0.06%
CC	2.35	7.84	13.27	18.63	27.87	32.60	47.03
CM	2.35	9.70	16.27	24.67	35.73	45.60	53.67
CCM	2.35	8.39	9.44	13.30	17.70	25.27	25.67

CC: calcium citrate CM: calcium malate CCM: calcium citrate malate

Table 11. Calcium ion concentration in soy milk fortified with various forms of calcium additives in 0.1N HCl/Pepsin (ppm)

	control	0.01%	0.02%	0.03%	0.04%	0.05%	0.06%
CC	2.78	10.18	11.57	20.93	27.53	34.63	42.80
CM	2.78	10.98	18.30	24.37	34.23	48.50	50.60
CCM	2.78	9.52	13.43	13.00	19.73	23.77	28.27

CC: calcium citrate CM: calcium malate CCM: calcium citrate malate

Table 12. Calcium ion concentration in soy milk fortified with various forms of calcium additives in 0.05N HCl/Pepsin (ppm)

	control	0.01%	0.02%	0.03%	0.04%	0.05%	0.06%
CC	3.27	8.94	12.22	17.63	22.80	30.13	38.47
CM	3.27	9.47	14.67	22.63	33.77	43.60	56.07
CCM	3.27	8.04	9.15	12.77	16.00	21.17	23.73

CC: calcium citrate CM: calcium malate CCM: calcium citrate malate

. 가

가 (pH 7) (pH 1.0)

가

가

.

**Table 13. Calcium ion concentration in soy milk fortified with various forms of calcium additives in 0.1N HCl/Pepsin/Pancreatin/Bile (ppm)**

	control	0.01%	0.02%	0.03%	0.04%	0.05%	0.06%
CC	0.13	0.57	0.96	3.03	5.94	8.73	11.63
CM	0.13	0.56	1.43	4.17	7.58	11.72	14.57
CCM	0.13	0.35	0.64	0.98	2.57	3.64	5.45

CC: calcium citrate      CM: calcium malate      CCM: calcium citrate malate

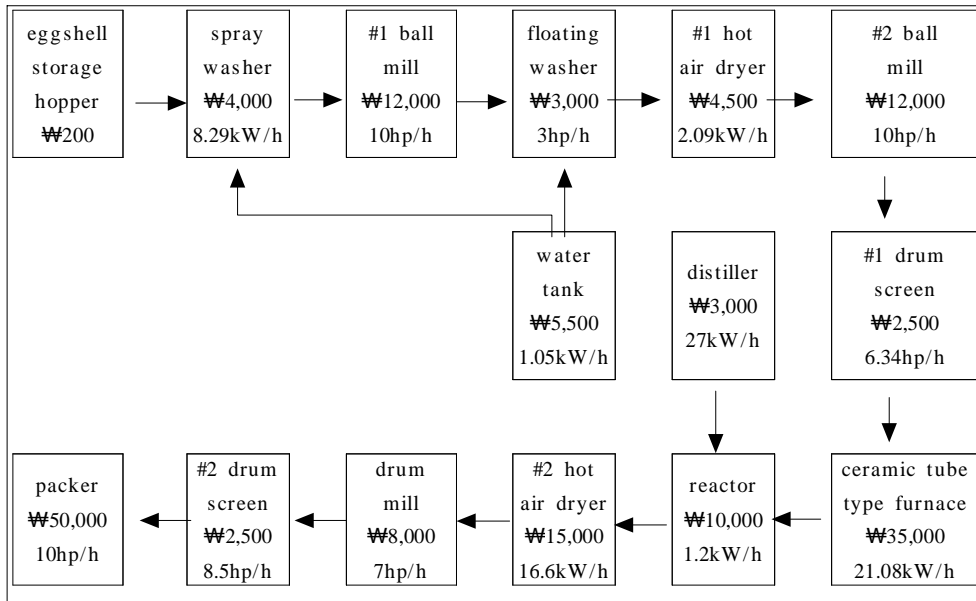
**Table 14. Calcium ion concentration in soy milk fortified with various forms of calcium additives in 0.05N HCl/Pepsin/Pancreatin/Bile (ppm)**

	control	0.01%	0.02%	0.03%	0.04%	0.05%	0.06%
CC	2.18	2.68	4.19	5.87	8.35	11.30	15.97
CM	2.18	3.46	4.64	7.13	10.57	15.07	19.33
CCM	2.18	3.02	3.29	4.35	5.62	6.86	8.70

CC: calcium citrate      CM: calcium malate      CCM: calcium citrate malate

1.

가.

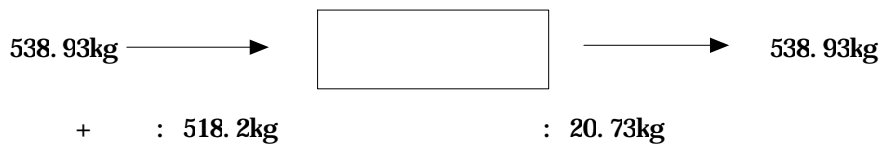


(block 가 :₩ )

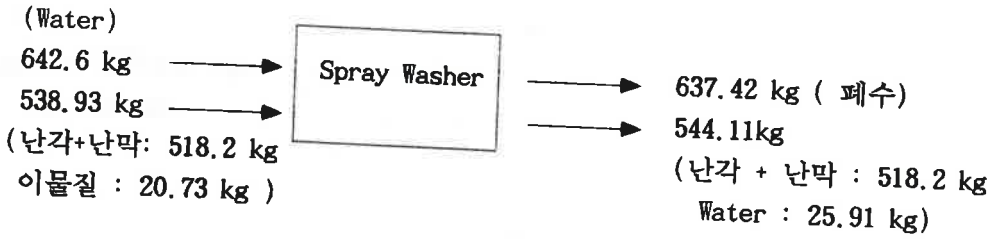
. Mass Balance

가 ) 1 ( + ) = 6.4g (6.1g + 0.3g)  
 $\text{CaCO}_3 = 5.8\text{g}$

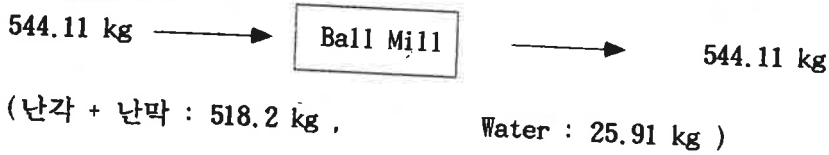
1)



2)



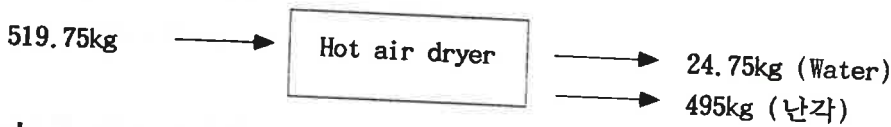
3)



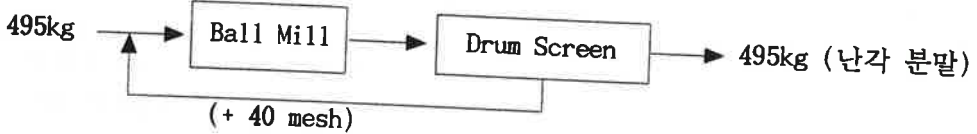
4)



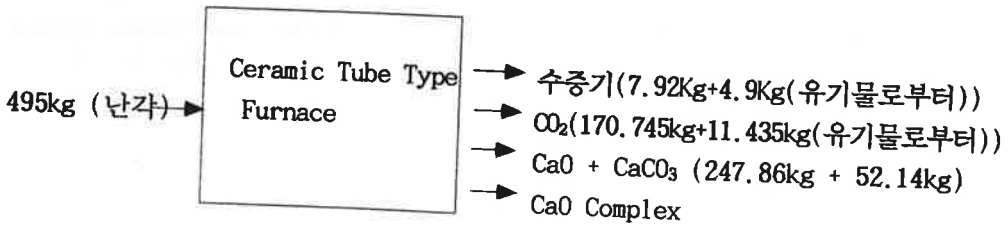
5)



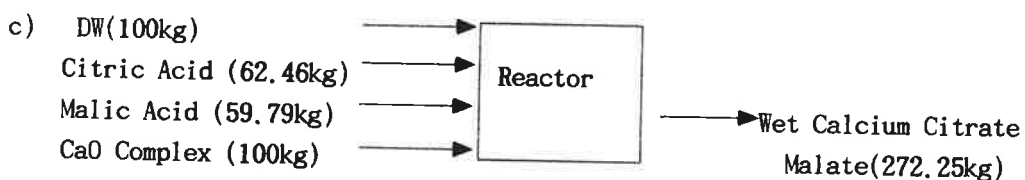
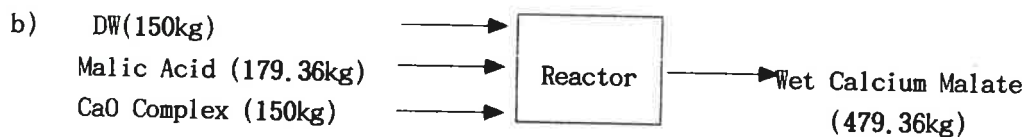
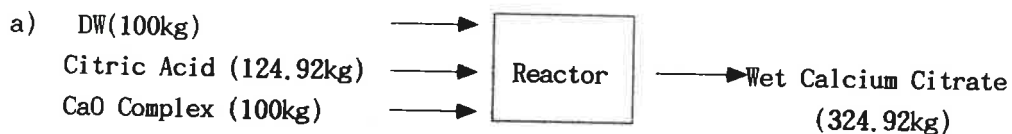
6) & 7)



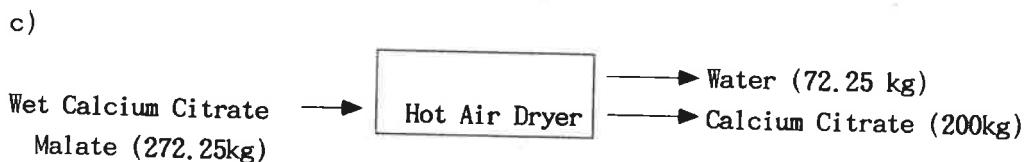
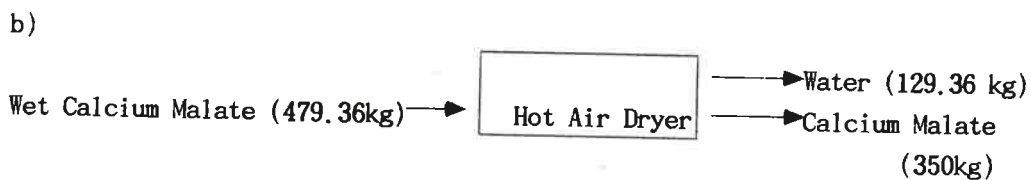
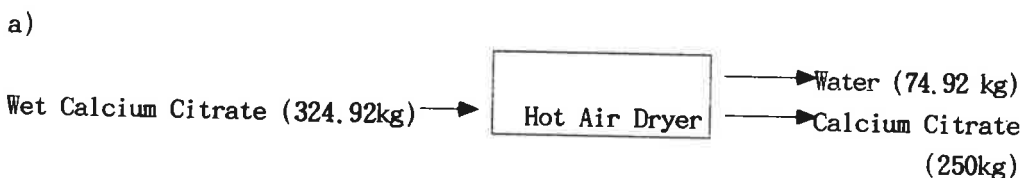
8)



9)

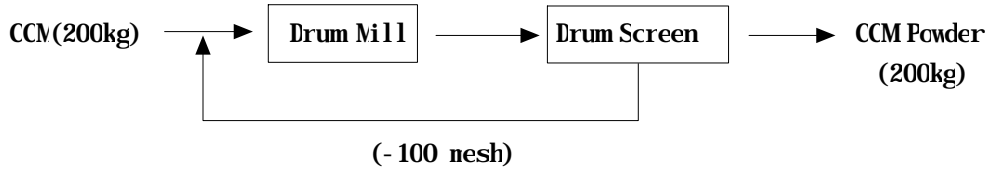
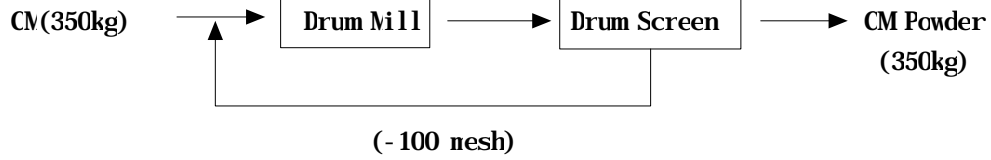
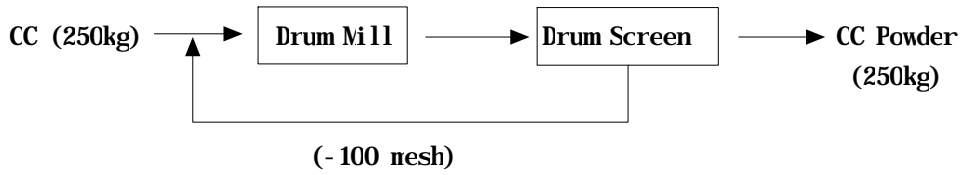


10)



11) & 12)

a)



13)

a)



b)



c)





2.

( : )

		177, 200	177, 200
	43%)	76, 196	
	14%)	24, 808	
	15%)	26, 580	
	35%)	62, 020	
	13%)	23, 036	
	20%)	35, 440	
2)		100, 000	525, 280
	30%)	53, 160	578, 440
	7%)	40, 490. 8	
	15%)	86, 766	705, 697. 8
			705, 697. 8

1) : % 2) : 가

3) : %

3. 가

가.

1)

	(kg/day)	가 ( /kg)	( )
	518. 20	-	-
citric acid	187. 38	5, 750	1, 077, 435
malic acid	238. 75	5, 300	1, 265, 375
1			2, 342, 810
1 (300 )			702, 843, 000

2)

	(n/ )	가( /n)	( / )	
	76, 800	160	12, 288, 000	(5kg) 256

3)

	(2600 /h)	(1770 /h)	(2700 /h)	
spray washer #1 ball mill floating washer		1		
#1 hot air dryer #2 ball mill #1 drum screen ceramic tube type furnace	1		1	
reactor #2 hot air dryer drum mill #2 drum screen	1			
packer		1		
	2	2	1	5
	5, 200	3, 540	2, 700	11, 440
	12, 480, 000	8, 496, 000	6, 480, 000	27, 456, 000
(1)				10, 982, 400
(2)				9, 152, 000
				47, 590, 400

(1) : 40%

(2) : 400%

4)

		( / )	가 ( )	( / )
	kW/h	323, 194	46	14, 866, 906
	ton	1, 981	200	700
	ton	1, 015	700	710, 289
				15, 973, 384

5)

	1	680, 000	8, 160, 000
	1	750, 000	9, 000, 000
	2		17, 160, 000
(1)			6, 864, 000
(2)			5, 720, 000
			29, 744, 000

(1) : 40%

(2) : 400%

6)

가

		( / )	가 (5kg) ( )
		702, 843, 000	14, 643
		12, 288, 000	256
		47, 590, 400	991
		15, 973, 384	333
	가 1)	544, 917, 200	11, 352
	2)	34, 790, 750	725
	3)	13, 916, 300	290
		29, 744, 000	620
		1, 402, 063, 034	29, 210
	4)	280, 412, 607	5, 842
		1, 682, 475, 641	35, 052

1) : 10% 2%

2) : 5% 3) : 2% 4) : 20%

## 4

1. Fenera, O. R., (1985) 'Food chemistry' 2nd Ed., 521 - 525, 675, Marcel Dekker, New York, USA.
2. , , , (1991) ' ' 216 - 215, .
3. Rivka, L. G. and S. Edelstein, (1996) Calcium citrate : A revised look at calcium fortification. *J Food Tech.*, 50, 96 - 98.
4. Heaney, R. P. and N. J. Barger-Lux, (1991) Calcium in nutrition and prevention of disease. *Food Nutr. News*, 63, 7 - 10.
5. , (1989) .
6. , (1991) .
7. Bronner, F., (1990) 'Intracellular calcium regulation' 2nd Ed., Wiley-Liss, New York, USA.
8. Smith, K. T., (1988) Calcium and trace mineral interactions. *Cereal Foods World*, 33, 776 - 777.
9. Lucinda L. W. and D. A. McCARRON, (1986) Dietary calcium : An assessment of its protective action in human and experimental hypertension. *J. food Tech.*, 40, 93 - 95.
10. , , (1995) ' ' 80 - 86, 681 - 686, .
11. , , (1996) ' ' 166 - 182, .
12. , , , (1974) Texturometer texture . *Korean J. Food Sci. Tech.*, 6, 42 - 54.

13. Finkowski, J. W. and Peleg, M., (1980) Some rheological characteristics of soy extrudates in tension. *J. Food Sci.*, 45, 200-207.
14. Moskowitz, H. R., (1987) 'Food texture' 1st Ed., 333-335, Marcel Dekker, New York, USA.
15. Williams, M., (1989) 'Foods' 2nd Ed., 435, Macmillan, New York, USA.
16. , , (1988) . *J. Korean Soc. Food Nutr.*, 17, 25-31.
17. 中嶋初吉, (1993) 鶏卵を原材料とした健康食品製造法. 日本特許庁 特開平 5-76313.
18. , , (1996) . *Korean J. Food Sci. Tech.*, 28, 995-1000.
19. 小畑紘一, (1981) 鶏卵殻の灰化物ならびにえの生肉の接着剤および麵類のゆでのび防止剤としての用途. 日本特許庁 昭 56-96681
20. , (1994) . , 27, 33-42.
21. Smith, K. T. and S. M. Henders, (1987) Calcium absorption from a new calcium delivery system(CC). *Calcif. Tissues*
22. Boyle, E. A. E., P. B. Addis and R. J. Epley, (1994) Calcium fortified, reduced fat beef emulsion product. *J. Food Sci.*, 59, 928-932.
25. Vidal, S. and F. Z. Saleeb, (1993) Calcium citrate anticaking agent. US patent 5, 208, 372.
26. 猪股哲二, (1984) 卵殻の処理方法. 日本特許庁 昭59-71667.

6 .

1

1

1.

가 2000

2300

.

, ,  
가

(Potts 1973, 1994).

가 가 가 가 ‘ ,

1970 (Wei & Nikolov 1992).

30

.

가

.

가

가 ,

.

가 .

( 1997),

starch/polymer interface

(Maddever 1989).

Griffin(1974) , maize, , , , arrowroot

polyethylene polyethylene

가 . Otey (1977, 1980, 1987)

EAA(ethylene-co-acrylic acid)

, polyethylene silian

octenyl succinate group polyethylene

. polyethylene

oxidized polyethylene 가 .

, esterification, etherification,  
oxidation (Loomis 1993).

(Johnson 1987).

PVA (polyvinyl alcohol)

(Otey 1977), Swanson (1988)

가

가 .

2.

가 ( & 1994).

가 .

가

.

가

가

가

.

가

가

.

가

.

,

가 .



## 2

### 1

1.

1 kg

3

가

( 1984)

40

48

100

2.

가. 가

가

Jane (1992)

100 g

166 mL

2

0. 1%, 0. 5%, 1%, 2% epichlorohydrin 가

1M NaOH

가

pH

10. 5

24

가

acetic acid

pH 5. 5

Whatman No. 2

2 , 95% 1 가  
40 48 100 .

Wootton Manatsathit (1983)  
40  
100 .

3.

가. 가

epi chlorhydrin 가  
Hamerstrand (1960) 가  
(anhydroglucose units per crosslink, AGU/CL) 가 (molar  
degree of crosslinking, MDC) .

Johnson (1969)  
ninhydrin .

4.

가.

(Model whiteness checker RF-1, Nippon  
Denshoku Kogyo Co., Japan) L, a, b .

L 92.5, a 0.7, b 3.0 .

.

Medcal f Gilles(1965) .

. 가

Gilbert Spragg(1964) 가 .

.

(Differential Scanning Calorimeter, Rheometric  
Scientific SP+, England) (1997)

.

.

(1997) 0.2% .

Schoch(1964)

(1997)

X-

X- (Philips, X'pert PW3710, Nedalland)

target: Cu-k , scanning speed:  $0.04^\circ 2 /s$ , voltage: 30kV,  
current: 20mA  $2 : 5-40^\circ$

(Scanning electron microscope,

Hitachi S-4200, Japan)

700

## 1. Starch-Polyethylene

가 5% 10%  
 linear low density polyethylene, prooxidant (IR1025, Novon  
 International, INC., NY, USA) (Table 1, Table  
 2). 가

0.3% , polyethylene  
 150 , 50 rpm kneader (Haake Rheonex 3000) kneading  
 50% master batch . Haake Rheocord 90 (Germany)  
 single screw extruder (Rheonex 254) barrel 150 ,  
 145 , 150 , 150 , screw speed 20 rpm 가  
 5% 10% 50% master  
 batch polyethylene prooxidant 0.5cm  
 . single screw extruder barrel 120 ,  
 140 , 150 , 150 , screw speed 55 rpm 10 cast

Fig. 2 .

2. Starch-Polyethylene film

(Model whiteness checker RF-1, Nippon

Denshoku Kogyo Co., Japan) L, a, b .

(standard plate) L 92.5, a 0.7, b 3.0 .

3. Starch-polyethylene

가

0.12mm 가 1cm × 3cm strip

Instron(Shinadzu, Japan) load cell: 50 kgf, load range:

5, speed: 100mm/min tensile strength, percent

elongation, strain energy . Instron

50% 40

10 .

**Table 1. Composition of the crosslinked potato starch-poly  
-ethylene films**



Type of film	Concentration of Propylene oxide (%)	Starch content (g)	Prooxidant <sup>1)</sup> (g)	Polyethylene (g)
5% - Native/PE	0	50	50	900
10% - Native/PE	0	100	50	850
5% - 0.1 CL/PE	2.5	50	50	900
10% - 0.1 CL/PE	2.5	↓ 100	50	850
5% - 0.5 CL/PE	5.0	50	50	900
10% - 0.5 CL/PE	5.0	100	50	850
5% - 1.0 CL/PE	7.5	50	50	900
10% - 1.0 CL/PE	7.5	100	50	850
5% - 2.0 CL/PE	10.0	50	50	900
10% - 2.0 CL/PE	10.0	100	50	850

<sup>1)</sup>Prooxidant contained native starch(10%), unsaturates content(8.0%), and transition metal compounds(0.2%) in linear low density polyethylene

**Table 2. Composition of the hydroxypropylated potato starch- polyethylene films**

Type of film	Concentration of Propylene oxide (%)	Starch content (g)	Prooxidant <sup>1)</sup> (g)	Polyethylene (g)
5% - Native/PE	0	50	50	900
10% - Native/PE	0	100	50	850
5% - 2.5HP/PE	2.5	50	50	900
10% - 2.5HP/PE	2.5	100	50	850
5% - 5.0HP/PE	5.0	50	50	900
10% - 5.0HP/PE	5.0	100	50	850
5% - 7.5HP/PE	7.5	50	50	900
10% - 7.5HP/PE	7.5	100	50	850
5% - 10.0HP/PE	10.0	50	50	900
10% - 10.0HP/PE	10.0	100	50	850

<sup>1)</sup>Prooxidant contained native starch(10%), unsaturates content(8.0%), and transition metal compounds(0.2%) in linear low density polyethylene

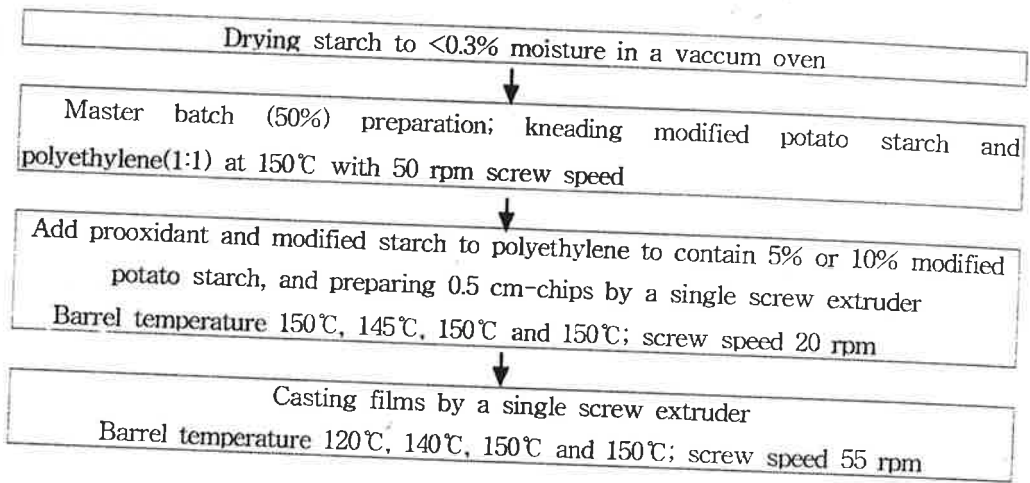


Fig. 2. Scheme of preparation for the modified potato starch-polyethylene films.



### 3 Starch-pol yethyl ene

1.

가.

1 × 3cm strip                      70   oven   12                      0. 12mm

. FT-IR

index(Kim 1994)                      FT- IR                      Carbonyl

Instron(Shinadzu, Japan)                      load  
cell: 50kgf, load range: 5, speed: 100mm/min                      tensile  
strength, percent elongation, strain energy

. Instron                      50%  
40                      4

2.

가. chemical disinfection

3cm , Universal disinfection solution 가 1x  
 1 2 . 1 L  
 1 . 95% 70% ,

. *Pseudomonas aeruginosa*

(1) nutrient broth가

35 , 100 rpm

*Pseudomonas aeruginosa*(KCTC 2651) 35 incubator 100  
 rpm 4 . *Pseudomonas*  
*aeruginosa* .

. FT-IR

4  
 , 70% 30 .  
 petri dish 45 8 .

FT-IR spectrum  
(Kim 1994).

Carbonyl index    Hydroxyl index

.

Instron

.

.

(Scanning electron

microscope, Hitachi S-4200, Japan)

2,000

.

### 3

#### 1

1.

가. 가

epi chlorohydrin 가 75% 91%  
85% (Table 3). epi chlorohydrin 85%  
가 Hollinger (1974) .  
epi chlorohydrin glycerol mono- diethers  
가  
. 가 가  
(Kasensuwan & Jane 1994).  
가 epi chlorohydrin 가  
( $r^2=0.9992$ ).

Propylene oxide 가 0, 2.5, 5.0, 7.5, 10.0%  
0, 0.0485, 0.1092, 0.1300, 0.2645 propylene oxide 가  
가 가 가 ( $r^2 = 0.93$ ) (Table 4).  
( 1991)

, propylene oxide

가

**Table 3. Degree of crosslinking and reaction yield in the crosslinked potato starches**

	Type of starch			
	0.1 CL	0.5 CL	1.0 CL	2.0 CL
Epichlorohydrin applied (g per 100 g of starch)	0.1	0.5	1.0	2.0
Unreacted epichlorohydrin (g per 100 g of starch)	0.02469	0.05935	0.14972	0.16439
Reacted epichlorohydrin (g per 100 g of starch)	0.07531	0.44065	0.85028	1.83561
Reacted epichlorohydrin (mole/ 100 g starch)	0.0008135	0.004760	0.009185	0.019829
Reaction yield(%)	75.31	88.13	85.03	91.78
AGU <sup>1)</sup> /crosslink	732	124	64	30
Crosslink/100AGU (MDC <sup>2)</sup> )	0.137	0.807	1.563	3.333

1) Anhydroglucose unit 2) Molar degree of crosslinking

**Table 4. Determination of hydroxypropyl group and degree of substitution in the hydroxypropylated potato starches**

Type of Starch	Propylene oxide concentration(%)	H.P <sup>1)</sup> (%)	DS <sup>2)</sup>
2.5 HP	2.5	1.7088	0.0485
5.0 HP	5.0	3.7625	0.1092
7.5 HP	7.5	4.4486	0.1300
10.0 HP	10.0	9.2363	0.2645

1)H.P : hydroxypropyl group 2)DS : degree of substitution

2.

가.

가

L, a

가 , b

(Table 5).

가

가

가

가

.

L

가

가

(Table 6).

a, b

가

.

.

가

가 가 가

가

(Table 5).

(1996)

가

가 가

가

가

가

.

(Lim & Seib

1993)

가

1980). 가 (Rasper  
 가 가 가

DS 0.2645

(Table 6).

. 가

가

가 가

가 가

(Table 5, Table 6).

가가

가

가

**Table 5. Some physicochemical properties of the crosslinked potato starches**

Property	Type of starch					
	Native	0.1 CL	0.5 CL	1.0 CL	2.0 CL	
Color)	L	93.2	93.1	93.3	93.9	93.5
	a	1.03	0.97	1.00	1.00	1.00
	b	2.27	2.07	2.13	2.13	2.03
Water binding capacity(%)	165.0	184.0	186.2	187.0	191.5	
Blue value	0.6741	0.0266	0.0092	0.0072	0.0068	

가

Table 7 . DSC thernogram

53.3 가

**Table 6. Some physicochemical properties of the hydroxypropylated potato starches**

Property	Type of starch					
	Native	2.5 HP	5.0 HP	7.5 HP	10.0 HP	
Color <sup>1)</sup>	L	92.1	93.5	93.5	93.3	93.8
	a	1.3	1.5	1.5	1.2	1.2
	b	1.7	1.3	1.5	1.2	1.2
Water binding capacity(%)	181.2	195.0	210.0	220.0	339.2	
Blue value	0.3986	0.3056	0.2814	0.2496	0.2230	

<sup>1)</sup>DS: degree of substitution

55.63 56.24 ,  
 59.31 , 가 60.80 61.32  
 가  
 , 가 .  
 Kartha (1985) 가 가  
 가

**Table 8**

. DSC thermogram  
 58 63 . 52 59  
 2.08 4.05 kcal/ng ,  
 ,  
 가 . Wootton (1984) .



**Table 7. DSC characteristics of crosslinked potato starches**

Variety	Type of starch				
	Native	0.1 CL	0.5 CL	1.0 CL	2.0 CL
TCI( )	53.3	55.97	55.63	56.24	56.02
TF2( )	59.31	61.32	60.80	61.16	61.23
HC3(mcal/mg)	3.46	3.68	3.02	2.97	3.31

1) TC: Onset temperature, 2) TF: Peak temperature, 3) HC: Enthalpy of gelatinization

**Table 8. DSC characteristics of hydroxypropylated potato starches**

Variety	Type of starch				
	Native	2.5 HP	5.0 HP	7.5 HP	10.0 HP
TCI( )	53.3	54.41	53.29	52.56	52.94
TF2( )	59.31	59.07	58.90	58.06	58.01
HC3(mcal/mg)	3.46	3.02	2.08	2.85	2.73

1) DS: degree of substitution, 2) TC: Onset temperature, 3) TF: Peak temperature  
4) HC: Enthalpy of gelatinization

epi chlorohydrin 0.5% 가 50 60  
 가 가 , 가 60  
 가 가 60 65  
 가 (Fig. 2). 가 가  
 . 2.5 HP, 5.0 HP 40 50  
 가 가 50 가  
 가 (Fig. 3). 7.5 HP, 10.0 HP 50

가 가 .  
가 가 .

가  
가 가 가 (Fig. 4).

60 가가 가 60 가  
가 , epichlorohydrin 0.1%, 0.5% 가 가  
70 가 가

. Epichlorohydrin 1%, 2% 가 가  
가가 . 가 가  
가 가  
가

가 가 ,  
Kasemsuwan (1994) 가 가  
가 . 가

가 가 가 (Fig. 5).

가가  
(Fig. 6). Fig. 7

. 50 가 60  
가 ,  
가 가 .

가

가

(Fig. 8).

가

가 가

(1996)

Kartha (1985)

가

가

chain

(A) 620nm

610nm

(Fig. 9).

가 가

가

El - Hi mnawy (1982)

가

가

. X-

가

2 5. 59, 14. 4, 17. 2,

22. 2

가

Zobel (1964)

B

, 가

2

14. 4, 17. 2 17. 5, 22, 24

가

(Fig. 10).

가

X-

. Koni ya(1986)

가 가

가

(Table 9).

2 5. 7, 14. 1 14. 4, 17. 1 17. 4, 22. 1 22. 6 가

B (Fig. 11). 2

5. 7, 17. 1 17. 4, 22, 24 가

가 (Table 10). French(1972)

Robin(1974)

X-

가

Fig. 12

. 가

가 . 가

(Kantha, 1985)

2% epi chlorohydrin 가 가

가 가 . 가

가 , 2. 5 HP(B)

5. 0 HP(C) 가

10. 0 HP(E) 가

(Fig. 13).

가

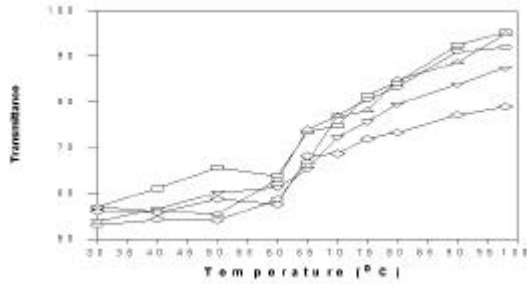


Fig. 2. Transmittance of 0.2% pastes of crosslinked potato starches. ○, native starch; □, 0.1 CL; ▽, 0.5 CL; △, 1.0 CL; ◇, 2.0 CL

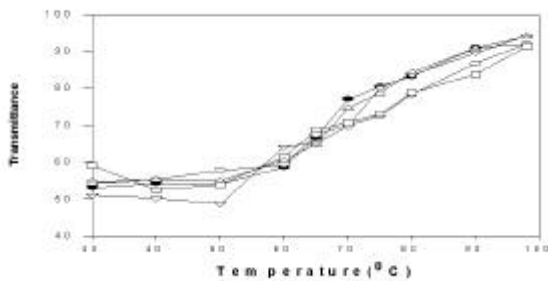


Fig. 3. Transmittance of 0.2% pastes of hydroxypropylated potato starches. ○, native starch; □, 2.5 HP; ▽, 5.0 HP; △, 7.5 HP; ◇, 10.0 HP

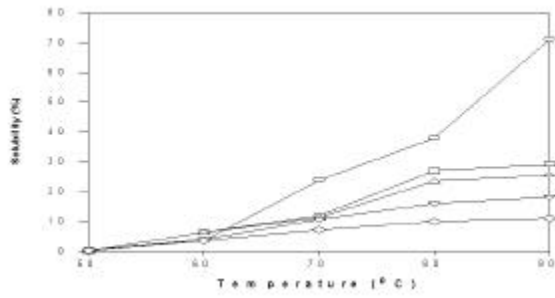


Fig. 4. Solubility of the crosslinked potato starches.

○, native starch; □, 0.1 CL; ▽, 0.5 CL; △, 1.0 CL; ◇, 2.0 CL

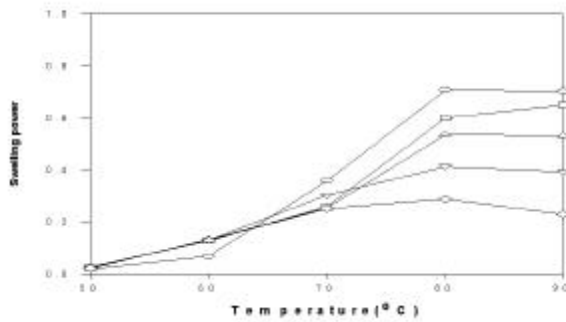


Fig. 5. Swelling power of the crosslinked potato starches.

○, native starch; □, 0.1 CL; ▽, 0.5 CL; △, 1.0 CL; ◇, 2.0 CL

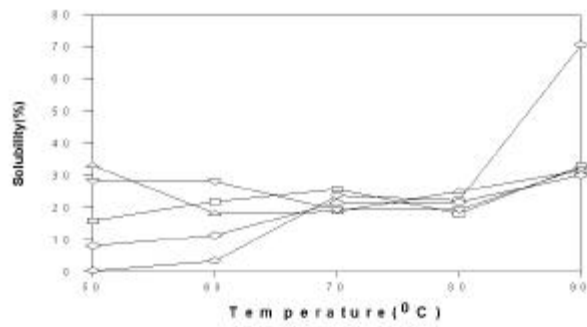


Fig. 6. Solubility of the hydroxypropylated potato starches.

○, native starch; □, 2.5 HP; ▽, 5.0 HP; △, 7.5 HP; ◇, 10.0 HP

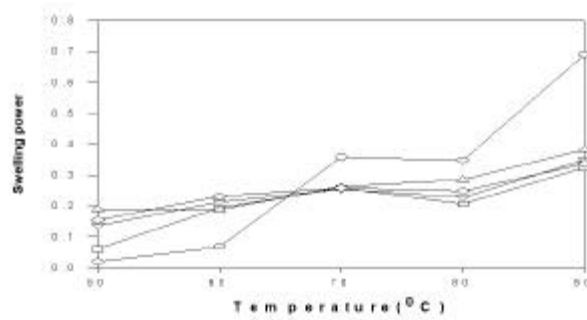


Fig. 7. Swelling power of the hydroxypropylated potato starches. ○, native starch; □, 2.5 HP; ▽, 5.0 HP; △, 7.5 HP; ◇, 10.0 HP

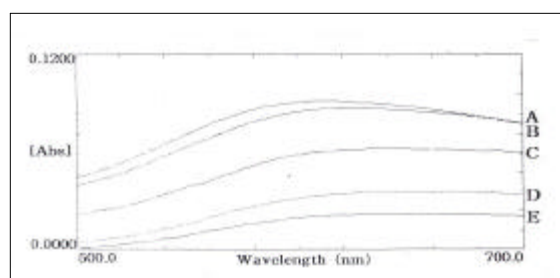


Fig. 8. Iodine absorption spectrum of crosslinked potato starches. A, Native ; B, 0.1 CL; C, 0.5 CL; D, 1.0 CL; E, 2.0 CL

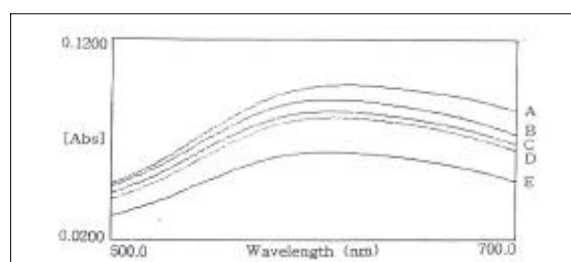


Fig. 9. Iodine absorption spectrum of hydroxypropylated potato starches. A, Native ; B, 2.5 HP; C, 5.0 HP; D, 7.5 HP; E, 10.0 HP

Table 9. Relative crystallinity of crosslinked potato starches

Property	Type of starch				
	native	0.1 CL	0.5 CL	1.0 CL	2.0 CL
Crystallinity (%) (Ac1)/(Ac+Aa2))	39.59	38.15	39.39	39.68	38.11

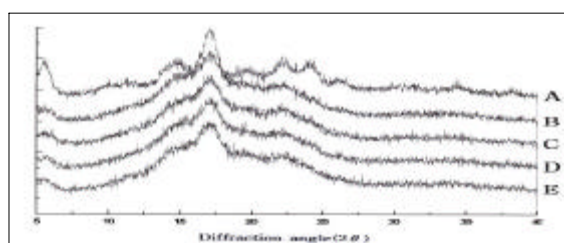
1) Ac: crystalline area 2) Aa: amorphous area



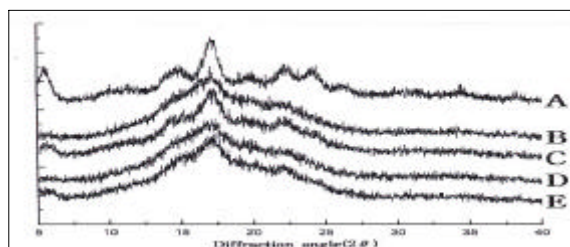
**Table 10. Relative crystallinity of hydroxypropylated potato starches**

Property	Type of starch				
	Native	2.5 HP	5.0 HP	7.5 HP	10.0 HP
Crystallinity (%) (Ac1)/(Ac+Aa2))	39.59	36.67	34.30	32.90	32.79

1) Ac: crystalline area    2) Aa: amorphous area



**Fig. 10. X-ray diffraction patterns of crosslinked potato starches.**  
A, Native ; B, 0.1 CL; C, 0.5 CL; D, 1.0 CL; E, 2.0 CL



**Fig. 11. X-ray diffraction patterns of hydroxypropylated potato starches.**  
A, Native ; B, 2.5 HP; C, 5.0 HP; D, 7.5 HP; E, 10.0 HP

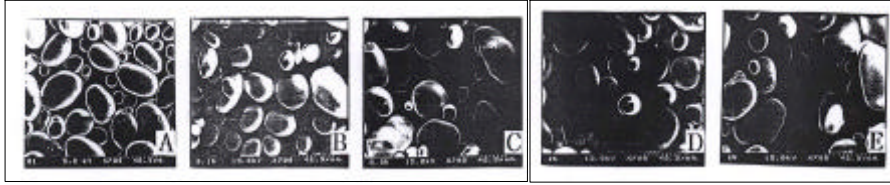


Fig. 12. Scanning electron micrographs( $\times 700$ ) of crosslinked potato starches. A, Native; B, 0.1 CL; C, 0.5 CL; D, 1.0 CL; E, 2.0 CL

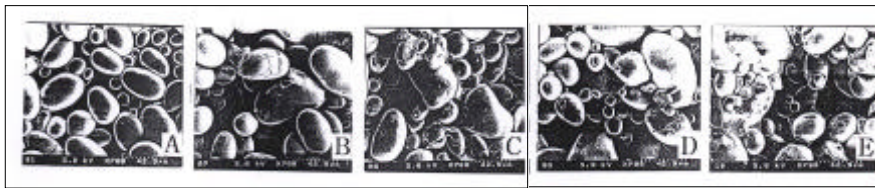


Fig. 13. Scanning electron micrographs( $\times 700$ ) of hydroxypropylated potato starches. A, Native; B, 2.5 HP; C, 5.0 HP; D, 7.5 HP; E, 10.0 HP

## 2 Starch-pol yethyl ene

### 1. Starch- pol yethyl ene

가 5%(wt) 가 L, b

가 a 2.0 CL 가

(Table 11, Table 12). 가

10%(wt) 가 a, b

가 L 가

가

5%(wt)

a, b

가

L

가

(Table 13, Table 14).

10%(wt)

a, b

가

L

10.0 HP

가

Table 11. Color properties of the crosslinked potato starch(5%)-polyethylene films

Type of film	L	a	b
5% - Native/PE	23.53 ± 0.90	- 0.07 ± 0.06tc	- 3.23 ± 0.38
5% - 0.1 CL/PE	23.63 ± 1.42	- 1.23 ± 0.45c	- 3.53 ± 0.64
5% - 0.5 CL/PE	24.67 ± 0.93	0.73 ± 0.72ab	- 4.37 ± 1.05
5% - 1.0 CL/PE	26.03 ± 0.12	- 0.83 ± 0.59tc	- 2.77 ± 0.75
5% - 2.0 CL/PE	25.07 ± 1.20	2.03 ± 1.64a	- 2.03 ± 2.93

Means ± SEM, Each value is mean for three replicates.

Means with different superscript within the same column are significantly different (p<0.05).

## 2. Starch- polyethylene

가

5%(wt)

tensile strength

1.0 CL

가

, percent

elongation strain energy 가

(Table

**Table 12. Color properties of the crosslinked potato starch(10%)- polyethylene films**

Type of film	L	a	b
10% - Native/PE	27.33 ± 0.38c	0.30 ± 1.23	- 3.30 ± 0.87
10% - 0.1 CL/PE	28.70 ± 1.04b	- 0.20 ± 2.36	- 4.07 ± 0.74
10% - 0.5 CL/PE	31.80 ± 0.53a	- 2.77 ± 1.00	- 3.80 ± 0.36
10% - 1.0 CL/PE	28.17 ± 0.47tc	- 2.47 ± 1.78	- 3.90 ± 0.56
10% - 2.0 CL/PE	29.33 ± 0.57b	2.23 ± 2.87	- 5.33 ± 0.97

Means ± SEM. Each value is mean for three replicates.

Means with different superscript within the same column are significantly different (p<0.05).

**Table 13. Color properties of the hydroxypropylated potato starch (5%)- polyethylene films**

Type of film	L	a	b
5% - Native/PE	23.53 ± 0.89a	- 0.07 ± 0.06	- 3.23 ± 0.38
5% - 2.5 HP/PE	21.07 ± 1.46b	- 0.93 ± 2.11	- 2.03 ± 1.42
5% - 5.0 HP/PE	21.93 ± 0.70ab	- 0.23 ± 1.78	- 4.57 ± 0.90
5% - 7.5 HP/PE	20.43 ± 1.29b	- 3.80 ± 1.4	- 4.03 ± 1.55
5% - 10.0 HP/PE	21.37 ± 0.64b	- 0.80 ± 0.87	- 2.93 ± 0.46

Means ± SEM. Each value is mean for three replicates.

Means with different superscript within the same column are significantly different (p<0.05).

**Table 14. Color properties of the hydroxypropylated potato starch (10%)- polyethylene films**

Type of film	L	a	b
10% - Native/PE	27.33 ± 0.38ab	0.30 ± 1.23	- 3.30 ± 0.87
10% - 2.5 HP/PE	27.80 ± 1.56a	- 1.00 ± 6.58	- 3.67 ± 0.93
10% - 5.0 HP/PE	28.90 ± 0.46a	4.47 ± 1.95	- 3.03 ± 0.50
10% - 7.5 HP/PE	28.03 ± 1.07a	- 1.97 ± 1.07	- 2.67 ± 0.25
10% - 10.0 HP/PE	25.77 ± 0.59b	- 1.17 ± 0.42	- 3.33 ± 1.27

Means ± SEM. Each value is mean for three replicates.

Means with different superscript within the same column are significantly different (p<0.05).

15, Table 16). 가 10%(wt)  
 tensile strength 0.5 CL/PE 가  
 strain energy 가  
 가 가  
 5%(wt) 10.0 HP  
 가 (Table 17,  
 Table 18).

**Table 15. Mechanical properties of the crosslinked potato starch (5%)- polyethylene films**

Type of film	Tensile strength (kgf/mm <sup>2</sup> )	Percentage elongation (%)	Strain energy (kgf- mm)
5% - Native/PE	1.0253 ± 0.04b	200.42 ± 19.71c	62.979 ± 8.34b
5% - 0.1 CL/PE	1.2320 ± 0.03ab	230.62 ± 32.78ab	89.975 ± 14.28a
5% - 0.5 CL/PE	1.0492 ± 0.03b	252.95 ± 35.78a	87.125 ± 13.41a
5% - 1.0 CL/PE	1.5831 ± 0.94a	217.11 ± 20.07bc	87.624 ± 8.77a
5% - 2.0 CL/PE	1.304 ± 0.05ab	227.35 ± 21.55b	91.418 ± 10.10a

Means ± SEM, Each value is mean for ten replicates. Means with different superscript within the same column are significantly different (p<0.05).

**Table 16. Mechanical properties of the crosslinked potatostarch (10%)- polyethylene films**

Type of film	Tensile strength (kgf/mm <sup>2</sup> )	Percentage elongation (%)	Strain energy (kgf- mm)
10% - Native/PE	1.0061 ± 0.05d	175.73 ± 16.00	52.203 ± 5.38c
10% - 0.1 CL/PE	1.0447 ± 0.04c	183.37 ± 10.90	59.350 ± 5.67tc
10% - 0.5 CL/PE	0.9159 ± 0.03e	191.21 ± 32.51	56.762 ± 10.90bc
10% - 1.0 CL/PE	1.2201 ± 0.04a	185.54 ± 23.45	70.476 ± 10.42a
10% - 2.0 CL/PE	1.1663 ± 0.03b	175.89 ± 13.22	63.958 ± 5.73ab

Means ± SEM, Each value is mean for ten replicates.  $\alpha$ -Means with different superscript within the same column are significantly different (p<0.05).

**Table 17. Mechanical properties of the hydroxypropylated potato starch(5%) -polyethylene films**

Type of film	Tensile strength (kgf/mm <sup>2</sup> )	Percentage elongation (%)	Strain energy (kgf- mm)
5% - Native/PE	1.0253 ± 0.04d	200.42 ± 19.71c	62.98 ± 8.33c
5% - 2.5 HP/PE	1.5385 ± 0.06a	235.37 ± 23.85b	107.53 ± 13.38a
5% - 5.0 HP/PE	1.3988 ± 0.07b	259.81 ± 26.83a	110.59 ± 14.48a
5% - 7.5 HP/PE	1.4995 ± 0.04a	200.70 ± 23.88c	88.68 ± 12.52b
5% - 10.0HP/PE	1.1082 ± 0.02c	121.78 ± 14.19d	37.12 ± 5.54d

Means ± SEM, Each value is mean for ten replicates.  $\alpha$ -Means with different superscript within the same column are significantly different (p<0.05).

**Table 18. Mechanical properties of the hydroxypropylated potato starch (10%)-polyethylene films**

Type of film	Tensile strength (kgf/mm <sup>2</sup> )	Percentage elongation (%)	Strain energy (kgf- mm)
10% - Native/PE	1.0061 ± 0.05d	175.73 ± 15.99a	52.20 ± 5.38b
10% - 2.5 HP/PE	1.2358 ± 0.04b	157.63 ± 14.30b	55.53 ± 6.90b
10% - 5.0 HP/PE	1.1671 ± 0.05c	160.24 ± 22.47b	53.89 ± 10.01b
10% - 7.5 HP/PE	1.4588 ± 0.06a	169.60 ± 9.17ab	71.28 ± 6.72a
10% - 10.0 HP/PE	0.9631 ± 0.05d	96.16 ± 8.78c	24.30 ± 3.40c

Means ± SEM, Each value is mean for ten replicates.  $\alpha$ -Means with different superscript within the same column are significantly different (p<0.05).

### 3 Starch-polyethylene

#### 1. Starch-polyethylene

##### 가. FT-IR

Starch-PE film ketone, aldehyde carbonyl  
 compounds가 FI-IR spectrum starch-PE film  
 (Wei & Nikolov 1992). aldehyde carbonyl, ketone carbonyl (1705 1740cm-1)  
 (Fig. 14). 가 5%(wt)  
 12 carbonyl index  
 가 , 0.1 CL 4  
 carbonyl index가 가 , 0.5 CL 2.0 CL  
 5 1.0 CL 6  
 carbonyl index가 가 (Fig. 15). 가  
 10%(wt) 가  
 8 carbonyl index가 가 , 0.1 CL  
 4 carbonyl index가 가  
 0.5 CL 2.0 CL 5 1.0  
 CL 6 carbonyl index가 가  
 (Fig. 15). 가  
 가

carbonyl index가 5%(wt) 10%(wt) 가

carbonyl index가 가

10.0 HP 7.5 HP 5%(wt) 7 ,

5.0 HP 8 , 2.5 HP

10 carbonyl index가 가 (Fig. 16) .

가

carbonyl index가 .

10%(wt)

8 carbonyl index가 가 , 7.5 HP

10.0 HP 6 가 7

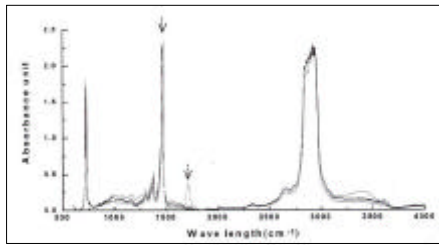
가 , 2.5 HP 5.0 HP 8

carbonyl index가 가 가

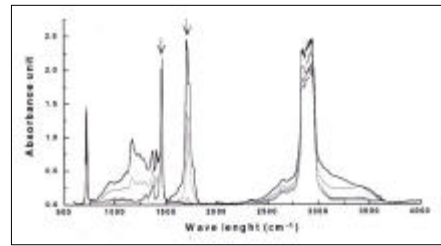
carbonyl index가

가 (Fig. 16).





(a)



(b)

Fig. 14. FT-IR spectra of starch-polyethylene films by the heat treatment at 70 for 12 weeks. (a) 10%-Native/PE film, (b) 10%- 10.0 HP/PE film. For each set of spectra the top-to bottom sequence is 12, 8, 7, 1, 0 week.

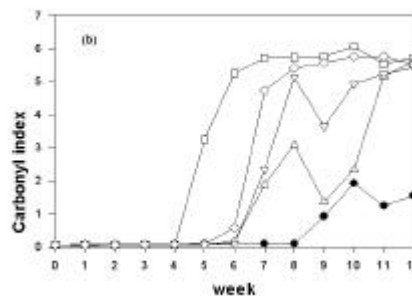
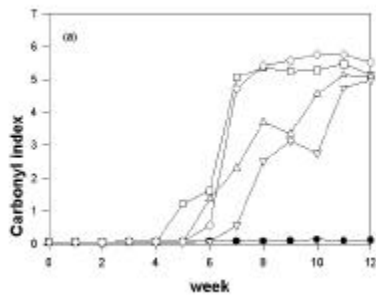


Fig. 15. Carbonyl index of crosslinked potato starch-polyethylene films by the heat treatment at 70 for 12 weeks. (a): CL(5% wt)/PE films, (b): CL(10% wt)/PE films. , Native/PE ; , 0.1 CL/PE ; , 0.5 CL/PE ; , 1.0 CL/PE ; , 2.0 CL/PE

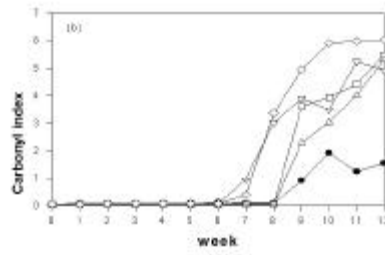
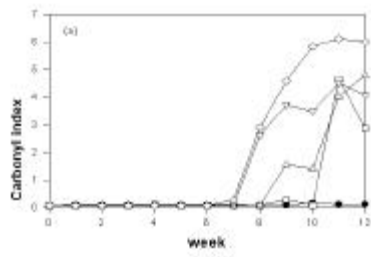


Fig.

16. Carbonyl index of hydroxypropylated potato starch-polyethylene films by the heat treatment at 70 for 12 weeks. (a): HP(5% wt)/PE films, (b): HP(10% wt)/PE films. , Native/PE ; , 2.5 HP/PE ; , 5.0 HP/PE ; , 7.5 HP/PE ; , 10.0 HP/PE

70

FT-IR

spectrum carbonyl index . 가

5%(wt)

tensile strength, percent elongation, strain energy

11 , 0.1 CL , 2.0 CL

5 , 0.5 CL 1.0 CL 7

( $p < 0.05$ ) (Table 19, Table 20,

Table 21). 가 10%(wt)

가 9

(Table 22, Table 23, Table 24), 0.1 CL

5 , 0.5 CL 1.0 CL 2.0 CL

6 tensile strength, percent elongation, strain

energy 가 ( $p < 0.05$ ). 가

. 0.1 CL 가  
 .  
 5%(wt) ,  
 tensile strength, percent elongation, energy 11  
 (p<0.05) (Table 25, Table 26, Table 27). 7.5 HP  
 10.0 HP 7  
 9  
 . 2.5 HP 5.0  
 HP 9 .  
 10%(wt)  
 5%(wt)  
 (Table 28, Table 29, Table 30).

**Table 19. Tensile strength of the crosslinked potato starch(5%)-polyethylene films by the heat treatment at 70 for 12 weeks**

Week	Tensile strength(kgf/mm <sup>2</sup> )				
	Type of film				
	Native/PE	0.1 CL/PE	0.5 CL/PE	1.0 CL/PE	2.0 CL/PE
0	C1.049a	E1.243a	C1.077a	A1.296a	A1.320a
1	A1.114a	A1.135ab	E0.942b	A1.161ab	A1.113b
2	C1.041a	E1.149ab	E0.952b	A1.222ab	EC1.098b
3	A1.071a	AE1.056b	C0.870b	AE1.069b	E1.032b
4	C1.031a	E1.112b	E0.895b	A1.200ab	EC1.077b
5	E0.978a	E0.873c	E0.878b	A1.124ab	E0.895c
6	E0.933a	C0.619d	E0.862b	A1.162ab	E0.853cd
7	A0.989a	-	EC0.702cd	AE0.887c	C0.590e
8	1.041a	-	0.757c	0.856c	0.790d
9	0.956a	-	0.728cd	0.782cd	-
10	0.969a	-	0.623d	0.647d	-
11	0.572b	-	-	-	-
12	0.136c	-	-	-	-

Each value is mean for four replicates. Means with different superscript are significantly different ( $p < 0.05$ ), *A-D* : Duncan's multiple range test for starch/PE film (column), *a-e* : Duncan's multiple range test for week (row)

**Table 20. Percent elongation of the crosslinked potato starch(5%)-polyethylene films by the heat treatment at 70 for 12 weeks**

Week	Percent elongation(%) Type of film				
	Native/PE	0.1 CL/PE	0.5 CL/PE	1.0 CL/PE	2.0 CL/PE
0	C210.5a	E253.1a	A279.8a	C227.4a	EC231.7a
1	197.2a	186.1b	211.2b	196.0bc	184.5tc
2	198.2a	192.6b	179.9b	210.9ab	181.7tc
3	183.6a	192.2b	188.5b	171.6c	199.2b
4	178.7a	206.0b	196.0b	193.1bc	168.4c
5	A155.3a	E46.8c	A184.3b	A174.8c	E59.0d
6	AE150.0a	C3.3d	E121.1c	A200.7abc	C15.3e
7	A148.2a	-	E22.1d	E30.7d	E3.6e
8	175.0a	-	7.6d	9.4d	6.4e
9	A157.5a	-	E4.4d	E5.9d	-
10	A145.1a	-	E3.1d	E5.6d	-
11	19.9b	-	-	-	-
12	3.5b	-	-	-	-

Descriptions for this table are the same as above.

**Table 21. Strain energy of the crosslinked potato starch(5%)- polyethylene films by the heat treatment at 70 for 12 weeks**

Week	Strain energy(kgf- mm)				
	Type of film				
	Native/PE	0.1 CL/PE	0.5 CL/PE	1.0 CL/PE	2.0 CL/PE
0	E67.5a	A99.3a	A97.8a	A92.5a	A95.8a
1	69.9a	68.0b	66.9b	73.2tc	65.0b
2	E67.0a	AE72.2b	E57.7b	A83.0ab	E63.7b
3	61.7a	65.7b	55.0tc	58.3d	65.5b
4	E58.5a	A73.7b	E58.9b	A74.3tc	E57.6b
5	A48.3a	E14.5c	A54.2tc	A63.1cd	E17.0c
6	E43.6a	C0.4c	E36.0c	A74.5tc	C3.7d
7	A47.0a	-	E5.3d	E8.4e	E0.5d
8	59.8a	-	1.7d	2.0e	1.2d
9	A52.0a	-	E0.7d	E1.4e	-
10	A48.2a	-	E0.5d	E1.1e	-
11	3.5b	-	-	-	-
12	2.6b	-	-	-	-

Descriptions for this table are the same as above.

**Table 22. Tensile strength of the crosslinked potato starch(10%)- polyethylene films by the heat treatment at 70 for 12 week**

Week	Tensile strength(kgf/mm <sup>2</sup> )				
	Type of film				
	Native/PE	0.1 CL/PE	0.5 CL/PE	1.0 CL/PE	2.0 CL/PE
0	E1.002ab	C1.039a	E0.939a	A1.242a	E1.174a
1	E0.982ab	E0.956a	C0.860ab	A1.104b	A1.077abc
2	E0.992ab	E0.989a	C0.885ab	A1.133b	A1.110ab
3	A1.038ab	E0.939a	C0.782tc	A1.059tc	A1.034tc
4	AE1.053a	CI0.921a	E0.868ab	A1.110b	EC0.983c
5	A1.025ab	E0.276b	A0.837ab	A1.071tc	A0.984c
6	A1.009ab	E0.777a	E0.728cd	A0.985c	E0.719d
7	A0.934ab	-	E0.637d	A0.800d	-
8	0.911b	-	0.631d	0.767d	-
9	0.716c	-	0.665d	0.747d	-
10	0.693c	-	-	-	-
11	0.675c	-	-	-	-
12	0.616c	-	-	-	-

Descriptions for this table are the same as above.

**Table 23. Percent elongation of the crosslinked potato starch(10%)-polyethylene films by the heat treatment at 70 for 12 weeks**

Week	Percent elongation(%)				
	Type of film				
	Native/PE	0.1 CL/PE	0.5 CL/PE	1.0 CL/PE	2.0 CL/PE
0	179.4a	188.4a	210.4a	196.9a	172.3a
1	AE165.8ab	AEC154.9b	EC153.2b	A178.3a	C133.3ab
2	AE164.0ab	EC150.1b	EC146.4b	A177.8a	C135.7ab
3	A167.8ab	E137.2b	E139.2b	A180.5a	E141.0ab
4	A166.4ab	AEC157.5b	EC144.5b	A183.5a	C123.4b
5	A166.3ab	C4.5c	AE136.8b	A169.6a	E106.3b
6	A138.1ab	E9.2c	E26.4c	A98.2b	E17.1c
7	A147.2ab	-	E9.3c	E18.0c	-
8	120.9b	-	4.4c	6.5c	-
9	45.5c	-	7.2c	7.5c	-
10	10.6c	-	-	-	-
11	7.3c	-	-	-	-
12	4.5c	-	-	-	-

Descriptions for this table are the same as above.

**Table 24. Strain energy of the crosslinked potato starch(10%)-polyethylene films by the heat treatment at 70 for 12 weeks**

Week	Strain energy(kgf- mm)				
	Type of film				
	Native/PE	0.1 CL/PE	0.5 CL/PE	1.0 CL/PE	2.0 CL/PE
0	E53.4a	E60.9a	E63.7a	A76.1a	E62.7a
1	E49.7ab	E46.5b	E42.7b	A62.2b	E42.5b
2	E49.7ab	E46.8b	E42.4b	A64.2b	E45.3b
3	A53.5a	E40.1b	E35.0b	A60.4b	E43.8b
4	AE54.0a	EC45.3b	C41.0b	A64.9b	C38.1b
5	A53.0a	C0.9c	E37.0b	A57.4b	E32.6b
6	A42.6ab	E1.9c	E6.6c	A30.6c	E3.7c
7	A42.2ab	-	E2.0c	E4.3d	-
8	35.9b	-	0.6c	1.3d	-
9	11.1c	-	1.5c	1.7d	-
10	2.2c	-	-	-	-
11	1.3c	-	-	-	-
12	0.7c	-	-	-	-

Descriptions for this table are the same as above.

**Table 25. Tensile strength of the hydroxypropylated potato starch(5%)-polyethylene films by the heat treatment at 70 for 12 weeks**

Week	Tensile strength(kgf/mm <sup>2</sup> )				
	Type of film				
	Native/PE	2.5 HP/PE	5.0 HP/PE	7.5 HP/PE	10.0 HP/PE
0	E1.049a	A1.554ab	E1.426atc	A1.518tc	C1.112a
1	E1.114a	E1.635a	C1.534a	A1.807a	E1.182a
2	E1.041a	E1.522ab	E1.487ab	A1.620atc	C1.140a
3	C1.071a	E1.425tc	E1.494ab	A1.751ab	C1.122a
4	E1.031a	A1.513ab	A1.484ab	A1.563atc	E1.134a
5	E0.978a	E1.464atc	AE1.529a	A1.607atc	C1.122a
6	E0.933a	E1.415tc	E1.439atc	A1.674atc	C1.147a
7	E0.989a	A1.488atc	A1.392tc	A1.445c	C0.700b
8	E1.041a	A1.321c	A1.344c	C0.574d	C0.127c
9	0.956a	0.733d	0.907e	0.210d	-
10	0.969a	0.535d	1.051d	-	-
11	0.572b	0.332e	-	-	-
12	0.136c	-	-	-	-

Descriptions for this table are the same as above.

**Table 26. Percent elongation of the hydroxypropylated potato starch(5%)-polyethylene films by the heat treatment at 70 for 12 weeks**

Week	Percent elongation(%)				
	Type of film				
	Native/PE	2.5 HP/PE	5.0 HP/PE	7.5 HP/PE	10.0 HP/PE
0	C210.5a	E246.8a	A270.8a	C193.4a	E123.9a
1	A197.2a	AE173.3b	E161.4c	E155.9a	C104.6b
2	A198.2a	EC163.6b	AE188.3tc	C142.8a	E111.0b
3	A183.6a	AE169.9b	A182.6tc	EC143.0a	C110.0b
4	E178.7a	C149.4b	A207.6b	EC161.6a	E112.8b
5	A155.3a	A177.8b	A175.4tc	A166.9a	E101.3b
6	E150.0a	A173.9b	A179.6tc	E145.8a	C102.4b
7	A148.2a	A173.4b	A185.3tc	E80.9b	E29.1c
8	A175.0a	A167.1b	A161.3c	E3.3c	-
9	A157.5a	E14.0d	E9.9d	E1.5c	-
10	145.1a	27.7c	11.2d	-	-
11	19.9b	1.8d	-	-	-
12	3.5b	-	-	-	-

Descriptions for this table are the same as above.

**Table 27. Strain energy of the hydroxypropylated potato starch(5%)-polyethylene films by the heat treatment at 70 for 12 weeks**

Week	Strain energy(kgf- mm)				
	Type of film				
	Native/PE	2.5 HP/PE	5.0 HP/PE	7.5 HP/PE	10.0 HP/PE
0	C67.5a	A114.1a	A116.9a	E85.6a	I37.8a
1	E69.9a	A86.6b	AE77.8tc	A85.4a	C34.6ab
2	E67.0a	E74.7tc	A88.1tc	E69.1a	C34.6ab
3	E61.7a	AE73.9tc	A86.3tc	AE74.2a	C34.0ab
4	C58.5a	EC66.0c	A97.1b	E76.2a	I35.1ab
5	E48.3a	A78.5tc	A84.4tc	A81.1a	E31.0b
6	E43.6a	A74.6tc	A81.4tc	A72.3a	C31.8b
7	E47.0a	A77.0tc	A81.0tc	E38.3b	C6.1c
8	A59.8a	A66.5c	A67.9c	E0.4c	E0.1d
9	A52.0a	E4.0ce	E2.7d	E1.1c	-
10	48.2a	9.3d	7.7d	-	-
11	3.5b	0.1e	-	-	-
12	2.6b	-	-	-	-

Descriptions for this table are the same as above.

**Table 28. Tensile strength of the hydroxypropylated potato starch(10%)-polyethylene films by the heat treatment at 70 for 12 week**

Week	Tensile strength(kgf/mm <sup>2</sup> )				
	Type of film				
	Native/PE	2.5 HP/PE	5.0 HP/PE	7.5 HP/PE	10.0 HP/PE
0	I1.002ab	E1.253b	C1.200ab	A1.467b	I0.991a
1	C0.982ab	E1.309ab	E1.275ab	A1.672a	C1.040a
2	I0.992ab	E1.369ab	C1.221ab	A1.558ab	I0.988a
3	C1.038ab	E1.420a	E1.323a	A1.595ab	C0.937a
4	I1.053a	E1.362ab	C1.203ab	A1.483b	E0.900a
5	C1.025ab	AE1.333ab	EC1.179ab	A1.447b	I0.702b
6	C1.009ab	E1.257b	E1.191ab	A1.472b	I0.431tc
7	E0.934ab	A1.331ab	E1.112tc	E0.809c	C0.513c
8	0.911b	0.978c	0.878d	0.602d	-
9	0.716c	-	0.959cd	-	-
10	0.693c	-	-	-	-
11	0.675c	-	-	-	-
12	0.616c	-	-	-	-

Descriptions for this table are the same as above.



**Table 29.** Percent elongation of the hydroxypropylated potato starch (10%)-polyethylene films by the heat treatment at 70 for 12 weeks

Week	Percent elongation(%)				
	Type of film				
	Native/PE	2.5 HP/PE	5.0 HP/PE	7.5 HP/PE	10.0 HP/PE
0	A179.4a	A162.8a	A172.0a	A168.5a	E102.4a
1	A165.8ab	E123.3b	E125.7b	E133.2b	C68.6b
2	A164.0ab	C120.8b	EC127.3b	AE145.9b	D93.4a
3	A167.8ab	E129.2b	E130.8b	E134.0b	C84.8ab
4	A166.4ab	E121.2b	E126.2b	E132.5b	C83.6ab
5	A166.3ab	E127.4b	E119.7b	E126.6b	C37.4c
6	A138.1ab	A119.0b	A120.2b	A132.1b	E14.6cd
7	A147.2ab	AE135.6b	E105.5b	C30.7c	C9.9d
8	A120.9b	E38.6c	E49.7c	E4.5d	-
9	45.5c	-	20.6c	-	-
10	10.6c	-	-	-	-
11	7.3c	-	-	-	-
12	4.5c	-	-	-	-

Descriptions for this table are the same as above.

## 2. Starch- polyethyl ene

### 가. FT-IR

가 5%(wt) ,

*P. aeruginosa* (Fig.

17) carbonyl index hydroxyl index 가 , 가

1.0 CL/PE *P. aeruginosa*

carbonyl index hydroxyl index가

. 가 10%(wt) ,

carbonyl index *P. aeruginosa*

가 hydroxyl index *F. aeruginosa*

Table 30. Strain energy of the hydroxypropylated potato starch(10%)-polyethylene films by the heat treatment at 70 for 12 weeks

Week	Strain energy(kgf- mm)				
	Type of film				
	Native/PE	2.5 HP/PE	5.0 HP/PE	7.5 HP/PE	10.0 HP/PE
0	E53.4a	E58.4a	E59.5a	A71.2a	C26.8a
1	E49.7ab	E47.0ab	E47.4abc	A65.9ab	C22.5ab
2	E49.7ab	E47.2ab	E45.1tc	A66.8a	C24.3ab
3	A53.5a	A53.3ab	A51.1ab	A62.5abc	E21.6ab
4	A54.0a	E48.0ab	C44.2tc	A57.0tc	E19.4b
5	A53.0a	A54.8ab	E41.3tc	A53.4c	C8.7c
6	E42.6ab	E43.1b	E41.9tc	A57.3tc	C2.5cd
7	A54.2ab	A52.1ab	E35.9c	C10.1d	C1.4d
8	A35.9b	E12.0c	A514.4d	E1.6e	-
9	11.1c	-	4.9d	-	-
10	2.2c	-	-	-	-
11	1.3c	-	-	-	-
12	0.7c	-	-	-	-

Descriptions for this table are the same as above.

가 . 가

*F. aeruginosa*

carbonyl index

hydroxyl index

가

(Fig. 18).

5%(wt)

*F. aeruginosa*

carbonyl

index

hydroxyl index

가

.

, 2.5 HP/PE

5.0 HP/PE

*F. aeruginosa*

carbonyl index

hydroxyl index가

가

,

7.5 HP/PE,

10.0 HP/PE

*F. aeruginosa*

carbonyl index

hydroxyl index가

가

(Fig. 19).

10%(wt)

carbonyl index *F. aeruginosa* 가

hydroxyl index *F. aeruginosa* 가

가

(Fig. 20).

*F. aeruginosa* carbonyl index

hydroxyl index가 5.0 HP/PE

.

Starch/PE film

carbonyl hydroxyl (1, 2 alcohol)가

. *Streptomyces*

FI-IR spectrum 가 *Streptomyces*

*setonii* *Streptomyces viridosporus* 900 1, 200cm<sup>-1</sup>(1,

2 alcohol ) 가 (Ponetto 1992).

,

가

(Fig. 11 , Table 10). 가

가 *F. aeruginosa*가

가 .

Octenyl succinate octenyl

succinate 가

(Evangelista 1991).

, 가 ,

. Gage(1990)

가

.

10%(wt) carbonyl index hydroxyl index가 가

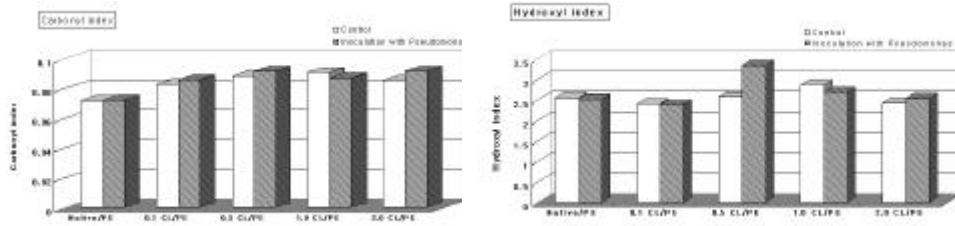


Fig. 17. Carbonyl index and hydroxyl index of the crosslinked potato starch(5%)-polyethylene films cultured with *Pseudomonas aeruginosa* for 4 weeks.

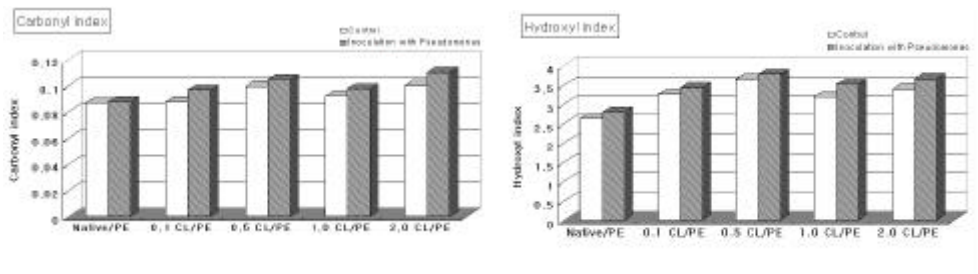


Fig. 18. Carbonyl index and hydroxyl index of the crosslinked potato starch(10%)-polyethylene films cultured with *Pseudomonas aeruginosa* for 4 weeks.

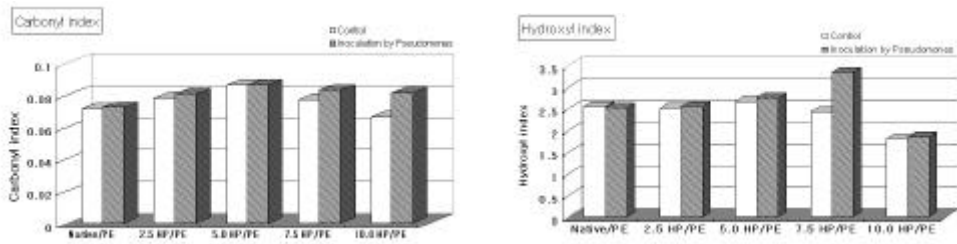


Fig. 19. Carbonyl index and hydroxyl index of the hydroxypropylated potato starch(5%)-polyethylene films cultured with *Pseudomonas aeruginosa* for 4 weeks.

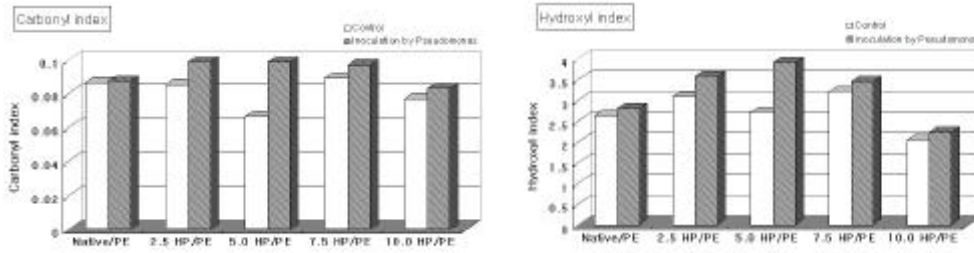


Fig. 20. Carbonyl index and hydroxyl index of the hydroxypropylated potato starch(10%)-polyethylene films cultured with *Pseudomonas aeruginosa* for 4 weeks.

*Pseudomonas aeruginosa*

*P. aeruginosa*

FT-IR spectrum

carbonyl index

hydroxyl index

가

5%(wt)

*P. aeruginosa*

tensile strength

strain energy

가

elongation

(Fig. 21).

가

0.5 CL/PE

*P. aeruginosa*

가

10%(wt)

percent elongation

*P. aeruginosa*

가

tensile strength

strain

energy

(Fig. 22),

가

0.1 CL

1.0 CL

*P. aeruginosa*

5%(wt)

*P. aeruginosa*

10.0

HP/PE

(Fig.

23).

10%(wt)

5.0 HP/PE tensile strength

tensile strength, percent

*P. aeruginosa*  
elongation, strain energy

가

7.5 HP/PE

(Fig. 24).

*P. aeruginosa*

(1994)

가

*P. aeruginosa*

*P. aeruginosa*

(Fig. 25, Fig. 26).

(A)

가

*P.*

*aeruginosa*

가

Wool (1991) polyethylene

가 1 10

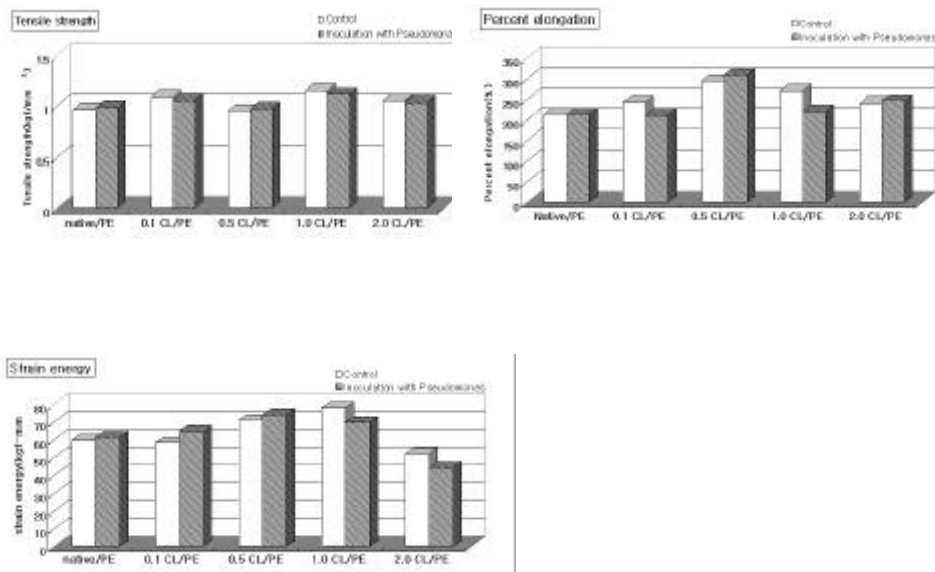
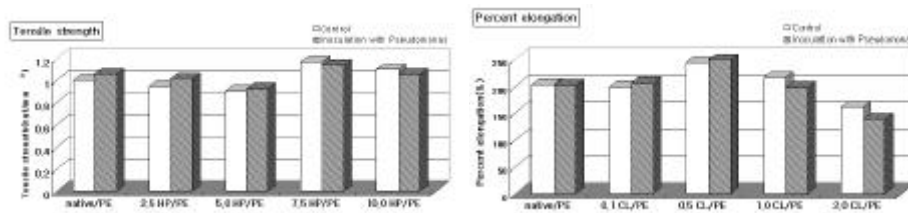


Fig. 21. Mechanical properties of the 5% crosslinked potato starch-polyethylene films cultured with *Pseudomonas aeruginosa* for 4 weeks. Each data represents mean for four replicates.



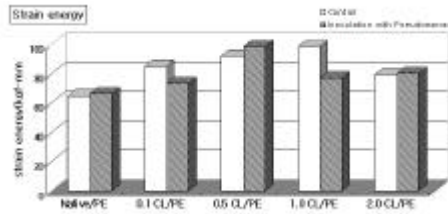


Fig. 22. Mechanical properties of 10% the crosslinked potato starch-polyethylene films cultured with *Pseudomonas aeruginosa* for 4 weeks. Each data represents mean for four replicates.

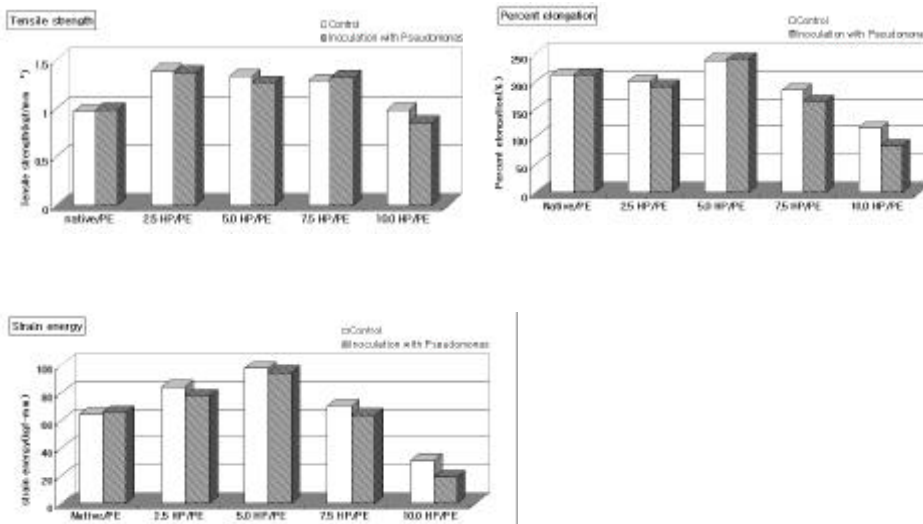


Fig. 23. Mechanical properties of the hydroxypropylated potato starch (5%)-polyethylene films cultured with *Pseudomonas aeruginosa* for 4 weeks. Each data represents mean for four replicates.



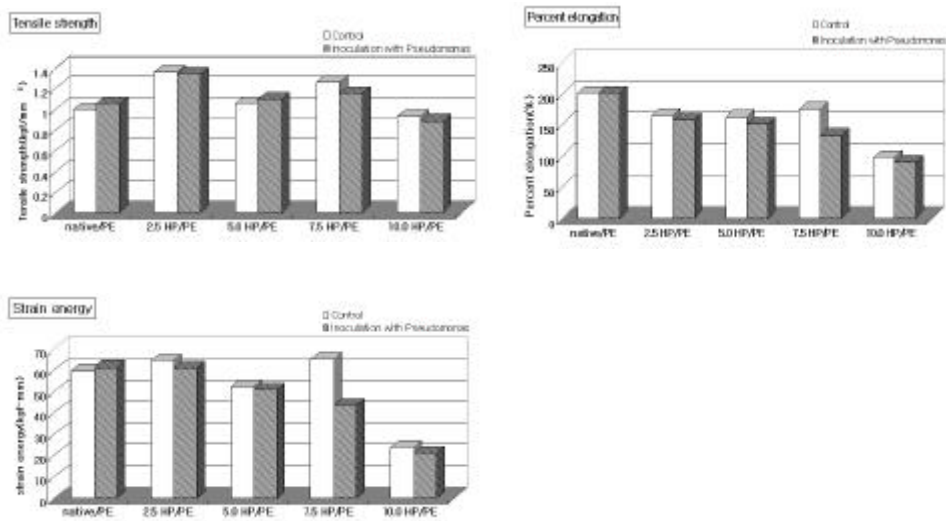


Fig. 24. Mechanical properties of the hydroxypropylated potato starch (10% wt)-polyethylene films cultured with *Pseudomonas aeruginosa* for 4 weeks. Each data represents mean for four replicates.

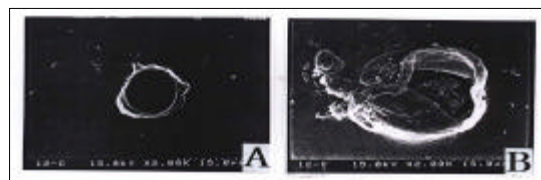


Fig. 25. Scanning electron micrograph ( $\times 2,000$ ) of the crosslinked potato starch-polyethylene films cultured with *Pseudomonas aeruginosa* for 4 weeks. A, Control (no inoculation); B, Inoculation by *Pseudomonas aeruginosa*

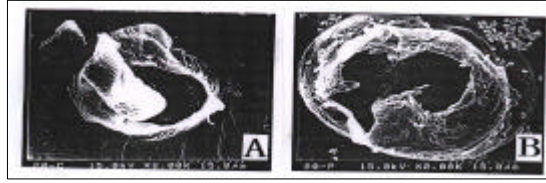


Fig. 26. Scanning electron micrograph( $\times 2,000$ ) of the hydroxypropylated potato starch-polyethylene films cultured with *Pseudomonas aeruginosa* for 4 weeks. A, Control(no inoculation); B, Inoculation by *Pseudomonas aeruginosa*

4

가

polyethylene

가 .

가가

. 가

가 ( 5%

) 가

가

가 가 .

## 4

- : Sodium hypochlorite ,  
 , 7(3):315-324, 1997.
- : , pp322-331, 1997.
- : , Polymer Science and  
Technology, 5(1):3-4, 1994.
- , : 가 ,  
 , 28:573, 1996.
- , , : Propylene oxide  
 , 7(4):519-526, 1997.
- , :  
 , Polymer(Korea), 18(4):613-621, 1994.
- , , :  
 , 23:175-182, 1991.
- , , , :  
 , 16:29, 1984.
- El-Hinnawy, S. I., El-Saied, H. M., Fahmy, A., El-Shirbeeney, A. E. and  
El-Sahy, K. M.: Viscosity and gelatinization characteristics of  
hydroxyethyl starch, Stärke, 34:112, 1982.
- Evangelista, R. L., Nikolov, Z. L., Wei, S., Jane, J. and Gelina, R.  
J. : Effect of compounding and starch modification on properties of  
starch-filled low-density polyethylene, Ind. Eng. Chem. Res.,  
30(8):1841-1846, 1991.

French, D. : Fine structure of starch and its relationship to the organization of starch granules, *J. Jpn. Soc. Starch Sci.*, 19:8, 1972.

Gage, P. : Degradable polyethylene film-the facts, *Tappl Journal*, 161-169, 1990.

Gilbert, G. A. and Spragg, S. P. : Iodometric determination of amylose. In "*Methods in carbohydrate chemistry*", Whistler, R. L. (ed.), Academic Press, New York, N.Y., vol. 4. p.168, 1964.

Goheen, S. M. and Wool, R. P. : Degradation of polyethylene-starch blends in soil, *J. of Appl. Poly. Sci.*, 42:2691-2701, 1991.

Griffin, J. L. : Biodegradable fillers in thermoplastics, *Advances in chem. series Amer. chem. Soci.*, Washington D. C., 159-170, 1974.

Griffin, J. L. : Synthetic resin sheet material, U. S. Patent, 4,021,388, 1977.

Hamerstrand, G. E., Hofretter, B. T. and Mehlretter, C. L. : Determination of the extent of reaction between epichlorohydrin and starch, *Cereal Chem.*, 37, 519, 1960.

Hollinger, G., Kuniak, L. and Marchessault, R. H. : Thermodynamic aspects of gelatinization and swelling of crosslinked starch, *Biopolymers*, 13:879, 1974.

Jane, T., Xu, A., Radosavljevic, M. and Seib, P. A. : Location of amylose in normal starch granules. . Susceptibility of amylose and amylopectin to cross-linking reagents, *Cereal Chem.*, 69:405, 1992.

Johnson, D. P. : Spectrophotometric determination of the hydroxypropyl group in starch ethers, *Anal. Chem.*, 41:859-860, 1969.

Kartha, K. P. R. and Srivastava, H. C., Ahmedabad. : Reaction of epichlorohydrin with carbohydrate polymers. part . Starch reaction mechanism and physicochemical properties of modified starch, *Stärke*, 37: 297, 1985.

Kasemsuwan, T. and Jane, J. : Location of amylose in normal starch granules. . Locations of phosphodiester cross-linking revealed by phosphorus-31 nuclear magnetic resonance, *Cereal Chem.*, 71:282, 1994.

Kin, M., Pometto, A. L., Johnson, K. E. and Fratzke, A. R. : Degradation studies of novel degradable starch-polyethylene plastics containing oxidized polyethylene and prooxidant, *J. Environ. Polym. Degrad.*, 2:27-38, 1994.

Koniya, T., Nara, S. and Tsu, M. : Changes in Crystallinity and Gelatinization phenomena of potato starch by acid treatment, *Stärke*, 38:9-13, 1986.

Lin, S. and Seib, P. A. : Preparation and pasting properties of wheat and corn starch phosphate. *Cereal Chem.*, 70:137, 1993.

Louis, G. L., Hopkins, A. R. and George, E. R. : Starch-based materials, In *biodegradable polymers and packaging*, ed. by Ching, C., Kaplan, D. L., Thomas, E. L., pp43-51, 1993.

Medcalf, D. G. and Gilles, K. A.: Wheat starches. . Comparison of physicochemical properties, *Cereal Chem.*, 42:558, 1965.

- Otey, F. H., Westhoff, R. P. and Russell, C. R. : Biodegradable films from starch and ethylene-acrylic acid copolymer, *Ind. Eng. Chem. Prod. Res. Dev.*, 16(4):305-308, 1977.
- Otey, F. H., Westhoff, R. P., Doane, W. M. : Starch-based blown film, *Ind. Eng. Chem. Prod. Res. Dev.*, 19:592-595, 1980.
- Otey, F. H., Westhoff, R. P., Doane, W. M. : starch-based blown film 2, *Ind. Eng. Chem. Res.*, 26:1659-1663, 1987.
- Potts, J. E., Clendinning, R. A., Ackart, W. B. and Niegisch, W. D. : "Polymer and Ecological problems", Plenum press, New York, p61, 1973.
- Rasper, V. F. and Deran, J. M. : Effect of Granule size of substituted starches on the rheological character of composite doughs. *Cereal Chem.*, 57:331, 1980.
- Robin, J. P., Mercier, C., Charbonniers, R. and Guilbort, A. : Lintnerized starches, Gel filtration and enzymatic studies of insoluble residues from prolonged acid treatment of potato starch, *Cereal Chem.*, 51:389-406, 1974.
- Schoch, T. J. : Swelling power and solubility of granular starches. In "*Methods in carbohydrate chemistry*", Whistler, R. L. (ed.), Academic Press New York, N. Y., vol. 4, p.106, 1964.
- Swanson, C. L., Westhoff, R. P., Doane, W. P. : Modified starches in plastic films, Proceedings of the corn utilization conference, Columbus, OH, 1988: National Corn Growers Association: St. Louis, Mo, 1988.

Wei, S., Nikolov, Z. L. : Accelerated degradation studies of starch-filled polyethylene films, *Ind. Eng. Chem. Res.*, 31(10):2332-2339, 1992.

Wootton, M. and Manatsathit, A.: The influence of molar substitution on the water binding capacity of hydroxypropyl maize starches, *Stärke*, 35:92-94, 1983.

Zobel, H. F.: X-ray analysis of starch granular starches, In *Methods in Carbohydrate Chemistry*, Whistler, R. L. (ed), Academic Press, New York, 4:109-113, 1964.