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L2934

밤바구미 유인제 개발

Development of Attractant(s) for
the Chestnut Weevils, *Curculio* spp.

산림청 임업연구원

농 립 부

1995

- .
: 1. 8 .
2. 1

1998. 12. .

:

: ()

:

“ ”

.

1998. 12. .

:

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:

.
 .
 1960 20 ha
 82,000ha가 , ,
 2,500 10
 1 13
 .
 20-30%,
 60%
 가 .
 가 , ,
 .
 가 가
 1998 3 11 ha . 3

가

가

가

.

.

.

1.

가 가 가

.

2.

issue

가

3

가

1.

가

Curculio robustus 3

C. robustus 2 1

1 1

. 4.3
 , 1 가 1 10 .
 . sensilla trichodea sensilla
 chati ca가 .
 . 5가 가 (GC)
 .
 .
 . Green volatile .

2.

. 8 9
 . 8 9 10
 .
 . 가 .
 . 가 .
 .
 .

Summary

The present study was conducted to investigate biological characteristics, and searching and identification of attractants for Chestnut weevils, *Curculio* spp. Chestnut weevils in the fields were identified three species, *Curculio sikkimensis*, *C. robustus* and *C. camelliae*.

Among three species, *C. sikkimensis* and *C. robustus*, are bivoltine, but the other species, *C. camelliae*, is univoltine in the fields. The peak of seasonal occurrences showed late August in *C. camelliae* and early-middle September in *C. robustus* and *C. sikkimensis*, respectively. The period of embryogenesis was 9.92 days under 25 ± 1 , 15L/9D and RH=60 \pm 10% regimes. The longevities of their female and male adults were 19.25- and 9.13-day for *C. robustus*, 7.16- and 6.18-day for *C. sikkimensis* and 5.83- and 3.70-day for *C. camelliae*, respectively. The *C. sikkimensis* females almost laid their eggs inside chestnut fruit, 0.4-0.8mm depth from the surface.

Copulation of *C. sikkimensis* male adults with their females occurs through a sequential mating behaviors followed: antennal movements, extension of genitalia and clasping female thorax and abdomen by the male legs. Otherwise, mating behavior of female adults is a simple comparing with that of the male. Female adults showed the highest rate of mating in 4-day-old of *C. robustus*, 3.21-day-old of *C. sikkimensis* and 2.71-day-old of *C. camelliae*, respectively. Mating time indicates a peak in the

middle part of photophase, from PM 2 to PM 4, among three species.

The late maturing chestnut varieties were most heavily damaged by the chestnut weevils. We had tried development of an artificial diet for the insect mass rearing, but came to a failure because the larvae survived for 4.3 days on the artificial diet. With scanning electromicroscopy structures of sensillia, odor receptors, showed the sensilla trichodea for pheromone and sensilla chaetica for attractants in *C. sikkimensis*.

Gas chromatography (GC), electroantennogram (EAG), olfactometry were employed in identification of active compounds in attracting chestnut weevil adults, *Curculio robustus*, *C. sikkimensis*, *C. camelliae*, from their own body and their host plant, chestnut, var. Choopa. Attraction of male and female adults was also investigated with sex pheromone lure indoors and outdoors. The retention time of detected compounds on GC gram from their own body were quite different from those of sex pheromone from the boll weevil, *Anthonomus grandis*, even though the two species belong to the same family. The sex pheromone of the boll weevil did not attract either sex of the chestnut weevil in the field. But chestnut weevil adults were significantly attracted to extract of chestnut burs damaged earlier by the yellow peach moth, *Dichocrosis punctiperalis*, another pest insect species on chestnut fruits. Their positive

behavioral response to the extract may be due to secondary metabolites of damaged burs, since both virgin and mated chestnut weevil adults of both sexes showed the same response. Hence more works were done on chemical composition of healthy and damaged chestnut fruits.

Methanol extracts of the chestnut burs, undamaged or damaged by the boll weevil, were analyzed to identify attractive green volatiles by GC, EAG and olfactometer. Attraction and EAG response of the adults were higher to chloroform fraction than any other fractions in the case of methanol extracts from burs undamaged. However, ethylacetate fraction showed the highest level in attracting and eliciting behavioral response in the adults in the case of methanol extracts from burs damaged. From these two fractions originated from methanol extracts of burs undamaged and damaged, eight compounds, (+)- α -pinene, E-2-hexanal, benzaldehyde, n-decyl aldehyde, nonyl acetate, terpineol, geraniol, 2-phenylethanol, were temporarily identified when compared their GC retention times with those of standard chemicals. Among these chemicals (+)- α -pinene showed the highest attractivity to chestnut weevil adults when bioassayed indoors.

With the present results, it may be suggested that chestnut weevil adults probably use, as their sex pheromone, compounds different from those of *A. grandis*. The weevil adults may use green volatiles, including (+)- α -pinene, from their host

plant, especially burs damaged by *D. punctiferalis*. However, further studies are required to identify their sex pheromone components and their composition, and green volatile compounds, from their host plant including burs, active in attracting adult weevils.

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3.	-----	62
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5 .	-----	77

Section 1: Biological characteristics of the chestnut weevils, *Curculio* spp.

SUMMARY: The present studies were conducted to investigate biological characteristics of three *Curculio* spp. and their seasonal occurrences in the chestnut orchards of three provinces. Chestnut weevils in the fields were identified three species, *Curculio sikkimensis*, *C. robustus* and *C. camelliae*.

Among three species, *C. sikkimensis* and *C. robustus*, are bivoltine, but the other species, *C. camelliae*, is univoltine in the fields. The peak of seasonal occurrences showed late August in *C. camelliae* and early-middle September in *C. robustus* and *C. sikkimensis*, respectively. The period of embryogenesis was 9.92 days under 25 ± 1 °C, 15L/9D and RH=60±10% regimes. The longeivities of their female and male adults were 19.25- and 9.13-day for *C. robustus*, 7.16- and 6.18-day for *C. sikkimensis* and 5.83- and 3.70-day for *C. camelliae*, respectively. The *C. sikkimensis* females almost laid their eggs inside chestnut fruit, 0.4-0.8mm depth from the surface.

Copulation of *C. sikkimensis* male adults with their females occurs through a sequential mating behaviors followed: antennal movements, extension of genitalia and clasping female thorax

and abdomen by the male legs. Otherwise, mating behavior of female adults is a simple comparing with that of the male. Female adults showed the highest rate of mating in 4-day-old of *C. robustus*, 3.21-day-old of *C. sikimensis* and 2.71-day-old of *C. camelliae*, respectively. Mating time indicates a peak in the middle part of photophase, from PM 2 to PM 4, among three species.

The late maturing chestnut varieties were most heavily damaged by the chestnut weevils. We had tried development of an artificial diet for the insect mass rearing, but came to a failure because the larvae survived for 4.3 days on the artificial diet. With scanning electromicroscopy structures of sensillia, odor receptors, showed the sensilla trichodea for pheromone and sensilla chatica for attractants in *C. sikimensis*.

1 .

1960

20 ha .

가 8 2 ha

10 ton . 2,500

13 1

2 .

1.

가.

1996 1998 3 , ,
, 1997 , 1998 . 1996

1995 10 17 468
: 25 ± 1 , : L/15: D/9, RH=60 \pm 10% 20

hole

..

(Vol t i n i s m)

(vol t i n i s m)

1995 10 , 1996 10 , 1997 10

,

(Ø 70cm H: 50cm)

2.

1997 9 10 10 19 (C. *sikkimensis*)
(100×60×60) 1995 1996

, , , .
()

3.

1996 1998 10 1 2

3 .

4.

1998

(*C. sikkimensis*)

hair sensor

sensilla trichodea

sensilla chatica

3 .

1.

가.

1).

1

10%

20%

(, 1996 , 1997, 19

98)

가 .

1.

			(%)
()	48	4	8.3
()	54	6	11.1
()	25	3	12.0
()	58	3	5.2
()	38	4	10.5
()	47	5	10.6
()	50	4	8.0
()	39	1	2.6
()	47	4	8.5
()	50	3	6.0
	456	37	-
	-	-	8.28 ± 2.97a
()	23	2	8.7
()	14	0	0.0
()	16	1	6.3
()	15	1	6.7
()	18	2	11.1
()	20	1	5.0
()	15	1	6.7
()	40	3	7.5
()	38	4	10.5
()	29	2	6.9
	228	17	-
	-	-	6.94 ± 3.09ab

			(%)
()	15	0	0.0
()	20	1	5.0
()	20	1	5.0
()	37	3	8.1
()	19	1	5.3
()	24	1	4.2
()	26	1	3.8
()	27	2	7.4
()	26	1	3.8
()	25	2	8.0
	239	13	-
	-	-	5.06 ± 2.43b
()	45	3	6.7
()	50	3	6.0
()	35	1	2.9
()	40	2	5.0
()	51	4	7.8
()	24	0	0.0
()	21	1	4.8
()	32	1	3.1
()	22	3	13.6
()	38	2	5.3
	358	20	-
	-	-	5.52 ± 3.59ab

2).

1996 10%
 1997 4
 9 *C. camelliae* C.
robustus 20-
 30% 500

4.

			(%)
()	136	40	29.4
()	144	43	29.9
()	161	51	31.7
()	154	42	27.3
()	158	37	23.4
()	146	39	26.7
()	142	45	31.7
()	141	29	20.6
()	170	54	31.8
()	163	49	30.1
	-	-	-
	-	-	28.3

			(%)
()	143	31	21.7
()	139	33	23.7
()	164	45	27.4
()	172	38	22.1
()	136	36	26.5
()	141	37	26.2
()	128	30	23.4
()	114	32	28.1
()	176	50	28.4
()	143	38	26.6
		-	-
	-	-	25.4
()	173	51	29.5
()	161	44	27.3
()	154	40	26.0
()	136	41	30.1
()	119	28	23.5
()	124	30	24.2
()	154	43	27.9
()	134	31	23.1
()	142	15	10.6
()	150	42	28.0
	-	-	-
	-	-	25.0

3).

5 1998
 58.9% 1996 1997 70%

5.

			(%)
	146.7	91.8	62.6
	158.8	70.0	44.0
	185.3	131.2	70.0
	163.6	97.6	58.9

4
 6 11
 48.5%가 가
 가 (dormancy)

가
가

6.

		(%)
10	168	5.01
11	302	9.01
11	1,625	48.49
11	516	15.40
12	465	13.88
12	199	5.94
12	76	2.27
	3,351	100

1).

1995 1 79.
8% 2 6.2%
1996 1995
36.2%
1997

(7).

7.

		1 (%)	2 (%)	3 (%)
1995	4, 200	3, 220(80)	260(6. 2)	-
1996	2, 650	960(36. 2)	-	-
1997	2, 600	-	-	-

8 10

(*C. sikkimensis*) *C. robustus*()
 '95 1 1 (uni vol ti ne) 6. 5%
 18. 2% , '96 2 12. 5% 2
 (bi vol ti ne) (*C. camelli*
 ae) 1995 1996 1 1
 2%

(9 11).

8. '95

	<i>sikkimensis</i>	<i>camelliae</i>	<i>robustus</i>	
1996	2	13	4	19
1997	29	-	18	47

9. '95

		()
1996	980	19(1.94%)
1997	2,960	47(1.58%)
	3,940	66(1.68%)

10. '96

	<i>sikkimensis</i>	<i>camelliae</i>	<i>robustus</i>	
1997	3	18	3	24
1998	-	-	-	-

13 14

(*C. sikkimensis*) *C. robustus*() 2 1 Bivo
 ltine (*C. camelliae*) 1 1 Univol tine
 . (1984)가 2
 1 .

13. '96

	<i>sikkimensis</i>	<i>camelliae</i>	<i>robustus</i>	
1997	3	18	3	24
1998	19	-	10	29

14. '97

	<i>sikkimensis</i>	<i>camelliae</i>	<i>robustus</i>	
1998	6	31	4	41

2).

'96

'97 4 21

George C. McGavin Mark Robinson

3가 가 (

15).

15.

Curculio spp.			(%)	
	'96	'97	'96	'97
<i>sikkimensis</i>	56	164	39.4	66.4
<i>camelliae</i>	58	51	40.8	20.6
<i>robustus</i>	28	32	19.7	13.0
Total	142	247	100	100

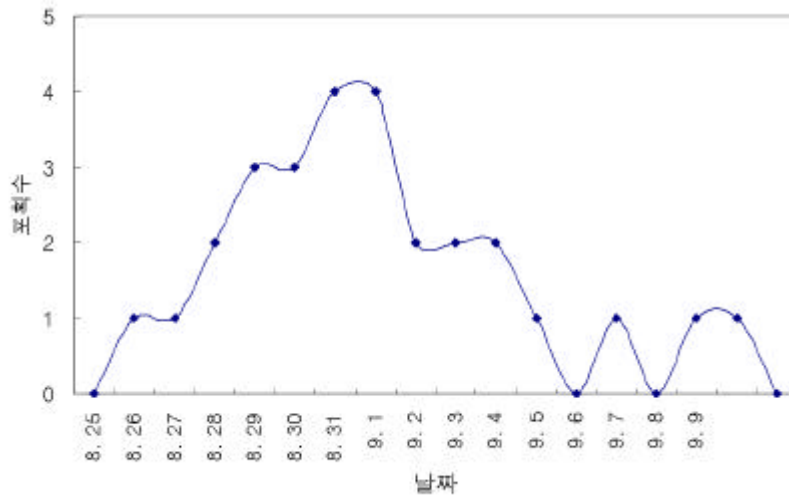
'97

1, 2, 3 .

*Curculio camelliae*가 가 8 25 ,

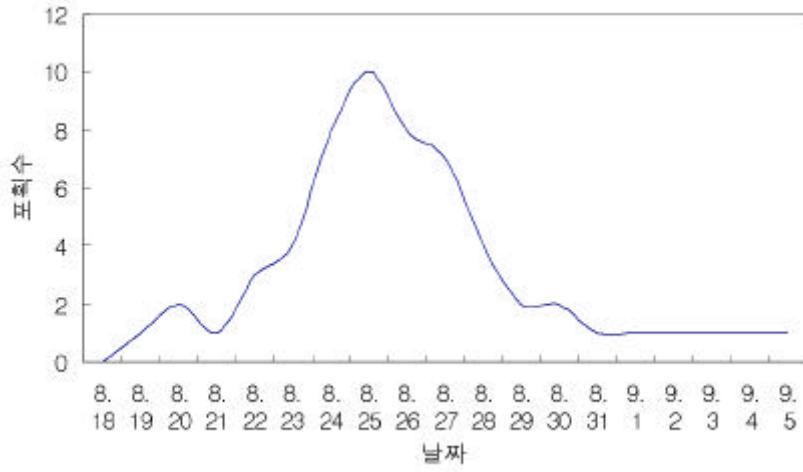
Curculio robustus 8 29 9 5

, *Curculio sikkimensis*() 8 10

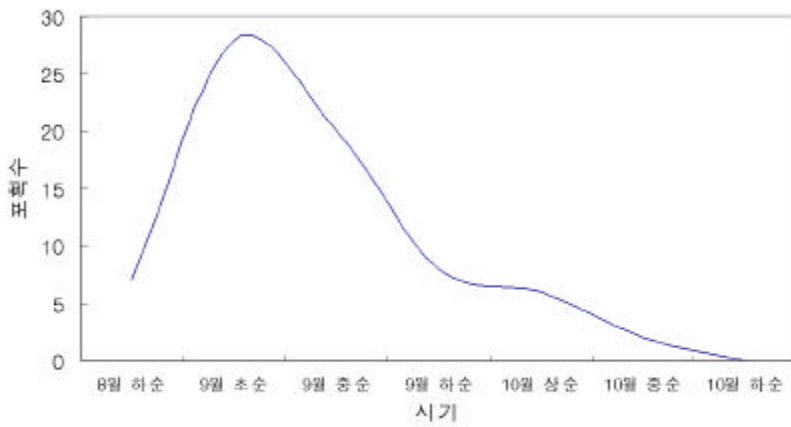


1. *Curculio robustus*

<i>Curculio</i>	8	9	
<i>camelliae</i> 가	8	25	C.
<i>robustus</i>	8	9	C.
<i>si kki mensi s</i> 가		, 9 5	C.
	10	가	
	가		10



2. *Curculio canelliae*

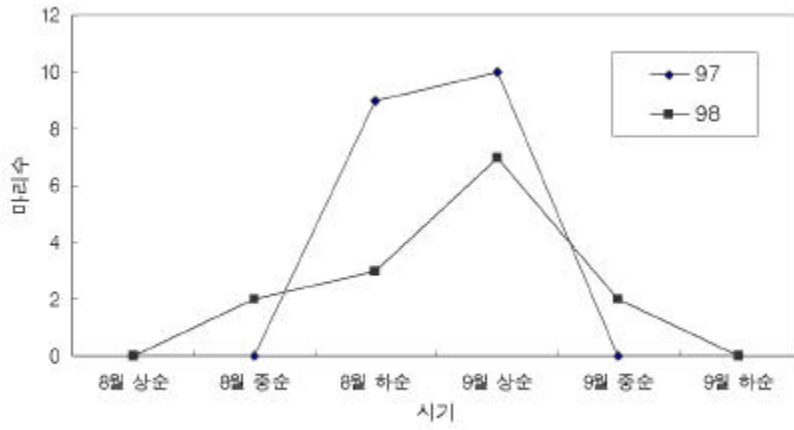


3. *Curculio sikkinensis*

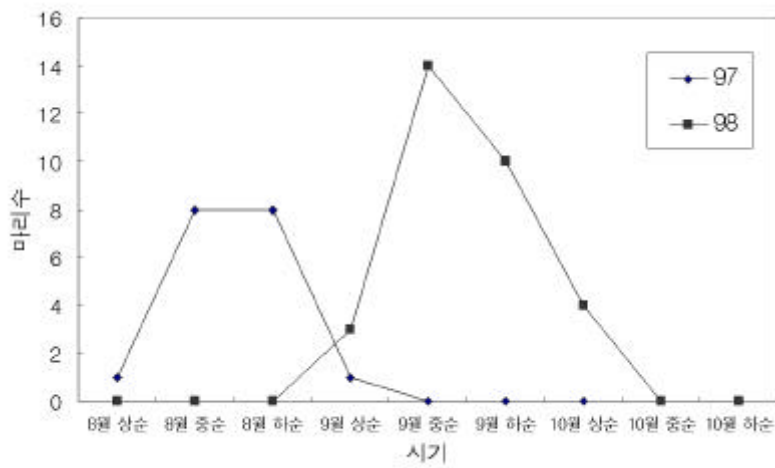
16
가 가 . '96
가 8 가
4, 5, 6 *C. robustus* 8 9
C. canelliae '97 '98 가
'98 . *C. sikkinensis*
9

16.

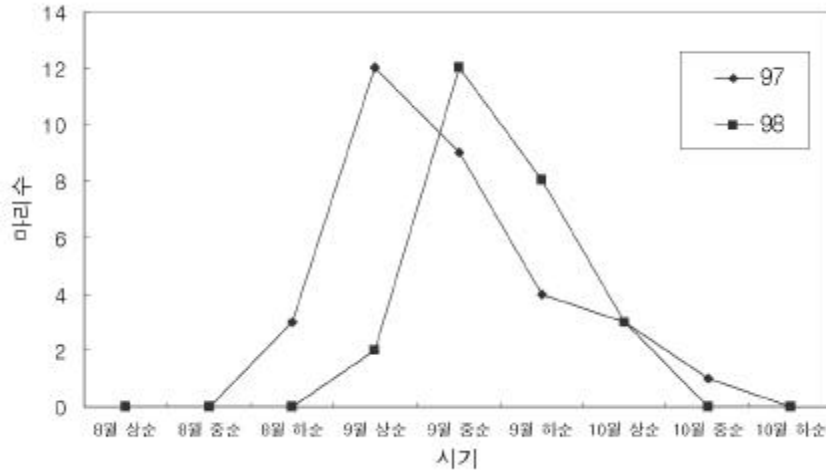
Curculio spp.				(%)		
	'96	'97	'98	'96	'97	'98
<i>sikkinensis</i>	56	164	77	39.4	66.4	41.4
<i>canelliae</i>	58	51	48	40.8	20.6	25.8
<i>robustus</i>	28	32	61	19.7	13.0	32.8
	142	247	186	100	100	100



4. *C. robustus*



5. *C. canelliae*



6. *C. sikkinensis*

25 , L: D=15: 9, 60%
 17 10 (1984) 5.9 40%
 3
 가 .

17. (:25 , L:D=15:9, :60%)

8	3
9	9
10	12
11	6
	30
	9.92

2.

가.

Upper : Lower = 2.4 : 2.9

가

Upper :

Lower = 3.9 : 2.1

(18).

18.

	(mm)		Upper(mm)		Lower(mm)	
	5.00	5.30	2.36	3.90	2.88	2.12
STD	0.45	0.68	0.24	0.50	0.17	0.15

. (*C. sikkimensis*)

1).

C. sikkimensis 64.3%, *C. robustus* 66.7%가 14:00-16:00 가

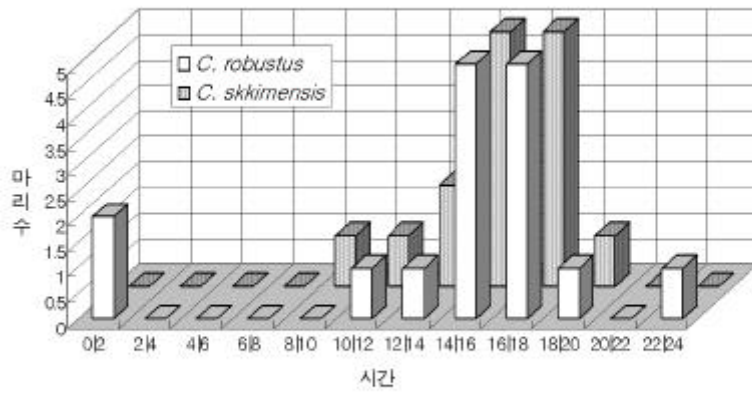
4 (photophase)

(scotophase)

C. sikkim

ensis *C. robustus*

(7).



7.

2).

가 cage 가 .
 ,
 , vibration()
 가
 가 2
 3 가 5-6 가 Hair Sensor head
 . 가

sex pheromone
 pheromone gland , ,
 .

3).

가 Food source feeding
 . 가 1
 .
 가
 feeding . 1-2
 .

4).

19

가 *C. robustus* 가 4.00 , *C. canelliae* 2.71 .

가

19.

<i>C. robustus</i>	12	4.00
<i>C. sikkinensis</i>	14	3.21
<i>C. canelliae</i>	11	2.71

5).

20

C. robustus

가 , 가
19.25 . (1984)
가 .
1 ,

20.

<i>C. robustus</i> ()	16	19.25
()	9	9.13
<i>C. sikkinensis</i> ()	19	7.16
()	16	6.18
<i>C. canelliae</i> ()	16	5.83
()	10	3.70

1 3-5
 (:) 가
 0.4 - 0.8mm 78.0% (21).

21.

(mm)	()	(%)
0.4	2	4.0
0.4 - 0.6	21	42.0
0.6 - 0.8	18	36.0
0.8	9	18.0
	50	100

3.

가
 , 1 , 2
 (22).
 1 . 2
 ,
 ,
 .
 2
 가 ().
 가 2-3
 가 .
 '96 Corn powder Chestnut powder 가
 (23) 가 가 가
 Sweet potato powder 가
 가
 . 2 1 1 가 1
 10
 .

22.

	1	2
Wheat germ	70g	30g
Cellulose	60g	-
Corn powder	55g	-
Casein	-	35g
Yeast	55g	-
Fructose	5g	-
Sucrose	-	35g
Vessen's salt nix.	0.6g	-
Agar	20g	25g
Cholesterol	-	0.5g
Corn oil	2.3mg	-
Choline chloride	-	1g
Inositol	-	0.4g
Ascorbic acid	6g	4g
Vitamine nix.	4g	5g
Distilled water	1,000Ml	850Ml
Methyl	2g	2g
p-hydroxyneboate	-	-
Sorbic acid	-	1g
Corn tee powder	2g	-
Malic acid	2g	-

23.

	1	2	3
Wheat germ	70g	30g	40g
Cellulose	60g	-	60g
Chestnut powder	55g	-	55g
Sweet Potato powder	-	45g	45g
Casein	-	35g	35g
Yeast	55g	-	55g
Fructose	5g	-	5g
Sucrose	-	35g	35g
Vessen' s salt nix.	0. 6g	-	0. 6g
Agar	20g	25g	20g
Cholesterol	-	0. 5g	0. 5g
Corn oil	2. 3mg	-	2. 3mg
Choline chloride	-	1g	1g
Inositol	-	0. 4g	0. 4g
Ascorbic acid	6g	4g	5g
Vitamine nix.	4g	5g	4g
Distilled water	1, 000Mℓ	850Mℓ	850Mℓ
Methyl p-hydroxynebzoate	2g	2g	2g
Sorbic acid	-	1g	1g
Corn tee powder	2g	-	2g
Malic acid	2g	-	2g

25.

	1	2	3	4	5
Wheat germ	70g	30g	40g	40g	40g
Cellulose	60g	-	60g	60g	60g
Chestnut powder	55g	40g	55g	65g	65g
Sweet potato powder	-	35g	45g	35g	35g
Casein	-	35g	35g	35g	35g
Yeast	55g	-	55g	55g	55g
Fructose	5g	45g	5g	5g	5g
Sucrose	-	35g	35g	35g	35g
Vessen's salt mix.	0.6g	-	0.6g	0.6g	0.6g
Agar	20g	25g	20g	20g	20g
Cholesterol	-	0.5g	0.5g	0.5g	0.5g
Corn oil	2.3Ml	-	2.3Ml	2.3Ml	2.3Ml
Choline chloride	-	1g	1g	1g	1g
Inositol	-	0.4g	0.4g	0.4g	0.4g
Ascorbic acid	6g	4g	5g	5g	5g
Vitamine nix.	4g	5g	4g	4g	4g
Distilled water	1,000Ml	850Ml	850Ml	850Ml	800Ml
Methyl p-hydroxyneboate	2g	2g	2g	2g	2g
Sorbic acid	-	1g	1g	1g	1g
Corntee powder	2g	-	2g	2g	2g
Malic acid	2g	-	2g	2g	2g
Niacin	-	-	-	7ng	7ng
Fe	-	-	-	-	0.3g
Ca	-	-	-	-	0.3g
P	-	-	-	-	0.5g

4.

8 9

sensilla trichodea(ST) sensilla chatica(SC)가 .

sensilla chatica(SC)

flagellonone

. sensilla trichodea(ST)

sensilla basiconica

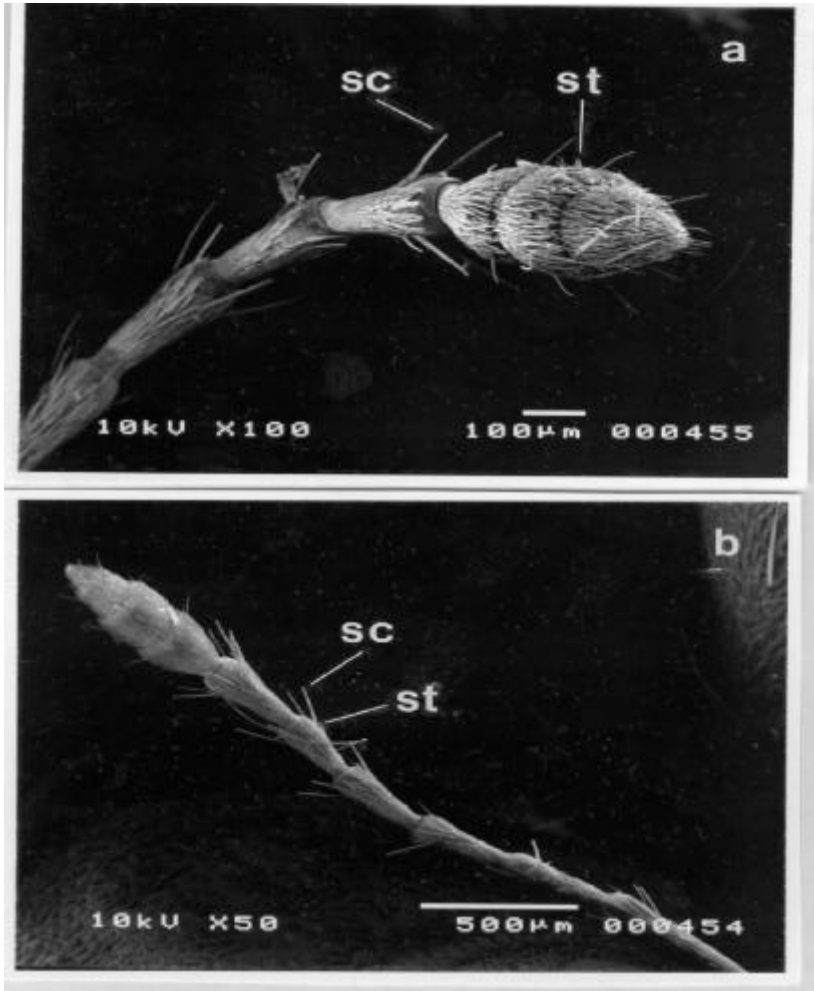
, single cell recording

9(a)

sensilla chatica(SC), sensilla trichodea(ST)

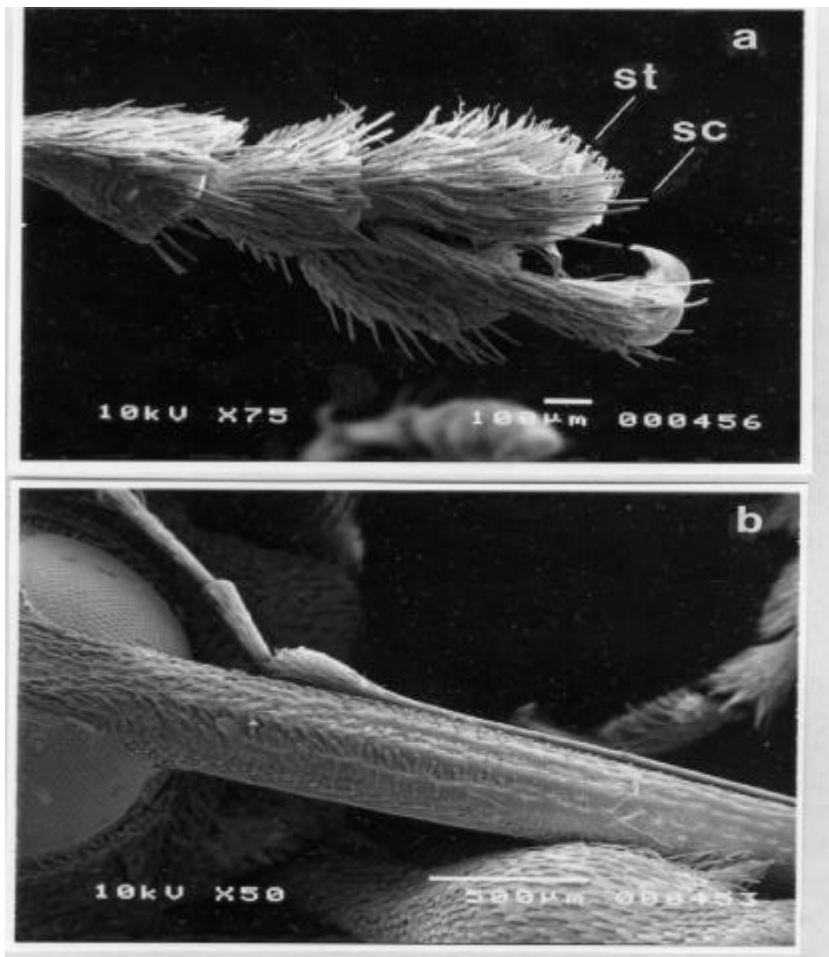
(9b)

modified scale



8.

a: , b:



9.

. a:

, b:

4 .

- , , , 1978. .
. 25:99 - 110.
- , . 1984.
. . 23; 132-136.
- . 1996. . 1
. 57 pp.
- . 1993. . 15
pp. 57-65.
- . 1997. . 19
pp. 111-124.
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2 .

Section 2: Searching and Identification of attractant for Chestnut weevil

SUMMARY

Gas chromatography (GC), electroantennogram (EAG), olfactometry were employed in identification of active compounds in attracting chestnut weevil adults, *Curculio robustus*, *C. sikkimensis*, *C. canelliae*, from their own body and their host plant, chestnut, var. Choopa. Attraction of male and female adults was also investigated with sex pheromone lure indoors and outdoors. The retention time of detected compounds on GC from their own body were quite different from those of sex pheromone from the boll weevil, *Anthonomus grandis*, even though the two species belong to the same family. The sex pheromone of the boll weevil did not attract either sex of the chestnut weevil in the field. But chestnut weevil adults were significantly attracted to extract of chestnut burs damaged earlier by the yellow peach moth, *Lichocrosis punctiperalis*, another pest insect species on chestnut fruits. Their positive behavioral response to the extract may be due to secondary metabolites of damaged burs, since both virgin and mated

chestnut weevil adults of both sexes showed the same response. Hence more works were done on chemical composition of healthy and damaged chestnut fruits.

Methanol extracts of the chestnut burs, undamaged or damaged by the boll weevil, were analyzed to identify attractive green volatiles by GC, EAG and olfactometer. Attraction and EAG response of the adults were higher to chloroform fraction than any other fractions in the case of methanol extracts from burs undamaged. However, ethylacetate fraction showed the highest level in attracting and eliciting behavioral response in the adults in the case of methanol extracts from burs damaged. From these two fractions originated from methanol extracts of burs undamaged and damaged, eight compounds, (+)- α -pinene, E-2-hexanal, benzaldehyde, n-decyl aldehyde, nonyl acetate, terpineol, geraniol, 2-phenylethanol, were temporarily identified when compared their GC retention times with those of standard chemicals. Among these chemicals (+)- α -pinene showed the highest attractivity to chestnut weevil adults when bioassayed indoors.

With the present results, it may be suggested that chestnut weevil adults probably use, as their sex pheromone, compounds different from those of *A. grandis*. The weevil adults may use green volatiles, including (+)- α -pinene, from their host plant, especially burs damaged by *L. punctiferalis*. However, further studies are required to identify their sex pheromone

components and their composition, and green volatile compounds, from their host plant including burs, active in attracting adult weevils.

1 .

가

가 가

.

, . 가 ,

, . ,

, 가

가

(, 1990),

가 .

가

가

가

. 가 .

, , .

, ,

가 ,
가 (Sorenson
, 1992; Toht , 1992),
가
(green
volatiles)
가
가 (Schoonhoven , 1988).

EAG single-cell recording
가

(Hunnel, 1984).

2 .

1.

가.

(30L) 20
Q(Signa Co.)

porapak

2500cc

hexane, ether

10 μ l 가

GC(Shinazu Co., HP 6890)

(Pherocon cap)

hexane

GC

Pherocon cap (McKibben, 1971)

wing trap (1), boll

weevil trap (2), flat trap (3) (Leggett, 1979;

Leggett Taft, 1979),

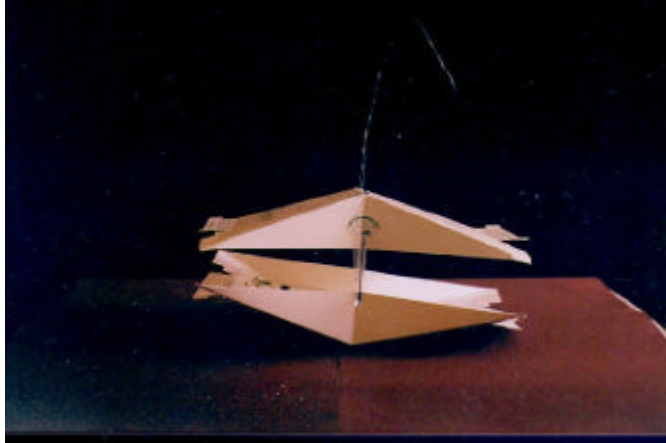
1996 8 24 1996 10 9

Pherocon cap boll trap

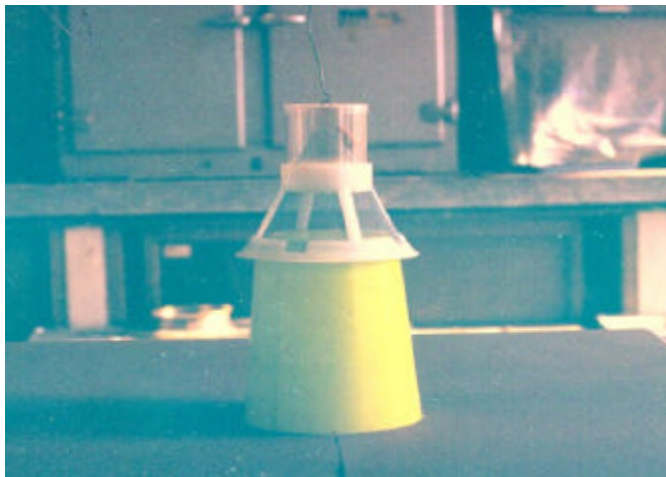
Pherocon cap

wing trap

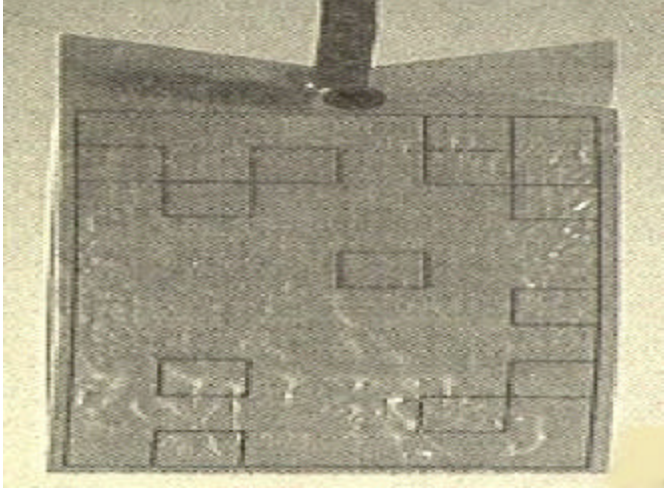
1997



1. Wing trap



2. Boll weevil trap



3. Flat trap

2.

()

.

가

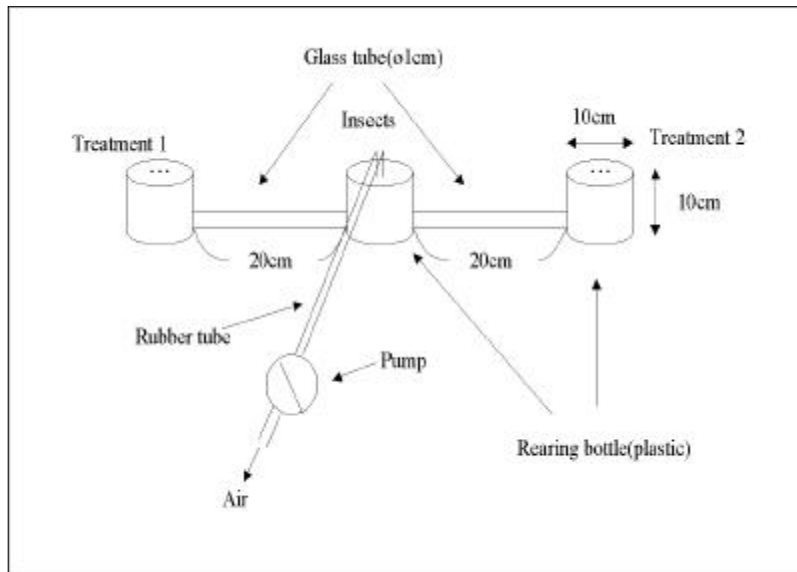
.

.

(4).

(Hardee ,

1967).



4.

3.

가.

(:)

hexane, chloroform, ethylacetate

Box of factometer

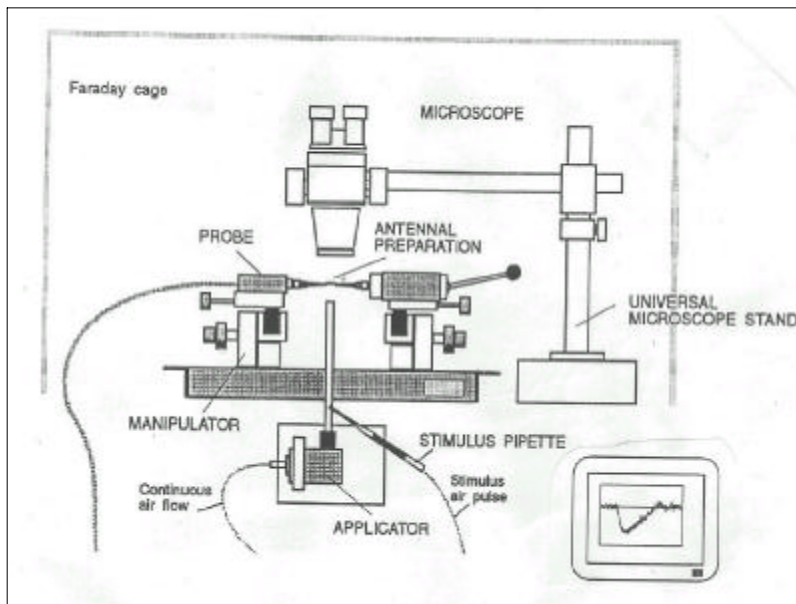
box (35cm x 25cm) 50M \emptyset

box *C. robustus* 10

2500cc/min

15L/9D, 25 \pm 1

10



5. Electroantennogram

Electroantennogram(EAG)
 CO₂
 (Ag) EAG
 (Syntech. Co) (5).

(:)

hexane, chloroform, ethylacetate, water

Box of factometer
 box 50M \emptyset
 box *C. robustus* 10

Y-tube ol factometer
 Y-tube(5cm) 가 50M \emptyset

가
Y-tube 가
가 5 가
2500cc/min
15L/9D, 25 ± 1

Electroantennogram(EAG)
CO₂
(Ag) EAG
(Syntech. Co)

. Green volatiles

Green volatiles
Green volatiles butyric acid
ethyl ester, hexanal, -myrcene, (+)-linonene, 2-heptanone,
E-2-hexanal, citral, eugenol, -humulene, (+)- -pinene,
(-)-E-caryophyllene, geraniol, 1-nonanol Signa

Electroantennogram(EAG)
green volatiles hexane
20ng hexane

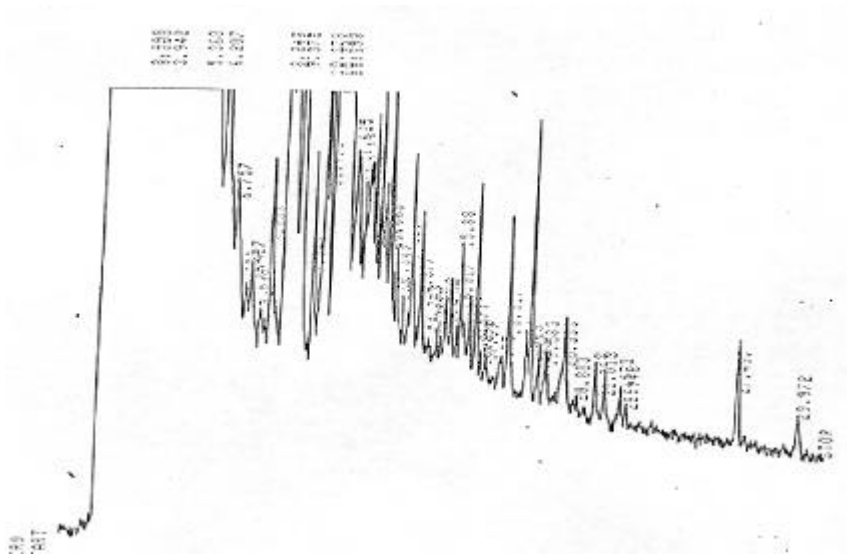
CO₂
 (Ag)
 EAG (Syntech. Co)

Box of factometer
 box 50M \emptyset
 green volatile
 box *C. robustus* 10

Glass tube of factometer
 Glass tube(7cm, 15cm) 50
 M \emptyset conical 50M \emptyset conical
 green volatile , glass
 tube tube
 10 tube Y

GC

가



6. Porapak-Q

decanal, hexadecanal, octadecanal, myrtenol, hexadecanyl acetate, octadecanyl acetate, Z9-octadecenyl acetate, E9-octadecenyl acetate, eicosanyl acetate (Kalo, 1979; Kalo Neders tron, 1983)

가 .

4

Old house beetle, *Ilyotrupes bayalus*

(Evans

Higgs, 1975).

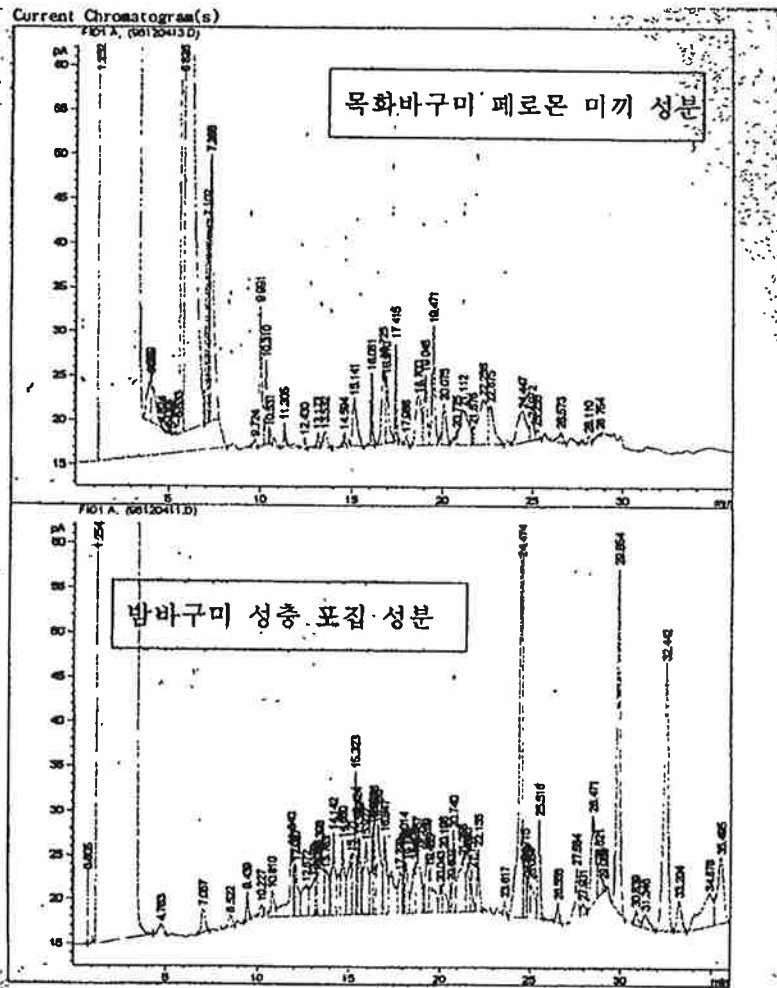


그림 8. 목화바구미(boll weevil) 페로몬 미끼성분 추출물과 밤바구미 발산 성분 비교 GC 그래프

dl), Z-3,3-dimethyl-1-cyclohexaneethanol .

GC

,
GC

가 .

Pherocon cap

가 .

가 (1),

1. Trap

Trap	
Wing trap	0
Boll trap	0
Flat trap	0

-. : ,

-. : 1996 8 24 - 1996 10 9

2.

	97. 8. 25	3	0
	97. 8. 26	3	0

* : boll weevil trap

3.

	97. 8. 25	10	1
	97. 8. 26	10	3

* : wing trap

3

가

가

가

.

2.

,

olfactometer

가

,

.

가

,

,

,

.

(4),

(5),

(6),

(7)

.

가

,

가

.

4.

Bait pair(1st/2nd)	1st bait	2nd bait	No response
CBU : UCF	21	1	5
CTL : CBU	1	21	8
: UCF	1	14	15
CBD : CTL	11	7	12
: CBU	16	2	12
: UCF	10	13	7

CBU :

UCF :

CTL :

CBD :

5.

Bait pair(1st/2nd)	1st bait	2nd bait	No response
CBU : UCF	5	4	21
CTL : CBU	1	8	21
: UCF	2	6	22
CBD : CTL	20	5	5
: CBU	23	2	5
: UCF	22	4	4

4

6.

Bait pair(1st/2nd)	1st bait	2nd bait	No response
CBU : UCF	4	2	24
CTL : CBU	3	9	18
: UCF	2	3	25
CBD : CTL	27	1	2
: CBU	28	0	2
: UCF	25	4	1

4

7.

Bait pair(1st/2nd)	1st bait	2nd bait	No response
CBU : UCF	6	3	21
CTL : CBU	3	5	22
: UCF	1	4	25
CBD : CTL	26	2	2
: CBU	24	1	5
: UCF	25	4	1

4

3.

가.

Box of factometer

,

가

가

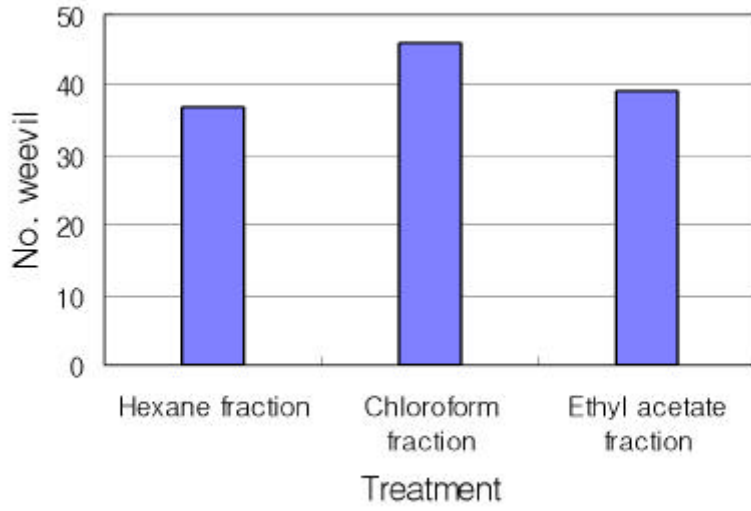
(9),

EAG

가

(10),

.



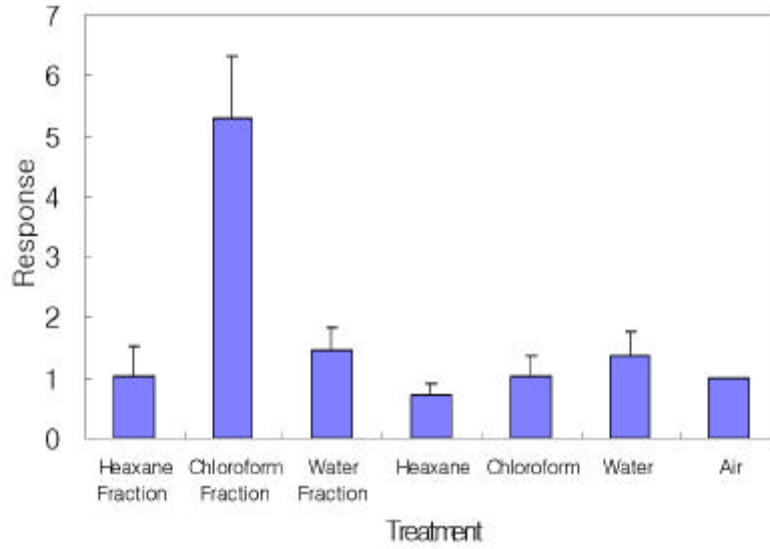
9. Box of factometer

Box of factometer

가

(11).

가



10.

EAG

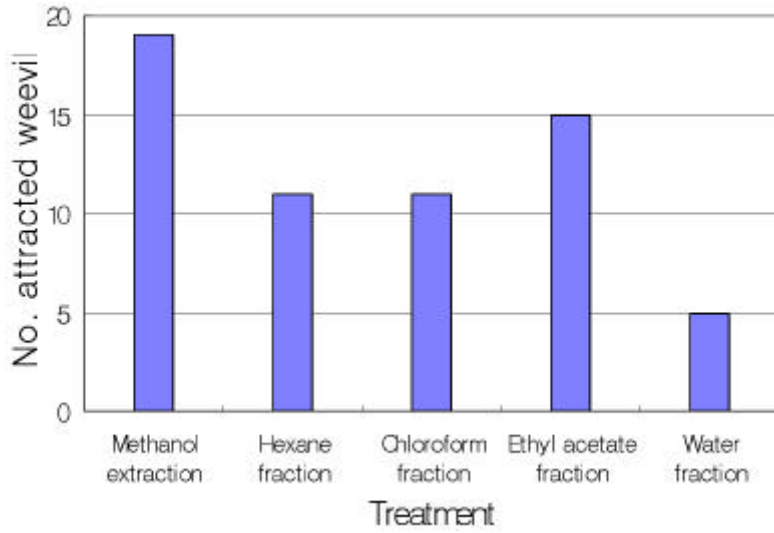
(air)

(Barnay Chapman, 1994; Turings , 1990; Turings Tunlinson, 1992)
가

Y-tube

가

(12). 가
(entered)

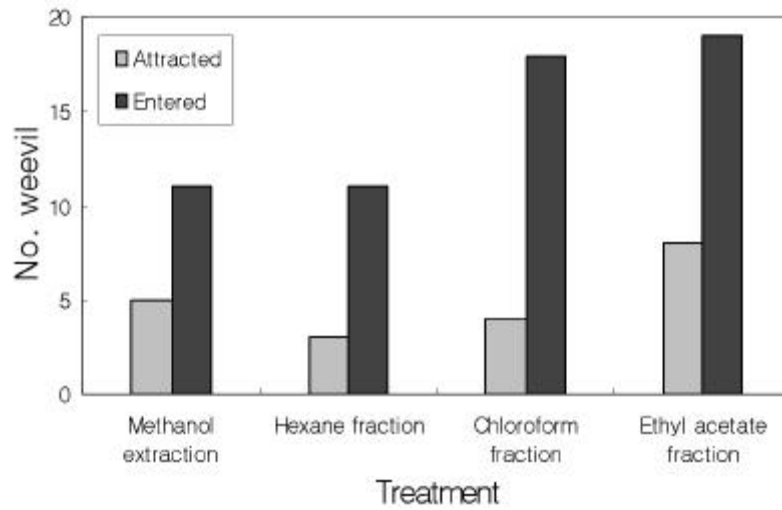


11. Box olfactometer

가 , 가
(attracted)
box olfactometer

가

가



12. Y tube olfactometer

EAG

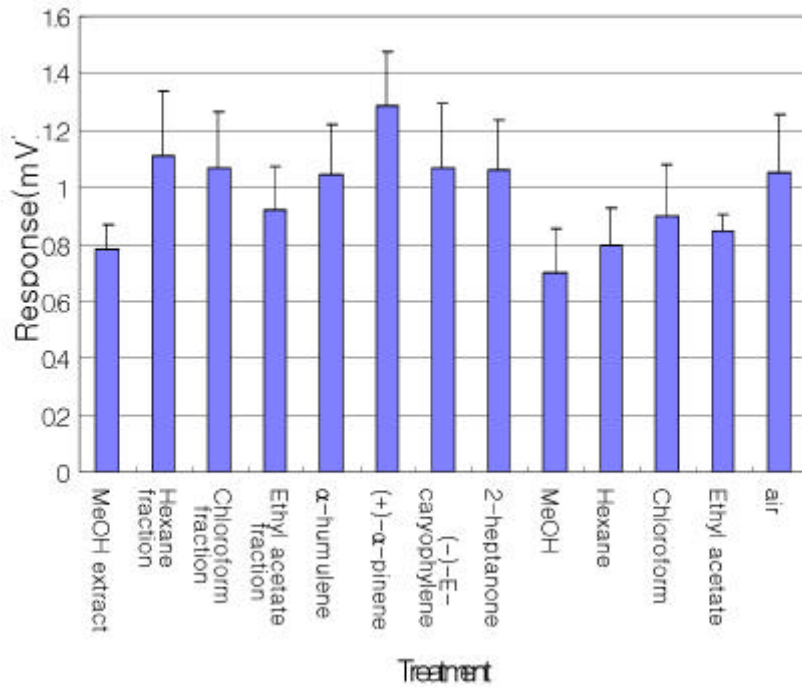
EAG

-pinene 가

(

13).

-humulene caryophyllene



13. green volatile

EAG

가 .

Pecan weevil, *Curculio caryae*, -humulene

caryophyllene

Z-3,3-dimethyl-1-cyclohexaneethanol

(Hedin 1979; Roseland, 1992).

. Green volatiles

green volatile

가

(+)- α -pinene, geraniol

가

box olfactometer

hexanal, 2-heptanone, E-2-hexanal,

(8).

가 ,

. Glass tube

olfactometer green volatile

(+)- α -pinene (-)-E-caryophyllene .

(+)- α -pinene EAG

가

가

pinene myrcene

8. Box olfactorimeter green volatile

Green volatile	
butyric acid ethyl ester	34
hexanal *	54
-myrcene	37
(+)-linonene	35
2-heptanone*	49
E-2-hexanal *	52
citral	34
eugenol	19
-humulene	34
(+)-pinene*	48
(-)-E-caryophyllene	32
geraniol *	45
1-nonanol	14
control	22

* : 가 green volatile

myrcene

caryophyllene

가

9. Glass-tube olfactometer green volatile

(/)

Green volatile	
butyric acid ethyl ester	0.92
-myrcene	0.80
(+)-limonene	0.50
2-heptanone	1.20
-humulene	1.31
(+)- α -pinene*	2.25
(-)-E-caryophyllene*	2.43
geraniol	0.80
1-nonanol	1.10

* : 가 green volatile

. Red sunflower seed weevil

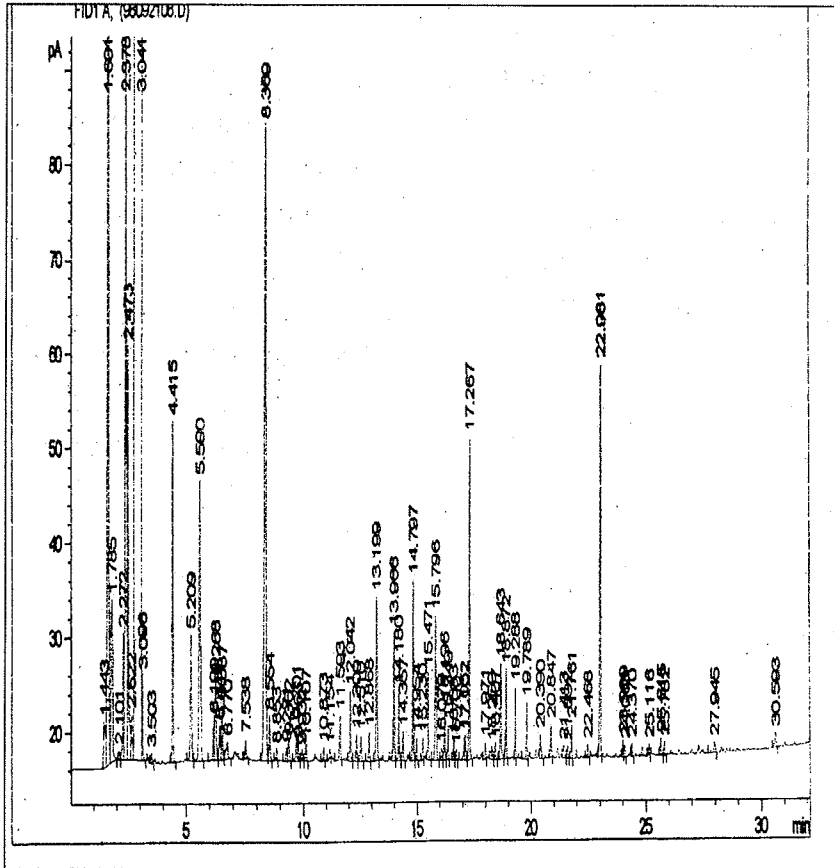
-pinene -pinene

.

GC

,

GC ,
 (+)-
 -pinene(5. 209), E-2-hexenal (9. 271), benzaldehyde(13. 966),
 n-decyl aldehyde(16. 700), nonyl acetate(17. 971),
 terpineol (18. 259), geraniol (20. 390) (14),
 (+)-
 -pinene(5. 167), benzaldehyde(13. 907), geraniol (20. 338)
 (15),
 (+)- -pinene(5. 503) terpineol (18. 377),
 2-phenylethanol (20. 274)
 (16).



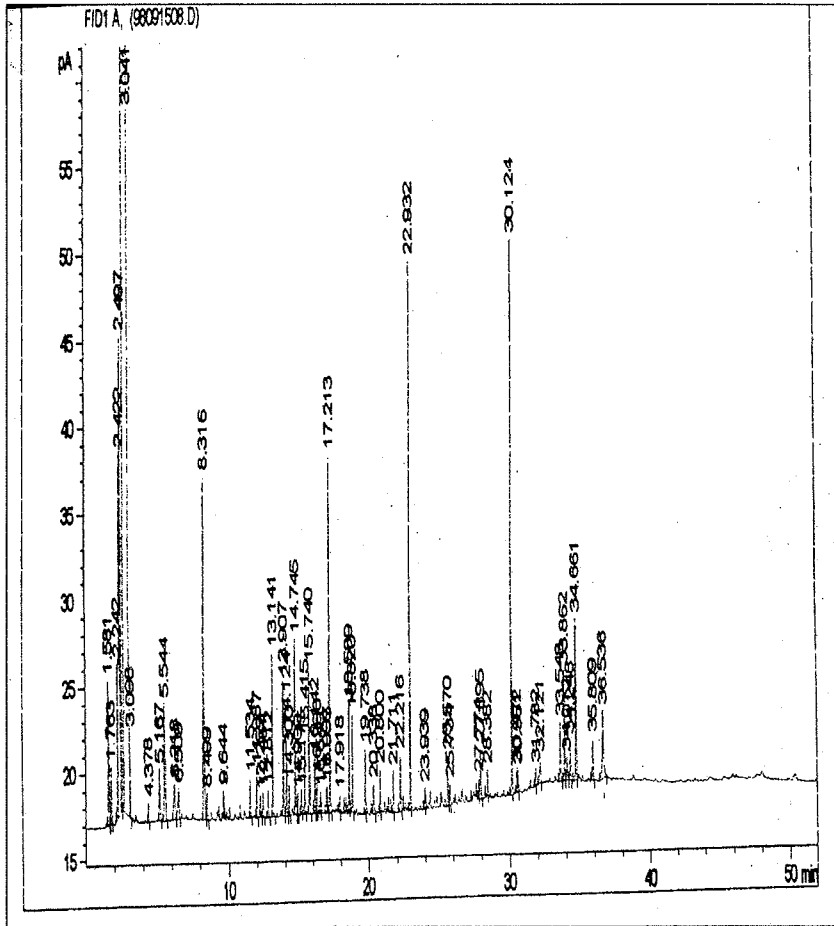


그림 15. 복숭아명나방피해 밤송이 메탄올추출물의 클로로포름 분획의 GC gram.(분석조건은 그림 14와 동일함)

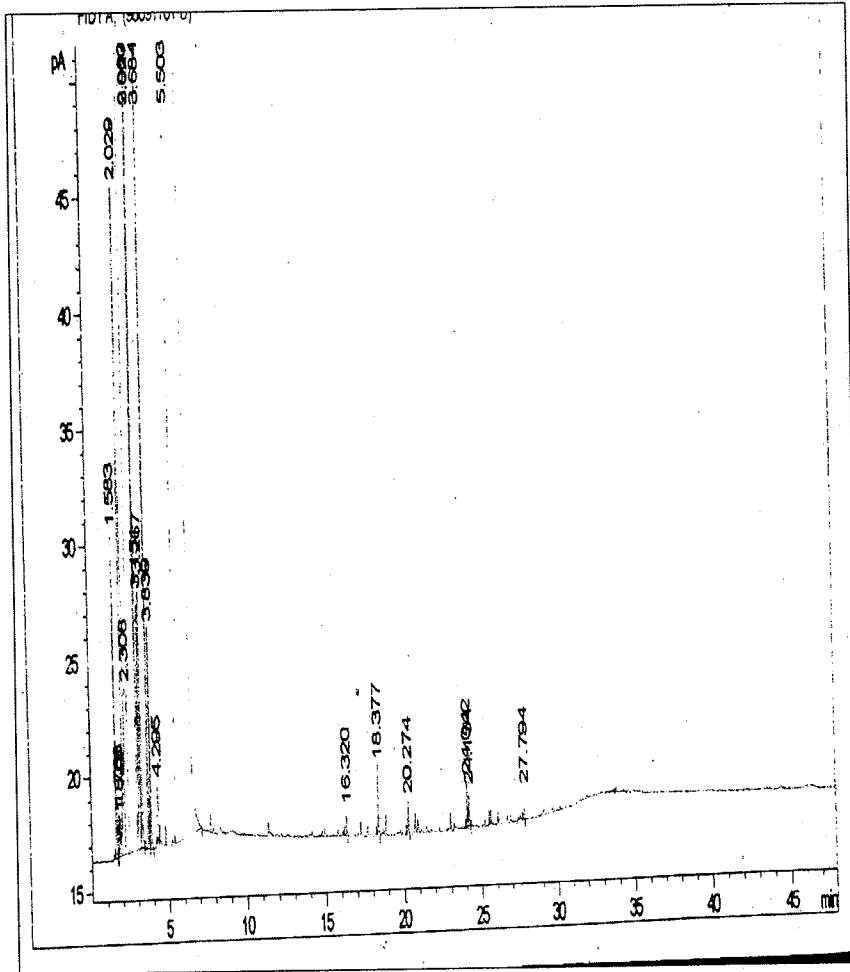


그림 16. 복숭아명나방 피해밤송이 메탄올추출물의
에틸아세테이트 분획의 GC gram (분석조건은 그림
14와 동일)

제 4 절. 적 요

1. 밤바구미에서 발산되는 화합물은 다수의 화합물이 검출되었고 근연종인 목화바구미(Boll weevil)의 성페로몬 조성과는 다른 성페로몬 체계를 갖고 있었으며 야외유인에서도 목화바구미의 성페로몬 성분으로는 전혀 유인되지 않았다.
2. 기주식물에 대해 밤바구미는 복숭아명나방 피해를 입은 밤송이에서 높은 유인력을 나타내었으며, 성별과 교미여부에 관계없이 같은 경향을 보여 복숭아명나방의 가해가 기주식물인 밤나무 특히 밤송이에서 이차대사산물의 변화를 야기하는 것으로 보이며 그 결과 밤바구미의 집합반응이 나타나는 것으로 여겨진다.
3. 기주식물의 방향성물질(green volatile) 중 유인성분을 탐색하기 위해 건전한 밤송이와 복숭아명나방 피해밤송이 추출물에 대한 olfactometer, EAG분석 결과 건전한 밤송이에서는 메탄올추출물의 클로로포름 분획에서 가장 높은 유인활성을 나타내었으며 EAG 반응도 가장 높았다.
4. 복숭아명나방피해 밤송이의 메탄올 추출물 중에서는 에틸아세테이트 분획에서 가장 높은 유인활성을 나타내었다.
5. 건전한 밤송이 메탄올추출물의 클로로포름 분획, 복숭아명나방피해 밤송이 메탄올 추출물의 클로로포름 분획과 복숭아명나방피해 밤송이 메탄올 추출물의 에틸아세테이트 분획에 대한 GC 분석 결과,

(+)- α -pinene, E-2-hexenal, benzaldehyde, n-decyl aldehyde, nonyl acetate, terpineol, geraniol, 2-phenylethanol 유지시간 비교를 통해 성분을 추정되었다.

6. 실내 생물검정으로 식물 유래 방향성화합물의 유인활성을 조사한 결과 (+)- α -pinene이 가장 높은 유인활성을 보여 유인제 성분으로 이용할 가능성이 있었다.

제 5 절. 인 용 문 헌

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