

원예 19412

최      중  
연구보고서

식물공장생산 방식에 의한 미나리의 재배기술 개발  
Development of water dropwort cultivation  
by plant factory production system

연구기관

서울대학교  
목포대학교

농림부  
농림부행정자료실



0005743

“

”

.

1997. 11.

:

:

(

)

:

(

)

.

.

가

.

,

.

가

가

가

가

가

.

.

,

,

.

,

.

paclobutrazol

, fog

CO2

CO2

, 가

NFT, DFT,

system

가

가

## SUMMARY

Conventionally, water dropwort has been cultivated in paddy fields after rice harvest. Thus, conventional method which was labor-intensive and usually conducted by old people in cold winter needs to be improved. Hydroponic culture with seed propagation is labor-saving compared to conventional method and can be conducted regardless of time and location which enables year-round production. Besides, hydroponic culture is safe from contamination by possible toxic elements like pesticides and heavy metals which can matter in the conventional method.

This research was conducted to investigate the optimum condition for seed propagation of water dropwort and to establish an efficient hydroponic system. The results were as follows.

1. Diverse variations were found among 119 lines from U.S.A., Japan and Korea, and their growth, flowering, seed harvesting and disease-tolerance were compared. Among 13 lines from Boseong local cultivar which produced more seeds than those from the other regions, 9 lines were selected for their high fertility and good seed harvesting characteristics. Paclobutrazol was effective in reducing lodging and stolon development, but delayed the flowering time and reduced the yield of seed harvesting.
2. pH in leachate from peat-moss, Baroker, and vermiculite bed soils were acidic, weak acidic, and alkaline, respectively. The germination percentage was highest in vermiculite or its mixture with the other bed soils. Growth of water dropwort seedlings were favorable with Baroker bed soil or its mixture with vermiculite but unsatisfactory with perlite or peat-moss.
3. The seed priming of water dropwort was effective in the rate of germination but the effect in PEG solution was not significantly higher than that in each salt solution. The seed priming in salt solutions was more effective in germination time and uniformity than that in PEG solution. The seed priming in 6%  $\text{Ca}(\text{NO}_3)_2$  solution for 3 days at 15 °C was the most effective in improving germination rate. The largest reduction in germination time by priming treatment occurred in 4%

KH<sub>2</sub>PO<sub>4</sub> solution for 3 days at 15 °C. While non-primed seeds did not emerge at all, primed seed in 4% KH<sub>2</sub>PO<sub>4</sub> solution for 3 days at 15 °C protruded the radicle in 3 days after sowing. The emergence time and emergence uniformity was not affected by priming. The emergence time was significantly reduced by priming in 4% KH<sub>2</sub>PO<sub>4</sub> and 4% KNO<sub>3</sub> solution for 3 days at 15 °C. The fresh weight of seedling increased by priming in 4% KNO<sub>3</sub> and 6% Ca(NO<sub>3</sub>)<sub>2</sub> solution for 3 days at 15 °C.

4. The effect of supplemental lighting on the growth of hydroponically grown water dropwort at the early or middle growth stage was not significantly different. Shading more than 50% excessively retarded growth, but control(0%) and 20% shading were effective. Fluorescent, incandescent and mercury lamp promoted the growth of water dropwort in the light quality experiment. Fog treatment was not effective in the growth of water dropwort grown hydroponically.
5. The CO<sub>2</sub> saturation concentration of *O. stolonifera* DC. seedling at 25 °C was proved to be 1500 µL/L. As CO<sub>2</sub> concentration increased up to 2200 µL/L, photosynthetic rate increased but stomatal conductance and transpiration rates decreased. The photosynthetic rate was significantly increased by CO<sub>2</sub> application under high light intensity of 320 µEm<sup>-2</sup>sec<sup>-1</sup>. The effect of CO<sub>2</sub> concentration on photosynthetic rate and transpiration rate was much higher in seedlings. As high concentration of CO<sub>2</sub> between 1500 and 2200 µL/L was applied, photosynthetic rate of water dropwort decreased in 100 µEm<sup>-2</sup>sec<sup>-1</sup>, while dark respiration increased. Under 2200 µL/L CO<sub>2</sub> application, plant height was longest through the whole phases of growth; T/R ratio, leaf area and specific leaf weight in CO<sub>2</sub> application were significantly higher than control during the late phase of growth.
6. The growth of hydroponically grown water dropwort was most favorable in An's solution but nitrate content was very high, which was 3,371 and 3,410mg/kg•FW in winter and summer, respectively. When cultured in conditioning solution, the nitrate content reduced to 840(24.9%) and 640mg/kg•FW(19.8%) in winter and summer, respectively. In tap water, severe nutrient deficiency appeared and foliar application of urea solution was not effective. In 6me/L chloride

and 3me/L ammonium substituted solution, growth was similar to control, but nitrate content was still 60% of control, while root was decayed because of sudden pH decline. Daylength extension during conditioning period was more effective on shoot growth than on root growth, while chlorophyll and nitrate content was not significantly changed. When chloride was added with ammonium during conditioning period, the higher was ammonium concentration in solution, the poorer the root growth. Nitrate content was not reduced significantly but chlorophyll content and visual quality were improved with 0.5me/L ammonium supply. Nitrate and chloride content in water dropwort showed significantly negative correlation and so did between nitrate content and fresh or dry weight.

7. With the decrease in planting distance, plant height increased and the number of stolons decreased. But adequate increase in planting distance helped photosynthesis due to enlarged canopy. Specifically, 15 × 5cm with 4 plants per hole in the spring and 10 × 10cm with 4 plants per hole in the autumn produced the highest yield in hydroponically grown water dropwort. Therefore, increase in planting distance within the range of optimum photosynthesis would be favorable for increasing the yield.
8. EC change was small for all systems except for deep aeroponics(80cm) while pH was more variable with hydroponics rather than with aeroponics. Aeroponics was more effective in growth and development of water dropwort than NFT and DFT. The growth of water dropwort was significantly higher in shallow bed(40cm) than deep bed in aeroponics. Growth and development of shoots were favorable in shallow bed but root growth was unsatisfactory in DFT. Chlorophyll content showed no differences among systems. Measured under natural condition, photosynthesis efficiency in NFT and deep aeroponics was highest at 10th day after planting, but measured under artificial light, it showed no difference between systems at 25th day after planting.

# CONTENTS

1. Introduction
  - 1.1. Purpose of research
  - 1.2. Contents and range of research
  - 1.3. A far-reaching influence of technique
2. Research for enhancement of seed harvesting efficiency
  - 2.1. Selection of desirable lines for seed harvesting and evaluation of seed harvesting efficiency by spray of paclobutrazol
    1. Introduction
    2. Materials and methods
    3. Results and discussion
  - 2.2. Selection of efficient substrate in raising seedling of seed-germinated water dropwort
    1. Introduction
    2. Materials and methods
    3. Results and discussion
  - 2.3. Establishment of efficient irrigation system and optimal fertilizing in culture of seed-germinated water dropwort for seed harvesting
    1. Introduction
    2. Materials and methods
    3. Results and discussion
  - 2.4. Effect of priming on stratificated seed germination and the early growth of water dropwort
    1. Introduction
    2. Materials and methods
    3. Results and discussion
3. Research for quality improvement of water dropwort
  - 3.1. Effect of supplement, shading, quality of light and fog spray on growth of hydroponically grown water dropwort



1. Introduction
2. Materials and methods
3. Results and discussion
- 3.2. Effect of CO<sub>2</sub> enrichment on the photosynthesis and growth of hydroponically grown water dropwort
  1. Introduction
  2. Materials and methods
  3. Results and discussion
- 3.3. Reduction of nitrate content in hydroponically grown water dropwort
  1. Introduction
  2. Materials and methods
  3. Results and discussion
4. Development of hydroponic system for water dropwort
  - 4.1. Clarification of optimum planting distance and planting number per cell in hydroponically grown water dropwort
    1. Introduction
    2. Materials and methods
    3. Results and discussion
  - 4.2. Clarification of optimum nutrient supplying method and irrigation interval in hydroponically grown water dropwort
    1. Introduction
    2. Materials and methods
    3. Results and discussion
  - 4.3. Effects of culture system on the growth of hydroponically grown water dropwort
    1. Introduction
    2. Materials and methods
    3. Results and discussion
5. Reference

1

1

2

3

2

1

pachlobutrazol

1.

2.

3.

2

1.

2.

3.

3

1.

2.

3.

4

priming

1.

2.

3.

3

1

, ,

fog 가

1.

2.

3.  
2 CO2

1.  
2.  
3.  
3

1.  
2.  
3.

4

1  
1.  
2.  
3.

2  
1.  
2.  
3.

3                    가  
1.  
2.  
3.

5

1

1

가

가

가

가

가

2

paclobutrazol

priming

, fog

CO2

CO2

NFT, DFT

system

3

가

1,200

300

1000

가

가

가

가

가 가 가

2

1

paclobutrazol

1.

가 .

(paclobutrazol)

2.

가.

119

( . 1991) 1/2

5 ,  
 , ,  
 .

Fig. 1. The culturing landscape of comparison among clones selected 'BoSung' local variety.

	가 가		13	
clone		가 4	29	. 1,000
kg/10a,	21.1,	15.0	가 20.0 kg/10a	,
				.
			가	, ,
,	. 13		가 가	



. Paclobutrazol

Fig. 2. The landscape of water dropwort treated by paclobutrazol.

plug tray 가 60 3 3  
4 29 .  
90cm .  
30cm, 1 .  
1,000 kg/10a, 21.1, 15.0 가 20.0 kg/10a  
,

1  
Paclobutrazol  
1 6 10 , 7 6  
0, 10, 25, 50, 100ppm  
10, 25, 50ppm . 7 6

3.  
가.

119  
1  
가  
130cm  
30cm  
1 15  
40-50 가  
1 가  
가 가  
가 가  
가

가

Table 1. Comparison of main characteristics among collected water dropwort local variety.

Characteristic	Local variety <sup>z</sup>	
Vigor	upright type	Kwangyang
	Z type	Sunchun
Stolon length	long( 130cm)	Kongju, Bosung, Seungju
	short( 30cm)	Samchuck, Yungwol, Okku
Superiority of resistance to virus and rust		Kimpo, Namyangju, Suwon, Yungwol, Yungdong, Yungchun, Woolsan, Pyung chang,
Time to blooming	early flowering	Kunsan, Namyangju, Myungju, Bongwha, Pusan, Buyu, Sokcho, Yungdong, Changsu, Chunju
	late flowering	Bongwha
No. of flower cluster	large	Kwangyang, Dangjin, Sunchang <sup>2</sup> , Ibaraki, Jaechun, Chungju
	small	Kunsan, Kimjae, Namyangju, Muan, Boeun, Buyu, Yungdong, Yungam, Yungwol <sup>2</sup> , Eumsung, Chunju, Cheju, Jinyang <sup>1</sup> , Jinyang <sup>2</sup> , Changnyung, Chunan, Hwasung <sup>2</sup>
No. of floret/flower cluster	large( 35)	Bosung, Sunchang <sup>1</sup> , Ibaraki, Hwasung <sup>1</sup>
	small( 15)	Koje, Namyangju, Woolleung, Jinan, Changnyung
Cluster diameter	big( 9cm)	Konju, Bosung, Seungju, Chilkok
	small( 3.5cm)	Samchuck, Yungwol, Okku
Earliness of seed harvesting ( 170 days)		Kongju, Bonghwa, Yungyang, Ckhulwon, Chungwon
Seed fertility rate	high( 50%)	Kunsan, Kimpo, Kimhae <sup>1</sup> , Kimhae <sup>2</sup> , Pusan, Buyu <sup>2</sup> , Changsu
	low( 50%)	Bonghwa, Suwon, Okku, Changnyung, Tongyung <sup>1</sup> , Tongyung <sup>2</sup> , Pyungchang
Low seed drop		Namyangju, Muan, Yungdong, Okku, Eusung, Cheju, Jinchun, Chungyang, Tongyung <sup>1</sup>
Superiority of seed maturation		Kunsan, Bonghwa, Okku, Woolleung, Woolsan, Eusung, Chungyang, Tongyung, Hadong

<sup>z</sup> Superscript number indicate selecting rank of local variety

가

가

2 , 13 4 9  
가 가

Table 2. The comparison of growth and seed-harvesting characteristics among water dropwort.

Local cultivar	Plant height (cm)	Stem length (mm)	Degree of overgrowingx	Degree of lodgingy	Degree of seed dropz	Amount of seed harvest (ml/m <sup>2</sup> )	1000 seed weight (g)
A7	107.3	13.6	2	3	3	107.3	1.42
A9	124.7	14.6	4	4	3	150.1	1.65
C3	115.3	14.1	3	2	1	110.2	1.46
E6	114.0	13.8	5	5	4	120.1	1.53
F1	160.0	14.7	3	2	3	140.6	1.50
S1	73.3	14.1	1	1	3	86.5	1.19
X1	88.3	14.3	2	2	2	135.5	1.63
Z7	114.0	13.8	3	3	3	145.4	1.50
U.S.A. derived	70.7	7.0	1	1	-	-	-

x, y, z 1: not severe 3: average 5: severe

F1 A9 , S1 가  
가 가  
S1 가  
S1

. 가 A9  
 E6 가 . C3 가  
 .  
 A9 F1 S1 가 .  
 . A9 X1 가  
 , S1 가 가  
 . 가  
 가 ,  
 가 .  
 , 가 ,  
 ,

. Paclobutrazol

3 6 10 paclobutrazol 0, 10, 25, 50  
 100 ppm , 7 6 10, 25 50ppm  
 , 9 20 .  
 6 10 , 가  
 7 6 , 6 10  
 가  
 가 . 7 .  
 .  
 가 , 가  
 . 6 10 7 6 .

Table 3. The effect of spray time and concentration of paclobutrazol on growth of water dropwort.

Paclobutrazol concentration ( ppm )	Plant height (cm)	Stem length <sup>x</sup> (cm)	Stem diametry (mm)	No. of internode	No. of stolons	Stolon length (cm)	Dry weight of stolon (g)
<u>10 June</u>							
0	131.2 az	37.8 a	14.6 a	17.1 a	4.2 a	144.1 ab	132.1 a
10	98.6 c	26.3 b	14.9 a	15.1 b	4.2 a	130.0 b	90.2 b
25	86.3 d	15.4 c	12.3 b	14.9 b	4.0 a	125.2 b	60.1 bc
50	84.6 d	12.9 cd	13.3 b	14.3 b	4.0 a	114.9 bc	53.4 c
100	74.6 d	10.6 d	12.9 bc	14.2 b	4.0 a	96.7 c	49.9 c
<u>6 July</u>							
10	110.1 bc	38.1 a	14.0 ab	16.5 ab	4.3 a	151.7 a	120.7 ab
25	112.0 b	37.3 a	14.0 ab	15.5 b	4.2 a	151.5 a	118.3 ab
50	103.8 bc	28.3 b	13.7 b	15.6 b	4.2 a	142.1 ab	95.2 b

<sup>x</sup> The length from first to 7th internode

<sup>y</sup> Second internode

<sup>z</sup> Duncan's multiple range test,  $p=0.05$

3 6 10 . 7 6 8 10 1  
7 13 20  
가

7 27 100% 10ppm 55% 가  
. 2 8 10 90%  
가 . 7 20 가 85% 25ppm,  
50ppm, 100ppm 6% , 8 10  
50%, 33%, 11.6%

4 7 6  
8 20 10ppm 35%, 25ppm 15%,  
50ppm 11.3%  
가 8 10 80%

5 6 10 8  
27 가 가  
100ppm 50% 가  
가 가  
gibberellin gibberellin 10ppm  
가 5  
1-6 7 6  
가 . chemical  
가





Fig. 1-4- -6

Table 4. The effect of Paclobutrazol treated at 10 June on fruit setting characteristics.

Paclobutrazol concentration ( ppm )	No. of fruits/flower cluster				No. of cluster /plant	Fruit amount (ml/m <sup>2</sup> )	1000 seed weight (g)
	1u	2w	3x	4y			
0	832.1az	542.9a	363.0a	333.9a	11.7a	113.4a	1.51a
10	635.9b	449.3ab	349.4a	283.9ab	10.5a	96.4b	1.50a
25	531.3bc	380.3b	357.9a	324.2a	7.8b	93.1b	1.49a
50	497.4c	273.2c	297.8b	251.2ab	5.7bc	85.1bc	1.51a
100	465.9c	336.8bc	262.9b	227.2b	4.6c	77.8c	1.52a

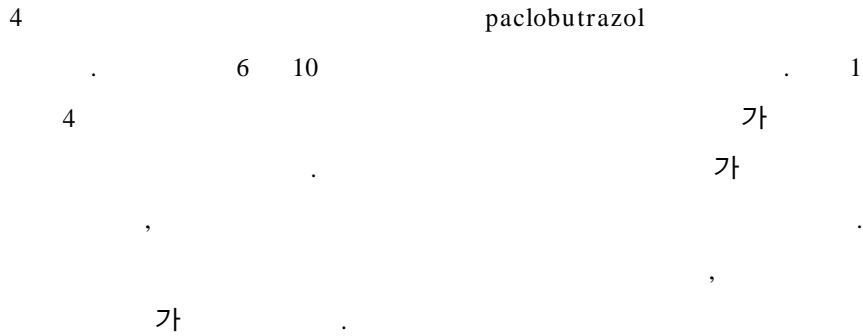
u First cluster

w Second cluster

x Third cluster

y Fourth cluster

z Duncan's multiple range test,  $p=0.05$



2

1.

가  
가  
가 ( )  
가

2.

가.

1995

, , vermiculite perlite

1)

1995 9

5:5

4cC 가 . , 2

2)

(PM), (B), (V), (P, 3  
5mm) PM:V, PM:P, PM:S, B:P, B:V, P:V 가 1 :

1

3)

1997 3 13 1997 5 10  
. 3 13 160 flug tray 5  
8 3 24 tray ,  
30 3  
1

2

4)

3 28 , 4 4 , 2 가  
4 11 . 5 10  
, , , , ,

Duncan .

3.

가.

1995

, , vermiculite perlite  
1 .

Table 1. Germination status of water-dropwort seed in various media.

Treatment <sup>z</sup>	Standing rate(%)	No. of seedlings/hole
PS	54.3	0.83
US	75.3	1.93
PS : US = 1 : 1	66.7	1.56
LM	81.6	2.53
S	81.3	2.47
L.M : S = 1 : 1	78.3	2.38
P	96.8	3.93
V	100	3.98
P : V = 1 : 1	100	3.96

<sup>z</sup> PS; Paddy soil, US; Upland soil, LM; Leaf mould, S; Sand, P; Perlite, V; Vermiculite,

50% 1  
75%  
2  
82% , 가 2.6 81%  
2.9 가  
vermiculite perlite 100%  
가 가  
perlite 가 vermiculite

가 가

plug tray

vermiculite perlite

perlite vermiculite

1/2

2

10cm

3.5

가

1.

3 28

가

( 2).

1

2

1

가

Table 1. Effects of substrates on germination rate and true leaf development of *Oenanthe stolonifera* DC.

Substratez	Germination rate (%)		Development of true leaf (4/11) (%)
	3/28	4/4	
PM : V	18.1	61.4	42.3
PM : P	8.6	45.6	11.0
PM : S	26.9	70.6	34.8
B	31.2	71.0	65.7
B : P	24.9	67.8	46.3
B : V	29.8	70.9	69.6
V	29.3	72.7	35.1
P : V	31.6	72.9	32.4

z PM ; peatmoss, V ; vermiculite, B ; Baroker, P ; perlite, S; sand

2.

Tisdale (1975)

가 pH

peat

pH 5

( 3).

pH가 6 7

pH

pH

가

Table 3. pH of each substrate

Leaching	PM:Vz	PM:P	PM:S	B	B:P	B:V	V	P:V
Distilled water	4.64	4.27	4.37	5.58	5.88	6.40	8.51	8.04
T a p water	4.84	4.65	4.74	5.85	6.68	6.70	7.43	7.39

z PM ; peatmoss, V ; vermiculite, B ; Baroker, P ; perlite, S ; sand

3. 가

가  
가

가

( 4).

Table 4. Effects of substrates on growth of *O. stolonifera* DC. seedlings.

Substrate <sup>z</sup>	Plant height (cm)	Root length (cm)	Fresh weight (g)		Dry weight (g)	
			Leaf	Root	Leaf	Root
Pm : V	15.4by	10.5a	0.558b	0.324c	0.041a	0.015bc
Pm : P	10.3d	9.2a	0.386cde	0.243d	0.030a	0.015bc
Pm : S	13.2c	8.7a	0.461bc	0.305c	0.036a	0.019b
B	18.4a	9.9a	0.715a	0.402b	0.054a	0.013c
B : P	13.7bc	10.3a	0.439cd	0.303c	0.036a	0.017bc
B : V	17.6a	9.7a	0.671a	0.477a	0.055a	0.027a
V	9.2d	9.2a	0.316e	0.259d	0.024a	0.013c
P : V	12.2c	10.5a	0.399cde	0.246d	0.031a	0.013c

<sup>z</sup> PM ; peatmoss, V ; vermiculite, B ; Baroker, P ; perlite, S; sand

<sup>y</sup> Mean separation within columns at p = 0.05 by Duncan's multiple range test.



. Schmilewski Gunther(1988) 가

Pivot(1988)

가

(Fakhri et al).

Spomer(1979)

72%(Manios , 1972)

가

가

3

1.

, , 가 ,

가

2.

가.

- 3

3.

가.

- 3

1 .

Table 1. Growth of water dropwort under various fertilizer level after 1, 2 and 3 months.

Treatment	Plant height (cm)			No. of branches		
	1	2	3(month)	1	2	3(month)
Standard (control)	26.5	65.3	102.3	2.8	4.1	9.2
NPK double	31.6	70.2	109.7	3.3	5.1	10.2
N double	30.8	71.3	108.6	3.2	4.8	9.9
P double	34.2	75.4	110.2	4.5	6.1	10.6
K double	36.3	81.2	112.4	4.2	5.2	10.3
NP double	32.5	70.4	107.3	4.1	5.4	10.5
NK double	25.8	66.2	103.6	4.5	6.2	10.2
PK double	30.3	68.7	104.4	4.0	5.6	10.3

K P

N K P 가

.

,

가 가

8 9

가 가 .

가

,

,

가 가

,

가 K

,

가

.

2

Table 2. Effect of various level of fertilizer on flowering and fruiting of water dropwort.

Treatment	Flow clousters /plant	No. of fruits /plant	Yield (ml/m <sup>2</sup> )	1000- grain weight (g)
Standard (control)	10.3	3,276	114.7	1.50
NPK double	11.2	3,475	121.7	1.51
N double	10.8	3,324	116.4	1.48
P double	10.9	3,316	116.1	1.52
K double	11.2	3,516	123.2	1.51
NP double	10.5	3,469	121.5	1.49
NK double	10.2	3,397	119.0	1.50
PK double	10.6	3,417	119.7	1.50

3

Table 3. Effects of irrigation type on the growth of water dropwort.

Treatment	Plant height(cm)			No. of branches		
	1	2	3 (month)	1	2	3 (month)
Furrow irrigation	22.3	58.4	98.3	2.4	3.2	8.9
Mini sprinkler	26.7	65.5	106.7	2.5	4.1	9.7
Below z	27.2	70.6	110.4	2.6	4.5	10.5
Above y	25.3	64.2	101.6	2.4	3.9	9.8

z Irrigation below the mulching film by watering hose

y Irrigation above the mulching film by watering hose

Table 4. Effects of irrigation type on the flowering and fruiting of water dropwort.

Treatment	No. of closters /plant	No. of fruits /plant	Yield (ml/m <sup>2</sup> )	1000- grain weight (g)
Furrow irrigation	9.2	3,087	108.1	1.51
Mini sprinkuler	9.9	3,294	115.4	1.49
Belowz	10.2	3,387	117.9	1.52
Abovey	9.7	3,254	114.0	1.51

z Irrigation below the mulching film by watering hose

y Irrigation above the mulching film by watering hose

가

가

4

## priming

1.

가

1986).

가

priming

Terry (1992) *Aquilegia canadensis*

priming

priming

Osmotic priming

priming

(Brocklehurst, 1983; Pill, 1986; Szafirowska, 1981).

priming

priming

2.

1993 9

5 : 5

2 4

가. Priming

Ca(NO<sub>3</sub>)<sub>2</sub>, NaH<sub>2</sub>PO<sub>4</sub>, NaNO<sub>3</sub>, KH<sub>2</sub>PO<sub>4</sub>, KNO<sub>3</sub>, mannitol

PEG(polyethylene glycol) 6000

Ca(NO<sub>3</sub>)<sub>2</sub>, NaNO<sub>3</sub>

KH<sub>2</sub>PO<sub>4</sub>, KNO<sub>3</sub> 2, 4 6%

NaH<sub>2</sub>PO<sub>4</sub> 3, 5 7%

mannitol 5, 10 15%

, PEG 6000 20, 30 40%

Priming 400mg

50ml

20ml

가

24

. Priming

15, 20 25 3, 7 10

25 가 2 3 가  
 , 4 ± 1

petri-dish (Whatman No.2) 1 , 50  
 5 5ml 가 25 ± 1 , 24  
 . Petri-dish 2 ,  
 1mm .  
 12

- GR(germination rate) = ( N / S ) × 100
- MGT(mean germination time) = ( t<sub>ni</sub> ) / N
- GU(germination uniformity) = [ (MGT - t<sub>i</sub>)<sup>2</sup>n<sub>i</sub> ] / N - 1

N : , S :  
 t<sub>i</sub> : , n<sub>i</sub> :

9 13 10 17

10/33 ( / )

)



- ER(emergence rate) = ( N / S ) × 100
- MET(mean emergence time) = (  $\sum t_{ni}$  ) / N
- EU(emergence uniformity) = [ (MET -  $t_i$ )<sup>2</sup> / N - 1

N : , S :

$t_i$  : ,  $n_i$  :

(

15 ) ( , 1991) ,

NO<sub>3</sub>-N 13, NH<sub>4</sub>-N 3, P 3, K 6, Ca 2, Mg 1me/l

Fe 3ppm, B 0.5, Mn 0.5, Zn 0.05, Cu 0.02 Mo

0.01ppm , Ca(NO<sub>3</sub>)<sub>4</sub> · 4H<sub>2</sub>O, NaNO<sub>3</sub> KNO<sub>3</sub>

NH<sub>4</sub>H<sub>2</sub>PO<sub>4</sub> MgSO<sub>4</sub> · 7H<sub>2</sub>O, EDTA · Na · Fe · 3H<sub>2</sub>O, H<sub>3</sub>BO<sub>3</sub> MnSO<sub>4</sub> ·

4H<sub>2</sub>O, ZnSO<sub>4</sub> · 7H<sub>2</sub>O, CuSO<sub>4</sub> · 5H<sub>2</sub>O Na<sub>2</sub>MoO<sub>4</sub> · 2H<sub>2</sub>O .

35 , , ,

Priming , 1 , 2

3 (SEM:scanning electron

microscope) .

(5% paraformaldehyde + 5% glutaraldehyde) 가 4 12

, buffer 3 ethanol series

, CO<sub>2</sub> coating

150 .

3.

가. Priming 가

15 7 priming

1 . KNO<sub>3</sub> 6% PEG 20%

58% , NO<sub>3</sub>-

. Haigh (1987) NO<sub>3</sub>-

, priming NO<sub>3</sub>-

. KH<sub>2</sub>PO<sub>4</sub> priming 4.2 4.7

, 가

PEG , priming

(Akers , 1987)가

가

가 (Mexal ,

1975) 가가 priming .

priming PEG

, .

, priming

NO<sub>3</sub>- Ca(NO<sub>3</sub>)<sub>2</sub> NaNO<sub>3</sub>

가 KH<sub>2</sub>PO<sub>4</sub> .

, priming

priming ( 2,

3, 4).

Table 1. Effect of osmotica treatment on the germination rate(GR), mean germination time(MGT) and germination uniformity(GU) of *O. stolonifera* DC. seeds.

Seed treatment <sup>w</sup>		GR(%)	MGT(days)	GU
Osmotica	Conc.(%)			
Ca(NO <sub>3</sub> ) <sub>2</sub>	2	65.5 bcdefz	5.8 bcd	3.37 a
	4	77.5 abc	5.9 bcd	3.89 a
	6	75.8 abcd	5.5 bcde	2.39 a
NaH <sub>2</sub> PO <sub>4</sub>	3	64.0 def	6.9 a	2.72 a
	5	78.2 ab	5.4 bcde	1.65 a
	7	65.5 bcdef	5.4 bcde	1.80 a
NaNO <sub>3</sub>	2	71.7 abcde	5.3 cdef	1.50 a
	4	73.1 abcde	5.8 bcd	3.63 a
	6	61.1 ef	6.1 abc	2.90 a
KH <sub>2</sub> PO <sub>4</sub>	2	63.5 def	4.4 fg	1.52 a
	4	66.0 abcdef	4.7 egf	2.56 a
	6	72.0 abcde	4.2 g	1.52 a
KNO <sub>3</sub>	2	60.1 ef	5.4 bcde	4.43 a
	4	64.4 cdef	5.6 bcd	5.17 a
	6	57.4 f	6.2 ab	3.31 a
Mannitol	5	73.2 abcde	5.6 bcde	3.06 a
	10	65.4 bcdef	5.3 bcde	2.22 a
	15	64.1 def	6.1 bc	3.88 a
PEG <sub>x</sub>	20	57.6 f	5.1 def	3.21 a
	30	78.7 a	5.1 def	2.25 a
	40	76.4 abcd	5.4 bcde	2.88 a
Control <sup>y</sup>		58.0 f	5.0 def	1.97 a

<sup>w</sup> Seeds were primed for 7 days at 15 °C in a growth chamber with light.

<sup>x</sup> PEG is polyethylene glycol 6000.

<sup>y</sup> Seeds were stratified with sand in the outdoors.

<sup>z</sup> Mean separation within columns by Duncan's multiple range test at 5% level

2 15  
 priming . NO<sub>3</sub>  
 Ca(NO<sub>3</sub>)<sub>2</sub> NaNO<sub>3</sub> KNO<sub>3</sub> priming

, KH<sub>2</sub>PO<sub>4</sub>  
 , priming  
 Ca(NO<sub>3</sub>)<sub>2</sub> 6%, NaNO<sub>3</sub> 4%,  
 KH<sub>2</sub>PO<sub>4</sub> 6%, KNO<sub>3</sub> 4% 가  
 3  
 priming 가 가  
 3 20 , priming  
 priming  
 가 KH<sub>2</sub>PO<sub>4</sub>  
 3 , 3  
 3 priming ,  
 Ca(NO<sub>3</sub>)<sub>2</sub> 6%, NaNO<sub>3</sub> 2%, KH<sub>2</sub>PO<sub>4</sub> 6%,  
 KNO<sub>3</sub> 4%  
 4 25 , priming  
 KNO<sub>3</sub> ,  
 2% , 3  
 . 25  
 KH<sub>2</sub>PO<sub>4</sub> KNO<sub>3</sub> 7  
 58% , 10 priming

Table 2. Effect of the duration and concentration of osmotica treatment on the germination rate (GR) of *O. stolonifera* DC. seeds primed at 15 in a lighted growth chamber.

Seed treatment		Duration of treatment(days)		
Osmotica	Conc.(%)	3	7	10
<i>Germination rate(%)</i>				
Ca(NO <sub>3</sub> ) <sub>2</sub>	2	76.0	65.2	65.6
	4	77.5	77.5	68.1
	6	81.1	76.5	80.8
NaNO <sub>3</sub>	2	78.8	71.7	72.9
	4	79.2	73.3	65.6
	6	75.2	61.1	51.5
KH <sub>2</sub> PO <sub>4</sub>	2	74.4	63.2	65.3
	4	73.2	66.0	65.1
	6	76.3	72.0	72.0
KNO <sub>3</sub>	2	76.1	60.1	56.2
	4	79.6	64.4	63.8
	6	79.2	57.4	55.2
Controlz		58.0		
Contrastsy				
Salt(A)			***	
Concentration(B)			NS	
Duration(C)			***	
A × B			***	
A × C			**	
B × C			NS	
A × B × C			NS	

y Contrasts were calculated by partitioning the treatment effects using SAS general linear models procedure. NS, \*, \*\*, \*\*\* mean non-significant and significant at P 0.01, 0.05, 0.001 respectively.

z Seeds were stratified with sand in the outdoors.

Table 3. Effect of the duration and concentration of osmotica treatment on the germination rate (GR) of *O. stolonifera* DC. seeds primed at 20 in a lighted growth chamber.

Seed treatment		Duration of treatment(days)		
Osmotica	Conc.(%)	3	7	10
<i>Germination rate(%)</i>				
Ca(NO <sub>3</sub> ) <sub>2</sub>	2	72.0	62.8	49.2
	4	66.5	73.5	68.8
	6	75.2	67.2	71.7
NaNO <sub>3</sub>	2	81.3	77.2	55.2
	4	73.5	74.2	57.1
	6	74.2	62.7	36.6
KH <sub>2</sub> PO <sub>4</sub>	2	77.7	70.6	59.9
	4	74.5	68.0	66.8
	6	78.0	71.4	57.6
KNO <sub>3</sub>	2	77.2	66.3	53.5
	4	70.5	72.8	55.2
	6	77.2	61.2	44.8
Control <sup>z</sup>		58.0		
Contrasts <sup>y</sup>				
Salt(A)			ns	
Concentration(B)			ns	
Duration(C)			***	
A × B			***	
A × C			***	
B × C			**	
A × B × C			ns	

<sup>y</sup> Contrasts were calculated by partitioning the treatment effects using SAS general linear models procedure. NS, \*, \*\*, \*\*\* mean non-significant and significant at P 0.05, 0.01, 0.001, respectively.

<sup>z</sup> Seeds were stratified with sand in the outdoors.

Table 4. Effect of the duration and concentration of osmotica treatment on the germination rate (GR) of *O. stolonifera* DC. seeds primed at 25 °C in a lighted growth chamber.

Seed treatment		Duration of treatment(days)		
Osmotica	Conc.(%)	3	7	10
<i>Germination rate(%)</i>				
Ca(NO <sub>3</sub> ) <sub>2</sub>	2	62.4	60.1	42.5
	4	71.3	67.6	50.8
	6	74.7	67.2	60.0
NaNO <sub>3</sub>	2	63.0	70.0	52.2
	4	74.1	65.5	51.2
	6	57.6	47.4	27.6
KH <sub>2</sub> PO <sub>4</sub>	2	64.6	47.0	44.8
	4	59.8	33.1	31.7
	6	59.8	35.8	22.1
KNO <sub>3</sub>	2	76.8	59.0	59.9
	4	73.6	60.6	52.2
	6	78.3	52.8	46.0
Controlz		58.0		
Contrastsy				
Salt(A)			***	
Concentration(B)			***	
Duration(C)			***	
A × B			***	
A × C			***	
B × C			*	
A × B × C			*	

y Contrasts were calculated by partitioning the treatment effects using SAS general linear models procedure. NS, \*, \*\*, \*\*\* mean non-significant and significant at P = 0.01, 0.05, 0.001, respectively.

z Seeds were stratified with sand in the outdoors.

fig 1.



1  
, priming .  
15 20 25  
, 가  
, KNO<sub>3</sub> priming 가  
, KNO<sub>3</sub> 2%  
가 .  
priming  
(15, 20, 25 ) pri-  
ming  
Haigh (1986) .  
Ca(NO<sub>3</sub>)<sub>2</sub> 6%  
, priming Ca(NO<sub>3</sub>)<sub>2</sub>가  
(1994) .  
, priming 5, 6, 7 .  
5 15  
priming .  
priming ,  
KH<sub>2</sub>PO<sub>4</sub> 가 , 3  
, priming  
가 , Ca(NO<sub>3</sub>)<sub>2</sub> 6% , NaNO<sub>3</sub>  
4% , KH<sub>2</sub>PO<sub>4</sub> 4% , KNO<sub>3</sub> 4% 가  
, 3  
가 .

Table 5. Effect of the duration and concentration of osmotica treatment on the mean germination time(MGT) of *O. stolonifera* DC. seeds primed at 15 in a lighted growth chamber.

Seed treatment		Duration of treatment(days)		
Osmotica	Conc.(%)	3	7	10
<i>Mean germination time (days after sowing)</i>				
Ca(NO <sub>3</sub> ) <sub>2</sub>	2	4.8	5.8	5.4
	4	4.8	5.9	5.2
	6	4.9	5.5	5.4
NaNO <sub>3</sub>	2	4.4	5.2	4.8
	4	4.4	5.8	5.4
	6	5.2	6.1	6.2
KH <sub>2</sub> PO <sub>4</sub>	2	4.2	4.5	4.5
	4	4.0	4.7	4.6
	6	4.4	4.2	4.7
KNO <sub>3</sub>	2	4.7	5.4	5.3
	4	4.9	5.6	4.6
	6	5.1	6.2	4.8
Controlz		5.0		
Contrastsy				
Salt(A)			***	
Concentration(B)			**	
Duration(C)			***	
A × B			***	
A × C			***	
B × C			ns	
A × B × C			**	

y Contrasts were calculated by partitioning the treatment effects using SAS general linear models procedure. NS, \*, \*\*, \*\*\* mean non-significant and significant at P 0.01, 0.05, 0.001, respectively.

z Seeds were stratified with sand in the outdoors.

Table 6. Effect of the duration and concentration of osmotica treatment on the mean germination time(MGT) of *O. stolonifera* DC. seeds primed at 20 in a lighted growth chamber.

Seed treatment		Duration of treatment(days)		
Osmotica	Conc.(%)	3	7	10
<i>Mean germination time(days after sowing)</i>				
Ca(NO <sub>3</sub> ) <sub>2</sub>	2	4.8	5.2	4.9
	4	4.9	4.8	4.2
	6	5.1	4.8	4.5
NaNO <sub>3</sub>	2	4.9	4.5	4.1
	4	5.0	5.1	4.9
	6	5.6	6.1	6.5
KH <sub>2</sub> PO <sub>4</sub>	2	4.5	4.9	4.9
	4	4.5	4.1	4.0
	6	4.5	4.3	4.3
KNO <sub>3</sub>	2	4.7	5.0	4.5
	4	4.6	4.8	4.0
	6	5.2	5.0	5.0
Controlz		5.0		
Contrastsy				
Salt(A)			***	
Concentration(B)			***	
Duration(C)			***	
A × B			***	
A × C			***	
B × C			***	
A × B × C			***	

y Contrasts were calculated by partitioning the treatment effects using SAS general linear models procedure. NS, \*, \*\*, \*\*\* mean non-significant and significant at P 0.01, 0.05, 0.001, respectively.

z Seeds were stratified with sand in the outdoors.

Table 7. Effect of the duration and concentration of osmotica treatment on the mean germination time(MGT) of *O. stolonifera* DC. seeds primed at 25 in a lighted growth chamber.

Seed treatment		Duration of treatment(days)		
Osmotica	Conc.(%)	3	7	10
<i>Mean germination time(days after sowing)</i>				
Ca(NO <sub>3</sub> ) <sub>2</sub>	2	5.6	6.6	6.9
	4	5.5	6.8	6.2
	6	5.6	7.3	7.1
NaNO <sub>3</sub>	2	5.8	5.6	6.7
	4	6.0	6.1	6.3
	6	6.1	5.7	7.0
KH <sub>2</sub> PO <sub>4</sub>	2	5.2	4.7	4.9
	4	6.0	5.1	5.6
	6	5.0	5.0	6.1
KNO <sub>3</sub>	2	5.1	6.0	5.4
	4	5.0	6.6	5.9
	6	5.8	6.6	6.7
Controlz		5.0		
Contrastsy				
Salt(A)			***	
Concentration(B)			***	
Duration(C)			***	
A × B			***	
A × C			***	
B × C			***	
A × B × C			**	

y Contrasts were calculated by partitioning the treatment effects using SAS general linear models procedure. NS, \*, \*\*, \*\*\* mean non-significant and significant at P 0.01, 0.05, 0.001, respectively.

z Seeds were stratified with sand in the outdoors.

fig 2.

6 20

priming

. 15

가 KH<sub>2</sub>PO<sub>4</sub>

, NaNO<sub>3</sub>

가 ,

5.0

가

NaNO<sub>3</sub> 15

가 .

7 25

priming

.

15

20

가 KH<sub>2</sub>PO<sub>4</sub>가

,

2

priming

.

20

Ca(NO<sub>3</sub>)<sub>2</sub> KNO<sub>3</sub>

, KH<sub>2</sub>PO<sub>4</sub>

1

5

가

. 25

가

KH<sub>2</sub>PO<sub>4</sub>

4%

가

KH<sub>2</sub>PO<sub>4</sub> priming

(1994)

priming

8, 9, 10

8 15

priming

. KH<sub>2</sub>PO<sub>4</sub>

가

,

KH<sub>2</sub>PO<sub>4</sub> 4%

priming

가

, KNO<sub>3</sub>

가

3

가

, Ca(NO<sub>3</sub>)<sub>2</sub> NaNO<sub>3</sub> 2% , KH<sub>2</sub>PO<sub>4</sub> KNO<sub>3</sub> 6%

Table 8. Effect of the duration and concentration of osmotica treatment on the germination uniformity(GU) of *O. stolonifera* DC. seeds primed at 15 in a lighted growth chamber.

Seed treatment		Duration of treatment(days)		
Osmotica	Conc.(%)	3	7	10
<i>Germination uniformityz</i>				
Ca(NO <sub>3</sub> ) <sub>2</sub>	2	1.93	3.38	3.30
	4	1.75	3.89	3.92
	6	2.44	3.08	3.48
NaNO <sub>3</sub>	2	2.55	1.50	2.12
	4	1.48	3.70	3.29
	6	2.36	2.90	2.91
KH <sub>2</sub> PO <sub>4</sub>	2	2.06	1.68	2.08
	4	1.19	2.56	3.20
	6	1.62	1.52	2.29
KNO <sub>3</sub>	2	1.92	4.43	3.75
	4	2.33	5.17	2.78
	6	2.46	3.31	2.51
Controlx		1.97		
Contrastsy				
Salt(A)			***	
Concentration(B)			ns	
Duration(C)			***	
A × B			***	
A × C			**	
B × C			*	
A × B × C			ns	

x Seeds were stratified with sand in the outdoors.

y Contrasts were calculated by partitioning the treatment effects using SAS general linear models procedure. NS, \*, \*\*, \*\*\* mean non-significant and significant at P 0.01, 0.05, 0.001 respectively.

z Germination uniformity =  $[(MGT - t_i) \sum n_i] / N - 1$

MGT:mean germination time, t<sub>i</sub>days after sowing, n<sub>i</sub>No. of germinate seeds on day t<sub>i</sub>

N:total no. of germinate seeds

Table 9. Effect of the duration and concentration of osmotica treatment on the germination uniformity(GU) of *O. stolonifera* DC. seeds primed at 20 in a lighted growth chamber.

Seed treatment		Duration of treatment (days)		
Osmotica	Conc.(%)	3	7	10
<i>Germination unimityZ</i>				
Ca(NO <sub>3</sub> ) <sub>2</sub>	2	1.58	4.34	3.32
	4	2.33	3.53	2.77
	6	1.78	2.91	3.67
NaNO <sub>3</sub>	2	2.04	2.43	2.53
	4	1.76	4.02	4.32
	6	2.16	4.08	3.51
KH <sub>2</sub> PO <sub>4</sub>	2	1.20	4.15	3.35
	4	1.26	1.10	2.08
	6	1.00	0.75	1.77
KNO <sub>3</sub>	2	1.22	3.11	2.58
	4	1.33	3.23	2.48
	6	1.91	2.60	3.76
ControlX		1.97		
ContrastsY				
Salt(A)			***	
Concentration(B)			ns	
Duration(C)			***	
A × B			**	
A × C			ns	
B × C			ns	
A × B × C			*	

x Seeds were stratified with sand in the outdoors.

y Contrasts were calculated by partitioning the treatment effects using SAS general linear models procedure. NS, \*, \*\*, \*\*\* mean non-significant and significant at P 0.01, 0.05, 0.001, respectively.

Z Germination uniformity =  $[(MGT - t_i)2n_i] / N - 1$

MGT:mean germination time, t<sub>i</sub>days after sowing, n<sub>i</sub>No. of germinate seeds on day t<sub>i</sub>

N:total no. of germinate seeds.



Table 10. Effect of the duration and concentration of osmotica treatment on the germination uniformity(GU) of *O. stolonifera* DC. seeds primed at 25 in a lighted growth chamber.

Seed treatment		Duration of treatment (days)		
Osmotica	Conc.(%)	3	7	10
<i>Germination uniformityz</i>				
Ca(NO <sub>3</sub> ) <sub>2</sub>	2	3.80	6.85	3.67
	4	4.39	5.75	4.73
	6	3.09	5.47	4.11
NaNO <sub>3</sub>	2	5.83	2.31	4.72
	4	4.34	3.30	4.91
	6	5.68	2.31	5.22
KH <sub>2</sub> PO <sub>4</sub>	2	3.12	3.65	2.22
	4	4.82	3.34	3.37
	6	3.16	3.82	4.29
KNO <sub>3</sub>	2	3.77	6.48	3.06
	4	2.79	6.14	4.97
	6	2.82	5.57	5.63
Controlx		1.97		
Contrastsy				
Salt(A)			**	
Concentration(B)			ns	
Duration(C)			***	
A × B			ns	
A × C			***	
B × C			ns	
A × B × C			ns	

x Seeds were stratified with sand in the outdoors.

y Contrasts were calculated by partitioning the treatment effects using SAS general linear models procedure. NS, \*, \*\*, \*\*\* mean non-significant and significant at P 0.01, 0.05, 0.001, respectively.

Z Germination uniformity =  $[(MGT - t_i)ni] / N - 1$

MGT:mean germination time, tidays after sowing, niNo. of germinate seeds on day ti

N:total no. of germinate seeds

fig 3.

9 20  
priming , 15 가 20  
KH<sub>2</sub>PO<sub>4</sub> 가 , 6%  
KH<sub>2</sub>PO<sub>4</sub> 3  
가  
가 .

10 25  
priming , KH<sub>2</sub>PO<sub>4</sub>  
가 , ,  
가 .  
3  
priming . 25  
가 , 20  
KH<sub>2</sub>PO<sub>4</sub> priming .

Khan(1992) priming priming  
, ,  
가 , ,  
priming Smith Cobb(1991)  
priming  
, ,  
priming 가 ,  
25 7 10 priming  
, ,  
가 가  
, priming  
15 , Ca(NO<sub>3</sub>)<sub>2</sub> 6% 가 ,  
15 , KH<sub>2</sub>PO<sub>4</sub> 4% 가

, priming 15 Ca(NO<sub>3</sub>)<sub>2</sub> 6%, NaNO<sub>3</sub>  
 4%, KH<sub>2</sub>PO<sub>4</sub> 4%, KNO<sub>3</sub> 4% 3 priming 가

fig 4.

Fig 4. Scanning electron microscope(SEM) micrograph( $\times 150$ ) of a longitudinal section along the flat of *Oenanthe stolonifera* DC. seeds. A-C; non-primed seeds. D-F; seeds were primed with 4% KH<sub>2</sub>PO<sub>4</sub> in the light at 15 for 3 days. R; radicle, E; endosperm and S; seed coat (A, D; 1 day • B, E; 2 days • C, F; 3 days after imbibition)

2. Priming

4 15 3 KH<sub>2</sub>PO<sub>4</sub> 4% priming .  
 1 , priming  
 , 3  
 priming  
 . PEG KH<sub>2</sub>PO<sub>4</sub>+  
 KNO<sub>3</sub> priming , embryo 가  
 Weibe Tissen(1979) .  
 ,  
 , (1995) priming  
 ,  
 . priming  
 , micropylar cavity  
 .

3. Priming 가

11  
 priming , priming  
 .  
 가 ,  
 priming 가 . ,  
 KNO<sub>3</sub> 4% 1 , KH<sub>2</sub>PO<sub>4</sub> 4%  
 가 . priming  
 .

Table 11. Effect of priming on the emergence rate(ER), mean emergence time(MET) and emergence uniformity(EU) of *O. stolonifera* DC. seedlings in a greenhouse

Seed treatmentz		ER(%)	MET(days)	EU
Osmotica	Conc.(%)			
Ca(NO <sub>3</sub> ) <sub>2</sub>	6	77.6 ax	11.1 a	2.01 a
NaNO <sub>3</sub>	4	80.0 a	11.0 ab	2.06 a
KH <sub>2</sub> PO <sub>4</sub>	4	79.6 a	10.6 bc	2.10 a
KNO <sub>3</sub>	4	76.0 a	10.3 c	1.59 a
Controly		73.6 a	11.2 a	2.20 a

x Mean separation within a column by Duncan's multiple range test at 5% level.

y Seeds were stratified with sand in the outdoors.

z Seeds were primed for 3 days at 15 °C in a growth chamber with light. Seeds were sowed in urethan sponge.

Table 12. Effect of priming on the seedling growth of *O. stolonifera* DC. in 35 days after sowing.

Seed treatmentz		Plant height(cm)	No. of leaves	Fresh weight(g)
Osmotica	Conc.(%)			
Ca(NO <sub>3</sub> ) <sub>2</sub>	6	8.7 ax	2.9 a	1.48 ab
NaNO <sub>3</sub>	4	8.9 a	3.2 a	1.41 b
KH <sub>2</sub> PO <sub>4</sub>	4	8.7 a	2.7 a	1.26 c
KNO <sub>3</sub>	4	8.8 a	3.0 a	1.50 a
Controly		8.6 a	3.1 a	1.30 bc

x Mean separation within a column by Duncan's multiple range test at 5% level.

y Seeds were stratified with sand in the outdoors.

z Seeds were primed for 3 days at 15 °C in a growth chamber with light. Seeds were sowed in urethan sponge. Seedlings were grown with An's solution in a greenhouse.

(Gray, 1976),

12 35 priming 가  
가 KNO3 4% 가 가 ,  
Ca(NO3)2 4% 가  
Ca(NO3)2 priming 가 ,  
(1995) , Brocklehurst  
Dearman(1984) KH2PO4 priming  
priming ,  
(Greenway, 1980). priming ,  
priming 가  
priming 가  
Priming  
, priming

3

1 , , fog 가

1.

,  
.  
가 .

,  
가 . , ,

2.

가.

30/15

6 30

8 25

9 1 . 1

2m × 1m

60W

3 1m .

PE film 가

가 .

10

3

1



7 30 9 15

, 9 25

2 1m  
90,000lux , 20% 70,000lux, 50% 46,000lux,  
80% 17,000lux 10

7 30 9 15

9 25 , 가

1 60W 3 ,  
40W 2 , 200W 1  
BCR film 2m × 1m × 1m  
BCR film 10

. Fog

6 30 8 25 12m  
20m , 9

1

, fog

Fog 10

3.  
가.

1 2 10 가  
 , , 가 ,  
 가 30 가 ,  
 가 10 , 30  
 가

Fig 3. The growth response of water dropwort treated with different supplemental lighting period

3 가  
 .  
 , .

Table 1. The effect of shading on growth of hydroponically grown water dropwort.

Rate of shading (%)	Sampling day	Plant height (cm)	Leaf width (mm)	Stem diameter (mm)	No. of stems
0	5 October	11.4	8.9	2.0	4.0
	15 October	22.3	10.9	4.0	4.0
	20 October	37.2	20.0	4.5	5.0
20	5 October	10.1	8.7	2.1	4.1
	15 October	18.4	11.9	3.2	4.1
	20 October	38.2	23.1	3.5	4.5
50	5 October	13.0	10.8	2.1	4.1
	15 October	20.9	12.4	3.2	4.5
	20 October	29.3	18.5	4.0	5.6
80	5 October	14.6	11.7	2.0	3.7
	15 October	24.7	12.6	2.4	4.0
	20 October	28.2	14.2	2.6	5.0

Fig 4. The growth response of water dropwort treated with different shading rate.

1 , 가  
50% 80% 가  
20% 가  
가 가  
80%  
50%  
가  
가  
가 5 BCR film 가 가  
6 가 가

2-6

7

가 가

가

가

가

8

가

,

가 BCR film

,

가

. Fog

Fig 9. The growth response of water dropwort due to fog treatment.

9 fog 가

.

가 fog

가 (250 )

,

가 가



가 ,

fog

가

가

7, 8

가

가

가

가

## 2 CO2

1.

가 , 가 가  
가 .  
가  
가  
가 .  
가 가 가 가  
가 , 가  
가 가  
가 1200 1800 ppm .  
가 가  
가 . 가  
가 가  
가 가  
가 .  
가 , ,  
가  
1000 1500ppm ,  
2000ppm 가  
1500 2500ppm .  
가 가  
1  
2 3 .  
가 .

2.

2 10

2 17 45 가

3

4 10

가

가

, 가

7:30

9:30

CO2 sensor(Horiba GH- 250E)

CO2

sensor Data logger(LI 1000)

274cm × 90.5cm

, 82cm × 60.5cm

42

, 10cm × 10cm

가

25

가

2

Photosynthesis system(LI 6400)

3.

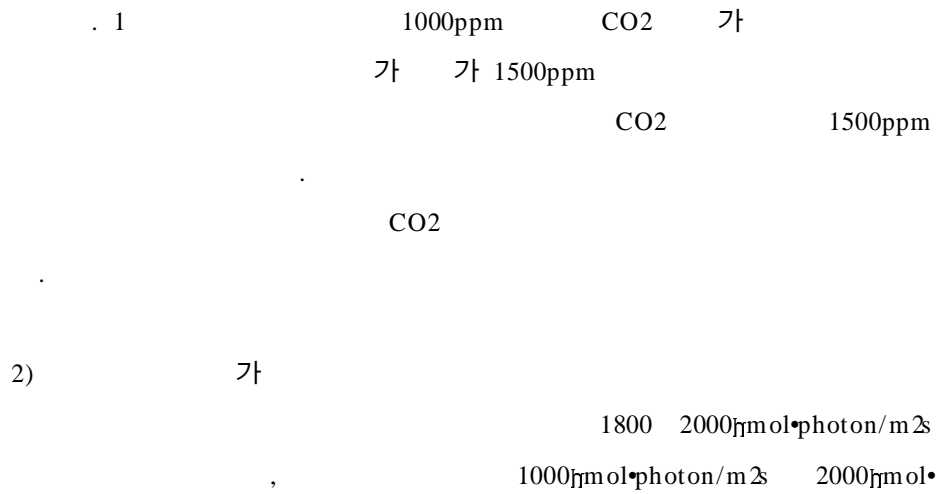
(가) CO2

1) CO2

, 가

( 1).

Fig. 1. The CO<sub>2</sub> saturation curve of *O. stolonifera* DC at , 2000 $\mu$ mol•  
 photon/m<sup>2</sup>•s



photon/m<sup>2</sup>s, 가 CO<sub>2</sub> 가  
 . CO<sub>2</sub> 가  
 .  
 3) 가  
 10 , 20 , 25 ,  
 . 2 ,  
 , 가 CO<sub>2</sub> 가  
 가 .

Fig. 2. The effects of temperature on photosynthesis of *O. stolonifera*  
 DC at 2000 μmol•photon/m<sup>2</sup>•s

( ) 가  
 CO<sub>2</sub> 20  
 . CO<sub>2</sub> 가  
 ,  
 . CO<sub>2</sub> , 가

( ) 가 , 1 가 , 가 2200ppm 가 가 1500ppm 750ppm 가 가 2200ppm 가 가 , 750ppm 가 가 가 CO2 가 CO2 가

Table 1. The effects of CO2 application on photosynthesis of *O. stolonifera* DC..

Date	Treatment	Plant height (cm)	Patiol length (cm)	No. of leaf	No. of runner	No. of internod	Fresh weight(g)		Dry weight(g)	
							leaf	root	leaf	root
4/29	2200ppm	27.1 az	11.1 a	4.6	2.4	-	13.9	5.51	0.89	0.33
	1500ppm	23.0 b	8.6 b	4.9	2.7	-	13.9	6.33	0.93	0.35
	750ppm	19.9 c	8.4 b	3.8	2.6	-	12.0	5.55	0.89	0.35
	Control	22.9 b	7.9 b	4.1	3.0	-	14.8	6.53	1.03	0.38
	F	17.96***	6.59**	1.59ns	1.05ns	-	0.94ns	1.53ns	0.68ns	1.70ns
5/6	2200ppm	39.8 a	20.9 a	7.0	1.9	2.0 a	32.9	8.10	2.10	0.42
	1500ppm	36.5 b	19.1 a	7.0	1.5	1.3 ab	28.1	6.73	1.89	0.38
	750ppm	33.3 c	16.0 b	7.0	1.5	0.9 b	25.1	6.85	1.80	0.39
	Control	38.1 ab	18.6 a	5.3	2.4	2.0 a	28.1	6.53	1.84	0.42
	F	6.80**	5.31**	2.33ns	1.18ns	4.61**	0.98ns	1.14ns	0.36ns	0.17ns
5/18	2200ppm	64.75 a	41.5 a	9.17	0.75 b	4.0	78.4	9.13	5.23 a	0.51
	1500ppm	61.58 a	38.0 a	9.58	0.67 b	4.1	71.2	9.44	5.22 a	0.53
	750ppm	55.0 b	32.4 b	9.50	1.08 b	3.6	57.5	10.3	3.83 b	0.52
	Control	62.42 a	37.6 a	8.50	2.08 a	3.7	68.9	9.7	4.53 ab	0.54
	F	9.02**	6.67***	0.87ns	6.43***	2.54ns	2.49ns	0.37ns	2.95*	0.12ns

z Mean separation within a column by duncan's multiple range at 5% level

\*, \*\*, ns Significant at P 0.05, 0.01 and nonsignificant respectively

가  
가

가

3

1.

( , 1987).

가

( , 1987).

가

가

가

nitrosamine (gastric cancer) (Forman, 1985; , 1992).

nitrite hemoglobin methemoglobin 8-10%가 ( ; methemoglobinemia, blue baby syndrome) (Wright Davison, 1964; Maynard, 1976; Sohn Oh, 1993).

(xylem)

(Blom-Zandstra, 1989).

1 143mg

83.9% 120mg

(World Health Organization)

3.75mg/kg( ), 0.13mg/kg



endive  
 4,500mg/kg 3,500mg/kg , 3,500mg/kg  
 2,500mg/kg , 3,000mg/kg ,  
 (Reinink Groenwold, 1987; Benoit Wiebe,  
 1992; Richardson Hardgrave, 1992; Reinink , 1994).

(solution conditioning)

2.

12 10 9 11  
 polyethylene(PE) film

25 ,  
 25%

3

Conditioning

( , 1991) ,

NO<sub>3</sub>-N 11, NH<sub>4</sub>-N 3, P 3, K 4, Ca 2, Mg 1me/L Fe 3ppm,

B 0.5, Mn 0.5, Zn 0.05, Cu 0.02 Mo 0.01ppm ,

NO<sub>3</sub>-N 13, K 6me/L

NaNO<sub>3</sub> KNO<sub>3</sub> Ca(NO<sub>3</sub>)<sub>2</sub> NH<sub>4</sub>H<sub>2</sub>PO<sub>4</sub> MgSO<sub>4</sub>·7H<sub>2</sub>O,

EDTA·Na·Fe·3H<sub>2</sub>O, H<sub>3</sub>BO<sub>3</sub> MnSO<sub>4</sub>·4H<sub>2</sub>O, ZnSO<sub>4</sub>·7H<sub>2</sub>O, CuSO<sub>4</sub>·5H<sub>2</sub>O

Na<sub>2</sub>MoO<sub>4</sub>·2H<sub>2</sub>O

pH EC 6.2 1.36

Hanna instruments) pH meter (HI8424, conductivity meter (HI8733, )

120 dry oven 1  
65 3 ,  
40mm 가

HClO<sub>4</sub>:H<sub>2</sub>SO<sub>4</sub>:H<sub>2</sub>O=90:5:55

indophenol- blue ( , 1990) , NO<sub>3</sub>-N 100mg 45 1  
Cataldo (Cataldo , 1975) , PO<sub>4</sub>-P  
vanadate ( , 1988)

Shimadzu UV-2201 UV-VIS spectrophotometer

665, 410 470nm . K, Ca

Mg automic absorption spectrophotometer(Perkin Elmer Model 303) ( , 1990) . Cl 1g 1/4

CaO 500 (M30A-2C, Blue M lab)

pH 6 7 .

5% potassium chromate 5 0.05N- AgNO<sub>3</sub>

(牛島 , 1981).

$Cl(mg/g \cdot DW) = 1.77 \times AgNO_3 (ml)$

2 2g

100% ethanol 24 5 649 665nm

(牛島 , 1981).

chlorophyll a (mg/l) = 13.7A65 - 5.76A69

chlorophyll b (mg/l) = 25.8A69 - 7.6A65

total chlorophyll (mg/l) = 6.1A65 + 20.04A69

, 3 , SAS

### 1. Conditioning NO3- 가

12 10 2 17

polyurethane sponge

, 4 5 ( 10cm)

cell 2 styrofoam(90 x 60 x 3cm, W x L x D) 4 1

, bed styrofoam

450L 가 3cm

, 25 가

2 300L 가

60W 2 17:00 24:00HR, 08:00 09:00HR

16 30

conditioning , KCl 4,

CaCl2 2, MgSO4 1 KH2PO4 3 me/L NO3-N

Cl-(6me/L) [ Cl ] , KCl

4, CaCl2 2, MgSO4 1, NH4H2PO4 3 (NH4)2CO 1 me/L

NO3-N NH4+ : Cl- = 3 : 6(me/L)가

3 4 1,000ppm .  
 , 450L  
 .  
 Conditioning 2 14 17 8 EC  
 pH ,  
 .

## 2. Conditioning

1995 3 18 5 28  
 . 3 cell 2 styrofoam  
 , 1 ,  
 . 36 conditioning  
 , 1 Cl  
 . 17:00 22:00<sub>HR</sub> , 08:00 09:00<sub>HR</sub> 200W  
 ( 110  $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ ) 2 14  
 , 17:00 04:00<sub>HR</sub> 08:00 09:00<sub>HR</sub>  
 20 150W ( 320  $\mu$   
 $\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ ) 20 ,  
 .  
 Conditioning 2 ,  
 , (total nitrogen), NO<sub>3</sub>-N, P, K, Ca, Mg Cl

3. Conditioning NH<sub>4</sub><sup>+</sup>

7 14 9 11 P.E. film plastic house  
 KNO<sub>3</sub> 2me/L가 가  
 가 가  
 3 8 8 styrofoam , 17  
 22 conditioning  
 1 Cl , 6 NH<sub>4</sub><sup>+</sup>  
 가 0.5, 1.0 2.0me/L NH<sub>4</sub>Cl 가 .  
 가 Cl-

pH, EC 8 25 8 30 9  
 11 3 conditioning  
 가 chroma meter(CR- 200, Minolta)

3.

1. Conditioning NO<sub>3</sub>- 가

1 conditioning EC pH  
 EC NH<sub>4</sub>-Cl 가  
 pH Cl ,  
 가 가  
 (Maynard , 1976; Ikeda Osawa,  
 1980; Ikeda Osawa, 1981; , 1991)

5 4.63 가 가

, Cl+NH4

3.59 .

가 OH-

, CO2 HCO3- pH ,

H+ pH

(Kirkby Mengel, 1967). NO3NH4

NH4 pH

(1991) .

30 7

, Cl 6me/L Cl 6me/L +NH4 3me/L

conditioning 1 .

가 ,

가

Cl+NH4 .

가

( $p=0.05$ ).

Cl 가 ,

가

가 .

가

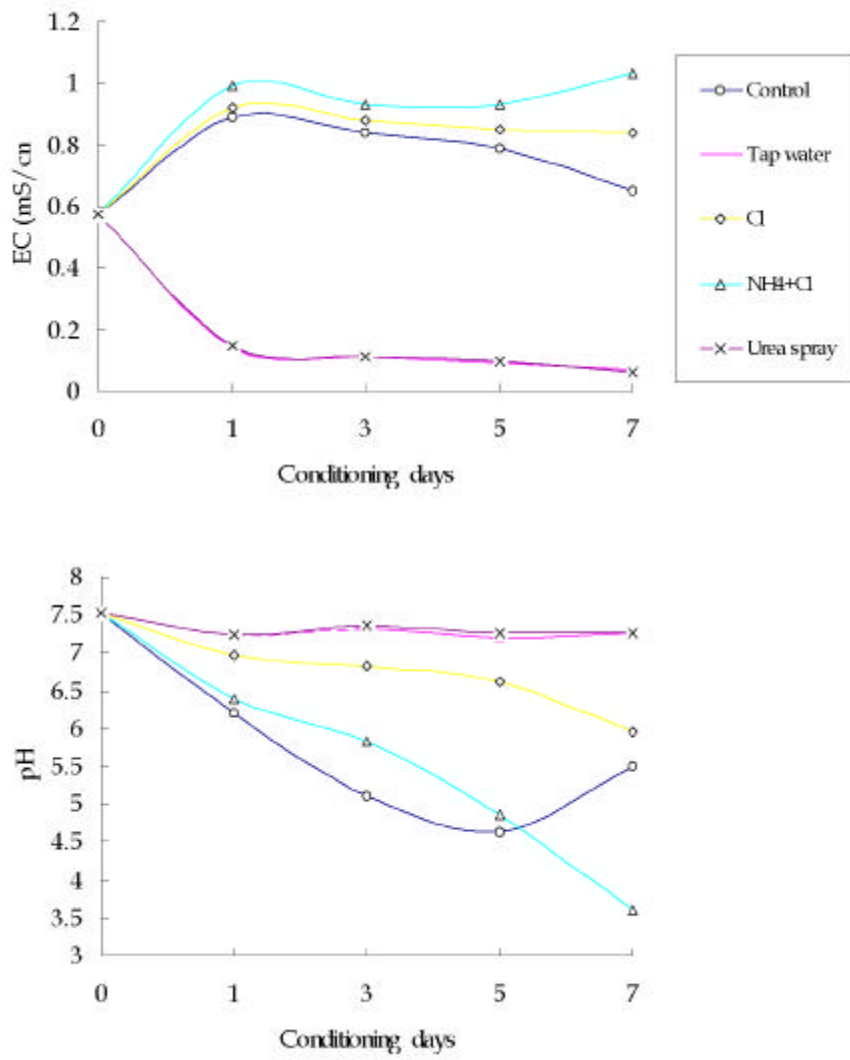


Fig. 1. The EC and pH changes of nutrient solution used for *O. stolonifera* DC. culture during conditioning period.

Table 1. The conditioning effects of nitrogen free nutrient solution applied for 7 days before harvest on the growth of hydroponically grown *Oenanthe stolonifera* DC.

Conditioning <sup>z</sup>	Plant height (cm)	Internode		Runner		Root length (cm)	Fresh weight(g)		Dry weight(g)		T/R ratio
		No.	length (cm)	No.	length (cm)		leaf	root	leaf	root	
Control	59.6 <sup>ay</sup>	5.9 <sup>a</sup>	0.80 <sup>b</sup>	6.8 <sup>a</sup>	53.1 <sup>a</sup>	39.88 <sup>ab</sup>	41.27 <sup>a</sup>	2.55 <sup>a</sup>	3.02 <sup>a</sup>	0.15 <sup>a</sup>	16.73 <sup>a</sup>
Tap water	57.8 <sup>a</sup>	4.9 <sup>c</sup>	1.46 <sup>a</sup>	5.8 <sup>ab</sup>	47.8 <sup>a</sup>	42.10 <sup>a</sup>	28.12 <sup>b</sup>	2.67 <sup>a</sup>	1.86 <sup>b</sup>	0.12 <sup>ab</sup>	11.53 <sup>b</sup>
Cl	57.3 <sup>a</sup>	5.1 <sup>bc</sup>	1.22 <sup>a</sup>	5.7 <sup>b</sup>	49.1 <sup>a</sup>	33.95 <sup>cd</sup>	31.10 <sup>a</sup>	2.53 <sup>a</sup>	2.17 <sup>b</sup>	0.14 <sup>ab</sup>	12.06 <sup>b</sup>
Cl + NH <sub>4</sub>	62.0 <sup>a</sup>	5.6 <sup>a</sup>	1.28 <sup>a</sup>	5.9 <sup>ab</sup>	47.5 <sup>a</sup>	28.63 <sup>d</sup>	30.20 <sup>a</sup>	2.32 <sup>a</sup>	1.99 <sup>b</sup>	0.11 <sup>ab</sup>	12.73 <sup>a</sup>
Tap water + urea spray	57.8 <sup>a</sup>	5.1 <sup>bc</sup>	1.43 <sup>a</sup>	6.3 <sup>ab</sup>	46.7 <sup>a</sup>	35.95 <sup>bc</sup>	28.12 <sup>b</sup>	2.21 <sup>a</sup>	2.08 <sup>b</sup>	0.10 <sup>b</sup>	12.80 <sup>a</sup>

<sup>z</sup> Control=An's solution; Cl=substitution of Cl(6me/L) for nitrogen; Cl+NH<sub>4</sub>=substitution of Cl-(6me/L) and NH<sub>4</sub>(3me/L) for nitrogen; tap water+urea spray=foliar spray of 1,000ppm urea soln.

<sup>y</sup> Duncan's multiple range test,  $p=0.05$

Kafkafi, 1980; Ruth Kafkafi, 1983), (Ruth Kafkafi, 1983), pH (Bennett , 1964; Warncke Barker, 1973).

가 , Cl Cl+NH<sub>4</sub> 가 . 가 + 가 가 , 가 .



가  
+ 가  
가 ,  
가 ,  
가  
가  
3  
가 가  
2 ,  
가  
가  
CI 가 CI+NH4  
5 ,  
가  
가  
가

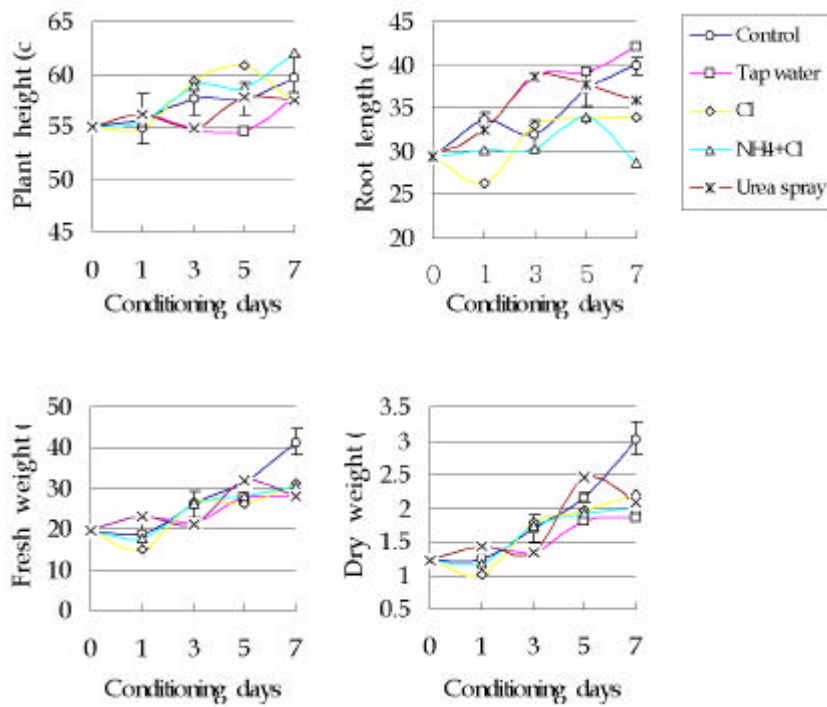


Fig. 2. The conditioning effects of nutrient substitution for 7 days before harvest on the growth changes of hydroponically grown *O. stolonifera* DC. Vertical bars indicate SE.

3

3,653ppm  
 4,388ppm      가      가  
 ,      3,300ppm  
 ,      7  
 1,949ppm  
 Cl      3



Cl				
가	1,597ppm		Cl	
			가	
2				
	3,371ppm		Cl	840ppm
	75.1%			
	1,539ppm	54.4%		

Table 2. The effects of nutrient solution substitution for conditioning solution for 7 days before harvest on nitrate and chloride contents of hydroponically grown *Oenanthe stolonifera* DC.

Conditioningz	Nitrate content (mg/kg FW)	Chloride content (mg/kg FW)	Percentage over control	
			nitrate	chloride
Control	3371.3ay	673.4d	100.0 %	100.0 %
Tap water	1949.0b	833.2c	57.8	123.7
Cl	839.9c	1596.9a	24.9	237.1
Cl + NH4	2023.2c	1133.9b	60.0	168.4
Tap water + urea spray	1538.9b	830.4c	45.6	123.3

zControl=An's solution; Cl=nitrogen was substituted by Cl(6me/L) in nutrient solution; Cl+NH4= nitrogen was substituted by Cl(6me/L) and NH4(3me/L); tap water+urea spray=foliar spray of 1,000ppm urea solution.  
yDuncan's multiple range test,  $p=0.05$

Cl+NH4 42.2 40%

Cl 2.4 가 ,  
 가 가 가 .  
 Veen Kleinendorst(1985) ,  
 PEG NaCl osmotic stress  
*Lolium multiflorum* , 가

4 conditioning

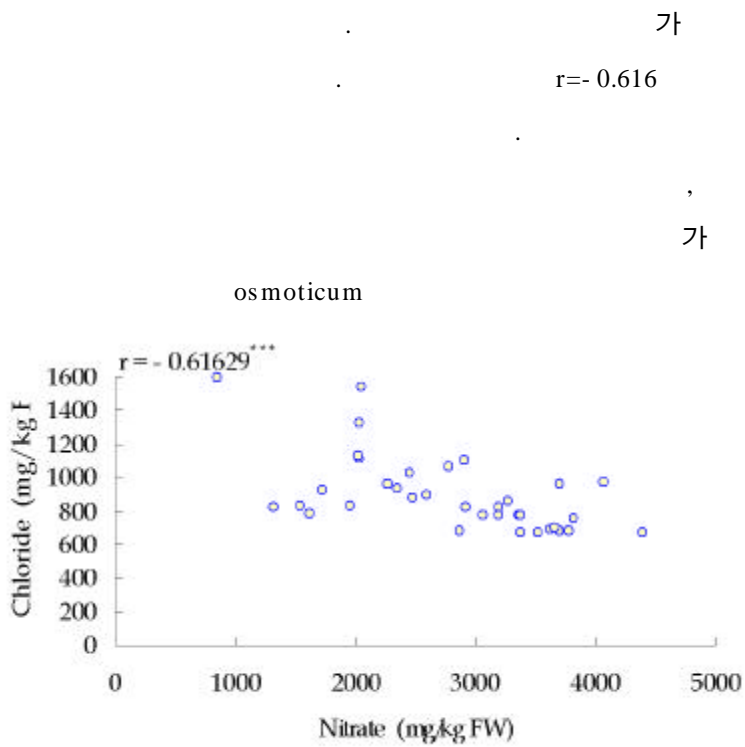


Fig. 4. The correlation between nitrate and chloride contents of hydroponically grown *O. stolonifera* DC. when nutrient solution was substituted by conditioning solution for 7 days.

Bloom- Zandstra Lampe(1985) , 52

가 가 plant sap

가

osmoticum

가

## 2. Conditioning

36 5 conditioning  
EC pH 5  
EC 0.54 1.0 가  
pH  
110  $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$  20 6.86  
6.5 6.6  
pH pH  
pH  
3

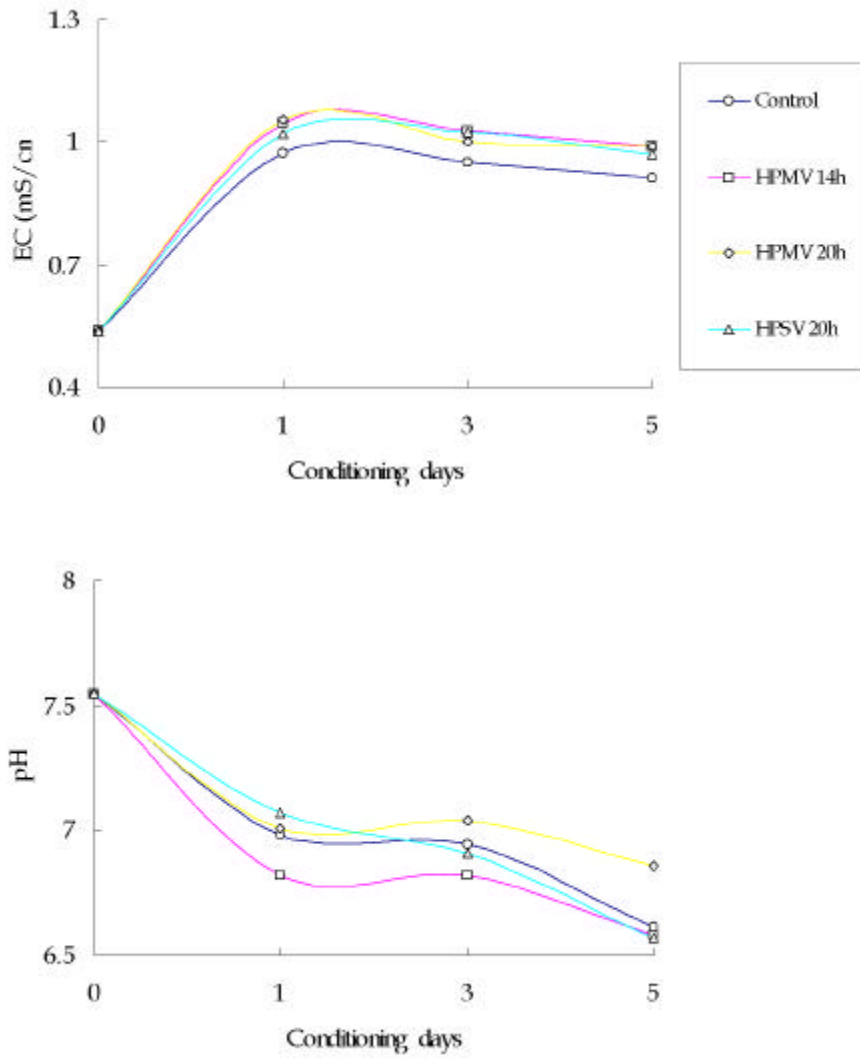


Fig. 5. The effects of photoperiod and photosynthetic photon flux on changes of EC and pH of nutrient solution used for *O. stolonifera* DC. culture during conditioning period. HPMV(high pressure mercury vapor lamp, 110  $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ ), HPSV(high pressure sodium vapor lamp, 320  $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ ).

Table 3. The effects of supplementary lighting with substitution of conditioning solution for nutrient solution for 5 days before harvest on growth of hydroponically grown *Oenanthe stolonifera* DC.

Photoperiod <sup>z</sup>	Plant height (cm)	Internode		Runner		Root length (cm)	Fresh weight(g)		Dry weight(g)		T/R ratio
		No.	length (cm)	No.	length (cm)		leaf	root	leaf	root	
Control	70.7aby	5.2a	5.17a	7.0a	69.8a	41.13a	64.61ab	4.11b	4.57a	0.46a	17.34a
14h HPMV	67.8b	4.7b	5.03a	6.8a	62.7b	32.89b	58.93b	6.19a	4.49a	0.38a	9.74c
20h HPMV	72.7a	5.4a	5.11a	6.4a	69.5a	38.18a	73.45a	6.16a	3.85a	0.40a	12.32b
20h HPSV	68.5ab	5.0a	5.02a	6.7a	69.5a	42.84a	70.97ab	4.22a	4.22a	0.39a	15.36a

<sup>z</sup> Control=natural daylength; HPMV=200W high pressure mercury vapor lamp(110 μmol/m<sup>2</sup>s); HPSV=150W high pressure sodium vapor lamp(320 μmol/m<sup>2</sup>s). nitrogen in all nutrient solution was substituted by Cl<sup>-</sup> 6me/L.

<sup>y</sup> Duncan's multiple range test, *p*=0.05

가

가

14

Nam Kung (1994)

가

14



가 , 14 가 .  
가 가 ,

가 .  
(T/R ratio)

20 ,  $320 \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$  가  
, 14  $110 \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$  가 . 14

14 가

5 25 14 .  
T/R ratio 가

가 , 14

( 6).

20  $110 \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$  PPF T/R ratio가 가  
가 ,  $320 \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$  PPF 가

가

가 가

가

(DWR)

가 . 가 가

가 ,

가

7 conditioning 2

, a, b

가 . 14

20 14

가 .

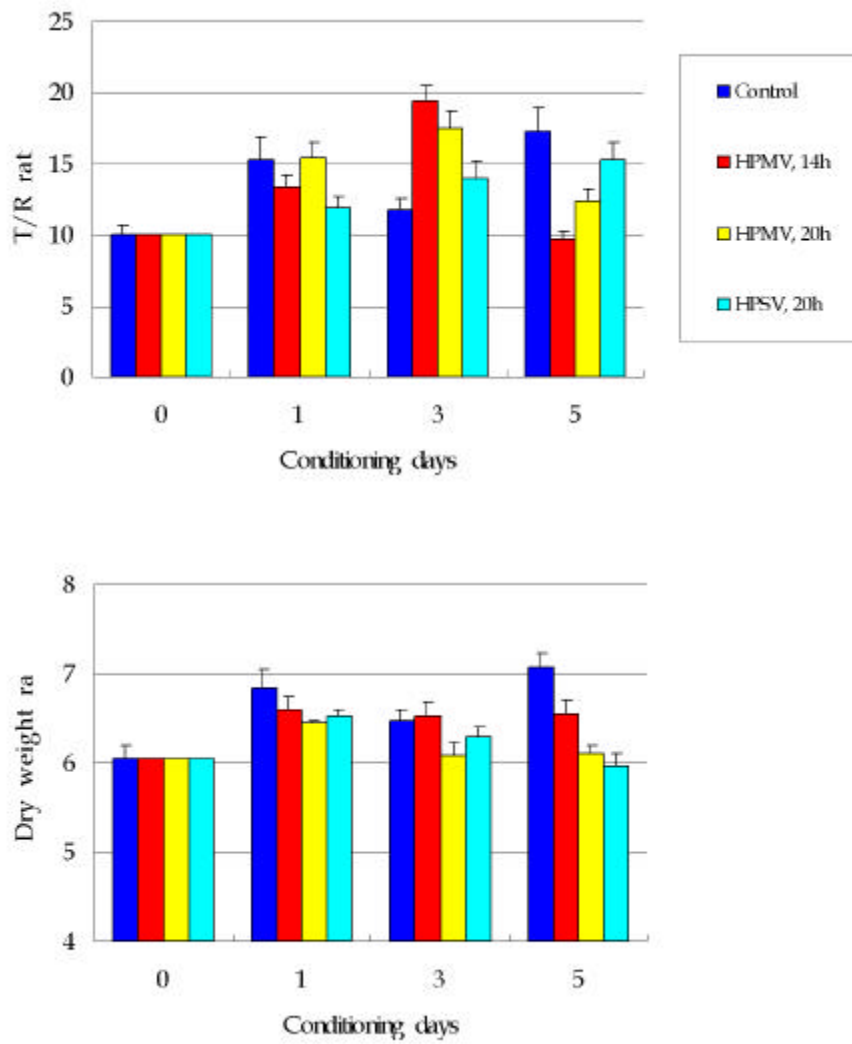


Fig. 6. The effects of supplementary lighting with  $110 \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$  and  $320 \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$  PPF and several photoperiods during conditioning period on growth changes of hydroponically grown *O. stolonifera* DC. Vertical bars indicate SE. HPMV and HPSV see Fig. 5.

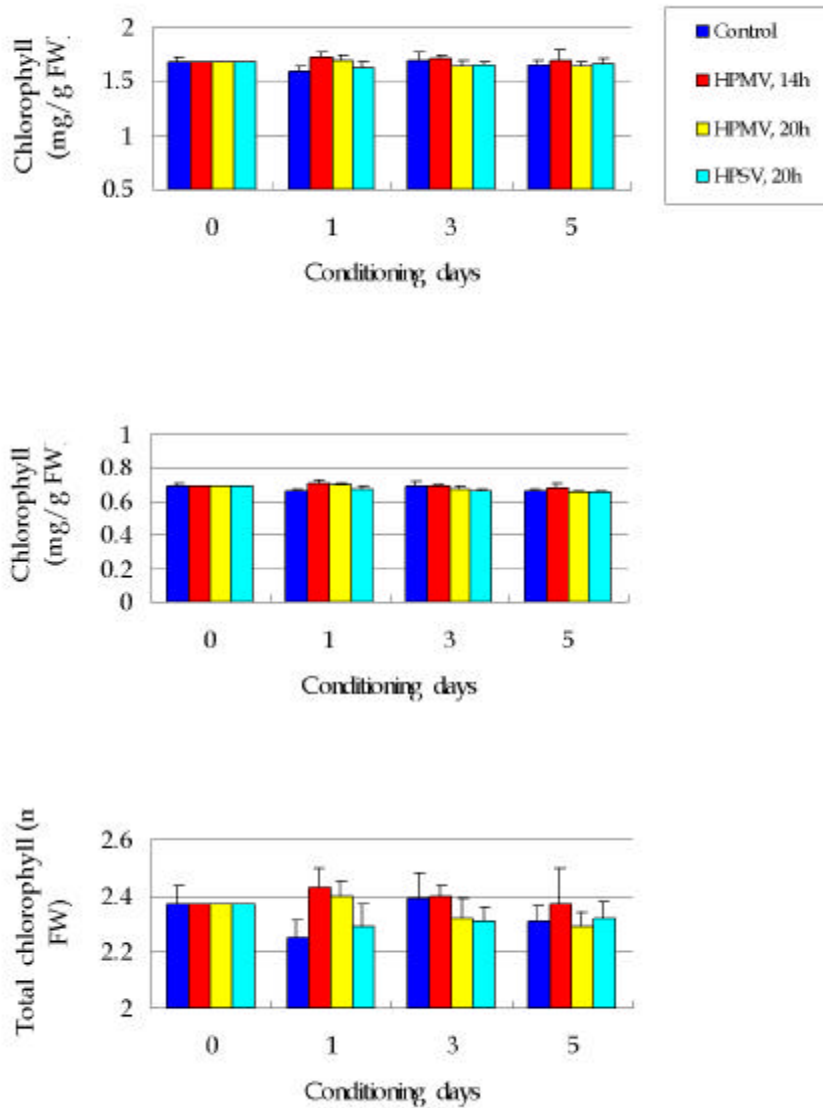


Fig. 7. The chlorophyll content changes of hydroponically grown *O. stolonifera* DC. during conditioning period when nutrient solution was substituted by conditioning solution under several photoperiods. Vertical bars indicate SE. HPMV and HPSV see Fig. 5.

가

11

가

14

3,514ppm

2,645ppm

24.7% 가

가

14

20

가

14

471ppm

3

1,072ppm

228%

가

3

5

가

4

가

14

6.7% 가

10% 가 20

3% 가

가

가

Cl

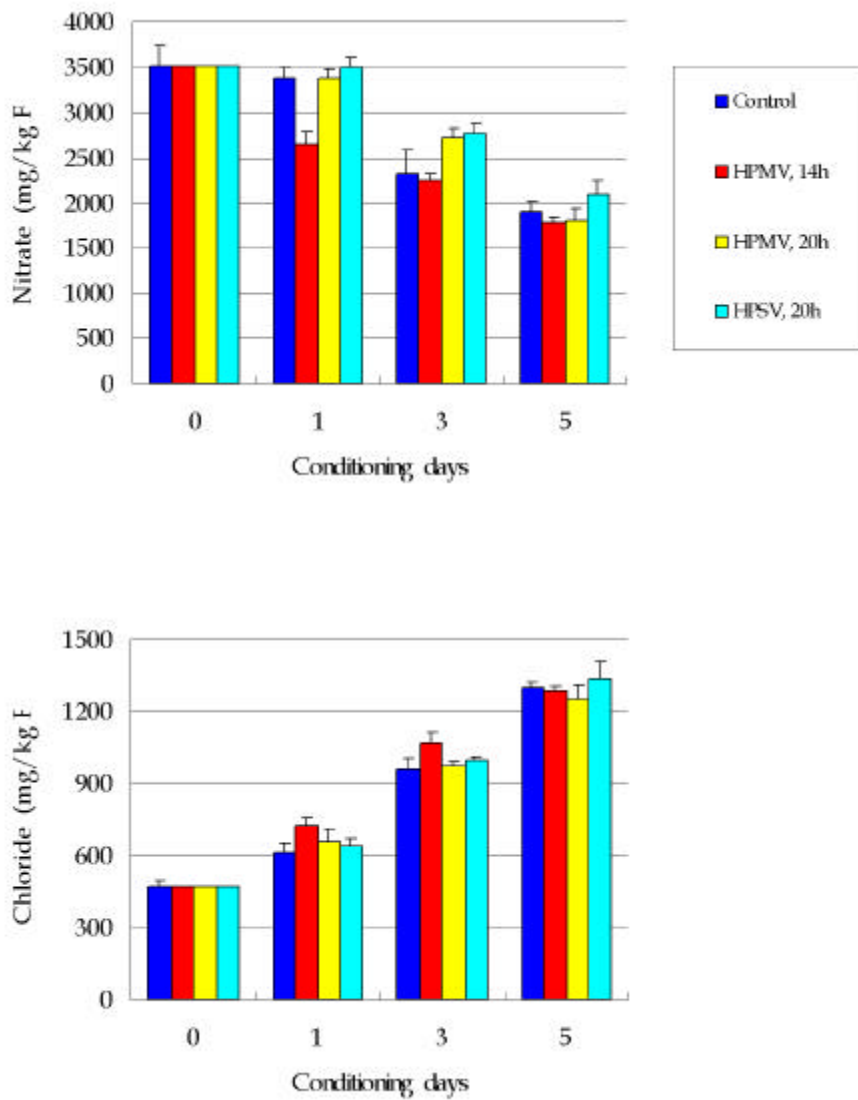


Fig. 8. The changes of nitrate and chloride contents of hydroponically grown *O. stolonifera* DC. during conditioning period when nitrogen was substituted by chloride in nutrient solution. Vertical bars indicate SE. HPMV and HPSV see Fig. 5.

Table 4. The effects of supplementary lighting when nutrient solution was substituted by conditioning solution for 5 days before harvest on nitrate and chloride contents of hydroponically grown *Oenanthe stolonifera* DC.

Photoperiod <sup>z</sup>	Nitrate (mg/kg FW)	Chloride (mg/kg FW)	Percentage over control	
			nitrate	chloride
Control	1903.7 <sup>ay</sup>	1298.9 <sup>a</sup>	100.0 %	100.0 %
14h HPMV	1776.0 <sup>a</sup>	1281.9 <sup>a</sup>	93.3	98.7
20h HPMV	1803.5 <sup>a</sup>	1250.5 <sup>a</sup>	94.7	96.3
20h HPSV	2097.2 <sup>a</sup>	1338.5 <sup>a</sup>	110.2	103.0

<sup>z</sup> Control=natural daylength; HPMV=200W high pressure mercury vapor lamp(110  $\mu\text{mol}/\text{m}^2/\text{s}$ ); HPSV=150W high pressure sodium vapor lamp(320  $\mu\text{mol}/\text{m}^2/\text{s}$ ). Chloride 6me/L in all nutrient solution was substituted for nitrogen.

<sup>y</sup> Duncan's multiple range test,  $p=0.05$

Gaudreau (1995)	50	100 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$	
	6	50 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$	28.9%
	5		
			(nitrate)

reductase;NR)가 , (Rouby , 1995).

가

가

Bergareche (1994) ,

phytochrome 가 ,

62% 가 가 ,

9

r=- 0.948

가 , 1

- 0.872

- 0.883

Reinink (1987)

가

가

가

Reinink

(1987)

가

가

14

가

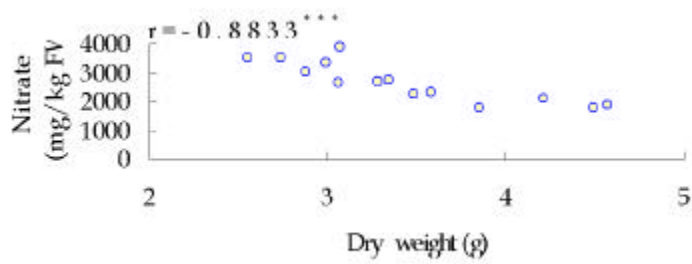
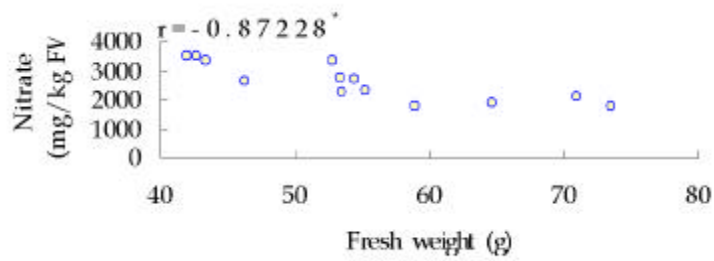
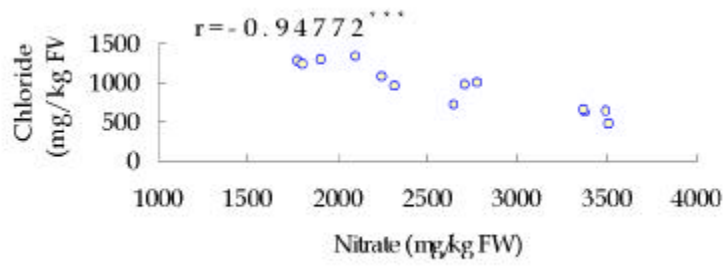


Fig. 9. The correlation between nitrate and chloride contents, fresh or dry weight during conditioning period when treated with supplementary lighting in hydroponically grown *O. stolonifera* DC.



5  
 가 , 가  
 K+ Mg2+ 14  
 Ca2+ 가

Table 5. The effects of supplementary lighting when conditioning solution was substituted for nutrient solution for 5 days before harvest on mineral content in hydroponically grown *Oenanthe stolonifera* DC.

Photoperiodz	Mineral content(mg/g DW)				
	Total nitrogen	K+	Ca2+	Mg2+	PO4 P
Control	30.00ay	83.50b	8.748a	3.174b	6.773b
14h HPMV	28.66a	85.29b	8.950a	3.444ab	7.843a
20h HPMV	29.37a	95.79a	9.823a	3.573a	8.155a
20h HPSV	30.34a	97.83a	9.458a	3.413ab	8.047a

z Control=natural daylength; HPMV=200W high pressure mercury vapor lamp(110  $\mu$  mol/m<sup>2</sup>/s); HPSV=150W high pressure sodium vapor lamp(320  $\mu$  mol/m<sup>2</sup>/s). Chloride 6me/L in all nutrient solution was substituted for nitrogen.

y Duncan's multiple range test,  $p=0.05$

10

, K+ PO4 P

20

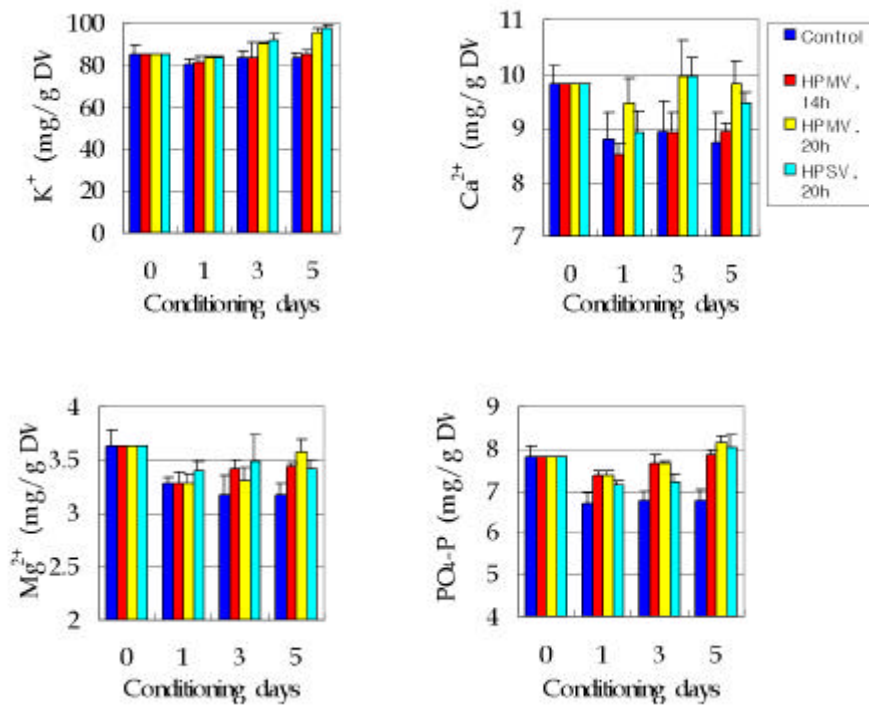


Fig. 13. The mineral content changes during conditioning period when treated with supplementary lighting in hydroponically grown *O. stolonifera* DC. Vertical bars indicate SE.

Ca<sup>2+</sup>

Mg<sup>2+</sup> 가 , 20

가 K+ PO<sub>4</sub>-P

가

Mg<sup>2+</sup> 가

3. Conditioning NH<sub>4</sub><sup>+</sup>가

22 6 Cl  
 8me/L 6  
 0, 0.5, 1.0 2.0me/L EC pH  
 11 .  
 EC 가 가 ,  
 가 가 가 가 pH  
 가 가 2.0me/L 3.16  
 가 .  
 6 , ,  
 가 , Cl  
 가 Cl  
 0.5 1.0me/L  
 2me/L  
 , 11B pH  
 Ruth Kafkafi(1980, 1983) H+  
 NH<sub>4</sub> 가  
 가

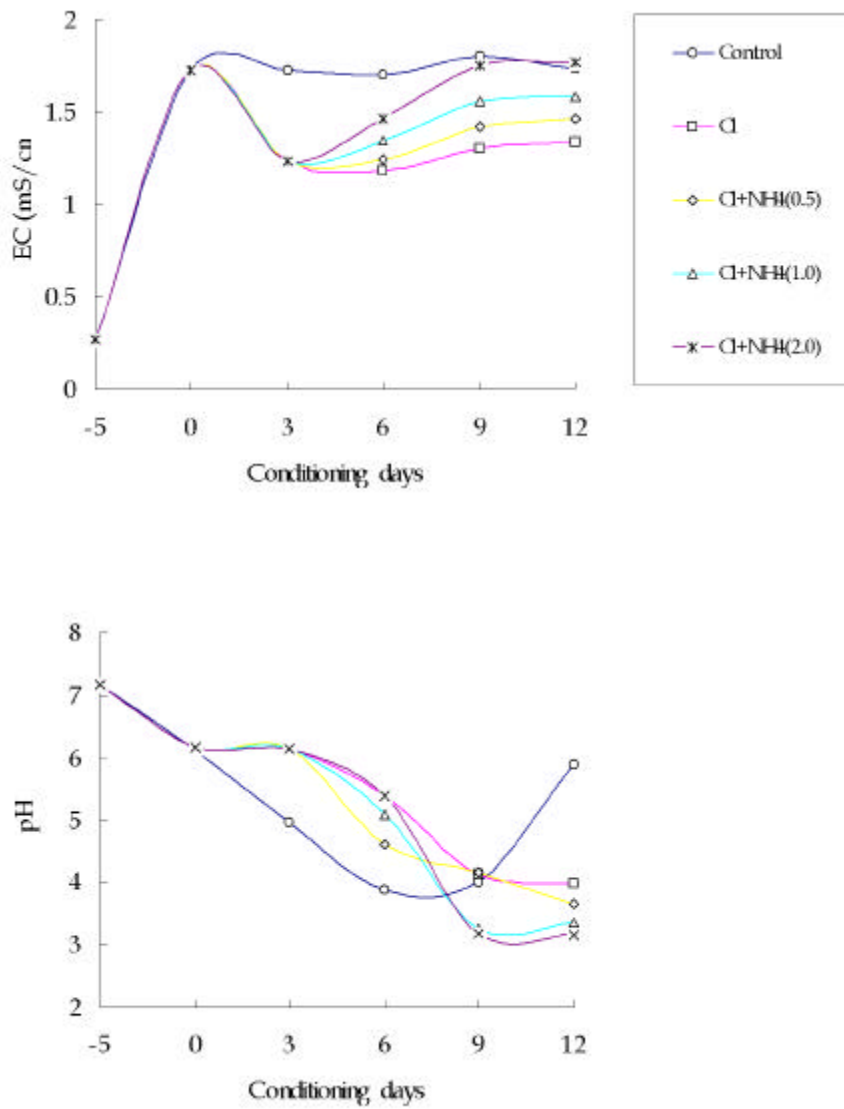


Fig. 11. The effects of substitution of chloride and/or ammonium for nitrogen in conditioning solution on changes of EC and pH in nutrient solution used for *O. stolonifera* DC. culture.

Table 6. The effects of substitution of chloride for nitrogen in nutrient solution for 6 days and subsequent ammonium chloride supply for 6 days before harvest on the growth of hydroponically grown *O. stolonifera* DC.

Conditioningz	NH <sub>4</sub> Cl conc. (me/L)	Plant height (cm)	Internode		Root length (cm)	Fresh weight(g)		Dry weight(g)		T/R ratio	DWRy
			no.	length (cm)		leaf	root	leaf	root		
Control	0	49.7ax	4.8a	37.1a	31.0b	14.87a	2.04bc	1.08ab	0.12b	7.4b	7.3c
Cl	0	51.6a	5.2a	41.2a	42.1a	13.53ab	3.95a	1.13a	0.19a	3.6d	8.3ab
Cl	0.5	49.4a	5.2a	37.7a	32.9b	12.77b	2.27b	1.06ab	0.13b	5.7c	8.3ab
Cl	1.0	48.3a	4.8a	40.3a	33.8b	11.93bc	1.55cd	0.95bc	0.10b	7.8b	7.9b
Cl	2.0	50.9a	4.8a	35.9a	22.2c	10.41c	1.05d	0.89c	0.09c	10.0a	8.6a

z Control=An's solution; Cl=substitution of Cl<sup>-</sup> 8me/L for nitrogen.

y Ratio of fresh weight over dry weight.

x Duncan's multiple range test,  $p=0.05$

Cl  
가 가  
가 2me/L  
T/R ratio Cl 가 , NH<sub>4</sub> 가  
가 NH<sub>4</sub> 1me/L

가

Table 7. The effects of nutrient substitution of chloride and/or ammonium ion for nitrogen applied for 12 days before harvest on chlorophyll content, leaf color, nitrate and chloride contents of hydroponically grown *O. stolonifera* DC.

Conditioningz	NH <sub>4</sub> Cl conc. (me/L)	Chlorophyll cont. (mg/g FW)			Leaf color (value)			Nitrate (mg/kg FW)	Chloride (mg/kg FW)
		a	b	total	L	a	b		
Control	0	1.74ax	0.67a	2.41a	44.2b	-19.8bc	+24.3b	3401a	1027c
Cl	0	1.40b	0.56b	1.96b	48.6a	-20.7d	+29.2a	640d	2278a
Cl	0.5	1.76a	0.67a	2.43a	46.4b	-19.2c	+25.4b	922c	2022b
Cl	1.0	1.68a	0.67a	2.34a	44.9b	-17.7ab	+22.5b	1029c	2225a
Cl	2.0	1.70a	0.70a	2.40a	42.7c	-16.6a	+20.7c	1287b	2324a

z Control=An's solution; Cl=substitution of Cl- 8me/L for nitrogen.

y L, lightness; a, redness; b, yellowness.

x Duncan's multiple range test,  $p=0.05$

( 7), Cl

가

가

가

0.5me/L

가

가 , L Cl  
 , 0.5~1.0me/L  
 . a b  
 16 Cl ,  
 가 가 2.0me/L  
 Cl 640ppm 가  
 81.2%가 , Cl+ NH4 0.5me/L 922ppm  
 ( 7).  
 가  
 가 ,  
 12 ,  
 , 가 가  
 ,  
 가  
 가 2,500ppm  
 ,  
 가

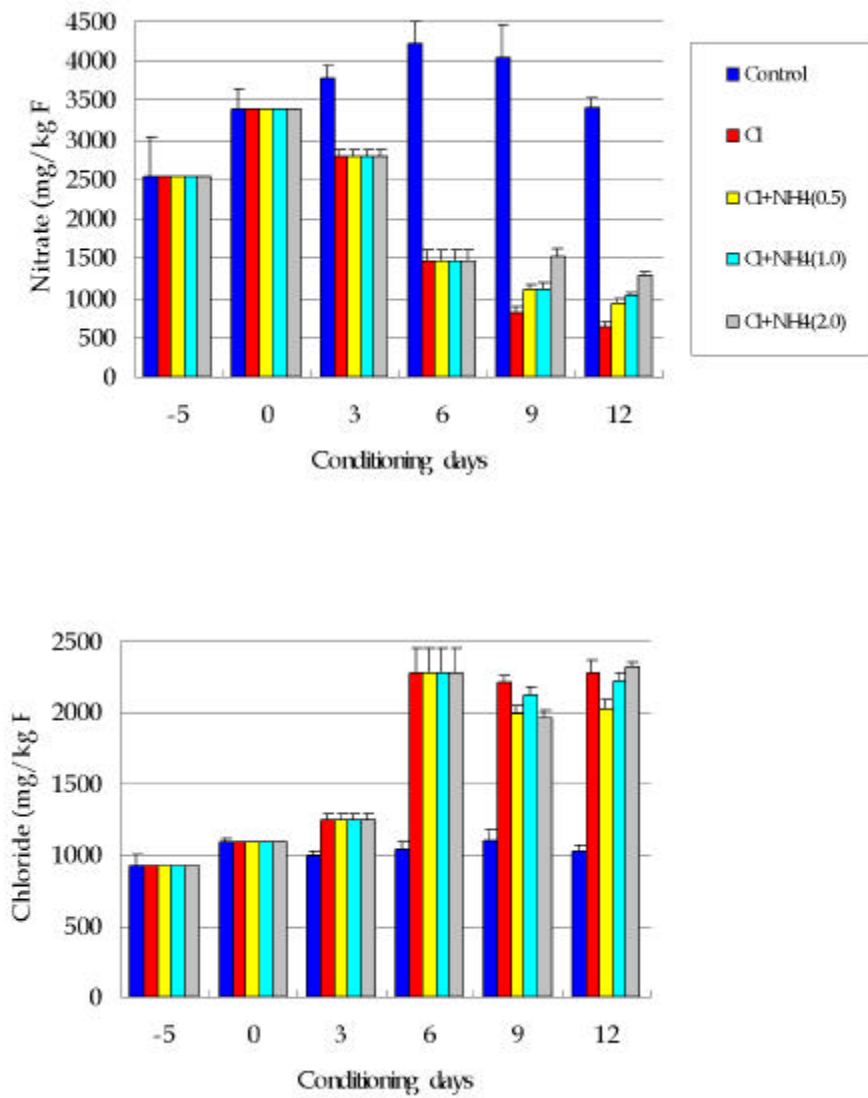


Fig. 12. The changes of nitrate and chloride contents during conditioning period in hydroponically grown *O. stolonifera* DC. Vertical bars indicate SE.



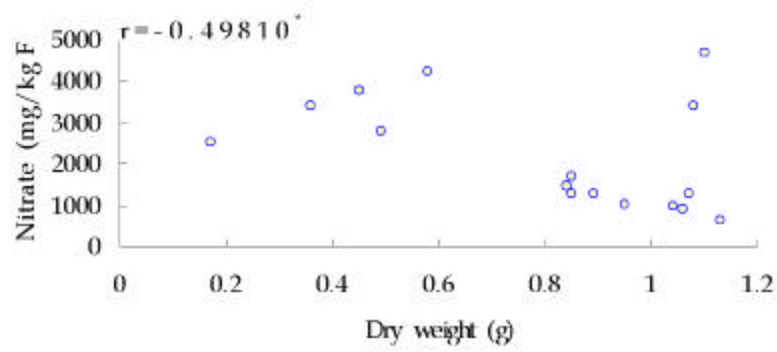
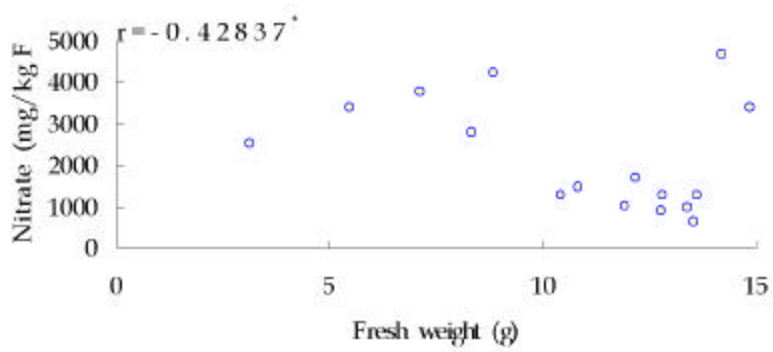
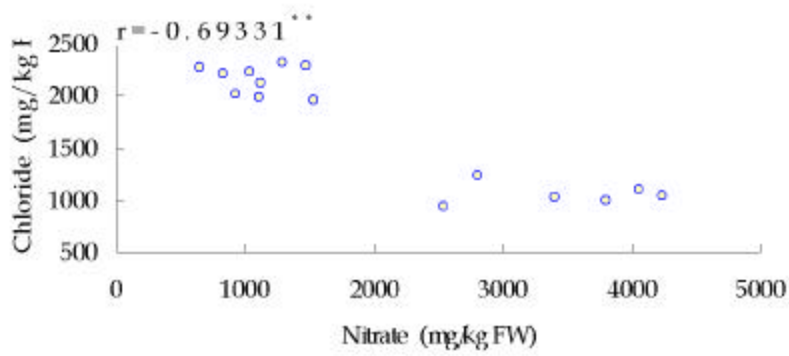


Fig. 13. The correlation between nitrate and chloride, fresh or dry weight during conditioning period of hydroponically grown *O. stolonifera* DC.

r=- 0.693

가

Conditioning

8

Table 8. The conditioning effects of nutrient substitution for 12 days before harvest on mineral content of hydroponically grown *Oenanthe stolonifera* DC.

Conditioningz	NH <sub>4</sub> Cl conc. (me/L)	Mineral content (mg/g DW)				
		Total nitrogen	K+	Ca <sup>2+</sup>	Mg <sup>2+</sup>	PO <sub>4</sub> P
Control	0	33.13ay	87.70a	6.886bc	3.895a	7.213a
Cl	0	19.84e	75.19b	8.455a	2.879c	6.267b
Cl	0.5	22.64d	74.08b	6.809c	2.756c	6.668b
Cl	1.0	28.51c	73.08b	7.385b	2.877c	6.635b
Cl	2.0	31.46b	71.42b	7.281bc	3.114b	6.397b

z Control=An's solution; Cl=substitution of Cl- 8me/L for nitrogen in nutrient solution.

y Duncan's multiple range test,  $p=0.05$

	가	,		가
		가	, Cl	
가		. K+ Mg2+ PO4 P		
	가	, Mg2+		
가		. Mg2+ NH4 2.0me/L		가

4

1

1.

가

2.

Fig. 1. The landscape of water dropwort to clarify the effect of optimum planting density and number of plants per hole.

5cm × 5cm, 7cm × 7cm , 10cm × 10cm

5cm × 15cm

4가

4 6 , 10 12 . 30

2

35

3.

, 1 4 7

가

가

가

5

가

가

. 7 × 7cm 4

15 × 5cm

4

5

7 × 7cm 4

15 ×

5cm 4

가

가

가

Table 1. The effect of planting density and number of plants per hole on the growth characteristics of water sropwort(April July).

Treatment		調 査 內 容					
Planting density	No. of plant	Plant height (cm)	No. of leaves	No. of stolons	Stolon length (cm)	Fresh weight (g/5plant)	Amount /Pyung (kg)
7 × 7cm	1	38.0	4.7	3.5	28.8	49.3	5.1
	2	40.5	4.5	2.7	24.1	41.7	8.6
	3	43.9	4.2	2.3	21.2	36.3	11.2
	4	43.8	4.3	2.3	18.6	34.0	14.0
	mean	41.55	4.43	2.70	23.18	40.33	9.73
10 × 10cm	1	33.7	4.3	4.3	29.6	56.7	4.2
	2	36.8	4.4	3.6	26.1	42.3	6.2
	3	39.6	4.6	3.2	27.8	37.3	8.2
	4	37.8	4.1	3.1	23.6	43.0	12.6
	mean	37.0	4.4	3.6	26.8	44.8	7.8
15x5cm	1	37.2	4.7	4.1	28.4	54.0	4.8
	2	37.3	4.5	3.7	26.8	43.7	7.7
	3	39.8	4.1	3.2	27.6	41.0	10.9
	4	38.1	4.6	3.5	26.4	48.0	16.9
	mean	38.1	4.5	3.6	27.3	46.7	10.1
15 × 15cm	1	29.4	4.7	4.5	24.3	51.0	1.5
	2	31.6	4.3	4.5	25.1	51.0	3.0
	3	34.9	4.9	4.3	28.5	68.3	6.0
	4	32.5	4.8	5.1	26.7	68.3	7.4
	mean	32.1	4.7	4.6	26.2	59.7	4.5
Planting density(A)z		***	ns	**	ns	**	***
No. of plants(B)		**	ns	**	ns	ns	***
A × B		ns	ns	ns	ns	ns	ns

z Mean seperation within a column by duncan's multiple range at 5% level

\*, \*\*, ns Significant at P 0.05, 0.01 and nonsignificant respectively

2 10 12

4 7

가 , 가  
. 가  
가 . ,  
가 가 .  
가 가 ,  
가 가 , 10 × 10cm  
4 7 × 7cm 4  
가 가  
가 가 ,  
가 7 × 7cm  
. 10 × 10cm ,  
15 × 5cm 가 1 2  
가 . 15 × 15cm 2  
3  
T/R 7 × 7cm 가  
가  
, 15 × 15cm 15 × 5cm T/R  
가 가 T/R  
가

Table 2. The effect of planting density and number of plants per hole on the growth characteristics of water dropwort(Oct. Dec.).

Treatment		Contents						
Planting density (cm)	No. of plants	Leaf length (cm)	No. of leaves	NO. of stolons	Stolon length (cm)	Fresh weight (mg)	Amount of edible part (kg/Pyung)	T/R ratio
7 × 7	1	41.0	5.2	5.0	30.9	14.7	7.6	5.0
	2	45.5	4.8	3.2	23.9	22.4	11.5	6.3
	3	45.3	4.4	2.6	29.0	21.7	11.2	6.5
	4	48.3	4.7	2.1	16.5	24.2	12.4	6.6
	mean	45.0	4.8	3.2	25.1	20.8	10.7	6.1
10 × 10	1	41.2	4.7	6.0	32.7	19.8	7.3	5.2
	2	43.9	4.6	3.8	25.1	21.1	7.8	6.3
	3	43.3	4.6	3.8	23.6	26.3	9.7	6.2
	4	47.9	5.2	2.9	26.1	39.2	14.4	6.9
	mean	44.1	4.8	4.1	26.9	26.6	9.8	6.2
15 × 5	1	38.1	5.2	5.2	28.8	14.6	6.4	4.9
	2	38.7	4.4	3.2	19.7	15.5	6.8	4.5
	3	42.6	5.1	3.0	23.7	24.2	10.6	5.2
	4	45.5	4.8	2.6	24.4	28.6	12.6	6.3
	mean	41.2	4.9	3.5	24.2	20.7	9.1	5.2
15 × 15	1	41.3	4.8	5.6	30.6	20.3	3.0	5.5
	2	39.9	4.8	4.3	27.1	24.1	3.5	5.3
	3	40.3	4.4	4.0	24.6	25.2	3.7	5.7
	4	42.6	5.0	3.0	19.4	46.8	6.9	5.9
	mean	41.0	4.8	4.2	25.4	29.1	4.3	5.6
Planting density(A)z		***	ns	**	ns	***		**
No. of Plants(B)		***	ns	***	***	***		***
A × B		ns	ns	ns	ns	**		ns

z \*\*, \*\*\* is significant at  $p=0.01$ ,  $0.001$  and ns is not significant.





7 27 timer . 8 1 . . 3  
 pH, EC, DO pH 5.5 6.5, EC 1.0 1.2  
 8 20 , 8 30

3.

가) 가

가 가

, , .  
 .  
 ,  
 .

( 1).

Table 1. Effect of pumping times on the growth of water dropwort.

Pumping method	Plant height (cm)	No. of leaves (ea.)	Petiole length (cm)	No. of stolons	Runner length (cm)	Root length (cm)	Fresh weight(g)		Dry weight(g)	
							leaf	root	leaf	root
1stx										
Continuez	32.9	5.8	10.0	4.9	29.7	24.7	15.83	4.05	0.90	0.33
Day onlyy	37.0	6.1	11.4	5.4	34.0	25.4	21.74	5.01	1.26	0.39
F	22.26***	2.88ns	15.01***	6.34*	11.95***	0.33ns	12.59***	8.08**	13.94***	12.09***
LSD005	1.709	0.312	0.726	0.342	2.482	2.299	3.302	0.666	0.190	0.033
LSD001	2.263	-	0.961	0.452	3.286	-	4.371	0.881	0.252	0.044
2ndw										
Continuez	53.0	6.6	16.2	6.4	53.8	32.2	51.42	6.63	-	-
Day onlyy	52.2	6.5	13.5	6.4	54.3	29.3	44.74	5.60	-	-
F	0.39ns	0.39ns	8.64**	0.00ns	0.10ns	3.98*	4.05*	9.32**	-	-
LSD005	2.509	0.477	1.794	0.371	3.366	2.857	6.607	0.675	-	-
LSD001	-	-	2.380	0.492	-	3.789	8.763	0.895	-	-

z Pumping continuously

y Pumping during day and no pumping during night

x 25days after planting

w 35days after planting

\*, \*\*, \*\*\*, ns Significant at p 0.05, 0.01, 0.001 and nonsignificant, respectively

가

( )

, 가 , 15 /15 , 15 /30 , 15

/60

( 2).

Table 2. Effects of pumping periods on the growth of water dropwort

Pumping method	Plant height (cm)	No. of leaves (ea.)	Petiole length (cm)	No. of stolons	Stolon length (cm)	Root length (cm)	Fresh weight(g)		Dry weight(g)	
							leaf	root	leaf	root
1stw										
Continue	34.8	5.8	10.7	5.1	31.6	23.4	18.10	4.39	1.03	0.35
15/15Z	33.6	6.0	10.3	5.1	29.9	23.1	17.00	4.27	1.00	0.35
15/30Y	34.8	6.0	11.0	5.2	31.5	25.3	19.10	4.45	1.03	0.36
15/60x	35.5	5.7	11.1	5.0	32.9	25.4	19.50	4.54	1.18	0.36
F	0.46ns	0.80ns	1.45ns	0.24ns	0.59ns	2.22ns	0.32ns	0.08ns	0.46ns	0.07ns
LSD005	2.950	0.490	1.190	0.730	5.410	4.590	7.260	1.440	-	0.074
2ndv										
Continue	53.0	6.5	15.8	6.3	53.7	28.9	48.53	6.23	-	-
15/15z	51.2	6.7	14.3	6.1	54.4	29.8	43.58	6.05	-	-
15/30Y	52.1	6.4	15.0	6.6	54.6	29.4	47.28	5.84	-	-
15/60x	54.2	6.6	14.5	6.6	53.6	34.9	52.94	6.32	-	-
F	1.04ns	0.18ns	0.49ns	1.80ns	0.08ns	3.98*ns	1.32ns	0.34ns	-	-
LSD005	3.530	0.682	-	0.514	4.820	3.902	9.469	1.017	-	-
LSD001	-	-	-	-	-	5.180	-	-	-	-

z Pumping during 15minuites and no pumping during 15minuites

y Pumping during 15minuites and no pumping during 30minuites

x Pumping during 15minuites and no pumping during 60minuites

w 25days after planting

v 35days after planting

\*, \*\*, \*\*\*, ns : Significant at p 0.05, 0.01, 0.001 and nonsignificant, respectively

( )

1) 15

15

15

가 ( 3).

Table 3. Effect of intermittent pumping method and general pumping method on the growth of water dropwort ( ).

Pumping method	Plant height (cm)	No. of leaves	Petiole length (cm)	No. of stolons	Stolon length (cm)	Root length (cm)	Fresh weight(g)		Dry weight(g)	
							leaf	root	leaf	root
15/15z	31.5	5.6	9.3	4.7	28.2	22.6	13.70	3.61	0.84	0.36
Inter.15y	36.7	6.4	10.2	5.7	33.6	30.8	21.60	5.15	1.21	0.40
F	15.03**	8.47**	2.27ns	5.56*	6.56*	9.85**	3.55ns	3.42ns	2.72ns	5.96*
LSD005	2.851	0.578	1.366	0.890	4.430	5.542	8.810	1.750	0.470	0.079
LSD001	3.906	0.791	-	1.220	6.069	7.593	-	-	-	0.109

z Pumping during 15 minutes and no pumping during 15 minutes

y Intermittent system with drain completely during 15 minutes

\*, \*\*, \*\*\*, ns Significant at p 0.05, 0.01, 0.001 and nonsignificant, respectively

2) 30

15

30

30

( 4).

Table 4. Effects of intermittent pumping system and general pumping method on the growth of water dropwort ( ).

Pumping method	Plant height (cm)	No. of leaves	Petiole length (cm)	No. of stolons	Stolon length (cm)	Root length (cm)	Fresh weight(g)		Dry weight(g)	
							leaf	root	leaf	root
15/30 z	32.6	6.1	9.8	5.0	30.0	23.4	15.34	3.81	0.87	-
Inter.30 y	31.3	5.7	9.5	4.7	28.5	24.5	13.03	3.91	0.78	-
F	0.37ns	4.80*	0.32ns	1.98ns	0.26ns	0.33ns	1.18ns	1.98ns	0.58ns	-
LSD005	4.406	0.384	1.122	2.100	6.392	4.179	4.460	1.137	0.248	-
LSD001	-	0.526	-	3.897	-	-	-	-	-	-

z Pumping during 15minuites and no pumping during 30minuites

y Intermittent system that drained completely during 30minuites

\*, \*\*, \*\*\*, ns Significant at p 0.05, 0.01, 0.001 and nonsignificant, respectively

3

가

1.

가

가

가 .

가

NFT, DFT

가  
deep flow

2.

8 25 10 21

3

, 3 ,  $6.2 \pm 0.3\text{cm}$  .

( , 1991)

NO<sub>3</sub>-N 16, NH<sub>4</sub>-N

3, P 3, K 4, Ca 2, Mg 1, SO4 S 1mg/L Fe 3ppm, B

0.5, Mn 0.5, Zn 0.5, Cu 0.02 Mo 0.01ppm .

pH EC 6.5 1.60

, pH meter(HI8284,  
Hanna instruments) conductivity meter(HI8733, )

CR10X data logger(Campbell Sci.)

30

18:00 23:30 60W

1m 16

60 × 50cm styrofoam 3

(Deep- flow) 가 3cm 7cm

, (Mist- spray)

10cm 40

80cm 3 .

가 . 06:00

11:00 30/15 ( / ) , 11:00 18:00 45/30 ( /

), 15/45 ( / ) .

15/15

5 , K, Mg Ca

atomic absorption spectrophotometry(AA6401,

Shimadzu Corp.) NH4 N

indophenol- blue ( , 1990) , NO3- N

Cataldo (Cataldo , 1975) , PO4 P vanadate (

, 1988) Shimadzu UV- 1601 UV- VIS



spectrophotometer 665, 410 470nm

6, 16, 26

9

, K, Ca, Mg

PO4 P

HClO4 : H2SO4

: H2O = 90 : 5 : 55

LI- 6400 portable photosynthesis

system(LI- COR)

10

( ) , (25 )

25 , 2000 μ

mol•m<sup>-2</sup>s<sup>-1</sup>

2

0.5g

100% ethanol 24

3

649

665nm

( , 1981).

3.

1

EC

pH

80cm mist- spray

EC

1.60mS/cm

가

가 가

80cm mist- spray

EC 가

가

40cm mist- spray

가

( 2).

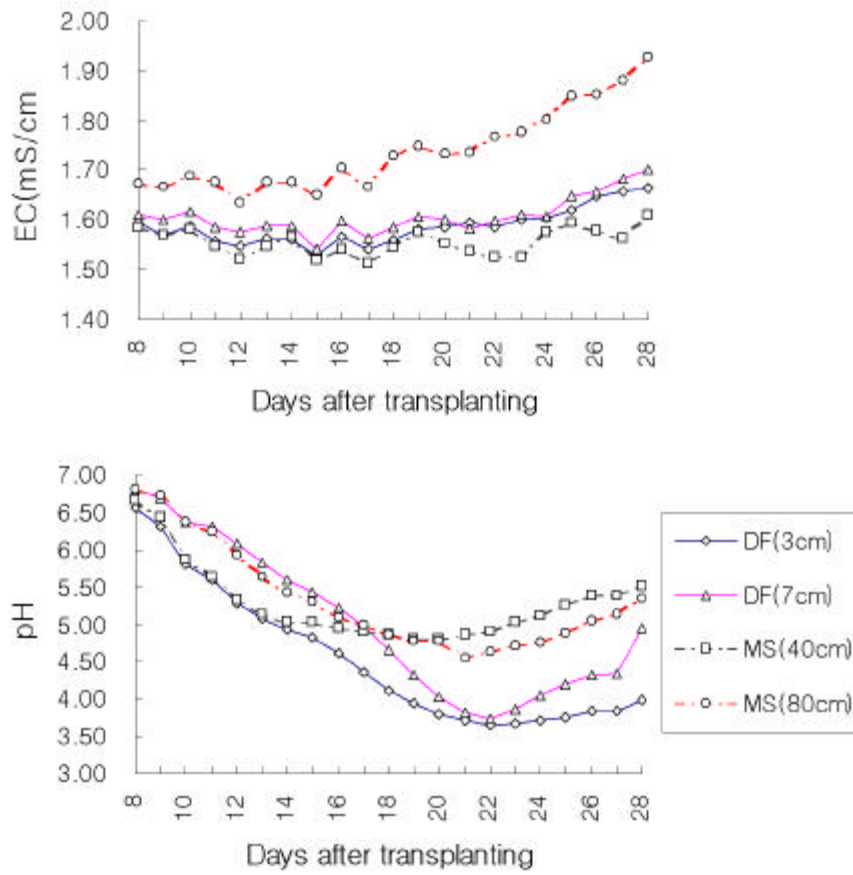


Fig. 1. The changes of EC and pH of nutrient solution during culture period of *Oenanthe stolonifera* DC. among different hydroponic systems. In legend, DF(3cm), DF(7cm), MS(40cm) and MS(80cm) indicate deep-flow with 3cm, 7cm nutrient solution level and MS with 40 and 80cm depth of bed, respectively.

pH  
가

(Maynard , 1976; Ikeda Osawa, 1980; Ikeda Osawa, 1981; , 1991).

deep- flow

mist- spray

deep- flow

가 pH 가 .  
2

7cm deep- flow

40cm mist- spray

3cm deep- flow 80cm

mist- spray 가 . 3

가

7cm deep- flow 40cm

mist- spray

가

80cm mist- spray 2

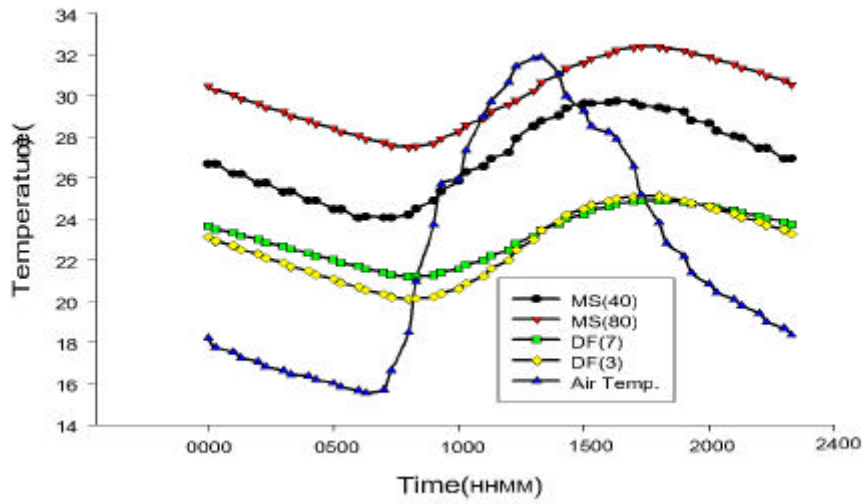


Fig. 2. The changes of temperature inside the bed, air and relative humidity of air. Legends see fig. 1.

가

7cm deep-flow

1 2

7cm

deep-flow

3cm deep-flow

3cm deep-flow

가



deep- flow

3 , 가  
40cm mist- spray 3cm deep- flow

40cm mist- spray

, 7cm deep- flow 가 80cm

deep- flow

가

7cm deep- flow

80cm mist- spray

가

가

Table 1. The effects of different hydroponic systems on the growth of *Oenanthe stolonijera* DC. at harvest.

Culture system		Plant height (cm)	No. of leaves	No. of internodes	Length of main internode	No. of stolons	Length of main stolon
Depth(cm)							
Deep flow	Low(3)	25.40 cz	7.44 a	7.22 ab	4.08 a	4.78 bc	24.84 b
	Deep(7)	33.02 b	7.56 a	7.67 ab	4.92 a	5.67 a	32.51 a
Mist spray	Low(40)	39.84 a	6.78 a	7.89 a	4.58 a	5.56 ab	25.40 b
	Deep(80)	24.40 c	7.11 a	6.89 b	3.71 a	4.67 c	33.02 a

z Duncan's multiple range test,  $p=0.05$

2

40cm mist- spray

가

7cm deep- flow, 3cm deep- flow 80cm

mist- spray

7cm deep-flow                      40cm

mist-spray

40cm mist-spray                      가                      3cm deep-flow

가

가

Table 2. The effects of different hydroponic systems on the fresh weight, dry weight, chlorophyll contents and root growth, respectively, in *Oenanthe stolonifera* DC.

Culture system	Fresh weight(g)	Dry weight(g)		Chlorophyll content(mg/g · F.W.)			Root length (cm)	Volume of root (ml)		
		Leaf	Root	a	b	Total				
		Leaf	Root	a	b	Total				
Deep flow	Low(3)	19.50 cz	2.02 c	3.55 b	0.29 c	1.38 a	0.54 a	1.92 a	20.78 b	5.22 c
	Deep(7)	27.93 b	3.33 b	4.21 b	0.41 bc	1.48 a	0.59 a	2.07 a	26.09 a	8.44 b
Mist spray	Low(40)	36.02 a	6.70 a	5.07 a	0.64 a	1.52 a	0.62 a	2.14 a	30.11 a	14.22 a
	Deep(80)	18.65 c	3.32 b	3.83 b	0.46 b	1.48 a	0.61 a	2.09 a	18.08 b	7.78 b

z Duncan's multiple range test,  $p=0.05$

3

80cm mist-spray

가                      40cm

mist-spray                      7cm deep-flow

가  
 ,  
 가  
 ,  
 7cm deep-flow                      80cm mist-spray  
 ,  
 가  
 가

Table 3. The influence of different hydroponic systems on photosynthesis rate and stomatal conductivity of *Oenanthe stolonifera* DC. at 10 and 25 days after transplanting.

Culture system		Photosynthesis rate ( $\mu\text{molCO}_2\text{m}^{-2}\text{s}^{-1}$ )		Stomatal conductivity ( $\text{molH}_2\text{O}\text{m}^{-2}\text{s}^{-1}$ )	
		Days after transplanting			
		10	25	10	25
Deep flow	Low(3)	19.63 bz	7.42 a	0.223 ab	0.077 a
	Deep(7)	16.28 c	6.32 a	0.766 bc	0.092 a
Mist spray	Low(40)	16.63 c	8.31 a	0.136 c	0.135 a
	Deep(80)	22.18 a	6.81 a	0.243 a	0.092 a

z Duncan's multiple range test, p=0.05

K<sup>+</sup>, NO<sub>3</sub><sup>-</sup> N      PO<sub>4</sub><sup>-</sup> P  
 4  
 가  
 ,  
 80cm mist-spray  
 가  
 deep-flow



mist-spray

가

K+

3cm

deep-flow

가

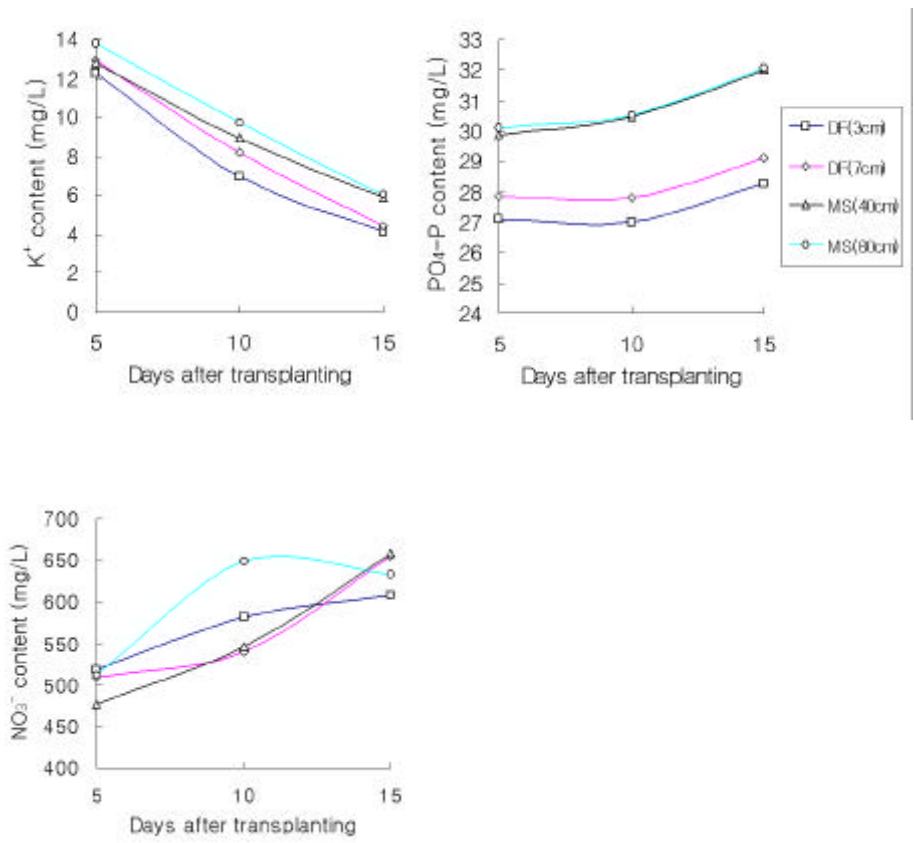


Fig. 4. The ion compositions in nutrient solution among different hydroponic systems at 5, 10 and 15 days after transplanting. In legend, DF(3cm), DF(7cm), MS(40cm) and MS(80cm) see fig. 1.

pH

7cm , 3cm

pH EC

가 가 가

가 pH EC



- The role of light and nitrate in the induction of nitrate reductase in radish cotyledons and maize seedlings. *Plant Physiology* 40 : 691-698.
- Bennett, W. F., J. Pesek and J. J. Hanway. 1964. Effect of nitrate and ammonium on growth of corn in nutrient sand culture. *Agron. J.* 56 : 342-345.
- Bennett, M. A., and L. Waters, Jr. 1987a. Germination and emergence of high sugar sweet corn is improved by presowing hydration of seed. *HortScience* 22 : 236-238.
- Bennett, M. A., and L. Waters, Jr. 1987b. Seed hydration treatments for improved sweet corn germination and stand establishment. *J. Amer. Soc. Hort. Sci.* 112 : 45-49.
- Benoit, F. and C. R. Wiebe. 1992. Relation between photosynthesis and nitrate content of lettuce cultivars. *Scient. Hort.* 49 : 175-179.
- Benoit, F. and N. Ceustermans. 1995. Horticultural aspects of ecological soilless growing methods. *Acta Horti.* 396 : 11-24.
- Bergareche, C. and E. Simon. 1988. Nitrate reductase activity and nitrate content under two forms and three levels of nitrogen nutrition in *Lolium perenne* L. *Plant Physiol.* 64 : 358-363.
- Blair, G. J., M. H. Miller and W. A. Mitchell. 1970. Nitrate and ammonium as sources of nitrogen for corn and their influence on the uptake of other ions. *Agron. J.* 62 : 530-532.
- Blom-Zanstra, M. and J. E. M. Lampe. 1985. The role of nitrate in the osmoregulation of lettuce (*Lactuca sativa* L.) grown at different light intensities. *J. Exp. Bot.* 36 : 1043-1052.
- Boon, J. and J. W. Steenhuizen. 1986. Nitrate in lettuce on recirculating nutrient solution. *Acta Horticulturae* 178 : 67-72.
- Bradford, K. J. 1985. Seed priming improves germination and emergence of cantaloupe at low temperatures. *HortScience.* 21 : 1105-1112
- Bradford, K. J. 1986. Manipulation of seed water relations via osmotic priming to improve germination under stress conditions. *HortScience* 21 : 1105-1112.
- Bradford, K. J., C. A. Argerich, D. Peetambar, O. Somasco, A. Tarquis, and G. E. Welbaum. 1988. Seed enhancement and seed vigor. *Proc. Intl. Conf. Stand. Estab. Hort. Crops.* Lancaster, PA. pp. 1-35.
- Bradford, K. J., J. J. Steiner, and S. E. Trawatha. 1990. Seed priming influence on germination and emergence of pepper seed lots. *Crop.*

- Sci. 30 : 718 721.
- Bray, C. M., P. A. Davison, M. Ashraf, and R. M. Taylor. 1989. Biochemical changes during osmopriming of leek seeds. *Ann. Bot.* 63 : 185 193.
- Brocklehurst, P. A., J. Dearman, and R. L. K. Drew. 1984. Effects of osmotic priming on seed germination and seedling growth in leek. *Scient. Hortic.* 24 : 201 210.
- Brocklehurst, P. A., and J. Dearman. 1983a. Interactions between seed priming treatments and nine seed lots of carrot, celery and onion: . Laboratory germination. *Ann. Appl. Biol.* 102 : 577 584.
- Brocklehurst, P. A., and J. Dearman. 1983b. Interactions between seed priming treatments and nine seed lots of carrot, celery and onion: . Seedling emergence and plant growth. *Ann. Appl. Biol.* 102 : 585 593.
- Brocklehurst, P. A., and J. Dearman. 1984. A comparison of different chemicals for osmotic treatment of vegetable seed. *Ann. Appl. Biol.* 105 : 391 398.
- Bruning-Fann, C. S. and J. B. Kaneen. 1993. The effects of nitrate, nitrite and N-nitroso compounds on human health : A Review *Vet. Human Toxicol.* 35 : 521 538.
- Buczek, J. 1985. Regulation of nitrate and nitrite reductase activities in whole cucumber plants by endogenous level of nitrate supply. *Acta Physiologiae Plantarum* 7 : 21 30.
- Bujalski, W., A. W. Nienow, and D. Gray 1989. Establishing the large scale osmotic priming of onion seeds by using enriched air. *Ann. Appl. Biol.* 115 : 171 176.
- Candela, M I., E. G. Fisher and E. J. Hewitt. 1957. Molybdenum as a plant nutrient. . Some factors affecting the activity of nitrate reductase in cauliflower plants grown with different nitrogen sources and molybdenum levels in sand culture. *Plant Physiol.* 32 : 280 288.
- Cano, E. A., M. C. Bolarin, F. Perez-Alfocea, and M. Caro. 1991. Effects of NaCl priming on increased salt tolerance in tomato. *J. Hort. Sci.* 66 : 621 628
- Canovas, F. M., C. Avila, J. R. Botella, V. Valpuesta and I. N. de Castro. 1986. Effect of light-dark transition on glutamine synthetase activity in tomato leaves. *Physiol. Plant.* 66 : 648 652.

- Cantliffe, D. J. 1972. Nitrate accumulation in vegetable crops as affected by photoperiod and light duration. *J. Amer. Soc. Hort. Sci.* 97 : 414-418.
- Cantliffe, D. J., J. M. Fischer, and A. C. Nell. 1984. Mechanism of seed priming in circumventing thermodormancy in lettuce. *Plant Physiol.* 75 : 290-294.
- Cantliffe, D. J., K. D. Schuler, and A. C. Guedes. 1981. Overcoming seed dormancy in heat sensitive romaine lettuce by seed priming. *J. Amer. Soc. Hort. Sci.* 100 : 65-71.
- Chalifour, F-P. and L. M. Nelson. 1987. Effects of time of nitrate application on nitrate reductase activity, nitrate uptake and symbiotic dinitrogen fixation in faba bean and pea. *Can. J. Bot.* 66 : 1646-1652.
- Chen, T. M. and S. K. Ries. 1969. Effect of light and temperature on nitrate uptake and nitrate reductase activity in rye and oat seedlings. *Can. J. Bot.* 47 : 341-343.
- Chen, T. M., S. K. Ries, and J. M. Cantliffe. 1992. Effect of nitrate reductase activity on seed priming in lettuce. *Plant Physiol.* 98 : 84-85.
- Chen, T. M., S. K. Ries, and J. M. Cantliffe. 1994. Priming of lettuce seeds with nitrate. *Plant Physiol.* 105 : 35(6) : 574-580.
- Chen, T. M., S. K. Ries, and J. M. Cantliffe. 1994. Effect of nitrate reductase activity on seed priming in lettuce. *Plant Physiol.* 105 : pp 356-358.
- Classen, M. E. T. and G. E. Wilcox. 1974. Effect of the nitrogen form on growth and composition of tomato and pea tissue. *J. Amer. Soc. Hort. Sci.* 99 : 171-174.
- Claussen, W. 1986. Influence of fruit load and environmental factors on nitrate reductase activity and on concentration of nitrate and carbohydrates in leaves of eggplant (*Solanum melongena*). *Plant. Physiol.* 62 : 358-362.
- Cohen, R., O. Yarden, J. Katan. 1987. Paclobutrazol and other plant growth-retarding chemicals increase resistance of melon seedlings to fusarium wilt. *Plant Pathology.* 36 : 558-564.
- Cooper, A. J. 1973. Root temperature and plant growth. *Commonwealth Bureau of Horticulture and Plantation Crops (Research review)* 4 : 1-73.
- Dalzie., J. and D. K. Lawrence, 1984. Biochemical and biological effects

- of kaurene oxidase inhibitors, such as paclobutrazol. In biochemical aspects of synthetic and naturally occurring plant growth regulators (R. Mengennett and D. K. Lawrence, eds), pp. 43-57. British Plant Growth Regulator Group, Monograph No. 11. Wantage. ISBN 90-247-3198-4.
- Danneberger, T. K., M. B. McDonald, Jr. C. A. Geron, and P. Kumari. 1992. Rate of germination and seedling growth of perennial ryegrass seed following osmotic conditioning. *HortScience* 27 : 28 30.
- Dearman, J., P. A. Brocklehurst, and R. L. K. Drew. 1987. Effects of osmotic priming and ageing on the germination and emergence of carrot and leek seed. *Ann. Appl. Biol.* 111 : 717 722.
- Dijak, M. and D. P. Ormrod. 1985. Responses involved in increased dry matter production with supplementary incandescent radiation in growth chambers. *Environ. Expt. Bot.* 25 : 195 201.
- Dirr, M. A. 1975. Nitrate reductase activity in the leaves of the highbush blueberry and other plants. *J. Amer. Soc. Hort. Sci.* 97 : 329 331.
- Djurhuus, R. 1985. The effect of photoperiod and temperature on growth and development of *Begonia X Tuberhybridia* 'Karelsk jomfru'. *Scientia Hort.* 27 : 123 131.
- Dodge, C. M. and C. E. Wilcox. 1972. Relationship of pH to ion uptake imbalance by varieties of wheat. *Agron. J.* 64 : 476 481.
- Duke, S. H and S. O. Duke. 1978. In vitro nitrate reductase activity and in vivo phytochrome measurements of maize seedlings as affected by various light treatments. *Plant Cell Physiol.* 19 : 481 489.
- Durrant, M. J., P. A. Payne, and J. M. Maclaren. 1983. The use of water and some inorganic salt solutions to advance sugar beet seed: . Experiments under controlled and field conditions. *Ann. Appl. Biol.* 103 : 517 526.
- Early, J. D. and G. C., Martin. 1988. Translocation and breakdown of <sup>14</sup>C-labelled paclobutrazol in 'Nemaguard' peach seedlings. *Hortscience* 23(1) : 196 200.
- Evans, T. A., and W. G. Pill. 1989. Emergence and seedling growth from osmotically primed or pregerminated seeds of asparagus (*Asparagus officinalis* L.). *J. Hort. Sci.* 64 : 275 282.
- Figliolia, A., M. T. Cale, G. Camele and S. Pagliara. 1978 1979. Effettidella nutrizione nitrica e ammoniacale su alcunisistemi enzimatici di *Nicotiana tabacum* cv. Virginia Bright. *J. Agr. Sci.* in

- Finland. 56 : 239 243.
- Finch-Savage, W. E., D. Gray, and G. M. Dickson. 1991. The combined effects of osmotic priming with plant growth regulator and fungicide soaks on the seed quality of five bedding plant species. *Seed Sci. & Technol.* 19 : 495 503.
- Fischer, R. A. and Stapper, M. 1987. Lodging effects on high-yielding crops of irrigation crops of irrigated semidwarf wheat. *Field Crops Research.* 17: 245 258.
- Foster, E. F. and C. A. Stutte. 1986. Glutamine synthetase activity and foliar nitrogen volatilization in response to temperature and inhibitor chemicals. *Ann. of Bot.* 57 : 305 307.
- Frett, J. J., W. G. Pill, and D. C. Morneau. 1991. A comparison of priming agents for tomato and asparagus seeds. *HortScience* 26 : 1158 1159.
- Frett, J. J., and W. G. Pill. 1989. Germination characteristics of osmotically primed and stored *Impatiens* seeds. *Scient. Hortic.* 40 : 171 179.
- Fricke, W. 1993. Glutamine synthetase and glutamate synthetase activities in high ammonium grown wheat cells. *Phytochemistry* 34 : 637 644.
- 藤重宣昭, 杉山直儀. 1968. 果菜苗の生長におよぼす地温の影響. *日園學雜.* 37 : 37 42.
- Gaudreau, L., J. Charbonneau , L.-P. Vezina and A. Gosselin. 1995. Effects of photoperiod and photosynthetic photon flux on nitrate content and nitrate reductase activity in greenhouse-grown lettuce. *J. Plant Nutri.* 18 : 437 453.
- Gebauer, G. and J. Stadler. 1990. Nitrate assimilation and nitrate content in different organs of ash trees(*Fraxinus excelsior*). *Plant Nutrition-Physiology and Application.* Pp. 101 106.
- Gebauer, G. 1990. Diurnal changes of nitrate content and nitrate reductase activity in different organs of *Atriflex hortensis*(C3 plant) and *Amaranthus retroflexus*(C4 plant). *Plant Nutrition - Physiology and Application.* Pp. 93 99.
- Gertsson, U. E. 1984. Effect of temperature, day length and light intensity on growth and development of *Dipladenia sanderi* Hemsl. 'Rosca'(Ledd) Hiern.). *Scientia Hort.* 32 : 217 305.
- Globerson, D., and Z. Feder. 1987. The effect of seed priming and fluid



- drilling on germination, emergence and growth of vegetables at unfavorable temperatures. *Acta Hortic.* 198 : 15 21.
- Gray, D. 1976. The effects of time to emergence on head weight and variation in head weight at maturity in lettuce (*Lactuca sativa*). *Appl. Biol.* 82:569 575.
- Gray, D., P. A. Brocklehurst, J. R. A. Steckel, and J. Dearman. 1984. Priming and pregermination of parsnip (*Pastinaca sativa* L.) seed. *J. Hort. Sci.* 59 : 101 108.
- Greenway, H., and Munns, R. 1980. Mechanisms of salt tolerance in nonhalophytes. *Ann. Rev. Plant Physiol.* 31: 140 190.
- Guedes, A. C., and D. J. Cantliffe. 1980. Germination of lettuce seeds at high temperature after seed priming. *J. Amer. Sci.* 105 : 777 781.
- Hageman, R. H. and D. Fleisher. 1961. Nitrate reductase activity in corn seedlings as affected by light and nitrate content of nutrient media. *Plant Physiol.* 37 : 700 708.
- Haigh, A. M., E. W. R. Barlow, F. L. Milthrope, and P. J. Sinclair. 1986. Field emergence of tomato (*Lycopersicon esculentum*), carrot (*Daucus carota*) and onion (*Allium cepa*) seeds primed in an aerated salt solution. *J. Amer. Soc. Hort. Sci.* 111 : 660 665.
- Haigh, A. M., and E. W. R. Barlow. 1987a. Germination and priming of tomato, carrot, onion and sorghum seeds in a range of osmotica. *J. Amer. Soc. Hort. Sci.* 112 : 202 208.
- Haigh, A. M., and E. W. R. Barlow. 1987b. Water relation of tomato seed germination. *Aust. J. Plant Physiol.* 14 : 485 492.
- Hallgren, S. W. 1989. Effects of osmotic priming using aerated solution of polyethylene glycol on germination of pine seeds. *Ann. Sci. For.* 46: 31 38.
- Harper, J. E., J. C. Nicholas and R. H. Hageman. 1972. Seasonal and canopy variation in nitrate reductase activity of soybean (*Glycine max* K. Merr.). *Crop Science* 12 : 382 386.
- Harper, J. E., and R. H. Hageman. 1972. Canopy and seasonal profiles of nitrate reductase in soybeans (*Glycine max* L. merr.). *Plant Physiol.* 49 : 146 154.
- Hart, J. W. 1988. Light and plant growth. Urwin Hyman, London. p.41.
- Hartmann, T. 1982. Die Ammonium-Assimilation im N-Stoffwechsel Pflanze. *Biologie in unserer Zeit.* 12 : 9 19.
- Hatam, M. 1980. Seasonal and diurnal variations in nitrate reductase

- activity of soybean(*Glycine max* L. Merr.). *Plant and Soil* 56 : 27  
32.
- Haugh, P. A. 1988. Sterol requirements and paclobutrazol inhibition of a celery cell culture. *Phytochemistry* 27(8) : 2491-2500.
- Healy, W. E., R. D., Heins and H. F., Wilkins. 1980. Influence of photoperiod and light quality on lateral branching and flowering in selected vegetatively propagated plants. *J. Amer. Soc. Hort. Sci.* 109 : 812-816.
- Heimer, Y. and P. Filner. 1971. Regulation of the nitrate assimilation pathway in cultured tobacco cells. The nitrate uptake system. *Biochem. Biophys. Acta.* 230 : 362-372.
- Helsel, D. G., D. R. Helsel, and H. C. Minor. 1986. Field studies on osmoconditioning soybeans, *Glycine max*. *Field Crops Res.* 14 : 291-298.
- Heuer, B., Z. Plaut and E. Federman. 1979. Nitrate and nitrite reduction in wheat leaves as affected by different types of water stress. *Physiol. Plant.* 46 : 318-323.
- Heydecker, W. 1973/74. Germination of an idea: The priming of seeds. Univ. of Nottingham School of Agriculture Report. Pp. 50-67.
- Heydecker, W., J. Higgins, and Y. J. Turner. 1975. Invigoration of seeds. *Seed Sci. & Technol.* 3 : 881-888.
- Heydecker, W., and P. Coolbear. 1977. Seed treatments for improved performance survey and attempted prognosis. *Seed Sci. & Technol.* 5 : 353-425.
- . 1993.
- 久村敦彦 外. 1973. 作物の光合成と物質生産. 養賢堂.  
. 1995.  
. 36 : 57-61.
- 池田英男, 大澤孝也. 1980. 施用窒素形態とそ菜の適應性(第2報). 水耕栽培において硝酸, アンモニア, 亞硝酸を窒素源とした葉菜の生育 びにアンモニア態及び硝酸態窒素蓄積の差異. *日園學雜.* 48 : 435-442.
- 池田英男, 大澤孝也. 1981. 施用窒素形態とそ菜の適應性(第3報). 水耕栽培においてNO<sub>3</sub>, NH<sub>4</sub>, NO<sub>2</sub>をN源とした根菜の生育 びにNH<sub>4</sub>-N及びNO<sub>3</sub>-N素蓄積の差異. *日園學雜.* 49 : 563-570.
- 池田英男, 大澤孝也. 1983. 水耕培養液中のNO<sub>3</sub>-とNH<sub>4</sub>-の濃度 びに比率がそ菜の生育, 葉中N成分及び培養液のpHに及ぼす影響. *日園學雜.* 52 :

159 166.

- 池田英男, 大澤孝也. 1985. 培養液中のNO<sub>3</sub>とNH<sub>4</sub>の比率及び液温がミツバ, シュンギク びにネギの生育に及ぼす影響. 日園學雜. 54 : 58 65.
- 今津正, 織田彌三郎. 1965. せりの形態および生態に関する研究(第1報). 營養生長期における栽培および野生せりの形態的差異. 日本園藝學會誌 34(4) : 297 304.
- Iversen, K. V., R. H. Fox, and P. Piekielek. 1985. Diurnal, shade and hybrid effects on nitrate content of young corn stalks. Commun. in Soil Sci. Plant Anal. 16 : 837 852.
- 岩田正利, 小野泰. 1968. 窒素形態の差異とそ菜の生育 (第4報)生育段階との關係. 日園學雜. 38 : 44 51.
- Jeswani, L. M., B. R. Murty and R. B. Mehara. 1970. Divergence in relation to geographical origin in a world collection of linseed. Ind. J. Gen. and Plt. Breed. 30 : 11 25.
- Kaiser, J. J., and O. A. M. Lewis. 1983. Nitrate reductase and glutamine synthetase activity in leaves and roots of nitrate-fed *Helianthus annuus* L. Plant and Soil 77 : 127 130.
- Kallio, H., R. Rousku, A. Salminen, and E. Tikanmaki. 1984. Diurnal variations in nitrate content of red beets. J. Agri. Sci. in Finland 56 : 239 243.
- Kamachi, K., Yamaya, T., Mae, T., and Ojima, K. 1991. A role for glutamine synthetase in the remobilization of leaf nitrogen during natural senescence in rice leaves. Plant Physiol. 96 : 411 417.
- , , . 1995a. priming (abstract). 13(1 ) : 82 83.
- , , . 1995b. priming 가 . Priming 가 (Abstracts). 13(1) : 80 81.
- Khan, A. A. 1980/81. Hormonal regulation of primary and secondary seed dormancy. Israel J. Bot. 29 : 207 224.
- Khan, A. A. 1992. Preplant physiological seed conditioning. Hort. Rev.13 : 131 181.
- Khan, A. A., K. Tao, J. S. Knypl, B. Borkowska, and L. E. Powell. 1978. Osmotic conditioning of seeds: Physiological and biochemical changes. Acta Hort. 83 : 267 278.
- Khan, A. A., N. H. Peck, A. G. Taylor, and C. Samimy. 1983. Osmoconditioning of beet seeds to improve emergence and yield in

- cold soil. *Agron. J.* 75 : 788 794.  
 . 1986.
- , . 1991. Paclobutrazol 가 , , . 32(1) : 111 116.  
 . 1977. . 17  
 : 327 331.  
 . 1979. . 17 : 260 263.
- Kim, H. Y. and K. R. Kim, 1986. Relationship between organic compound contents and freezing tolerance in kiwifruit plant. *Res. Rep. Rural Devel. Admin. (Suweon)*. 28 : 95 103.
- Kirkby, E. A., and Mengel, K. 1967. Ionic balance in different tissues of the tomato plant in relation to nitrate, urea, or ammonium nutrition. *Plant Physiol.* 42 : 6 14.
- Knight, S. L. and C. A. Mitchell. 1988. Effects incandescent radiation on photosynthesis, growth rate and yield of 'Weldmann's Green' leaf lettuce. *Scientia Hort.* 35 : 37 49.
- Knypl, J. S., and A. A. Khan. 1981. Osmoconditioning of soybean seeds to improve performance at suboptimal temperatures. *Agron. J.* 73 : 112 116.
- 位田藤久太郎. 1982. 水耕液のpH管理. *農及園*. 57 : 327 331.
- Larouche, R., A. Gosselin, and L. P. Vezina. 1989. Nitrogen concentration and photosynthetic photon flux in greenhouse tomato production : . *Growth and development. J. Amer. Soc. Hort. Sci.* 114 : 458 461.  
 1 . 1986. .  
 ( ) 267 280.  
 1 . 1988. , .  
 29(3) : 191 200.  
 1 . 1990.  
 . 31(1) : 15 21.  
 1 . 1991. I.  
 . 32(1) :  
 29 42.  
 1 . 1991. II.  
 . 32(4) : 425 433.  
 1 . 1992. Induction and morphological characteristics

- associated with embryogenesis of *Oenanthe stolonifera* DC., 89th Annual Meeting of the American Society for Horticultural Science.
- 1 . 1993. I. . 34 (2) : 108 114.
- Lee, B. Y. et al. 1994. Preliminary studies on the establishment of seedling production system by somatic embryogenesis in *Oenanthe stolonifera* DC., XXIV International Horticultural Congress, Kyoto Japan.
- 1 . 1995. , 36(1) : 38 45.
- 2 . 1987. III. 28(4) : 300 308.
- 2 . 1987. I. 12(1) : 15 20.
- 2 . 1987. IV. 가 28(4) : 309 315.
- 2 . 1987. II. 가 28(4) : 289 299.
- 4 . 1992. 1(2) : 123 134.
1991. 32(1) : 29 42.
- Lee, E. H., J. K. Byun and J. W. Stephanie. 1986. Protection of crop plants from sulfur dioxide, chilling, and heat-induced injury with paclobutrazol. *Perspetive Environ. Bot.* Vol. 2, in press.
- Lee, E. H., J. K. Byun and S. J. Wilding. 1985. A new gibberellin biosynthesis inhibitor, paclobutrazol(PP 333); confers increase SO<sub>2</sub> tolerance on snap bean. *Plant Environ. Exp. Botany* 25 : 265 275.
- Lillo, C. 1993. Magnesium and calcium inhibition of squash leaf nitrate reductase. *Plant Cell Physiol.* 34:1181 1185.
- Lillo, C. and A. Hendrikson. 1984. Comparative studies of diurnal variations of nitrate reductase activity in wheat, oat and barley. *Physiol. Plant.* 62 : 89 94.
- Lin, W. H. and C. H. Kao. 1980. Factors affecting in vivo nitrate

- reductase activity in triticale. *Physiol. Plant.* 48 : 361 364.
- Lowry, O. H., N. I. Rosebrough, A. L. Farr and R. J. Randall. 1951. Protein measurement with the folin phenol reagent. *J. Biol. Chem.* 193 : 265 275.
- Maanen, J. M. S., A. Dijk, K. Mulder, M. H. Baets, P. C. A. Menheere, D. Heide, P. L. J. M. Mertens and J. C. S. Kleinjans. 1994. Consumption of drinking water with high nitrate levels causes hypertrophy of the thyroid. *Toxicology Letters* 72 : 365 374.
- Macduff, J. H. and F. E. Trim. 1986. Effects of root temperature and form of nitrogen nutrition on nitrate reductase activity in oilseed rape (*Brassica napus* L.). *Ann. Bot.* 57 : 345 352.
- Malnassy, P. G. 1971. Physiological and biochemical studies on a treatment hastening the germination of seeds at low temperatures. Ph. D. Diss. Rutgers Univ., New Brunswick, N. J.
- Martinoia, E., U. Heck and A. Wiemken. 1980. Vacuoles as storage compartments for nitrate in barley leaves. *Nature* 289 : 292 293.
- Mehta, P. and H. S. Srivastava. 1980. Comparative stability of ammonium and nitrate induced nitrate reductase activity in maize leaves. *Phytochem.* 19 : 2527 2530.
- Mengel, K., P. Robin and L. Salsac, 1983. Nitrate reductase activity in shoots and roots of maize seedlings as affected by the form of nitrogen nutrition and the pH of the nutrient solution. *Plant Physiol.* 71 : 618 622.
- Mexal, J., J. T. Fisher, J. Osteryoung, and C. P. Reid. 1975. Oxygen availability in polyethylene glycol solutions and its implication in plant-water relations. *Plant Physiol.* 55 : 20 24.
- Michel, B. E., and M. R. Kaufman. 1973. The osmotic potential of polyethylene glycol 6000. *Plant Physiol.* 51 : 914 916.
- Minotti, P. L., D. C. Williams and W.A. Jackson. 1969. Nitrate uptake by wheat as influenced by ammonium and other cations. *Crop Sci.* 9 : 9 14.
- Mohanty, B. and J. S. Fletcher. 1976. Ammonium influence on the growth and nitrate reductase activity of paul's scarlet rose suspension cultures. *Plant Physiol.* 58 : 152 155.
- Money, N. P. 1989. Osmotic pressure of aqueous polyethylene glycols. Relationship between molecular weight and vapor pressure deficit. *Plant Physiol.* 91 : 766 769.



- Park, K. W. and D. Fritz. 1984. Effects of fertilization and irrigation on the quality of radish (*Raphanus sativus* L. var. Niger) grown in experimental pots. *Acta Horticulturae* 145 : 129-132.
- Passam, H. C., P. I. Karavites, A. A. Papandreou, C. A. Thanos, and K. Georghiou. 1989. Osmoconditioning of seeds in relation to growth and fruit yield of aubergine, pepper, cucumber and melon in unheated greenhouse cultivation. *Scient. Hortic.* 38 : 207-216.
- Perkins-Veazie, P., and D. J. Cantliffe. 1984. Need for high quality seed for effective priming to overcome thermodormancy. *J. Amer. Soc. Hort. Sci.* 109 : 368-372.
- Pill, W. G. 1986. Parsley emergence and seedling growth from raw, osmoconditioned and pregerminated seeds. *HortScience* 21 : 1134-1136.
- Pill, W. G., J. J. Frett, and D. C. Morneau. 1991. Germination and seedling emergence of primed tomato and asparagus seeds under adverse conditions. *HortScience* 26 : 1160-1162.
- Pill, W. G., and W. E. Finch-Savage. 1988. Effects of combining priming and plant growth regulator treatments on the synchronization of carrot seed germination. *Ann. Appl. Biol.* 114 : 383-389.
- Plaut, Z. 1974. Nitrate reductase activity of wheat seedlings during exposure and recovery from water stress and salinity. *Physiol. Plant.* 30 : 212-217.
- Puritch, G. S. and A. V. Barker. 1967. Structure and function of the tomato leaf chloroplasts during ammonium toxicity. *Plant Physiol.* 42 : 1229-1238.
- Quinche, J. P. 1982. Fluctuations des teneurs en nitrates des legumes au cours de la journee. *Revue suisse Vitic. Arboric. Hortic.* 14 : 85-87.
- Rabin, J., G. A. Berkowitz, and S. W. Akers. 1988. Field performance of osmotically primed parsley seed. *HortScience* 23 : 554-555.
- Radin, J. W., L. L. Parker and C. R. Cell. 1978. Partitioning of sugar between growth and nitrate reduction in cotton roots. *Plant Physiol.* 62 : 550-553.
- Rajasekhar, V. K. and H. Mohr. 1986. Appearance of nitrate reductase in cotyledons of the mustard (*Sinapsis alba* L.) seedlings as affected





- locust seedlings *via* pretreatment of seeds with paclobutrazol. Can. J. For. Res. 23 : 2548-2551.
- Simon, C. 1970. Die alimentäre Methämoglobinämie im Säuuglin GSA. Ernährungs-Umschau. 17 : 3-5.
- Smith, P. T., and B. G. Cobb. 1991a. Accelerated germination of pepper seed by priming with salt solutions and water. HortScience 26 : 417-419.
- Smith, P. T., and B. G. Cobb. 1991b. Physiological and enzymatic activity of pepper seeds (*Capsicum annum*) during priming. Physiol. Plant. 82 : 417-419.
- , E. Przemek. 1991.            glutamine synthetase            .            . 36 : 545-553.
- , M. J. Ems. 1992. NO<sub>3</sub>-            oxidative pentose phosphate pathway            [nitrate reductase, nitrite reductase, glutamine synthetase]            [glutamine synthetase]            .            . 37 : 468-475.
- ,            . 1993.            가            NO<sub>3</sub>-            가            .            . 2 : 2-15.
- . 1989.            glutamine synthetase            .            .            GS            "V"            .            . 34 : 98-105.
- Stance, G. M., G. Jenkins, and P. R. Hanson. 1979. Varietal responses in spring barley to natural and artificial lodging and to a growth regulator. J. Agric. Sci. 93 : 449-456.
- Steer, B. T. 1974. Control of diurnal variations in photosynthetic products.            . Nitrate reductase activity. Plant Physiol. 54 : 762-765.
- Steingrover, E., P. Ratering and J. Sissing. 1986. Daily changes in uptake, reduction and storage of nitrate in spinach grown at low light intensity. Physiol. Plant. 66 : 550-556.
- Stinchcombe, G. R., E. Copas, P. R. Williams, G. Arnold. 1984. The effect of paclobutrazol and daminozide on the growth and yield of cider apple trees. J. Hort. Sci. 59 : 323-327.
- Stokes, P. 1965. Temperature and dormancy. Encyclop. Plant. Physiol. 15(2) : 746-803.
- Street, J. E., J. H. Jordan, M. W. Ebelhar, and D. L. Boykin. 1986.

- Plant height and yield responses of rice to paclobutrazol. *Agron. J.* 77 : 288-291.
- Sugiyama, N. and S. Hanawa. 1992. Growth responses of rabbiteye blueberry plants to N forms at constant pH in solution culture. *J. Japan Soc. Hort. Sci.* 61 : 25-29.
- Szafirowska, A., A. A. Khan, and N. H. Peck. 1981. Osmoconditioning of carrot seeds to improve seedling establishment and yield in cold soil. *Agron. J.* 73 : 845-848.
- Tafazoli, E. and C. A. Beyl, 1993. Foliarily applied abscisic acid and its effect on cold hardiness and carbohydrates in *Actinidia arguta*. *Iran J. Agri.* (in press).
- Takács, E. and L. Técsi. 1992. Effects of NO<sub>3</sub>/NH<sub>4</sub><sup>+</sup> ratio on photosynthetic rate, nitrate reductase activity and chloroplast ultrastructure in three cultivars of red pepper (*Capsicum annuum* L.). *J. Plant Physiol.* 140 : 298-305.
- 高橋久光, C. Shennan and R. C. Huffaker. 1993. 亜鉛缺乏と遮光がトウモロコシ (*Zea mays* L.) の硝酸還元酵素の活性に及ぼす影響. *東農大農學集報.* 38 : 117-123.
- 高德錚. 1991. 温度・光照及 肥對水耕蔬菜品質之影響. 動態浮根式水耕系統之開發與利用 151-159.
- Tang, R. S. and Y. N. Wu. 1986. Growth retardant-first report of PP333 test. *Jiangsu Agric. Sci. Sin.* 2 : 12.
- Tannenbaum, S. R., J. S. Wishnok and C. D. Leaf. 1991. Inhibition of nitrosamine formation by ascorbic acid. *Am. J. Clin. Nutr.* 53 : 247-250.
- Taylor, A. A. and G. R. Stewart. 1980. The effect of ammonia and light/dark transitions on the level of glutamine synthetase activity in *Osmunda regalis*. *Plant Science Letters.* 20 : 125-131.
- Tayo, T. O. and D. G. Morgan. 1979. Factors influencing flower and pod development in oil-seed rape. *J. of Agric. Sci.* 92 : 363-373.
- Teare, I. D., R. Manam and E. T. Kanemasu. 1974. Diurnal and seasonal trends in nitrate reductase activity in field grown sorghum plants. *Agron. J.* 66 : 733-736.
- Terry, L. F., J. M. Zajicek, and M. A. Hussey. 1992. Use of seed priming to bypass stratification requirements of three *Aquilegia* species. *HortScience* 27(4) : 310-313.
- Terry, N. 1968. Developmental physiology of sugar beet. I. The influence

- of light and temperature on growth. J. Expt. Bot. 19 : 795 811.
- Travis, R. L. and J. L. Key. 1971. Correlation between polyribosome level and the ability to induce nitrate reductase in dark-grown corn seedlings. Plant Physiol. 48 : 617 620.
- Travis, R. L., R. C. Huffaker and J. L. Key. 1970. Light induced development of polyribosomes and the induction of nitrate reductase in corn leaves. Plant Physiol. 46 : 800 805.
- Upadhyay, M. K. and B. R. Murty. 1970. Genetic divergence in relation to geographical distribution in pearl millet. Ind. J. Gen. and Plt. Breed. 30 : 704 715.
- Valdes, V. M., K. J. Bradford, and K. S. Mayberry. 1985. Alleviation of thermodormancy in coated lettuce (*Lactuca sativa*) cultivar "Empire" by seed priming. HortScience. 20 : 1112 1114.
- Van der Toorn, P. 1989. Embryo growth in mature celery seeds. Ph.D. Dissertation, Agricultural University, Wageningen, The Netherlands, 1989. pp.95
- Vines, H. M. and R. T. Wedding. 1960. Some effects of ammonia on plant metabolism and a possible mechanism for ammonia toxicity. Plant Physiol. 35 : 820 825.
- Vlahos, J. C. 1990. Daylength influences growth and development of *Achimenes*. HortScience 25:1595-1596.
- Vlahos, J. C., E. Heuvelink and G. F. P. Martakis. 1991. A growth analysis study on 3 *Achimenes* cultivars grown under 3 light regimes. Scientia Hort. 46 : 275 282.
- Vlahos, J. C., G. R. P. Martakis and E. Heuvelink. 1992. Daylength, light quality and temperature influence growth and development of *Achimenes*. HortScience 27(12) : 1269 1271.
- Vollbrecht, P., E. Klein and H. Kasemir. 1989. Different effects of supplied ammonium on glutamine synthetase activity in mustard (*Sinapis alba*) and pine (*Pinus sylvestris*) seedlings. Physiol. Plant. 77 : 129 135.
- Wakhloo, J. L. and A. Staudt. 1988. Development of nitrate reductase activity in expanding leaves of *Nicotiana tabacum* in relation to the concentration of nitrate and potassium. Plant Physiol. 87 : 258 263.
- Wallace, W. 1973. The distribution and characteristics of nitrate reductase and glutamate dehydrogenase in the maize seedling. Plant Physiol. 52 : 191 196.

- Wample, R. L. and E. B. Culver. 1983. The influence of paclobutrazol, a new growth regulators on sunflower. *J. Amer. Soc. Sci.* 108(1) : 122-125.
- Wang, C. Y. 1982. Physiological and biochemical responses of plants to chilling stress. *Hortic.* 26 : 293-298.
- Wang, Y. T. 1991. Modification of *Hibiscus* growth by treating paclobutrazol. *J. Plant Growth Regul.* 10 : 47-51.
- Weibe, H. J., and H. Tissen. 1979. Effects of different seed treatments on embryo growth and emergence of carrot seeds. *Gartenbauwissenschaft* 44 : 280-284.
- Weibe, H. J., and T. Muhyaddin. 1987. Improvement of emergence by osmotic seed treatments in soils of high salinity. *Acta Hortic.* 198 : 91-100.
- Wiebe, H. -J. 1987. Einfluß der Tageslänge auf Entwicklung, Wachstum und Nitratgehalt von Spinatsorten. *Gartenbauwissenschaft*. 52.S. : 103-108.
- Wild, A., R. Zerbe and A. S. W. Leidig. 1981. A comparison of the activities of nitrate and nitrite reductase under weak and strong light conditions. *Ber. Deutsch. Bot. Ges. Bd.* 94S : 701-707.
- William, S. M. and A. M. Harry. 1978. Influence of  $\text{NO}_3/\text{NH}_4$  on growth, N absorption and assimilation by lima beans in solution culture. *Agron. J.* 70: 1027-1031.
- Willihan, E. F., R. G. Sharpless and W. L. Printy. 1977. Effect of pH on yield and leaf composition of hydroponic tomato. *HortSci.* 12 : 316-317.
- Wolfe, D. W., and W. L. Sims. 1982. Effects of osmoconditioning and fluid drilling of tomato seed on emergence rate and final yield. *HortScience* 17 : 936-937.
- Wolfe, J. A. 1978. Chilling injury in plants - The role of membrane lipid fluidity. *Plant Cell Envir.* 1 : 241-247.
- Wright, M. G. and K. L. Davison. 1964. Nitrate accumulation in crops and nitrate poisoning of animals. *Advances in Agron.* 16 : 197-247.
- 山崎肯哉. 1982. 養液栽培全編. 博友社.
- Yamaya, T., H. Tanno, N. Hirose, S. Watanabe and T. Hayakawa. 1995. A supply of nitrogen causes increase in the level of NADH-dependent glutamine synthetase protein and in the activity of the enzyme in roots of rice seedlings. *Plant Cell Physiol.* 36 :

- 1197-1204.  
 , , . 1989. (*Oenanthe stolonifera* DC)  
 , 30(3)  
 : 180-186.  
 . 1983.  
 . 2 : 193-206.  
 . 1984.  
 3 : 421-447.
- Yelle, S., A. Gosselin and M. J. Trudel. 1987. Effect of atmospheric CO<sub>2</sub> concentration and root-zone temperature on growth, mineral nutrition and nitrate reductase activity of greenhouse tomato. *J. Amer. Soc. Hort. Sci.* 112 : 1036-1040.  
 . 1979. (*Oenanthe stolonifera* DC.) .
- Zuo, W., C. H. Hang, and G. Zheng. 1988. Physiological effects of priming with SPP on seeds of pea, tomato and spinach. *Proc. Int. Conf. Stand. Estab. Hort. Crops*, Lancaster, PA. Pp.124-133.